

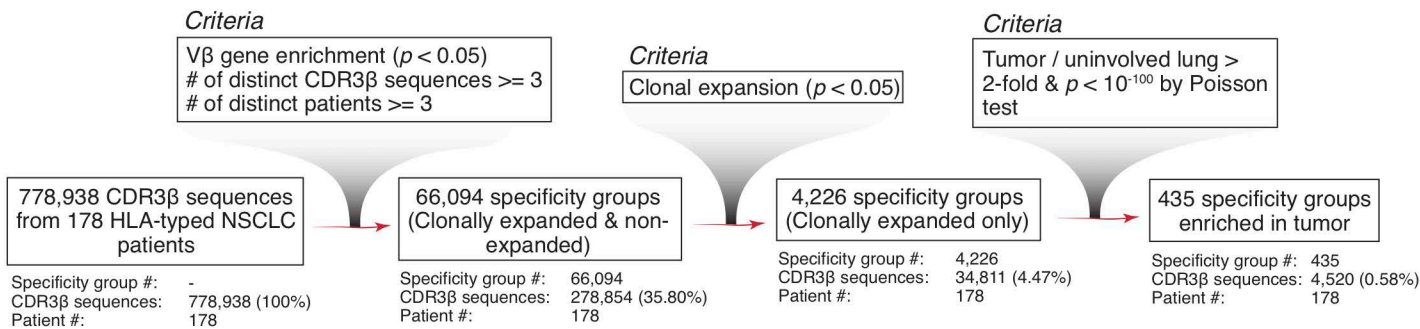
Supplemental information

Global analysis of shared T cell specificities in human non-small cell lung cancer enables HLA inference and antigen discovery

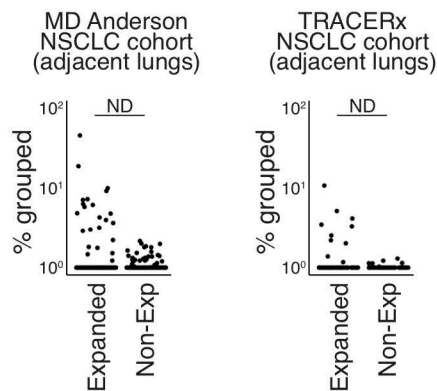
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Figure S1

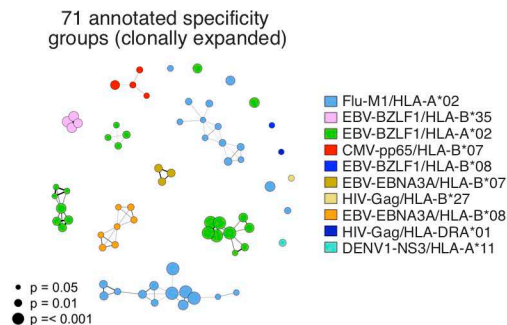
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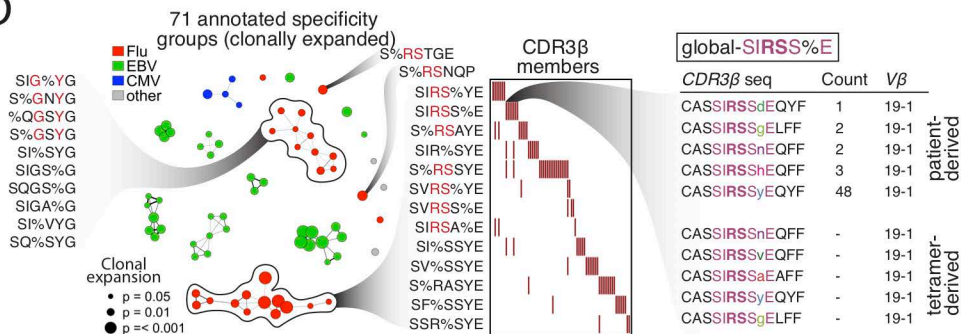
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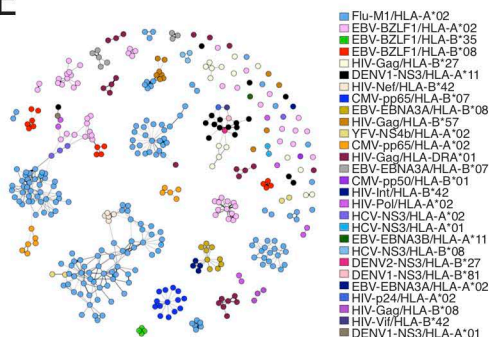
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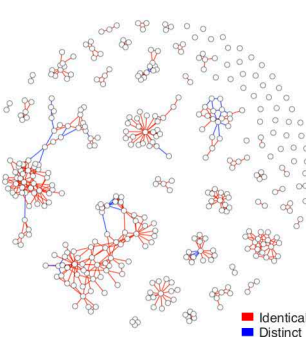
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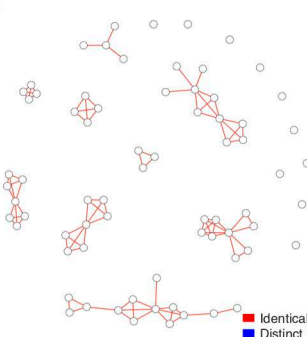
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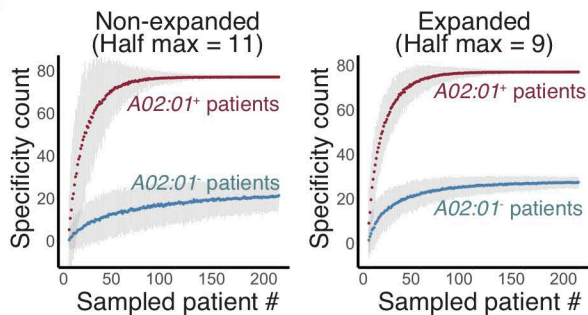
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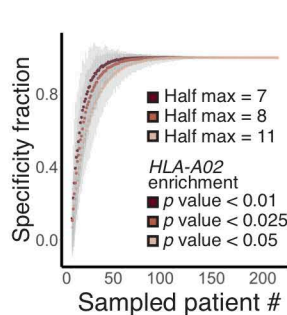
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H



I



J

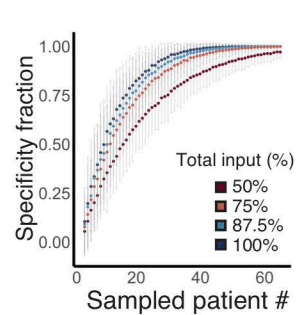


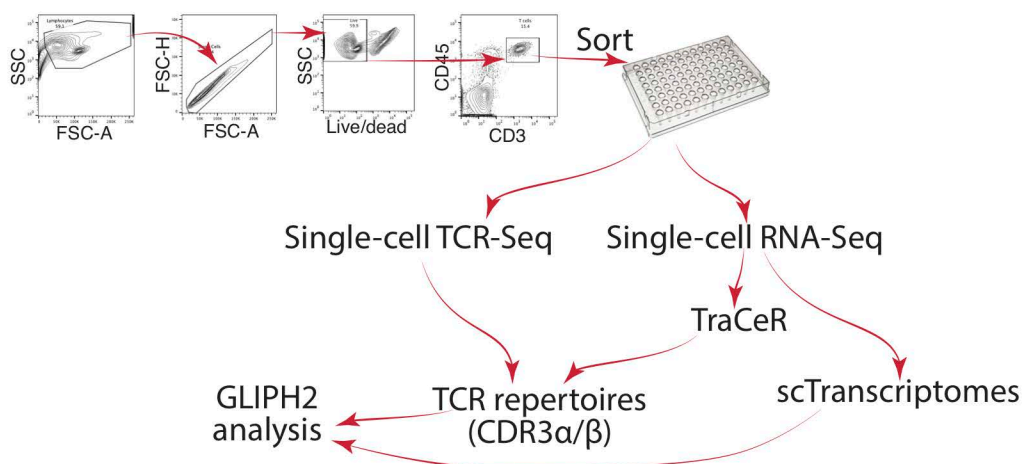
Figure S1, related to Figure 1

(A) Specificity inference pipeline. **(B)** Low percentages of TCR clonotypes from adjacent lung tissues are grouped into tumor-enriched specificity groups. The percentages of the top 20 most expanded CDR3 β clonotypes from the adjacent lung tissues of patients belonging to the MDACC NSCLC cohort (n=178 samples, left) and the TRACERx cohort (n=63 samples, right) were quantified for those that belonged to the 435 tumor-enriched specificity groups as in Figure 1B (% grouped, \log_{10} -converted). The same analysis was performed on the remainder of the CDR3 β clonotypes (Non-exp, non-expanded). ND, no statistically significant difference was found. **(C)** The 71 clonally expanded specificity groups annotated and colored with 10 indicated tetramers are shown in the network. **(D)** Left to right, network analysis of 71 clonally expanded specificity groups colored as in Figure 1C is shown; two large Flu-related communities (red) are circled and the CDR3 β members of the specificity groups are highlighted with the previously reported short motifs "RS" and "GxY" highlighted in red font; heatmap showing distinct CDR3 β members (columns) of the Flu-related (with the "RS" motif) specificity groups (rows) and the levels of shared CDR3 β members between specificity groups within the circled community; table showing an example of the "SIRSS%E" specificity group containing the short "RS" motif (bold) that is annotated with 5 Flu-specific tetramer sequences (bottom). The counts of distinct CDR3 β members from tumor and the $V\beta$ gene usage are shown (top). **(E)** 394 specificity groups annotated with indicated tetramers (key) were organized into distinct communities through shared CDR3 β sequence(s) as in Figure 1C. Thickness of edge represents numbers of shared CDR3 β sequence(s) between any two connected nodes. **(F)** Community plot as in **(E)**. Color of edge represents shared CDR3 β sequence(s) between specificity groups with identical (red) or distinct (blue) specificities defined by tetramer-derived sequences (labeled with distinct colors, **E**). 588 of all (n=634) connections (edges) are labeled in red (92.74%). **(G)** 71 clonally expanded specificity groups as in **(C)**. Color of edge represents shared CDR3 β sequence(s) between specificity groups with identical (red) or distinct (blue) specificities defined by tetramer-derived sequences (**C**). 92 of all (n=92) connections (edges) are labeled in red (100%). **(H-J)** TCR specificity group saturation is dependent of the level of clonal expansion, the absolute numbers of specificity groups, as well as the sequencing depth of the repertoires. **(H)** Bootstrapping for quantification of clonally expanded (right, n=77) and non-expanded (left, n=77) HLA-A*02:01-enriched specificity groups with CDR3 β sequences from either HLA-A02⁺ (red) or HLA-A02⁻ (blue) NSCLC patients. Bootstrapping was done by "sampling with replacement" and the X axis represents the number of patients randomly sampled (Sampled patient #, Methods) and the Y axis represents the numbers of specificity groups quantified with a given sampling event. Shades of error bars represent the 3X standard errors derived from 100 sampling events

for a given number of sampled patients. **(I)** Bootstrapping for quantification of *HLA-A*02:01*-enriched specificity groups with varying cutoffs for *HLA-A*02:01* enrichment ($p < 0.05$, $n=1267$; $p < 0.025$, $n=319$; $p < 0.01$, $n=72$). As in **(H)**, X axis represents the number of patients randomly sampled (Sampled patient #). Y axis represents the numbers of specificity groups quantified with a given sampling event that are normalized against the number of the total specificity groups used (Specificity fraction). Shades of error bars represent the 3X standard errors derived from 100 sampling events for a given number of sampled patients. **(J)** Bootstrapping for quantification of *HLA-A*02:01*-enriched specificity groups with varying input CDR3 β sequencing depth (50, 75, 87.5, or 100% of total input by random down-sampling). As in **(I)**, X axis represents the number of patients randomly sampled and Y axis represents the normalized numbers of specificity groups. Shades of error bars represent the 3X standard errors derived from 100 sampling events for a given number of sampled patients.

Figure S2

A



B

Flu-related		Flu-related		EBV-related		EBV-related	
Specificity group ID	Specificity group ID	Specificity group ID	Specificity group ID	Specificity group ID	Specificity group ID	Specificity group ID	Specificity group ID
SV%SNQP	SIRS%YE	S%RSTDT		RTG%GNT			
TCRβ	Vβ	TCRβ	Vβ	TCRβ	Vβ	TCRβ	Vβ
CASSVaSNQPQHF	TRBV9-1	CASSIRSIEQYF	TRBV19-1	CASSkRSTDTQYF	TRBV19-1	CATRTGgGNTIYF	TRBV6-8
CASSVdSNQPQHF	TRBV9-1	CASSIRsgYEQFF	TRBV19-1	CASSdRSTDTQYF	TRBV2-1	CASRTGpGNTIYF	TRBV9-1
CASSVeSNQPQHF	TRBV6-8	CASSIRsgYEQYF	TRBV19-1	CASShRSTDTQYF	TRBV19-1	CSARTGtGNTIYF	TRBV29-1
CASSVfSNQPQHF	TRBV19-1	CASSIRSdYEQYF	TRBV19-1	CASSsRSTDTQYF	TRBV19-1	CASRTGdGNTIYF	TRBV9-1
CASSVgSNQPQHF	TRBV9-1	CASSIRSeYEQYF	TRBV19-1	CASSIRSTDTQYF	TRBV19-1	CSARTGvGNTIYF	TRBV20-1
CASSVkSNQPQHF	TRBV9-1	CASSIRSaYEQFF	TRBV19-1	CASSeRSTDTQYF	TRBV10-2	CSVRTGaGNTIYF	TRBV29-1
CASSVISNQPQHF	TRBV6-8	CASSIRSaYEQYF	TRBV19-1	CASSsRSTDTQYF	TRBV6-8	CAYRTGsGNTIYF	TRBV30-1
CASSVnSNQPQHF	TRBV10-2					CASRTGsGNTIYF	TRBV6-8
CASSVsSNQPQHF	TRBV19-1						
CASSVtSNQPQHF	TRBV4-2						
CASSVySNQPQHF	TRBV19-1						
TCR12 clone		TCR13 clone		TCR14 clone		TCR15 clone	
α CATDDSGGFKTIF		α CAGPTGGGSQGNLIF		α CAVTYGGSQGNLIF		α CAEDLNARLMF	
β CASSVtSNQPQHF		β CASSIRSaYEQYF		β CASSsRSTDTQYF		β CSARTGVGNTIYF	

C

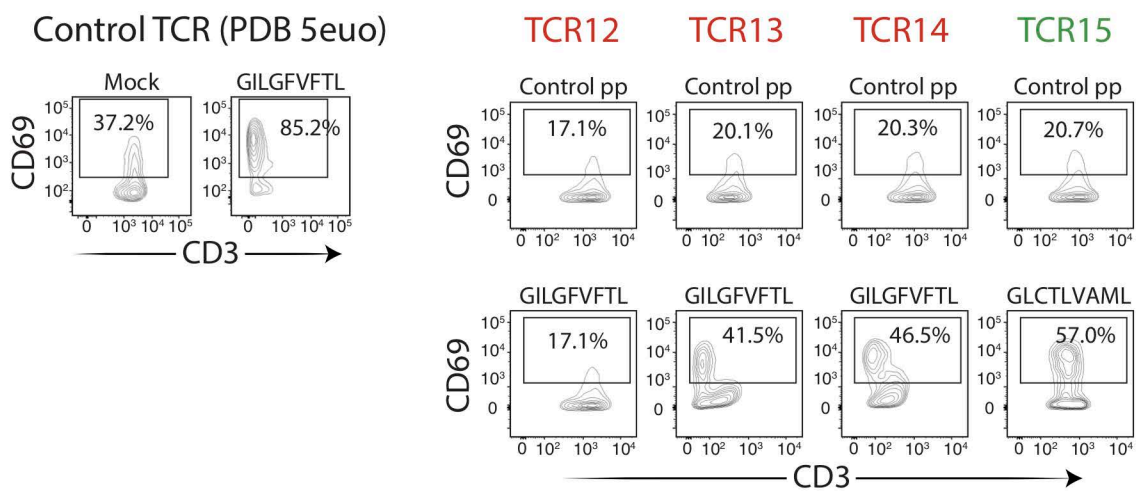


Figure S2, related to Figure 2

(A) Schematic of the combined single-cell TCR-Seq and single-cell RNA-Seq (scRNA-seq) procedures. CD45⁺CD3⁺ T cells were sorted from single-cell suspensions of lung tumor samples from patients with NSCLC at Stanford. Single-cell TCR-Seq was performed using nested multiplexed PCR as previously described [Han *et al.*, 2014. Nat. Biotechnol. 32, 684]. scRNA-seq was performed according to previous methods [Picelli *et al.*, 2014. Nat. Protoc. 9, 171] with modifications (STAR Methods). TCR repertoires were integrated from the single-cell TCR-Seq pipeline and from the scRNA-seq data with reconstruction using the TraCeR algorithm [Stubington *et al.*, 2016. Nat. Methods 13, 329] for GLIPH2 analysis. scTranscriptomes, single-cell transcriptomes derived from scRNA-seq. **(B)** GLIPH2 inferred clone TCR12, TCR13, and TCR14 to recognize Influenza virus M1 (FluM1) 9mer peptide "GILGFVFTL" in the context of HLA-A*02; clone TCR15 is inferred to recognize EBV BMLF1 9mer peptide "GLCTLVAML" in the context of HLA-A*02. Tables show subsets of CDR β members for each specificity group. The selected T cell clones with paired CDR3 α/β sequences are highlighted in bold in the tables and the CDR3 sequences of both TCR α/β chains are shown at the bottom. All four clones were found in tumors from 2 different NSCLC patients from the Stanford cohort. **(C)** Right, the TCR α/β sequences of the four chosen T cell clonotypes in **(B)** were ectopically expressed in TCR-deficient Jurkat-76 cells and stimulated with T2 (HLA-A02⁺) cells pulsed with indicated peptides (right, above FACS plots). CD69 expression quantified by FACS is shown. Left, Jurkat cells expressing the control TCR α/β chains (PDB 5euo) previously reported to recognize FluM1 in the context of HLA-A*02 and stimulated with or without (Mock) the 9mer peptide "GILGFVFTL" are shown. Control peptide (pp), CMV/pp65₄₉₅₋₅₀₃.

Figure S3

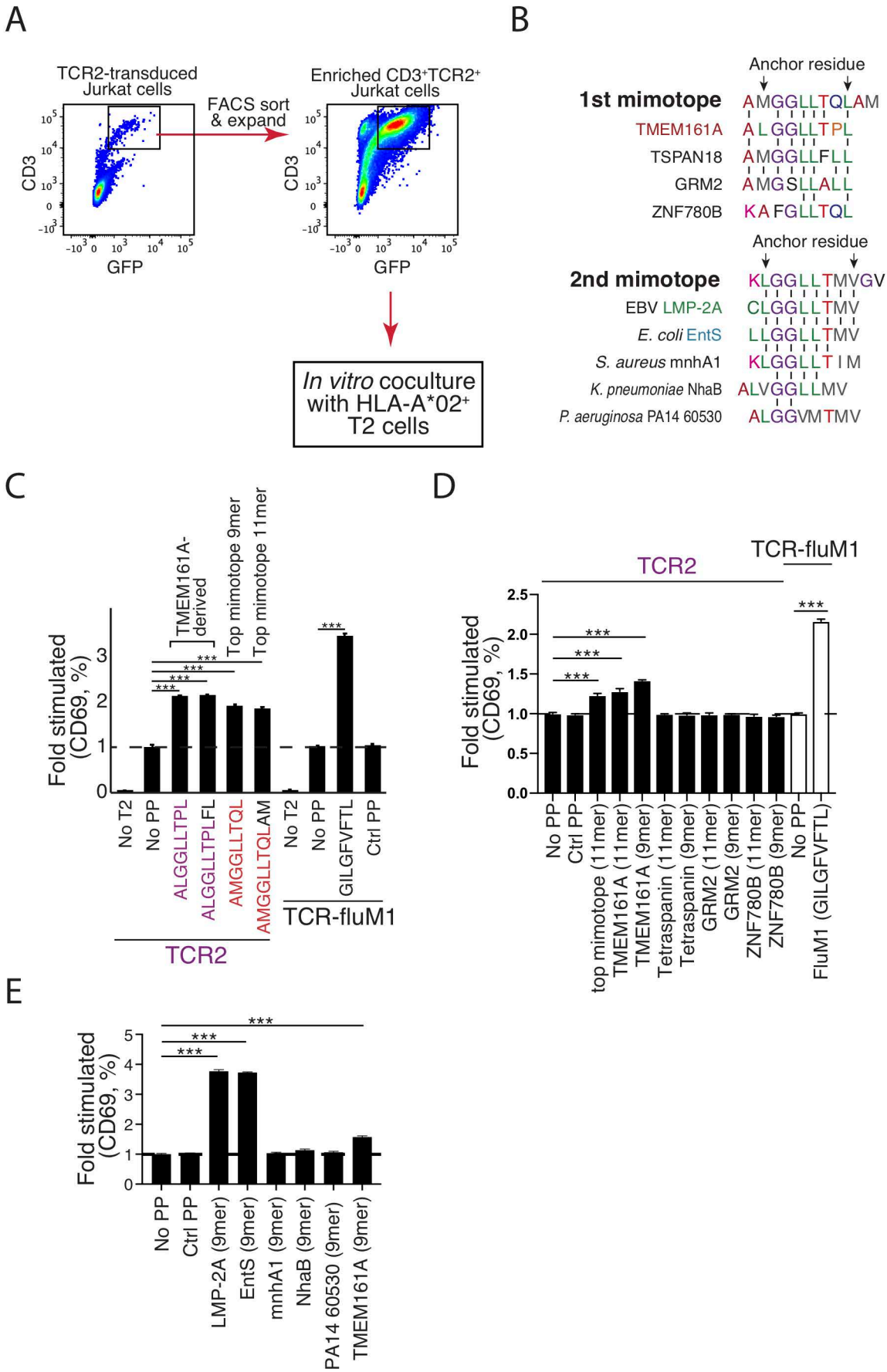
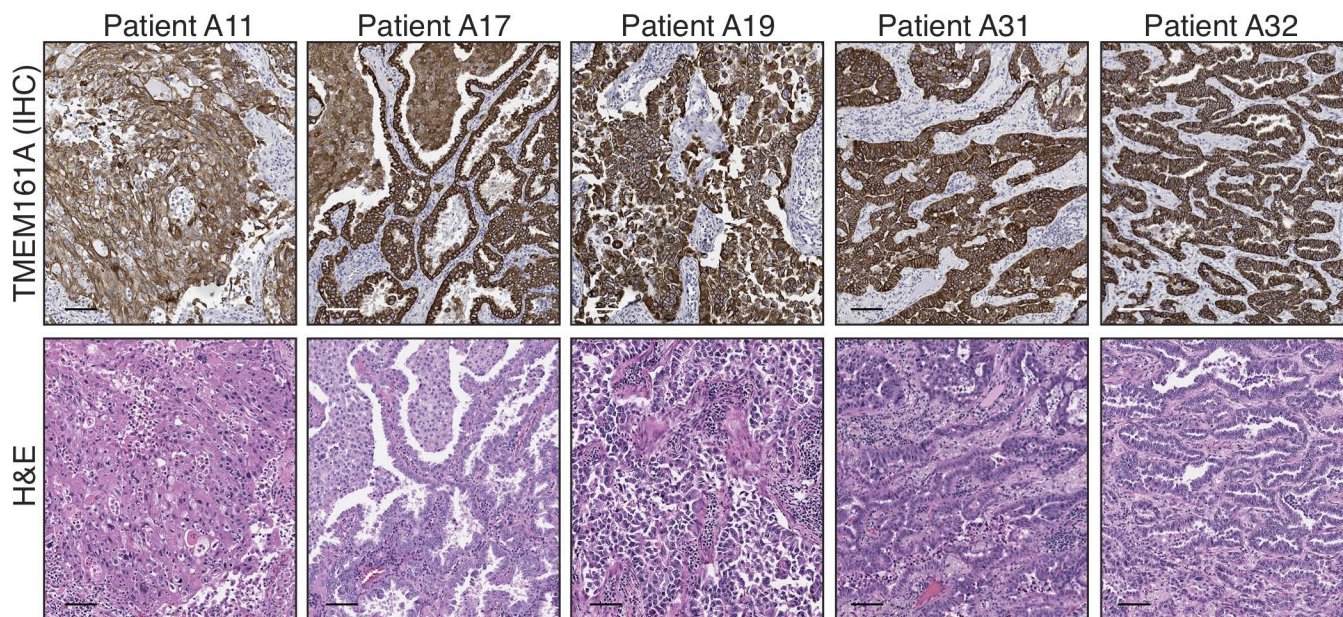


Figure S3, related to Figure 3

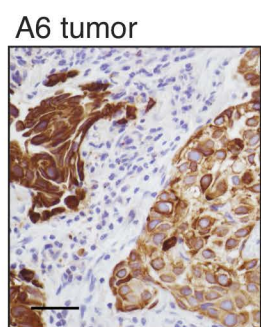
(A) TCR-deficient Jurkat cells were transduced with lentivirus carrying a composite coding region of TCR2 α chain, 2A peptide sequence (2Ap), TCR2 β chain, 2Ap, and GFP (Jurkat-TCR2). Transduced cells were subsequently sorted by FACS for the GFP⁺CD3⁺ population (left) and allowed to expand (top right) and used in *in vitro* stimulation experiments (bottom right). Similar strategies were used to make other stable TCR-Jurkat clones throughout the paper. **(B)** Protein database search showed partial matches of the top 2 mimotopes with candidate coding sequences from various species. All matches were 9mers and predicted to bind HLA-A*02 with high affinities by netMHCpan 4.0. **(C)** Jurkat-TCR2 was stimulated with T2 (HLA-A2⁺) cells pulsed with indicated peptides, including the top mimotope from the yeast screen (11mer, AMGGLLQLAM), the 9mer from the top mimotope predicted to bind HLA-A*02 with high affinity (AMGGLLQL), and both 9mer/11mer peptides from the TMEM161A coding region (ALGGLLTPL and ALGGLLTPLFL, respectively) with sequence homology to the top mimotope. CD69 upregulation was quantified by FACS. Jurkat cells expressing the control TCR α / β chains (TCR-fluM1, PDB 5euo) and stimulated with the cognate 9mer "GILGFVFTL", the control peptide (Ctrl PP, CMV/pp65₄₉₅₋₅₀₃), no peptide (No PP), or no co-cultured T2 cells are included as controls. ***, $p < 0.001$ by t test. **(D,E)** The endogenous peptides derived from human TMEM161A, LMP-2A of EBV, and EntS of *E. coli* were recognized by TCR2. Jurkat-TCR2 cells were cocultured with HLA-A02⁺ T2 cells and pulsed with indicated peptides: Ctrl PP (control peptide), Flu/M1₅₈₋₆₆ GILGFVFTL; top mimotope 11mer, AMGGLLQLAM; TMEM161A 11mer, ALGGLLTPLFL; TMEM161A 9mer, ALGGLLTPL; Tetraspanin 11mer, AMGGLLFLGF; Tetraspanin 9mer, AMGGLLFL; GRM2 (Glutamate receptor) 11mer, AMGSLALLAL; GRM2 9mer, AMGSLALL; ZNF780B (Zinc finger protein 780B isoform X1) 11mer, KAFGLLTQLAQ; ZNF780B 9mer, KAFGLLTQL in **(D)** and Ctrl PP (control peptide), Flu/M1₅₈₋₆₆ GILGFVFTL; LMP-2A 9mer, CLGGLLTMV; EntS 9mer, LLGGLLTMV; mnhA1 9mer, KLGGLLTIM; NhaB 9mer, ALVGGLLMV; PA14 60530 9mer, ALGGVMTMV; TMEM161A 9mer, ALGGLLTPL in **(E)**. Jurkat cells expressing the control TCR (TCR-fluM1, PDB 5euo) are stimulated with the cognate 9mer "GILGFVFTL" for comparison.

Figure S4

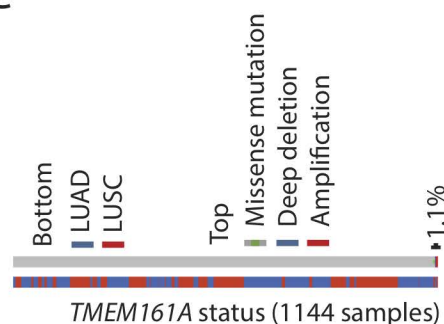
A



B

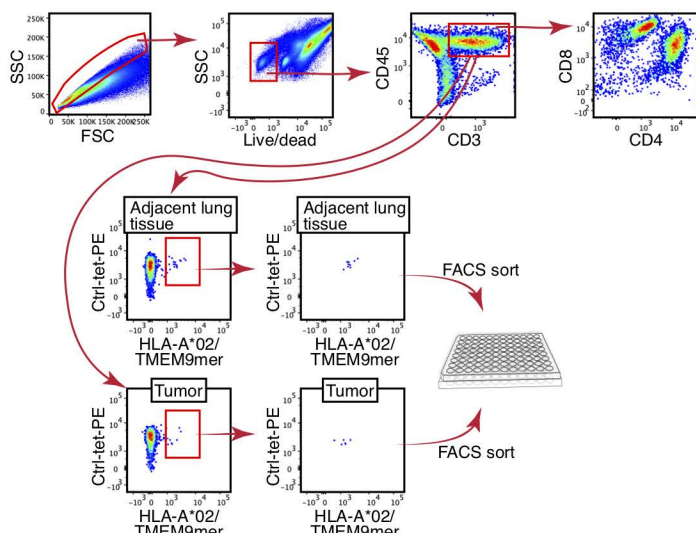


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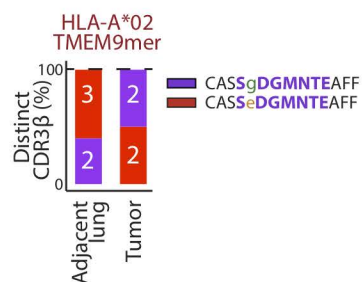


Mutation type	Count
Missense mutation	4
Deep deletion	2
Amplification	6
No alterations	1132

D



E



F

	%DGMNTE	LUSC	LUAD
+	21	10	
-	16	31	

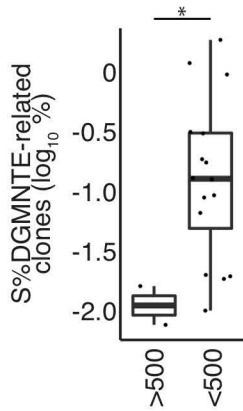
$p = 5.3 \times 10^{-3}$

Figure S4, related to Figure 4

(A) TMEM161A is broadly expressed on human NSCLC tumors. Representative images of TMEM161A immunohistochemistry on tumor (top) and hematoxylin & eosin (H&E) staining from adjacent sections (bottom) from Stanford patients A11, A17, A19, A31, and A32. Scale bar, 100 μm . **(B)** Strong and dim TMEM161A staining on a section from A6 patient tumor. Scale bar, 40 μm . **(C)** TMEM161A is a non-mutated tumor antigen. Left, percentages of all types of genetic alterations within the TMEM161A locus defined with whole-genome sequencing from the Cancer Genome Atlas (TCGA) project (pan-lung cancer cases, $n=1144$) are shown. Right, number of cases with indicated genomic alterations. **(D,E)** TMEM9mer/A02 tetramer⁺ sorted T cells from tumor and the adjacent lung tissue carry the "S%DGMNTE" motif, as predicted by GLIPH2. **(D)** Sorting HLA-A*02/TMEM9mer⁺CD8⁺ T cells from the resected tumor and the adjacent lung tissue of NSCLC patient A6 through FACS. Single cell suspensions were prepared and stained with anti-CD4, CD8, CD3, and CD45 antibodies, live/dead marker AquaZombie, PE-conjugated tetramer HLA-A*02/viral peptides (CMV-pp65₄₉₅₋₅₀₃ and EBV-BMLF1₂₈₀₋₂₈₈), and APC-conjugated tetramer HLA-A*02/TMEM9mer. Specific T cells were sorted onto 96-well plates based on the following criteria: non-doublers, live cells, CD3⁺CD45⁺, HLA-A*02/viral peptides⁻, and HLA-A*02/TMEM9mer⁺. **(E)** Percentages (%) of distinct CDR3 β sequences of tetramer-sorted CD8⁺ T cells as in **(D)** from the adjacent lung tissue and tumor are shown. Numbers in bars represent the counts of sorted cells. **(F)** Numbers of A02⁺ squamous cell lung carcinoma (LUSC) or lung adenocarcinoma (LUAD) patients from the MDACC cohort with or without detected T cells carrying the "S%DGMNTE" CDR3 β motif. p value = 5.3×10^{-3} by Fisher's Exact test.

Figure S5

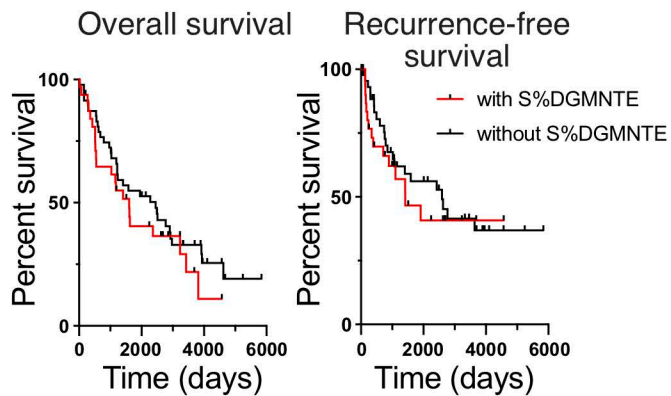
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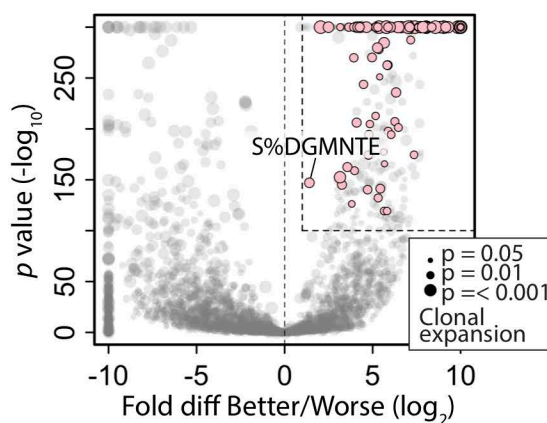
B

	With S%DGMNTE motif	Without S%DGMNTE motif	Fisher's exact test
# Patients	32	47	
Gender			
M	16/32 (50%)	29/47 (62%)	ns
F	16/32 (50%)	18/47 (38%)	ns
Race			
Caucasian	31/32 (97%)	43/47 (91%)	ns
Other	1/32 (3%)	4/47 (9%)	ns
Histology			
Adenocarcinoma	10/32 (31%)	31/47 (66%)	p < 0.01
Squamous cell carcinoma	21/32 (65%)	16/47 (34%)	p < 0.01
Smoking status			
Current smoker	16/32 (50%)	21/47 (45%)	ns
Former smoker	16/32 (50%)	21/47 (45%)	ns
Never smoker	0/32 (0%)	5/47 (11%)	ns
Stage			
I	13/32 (41%)	23/47 (49%)	ns
II	16/32 (50%)	18/47 (38%)	ns
III	3/32 (9%)	6/47 (12%)	ns
Primary tumor size (cm)	4.6	3.8	ns
Adjuvant chemotherapy	12/30 (40%)	14/46 (30%)	ns
Multifocal disease	5/32 (16%)	5/47 (11%)	ns

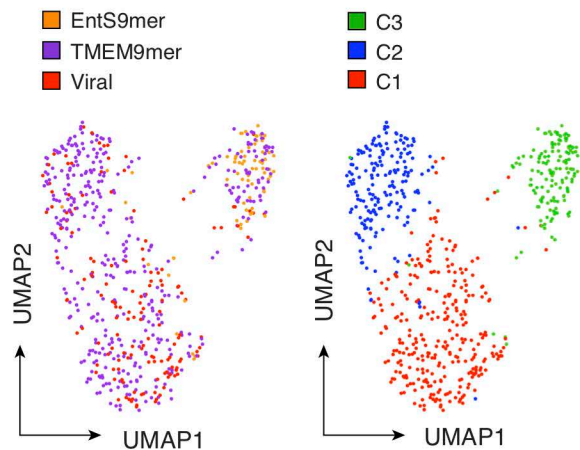
C



D



E



F

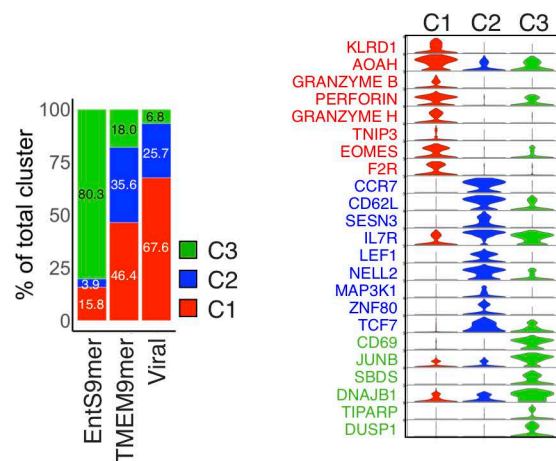
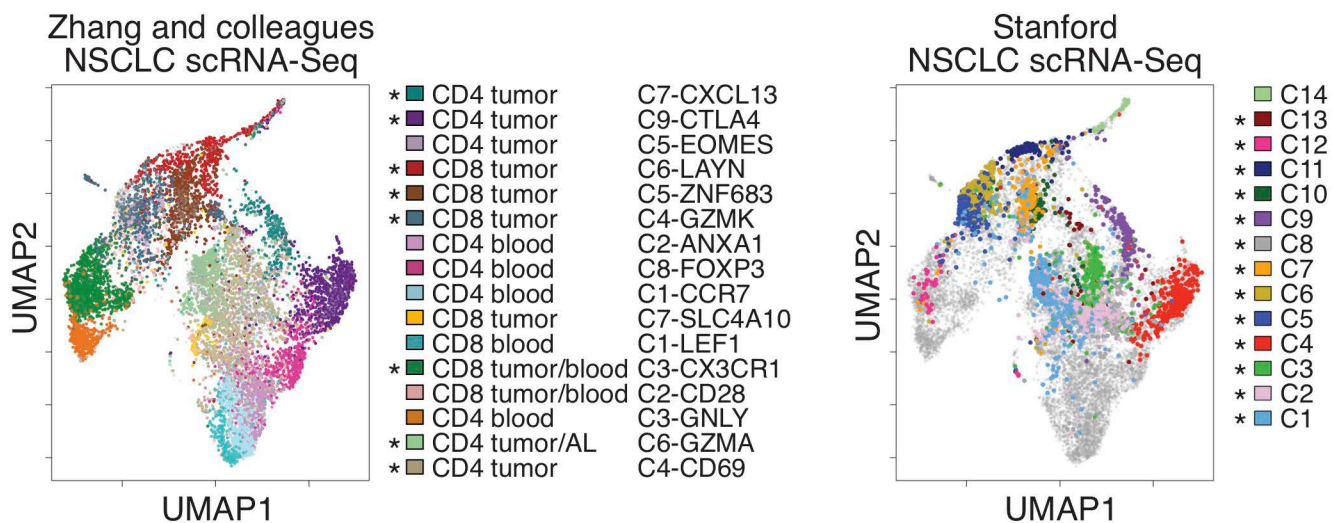


Figure S5, related to Figure 5

(A) Total percentage (\log_{10}) of T cells carrying the "S%DGMNTE" CDR3 β motif for A02⁺ patients from the MDACC cohorts stratified as having more than 500 (>500) or less than 500 (<500) total mutation counts (n = 34). **(B)** Break down of the clinical data of the MDACC NSCLC patient cohort stratified by detection of CDR3 β sequences carrying the "S%DGMNTE" sequence motif. **(C)** There are no differences in overall survival (left) or recurrence-free survival (right) among patients with tumor-infiltrating T cell CDR3 β containing the "S%DGMNTE" sequence motif (n = 32) vs patients without tumor-infiltrating T cell CDR3 β containing the same motif (n = 49). **(D)** Volcano plot showing the probabilities of clonal biases for the 4,226 clonally expanded specificity groups between the tumors from patients with (worse, n = 88) or without recurrence (better, n = 90) by Poisson test. Tumor-enriched specificity groups that show a significant clonal bias in patients without recurrence are highlighted in pink (n = 146). Specificity group S%DGMNTE is highlighted. **(E,F)** A subset of TMEM161A-specific T cells found in healthy donors' peripheral blood reveal effector phenotype. **(E)** Dimension reduction by Uniform Manifold Approximation and Projection (UMAP) of the scRNA-Seq results from the sorted CD45⁺CD8⁺CD3⁺ T cells from PBMC with indicated HLA-A*02 tetramers (left) as in Fig 5A identified 3 major cell states (right, # indicates percentage of each cell state). Viral, CD8 T cells sorted with HLA-A*02 tetramers loaded with Flu, EBV, or CMV peptides (Flu-M1₅₈₋₆₆, CMV-pp65₄₉₅₋₅₀₃, and EBV-BMLF1₂₈₀₋₂₈₈). **(F)** Stacked violin plot showing the differential genes expressed by the identified cell states as in **(E)**.

Figure S6

A



B

Cluster ID (Stanford)	Lineage (Stanford)	Cluster ID (Guo)	Lineage (Guo)	Location (Guo)
C1	CD4	C6-GZMA	CD4	T and AL
C2	CD4	C4-CD69	CD4	T
C3	CD4	C4-CD69	CD4	T
C4	CD4	C9-CTLA4	CD4	T
C5	CD8	C4-GZMK	CD8	T
C6	CD8	C4-GZMK	CD8	T
C7	CD8	C5-ZNF683	CD8	T
C8	CD4	C4-CD69	CD4	T
C9	CD4	C7-CXCL13	CD4	T
C10	CD8	C5-ZNF683	CD8	T
C11	CD8	C6-LAYN	CD8	T
C12	CD8	C3-CX3CR1	CD8	*PB and T
C13	CD4	C4-GZMK/C4-CD69	CD8/CD4	T/T
C14	CD4/CD8	-	-	-

C

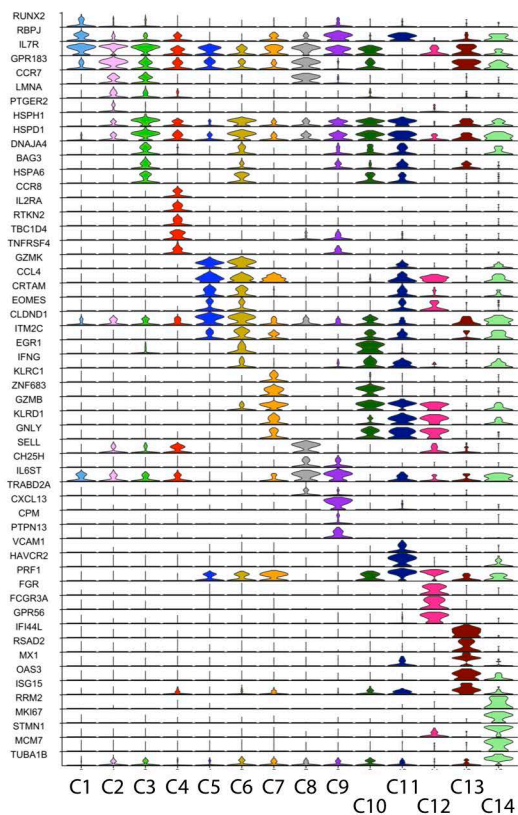


Figure S6, related to Figure 6

(A) Dimension reduction by UMAP of the published NSCLC scRNA-Seq results from Zhang and colleagues (Guo *et al.* 2018. Nat. Med. 24, 978). 12,346 sorted T cells from the report by Zhang and colleagues were combined with the 2,950 sorted T cells from the current study (Stanford cohort) for a joint analysis of dimension reduction by UMAP. Cells from Zhang and colleagues are colored according to the identified cell states as reported (left) in comparison with cells from the Stanford cohort colored with the 14 cell states identified in the current study (right). *, cell states identified by Zhang and colleagues (left) that mostly resembled at least one of the cell states identified in the current study (*, right). AL, adjacent lung. **(B)** Cross reference of the 14 cell clusters (C1-C14) identified in the Stanford lung cancer cohort with the clusters reported by Guo *et al.* (Guo *et al.* 2018. Nat. Med. 24, 978). *, CD8 T cells of the C3-CX3CR1 cluster could be found in both the peripheral blood and tumor (Guo *et al.* 2018. Nat. Med. 24, 978). T, tumor; AL, adjacent lung tissue; PB, peripheral blood. **(C)** Selected top differentially expressed genes for each of the identified clusters (n = 14). Expression of indicated genes were shown with the violin plot.

Figure S7

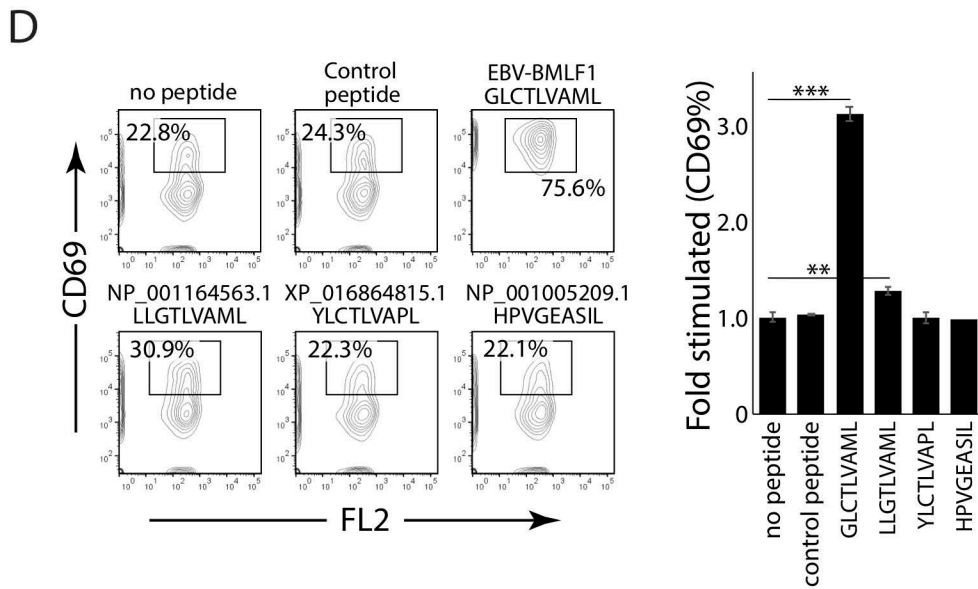
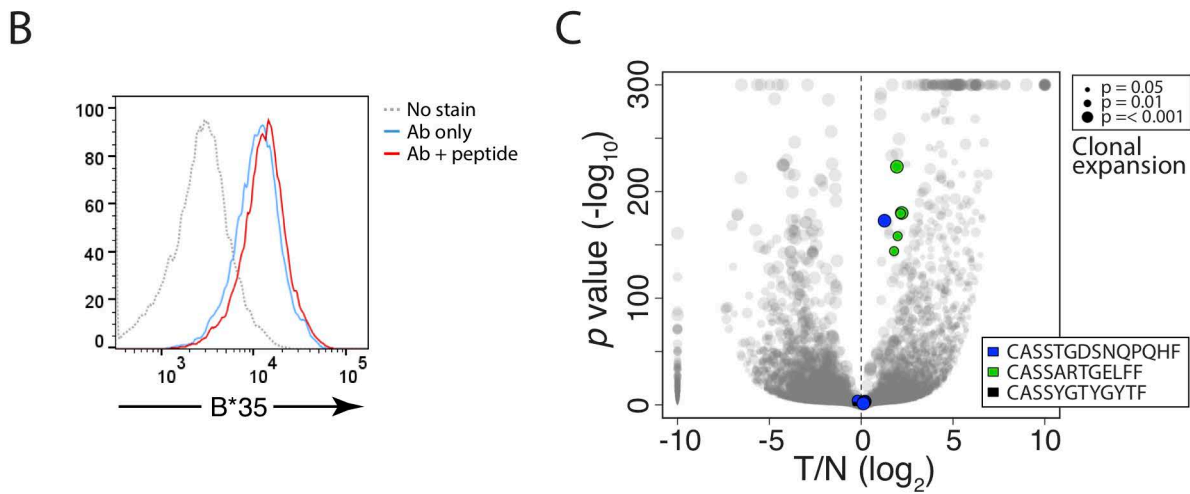
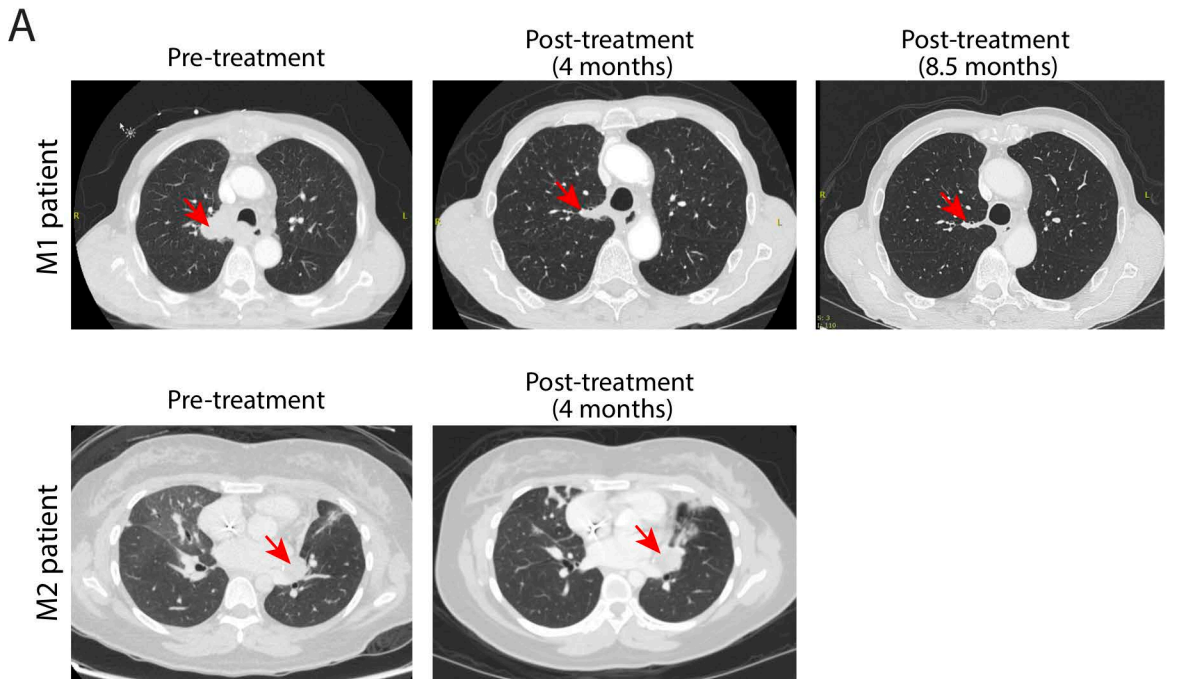


Figure S7, related to Figure 7

(A) CT scan images of pre- and post-treatment from NSCLC patient M1 (top panels) and M2 (bottom panels) treated with anti-PD1 therapy. Tumors are highlighted with red arrowheads. **(B)** T2 (174 x CEM.T2) cells were transduced with lentiviral vector encoding the full-length coding sequence of WT human *HLA-B*35:01*. Cells were selected with puromycin and the surface B*35 expression was quantified by FACS with or without the control peptide “LPFDFTPGY” reported previously (Takamiya *et al.*, 1994. *Int Immunol.* Vol. 6, 255). **(C)** Volcano plot showing the comparison of the 66,094 shared specificity groups between tumor (T) and the adjacent lung (N) by Poisson test. The y-axis represents the negative \log_{10} converted p values of the Poisson tests and the x-axis represents the \log_2 converted fold difference between tumor and the adjacent lung (T/N). Dot size represents levels of clonal expansion. Specificity groups annotated with pathogen-related tetramer CDR3 β sequences as in Figure 7B (n=11) are highlighted according to the respective CDR3 β sequences of the expanded clones (5th column, Figure 7B). **(D)** Left panels, representative FACS plots showing the stimulation of the Jurkat-TCR15 cells with 9mers from the EBV *BMLF1* locus (GLCTLVAML), uniprot NP_001164563.1 (*CLDN2* locus, LLGTLVAML), XP_016864815.1 (*SERINC5* locus, YLCTLVAPL), and NP_001005209.1 (*TMEM198* locus, GLLCGLVAML). Right, results of Jurkat-TCR15 cell stimulation in triplicate. Control peptide: flu M1 “GILGFVFTL”. ***, $p < 0.001$; **, $p < 0.01$ by student t test.

MDA-2054	ADC	F	African American	A*23:01	A*11:01	B*14:02	B*35:01	C*08:02	C*04:01
MDA-2057	SCC	M	Caucasian	A*31:01	A*36:01	B*08:12	B*40:01	C*03:04	C*07:01
MDA-2060	ADC	M	Caucasian	A*01:01	A*01:01	B*08:01	B*08:01	C*06:02	C*07:01
MDA-2061	SCC	F	Caucasian	A*68:01	A*68:01	B*51:01	B*08:01	C*01:02	C*07:01
MDA-2068	SCC	F	Caucasian	A*26:01	A*02:60	B*18:01	B*07:02	C*01:02	C*07:02
MDA-2070	ADC	M	Caucasian	A*03:56	A*03:01	B*51:01	B*35:01	C*04:01	C*15:02
MDA-2080	ADC	F	Caucasian	A*02:01	A*01:01	B*08:01	B*18:01	C*07:01	C*07:01
MDA-2082	ADC	M	Caucasian	A*02:01	A*01:01	B*07:02	B*15:01	C*03:04	C*07:02
MDA-2088	SCC	M	Caucasian	A*11:01	A*02:01	B*55:02	B*55:01	C*05:01	C*03:03
MDA-2090	SCC	M	Caucasian	A*01:01	A*03:56	B*07:02	B*15:17	C*07:01	C*07:01
MDA-2093	ADC	M	African American	A*11:01	A*02:01	B*15:01	B*35:01	C*03:03	C*04:01
MDA-2095	ADC	F	African American	A*02:05	A*03:01	B*50:01	B*07:02	C*06:02	C*07:02
MDA-2097	ADC	M	Caucasian	A*02:05	A*02:51	B*35:02	B*35:01	C*04:01	C*07:18
MDA-2108	SCC	M	Caucasian	A*02:05	A*30:01	B*18:01	B*57:01	C*12:03	C*06:02
MDA-2115	ADC	M	African American	A*03:04	A*03:63	B*51:02	B*51:01	C*15:02	C*03:04
MDA-2118	ADC	F	Asian or Pacific Islander	A*24:02	A*01:01	B*08:01	B*15:01	C*01:02	C*07:01
MDA-2121	SCC	F	Caucasian	A*03:01	A*23:01	B*35:01	B*35:01	C*04:01	C*04:01
MDA-2122	SCC	M	Caucasian	A*31:01	A*31:01	B*27:05	B*40:01	C*03:04	C*02:02
MDA-2139	SCC	M	Caucasian	A*02:01	A*03:26	B*27:02	B*07:02	C*07:02	C*02:02
MDA-2140	SCC	M	Caucasian	A*25:01	A*01:01	B*18:01	B*18:13	C*12:03	C*12:03
MDA-2150	ADC	F	Caucasian	A*03:01	A*03:01	B*27:05	B*07:02	C*07:02	C*01:02
MDA-2152	SCC	M	Caucasian	A*68:01	A*68:01	B*35:01	B*44:02	C*02:02	C*04:01
MDA-2155	SCC	M	Caucasian	A*02:01	A*02:01	B*08:01	B*14:02	C*08:02	C*07:01
MDA-2161	SCC	M	Caucasian	A*25:01	A*02:01	B*18:01	B*44:02	C*05:01	C*12:03
MDA-2175	ADC	M	Caucasian	A*02:01	A*03:01	B*07:02	B*15:01	C*03:04	C*07:02
MDA-2176	ADC	M	Caucasian	A*02:01	A*03:01	B*07:02	B*18:01	C*07:02	C*12:13
MDA-2195	SCC	F	Caucasian	A*33:01	A*03:01	B*14:02	B*08:01	C*08:02	C*07:01
MDA-2214	ADC	F	Caucasian	A*26:01	A*26:01	B*38:01	B*57:01	C*06:02	C*12:03
MDA-2221	ADC	M	Caucasian	A*24:02	A*02:01	B*08:01	B*14:02	C*08:02	C*07:01
MDA-2226	SCC	M	Caucasian	A*31:01	A*02:01	B*35:03	B*35:03	C*04:01	C*04:01
MDA-2230	ADC	M	Caucasian	A*30:02	A*03:01	B*53:01	B*07:02	C*07:19	C*07:01
MDA-2237	ADC	F	Caucasian	A*11:05	A*33:01	B*14:02	B*14:02	C*08:02	C*08:02
MDA-2240	ADC	F	Caucasian	A*26:01	A*29:02	B*27:05	B*44:03	C*01:02	C*16:01
MDA-2243	ADC	M	African American	A*03:01	A*24:02	B*27:05	B*07:02	C*07:02	C*01:02
MDA-2244	ADC	M	Caucasian	A*02:01	A*03:01	B*15:01	B*07:02	C*07:02	C*07:39
MDA-2254	ADC	M	Caucasian	A*26:01	A*11:01	B*51:01	B*15:01	C*15:02	C*04:01
MDA-2262	SCC	M	Caucasian	A*03:01	A*03:01	B*08:01	B*07:02	C*07:02	C*07:01
MDA-2264	SCC	F	Caucasian	A*02:03	A*02:03	B*07:02	B*07:02	C*06:02	C*07:02
MDA-2265	SCC	F	Caucasian	A*03:01	A*03:01	B*07:02	B*18:01	C*05:01	C*07:02
MDA-2271	ADC	F	Caucasian	A*11:01	A*11:01	B*08:01	B*44:02	C*05:01	C*07:01
MDA-2276	ADC	F	Caucasian	A*01:01	A*02:01	B*44:02	B*44:02	C*05:01	C*05:01
MDA-2286	ADC	M	Caucasian	A*24:02	A*02:03	B*40:01	B*51:01	C*03:04	C*01:02
MDA-2292	ADC	M	Caucasian	A*32:01	A*03:01	B*07:02	B*55:01	C*03:03	C*03:03
MDA-2294	ADC	F	Caucasian	A*03:01	A*03:01	B*51:01	B*08:01	C*01:02	C*07:01
MDA-2300	SCC	M	Caucasian	A*24:02	A*24:03	B*08:01	B*08:01	C*07:01	C*07:01
MDA-2307	ADC	M	Caucasian	A*02:01	A*01:01	B*15:01	B*51:01	C*16:02	C*03:04
MDA-2315	ADC	F	Caucasian	A*02:01	A*11:01	B*35:01	B*44:02	C*05:01	C*04:01
MDA-2316	ADC	M	African American	A*03:01	A*03:01	B*08:01	B*35:01	C*07:01	C*04:01
MDA-2318	ADC	F	Caucasian	A*30:26	A*03:01	B*39:01	B*13:02	C*06:02	C*07:02
MDA-2320	ADC	F	Caucasian	A*02:01	A*02:01	B*15:01	B*44:09	C*03:03	C*05:01
MDA-2325	SCC	F	Caucasian	A*26:01	A*24:10	B*18:01	B*13:02	C*06:02	C*07:01
MDA-2333	ADC	M	Caucasian	A*32:01	A*24:02	B*07:02	B*07:02	C*07:02	C*02:02
MDA-2335	ADC	F	Caucasian	A*25:01	A*25:01	B*08:01	B*57:01	C*06:02	C*07:01
MDA-2337	ADC	F	Caucasian	A*24:02	A*02:01	B*15:01	B*15:01	C*03:03	C*03:04
MDA-2343	ADC	M	Caucasian	A*68:02	A*30:02	B*15:10	B*35:01	C*06:02	C*03:02
MDA-2344	SCC	F	Caucasian	A*11:01	A*02:03	B*08:01	B*18:01	C*07:01	C*07:01
MDA-2345	ADC	M	Caucasian	A*01:01	A*01:01	B*08:01	B*08:01	C*07:01	C*07:01
MDA-2348	ADC	F	Caucasian	A*02:01	A*02:01	B*07:02	B*07:02	C*05:01	C*07:02
MDA-2352	ADC	F	Caucasian	A*03:01	A*03:01	B*51:01	B*07:02	C*07:02	C*14:02
MDA-3203	ADC	M	Caucasian	A*02:01	A*11:01	B*35:01	B*07:02	C*04:01	C*07:02
MDA-3212	ADC	F	Caucasian	A*30:01	A*02:01	B*13:02	B*15:01	C*03:61	C*06:02
MDA-3214	ADC	F	Caucasian	A*02:01	A*02:89	B*50:01	B*38:01	C*04:01	C*12:03
MDA-3234	SCC	F	Caucasian	A*03:01	A*02:01	B*51:01	B*35:01	C*02:02	C*04:01
MDA-3237	SCC	F	Caucasian	A*24:02	A*24:02	B*15:07	B*40:01	C*03:03	C*03:04
MDA-3238	ADC	M	African American	A*33:03	A*30:02	B*53:01	B*15:10	C*03:04	C*04:01
MDA-3242	ADC	M	Caucasian	A*25:01	A*25:01	B*15:17	B*18:01	C*07:01	C*12:03
MDA-3253	SCC	F	Caucasian	A*03:01	A*02:01	B*07:02	B*07:02	C*07:02	C*07:02
MDA-3259	ADC	F	Caucasian	A*01:01	A*01:01	B*08:01	B*07:02	C*07:01	C*07:01
MDA-3260	ADC	M	Caucasian	A*29:02	A*29:02	B*44:03	B*57:01	C*16:04	C*06:02
MDA-3273	ADC	M	Asian or Pacific Islander	A*24:02	A*02:01	B*40:06	B*40:01	C*15:02	C*08:01
MDA-3290	SCC	F	Caucasian	A*02:251	A*24:02	B*07:02	B*27:05	C*01:02	C*07:02
MDA-4309	ADC	F	Caucasian	A*02:01	A*02:01	B*27:07	B*08:01	C*01:02	C*07:01
MDA-4347	ADC	F	Caucasian	A*03:01	A*03:02	B*08:01	B*15:01	C*03:04	C*07:39
MDA-4361	ADC	M	Hispanic	A*68:03	A*11:01	B*39:05	B*40:138	C*03:04	C*07:02
MDA-4363	ADCA/SCC	F	Caucasian	A*11:01	A*02:01	B*35:01	B*57:01	C*06:02	C*04:01
MDA-4378	ADC	F	Caucasian	A*29:02	A*11:01	B*51:01	B*44:03	C*15:02	C*16:01
MDA-4382	SCC	M	Hispanic	A*31:01	A*11:01	B*52:01	B*39:06	C*07:02	C*12:02
MDA-4383	ADC	M	Caucasian	A*02:01	A*02:01	B*40:01	B*57:01	C*06:02	C*03:04
MDA-4385	ADC	F	Caucasian	A*26:01	A*26:01	B*37:01	B*08:01	C*06:02	C*07:01
MDA-4386	ADC	M	Caucasian	A*03:01	A*03:01	B*08:01	B*18:01	C*07:01	C*07:01

HLA class I genotypes and clinical attributes for the MDACC NSCLC cohort (Reuben *et al.*, 2020 Nat Commun). ADC, adenocarcinoma; SCC, squamous cell carcinoma.

Table S4, related to Figure 2.

Patient ID	scTCR-seq	scRNA-Seq	Histology (tumor)	Histology (adjacent lung tissue)	Exome seq	Age	Gender	HLA-A*02	Smoking (pk yr)	Stage	Histology
A2	yes					65	M	N	50	IB	SCC
A3	yes					71	M	Y	50	IB	SCC
A4	yes		yes			49	M	Y	0	IA	adeno
A5	yes		yes	yes		65	F	N	0	IB	adeno
A6*	yes		yes	yes		73	F	Y	45	IB	SCC
A7			yes	yes		64	M	Y	118	IB	adeno
A8			yes	yes		71	F	Y	60	IB	adeno
A9			yes			77	M	N	45	IB	adeno
A10			yes	yes		82	F	N	0	IIA	adeno
A11	yes	yes	yes	yes	yes	76	F	Y	50	IIIA	adeno
A14			yes	yes		62	F	Y	10	IA	adeno
A15	yes	yes	yes	yes	yes	71	F	N	3	IB	adeno
A16	yes	yes	yes	yes	yes	68	F	N	20	IB	adeno
A17	yes	yes	yes	yes	yes	65	F	Y	0	IA	adeno
A18	yes	yes	yes	yes		65	M	N	30	IB	adeno
A19	yes	yes	yes	yes	yes	70	F	N	37	IIA	adeno
A21			yes	yes		31	M	N	0	IIIA	adeno
A28	yes	yes	yes	yes	yes	64	M	N	24	IA	adeno
A29	yes	yes	yes	yes	yes	64	F	Y	0	IA	adeno
A31	yes	yes	yes	yes	yes	68	F	Y	0	IIIB	adeno
A32	yes	yes	yes	yes	yes	49	F	N	0	IB	adeno
Total # with yes	15	10	19	17	9	-	-	-	-	-	-

Data availability and clinical attributes of the Stanford NSCLC cohort. Single-cell TCR-seq was performed on 15 patients, single-cell RNA-seq was performed on 10 patients, histology was performed on tumors from 19 patients and the adjacent lung tissues from 17 patients, and 9 NSCLC patients have tumors assessed with whole-exome sequencing (yes, data available). A6*, the paired TCR CDR3 α/β sequences from the tumor of patient A6 were derived by using the Chromium Single Cell Immune Profiling Solution for human T cell repertoires from the 10x Genomics. M, male; F, female; N, HLA-A*02⁻; Y, HLA-A*02⁺; adeno, adenocarcinoma; SCC, squamous cell carcinoma.

Table S5, related to Figure 3.

Rank	Peptide seq (Naive)	#	Peptide seq (Round 1)	#	Peptide seq (Round 2)	#	Peptide seq (Round 3)	#	Peptide seq (Round 4)	#
1	SMQGLTRPPRV	43	VMRQLACSRRL	25	RLHVAPGQRRM	149	AMGGLLTQLAM	14764	AMGGLLTQLAM	70220
2	AMGGLLTQLAM	30	FLQNGRSSERM	24	RLREAAPIPLM	108	EMRVASAMWGM	1205	KLGLLTMVGV	23704
3	ALRTPRKKQTL	17	RLQSRKLPPLL	22	PMDTHLPTSVL	89	GMVEDICSTLL	712	GMVEDICSTLL	18950
4	CLTQSSQEWTL	16	TLARTHHTRRL	20	AMQERLGRQV	88	RLHEQVPEVTL	557	EMRVASAMWGM	11431
5	KLDSANAQIV	13	HMHRPRPSGRL	19	WLTAALPCGV	88	HLRVKEHRHGL	469	RLHEQVPEVTL	3165
6	PMLTRSPLSRM	10	DLLVQKIRHL	18	CLPWQKTESPL	81	HMHHHNEHNRM	468	KMKMKHHRPL	2760
7	WMRQMEPSGRL	10	RLNELRLAKGL	18	RLTRTQGSSETL	78	RLHVAPGQRRM	315	HMHHHNEHNRM	1229
8	GMLASLAARQV	10	RLHKAPRPAPL	18	NMEALGHCHRL	77	PLACAKRRGGM	280	ALNLPSSPSSL	1019
9	EMTLPTAKWL	9	DLQSTLEAWAV	17	RMEQVATARVL	73	NLKLMIWIEM	268	KMPHRPKHKVV	987
10	WLSLAAPPAL	9	RMSSRPSLLL	17	RLAEPHSWIRL	67	ALNLPSSPSSL	264	HLRVKEHRHGL	968
11	RLWQRPVTRL	9	GMRTGPRWRHL	17	RLHVRPGRPL	67	NLRHRHPSRFL	262	HMHEHRVKGKV	841
12	GLGIAPALTDL	9	HMTGKHRTHMV	17	MMRTGPLGLL	67	SMPKRGTQVTM	261	RLHVRPGRPL	834
13	RMPTRRPTKRM	9	AMHRNSRWHS	16	RMLTRSRLAL	67	HMHEHRVKGKV	260	WLETPTPLLL	818
14	PLAHTPPRRTL	9	ALSARLPQAL	16	HMSRTHAPGQL	67	RLHVRPGRPL	259	PLACAKRRGGM	573
15	TLMRHLHAGVVL	9	RLHAPTHHSL	16	MMSCPAQEPCL	67	ALREKPRLPRL	257	NLRHRHPSRFL	459
16	RMGPLALSPSM	9	ALSARLSRPV	16	TMKTQGGQESM	66	RLPARHPPEAL	255	PLTSARKPRM	396
17	AMPNPRVREHL	9	RLPPLPQWPL	15	RLAHRARQVQV	65	RLPEPPARPL	249	RLHGRPTRWNL	382
18	AMLTLTRGQQM	9	PLTPTPRPGSM	15	RMSGKPPQHPL	65	RLAKVPQGT	248	HMPRHRWAARV	359
19	PLPPPLRPPM	9	PLLATARTWV	15	NLLPWRTKLM	63	QLRLASTPRGL	247	SMGGLLTQLAM	350
20	RMTMTARRRPV	9	ALEASWEWTGM	15	VMRTPRRPVPL	63	QMTARGRVHLL	234	RMPAAARKRGV	326
21	GLLQMLMSLAV	9	VLPAHAPGGL	15	KLAAGKQGLAM	63	RMACGQHPGPL	218	RLPARHPPEAL	314
22	RLPLEPHRQCM	8	ELARSHRAWSM	14	RLRAHQHMFRV	61	RMPAAARKRGV	217	HMRGGRHSRLM	302
23	LLASPTGVTKM	8	EMTCQLGPKPL	14	RLKPPQVPAL	61	RMLTRSRLAL	216	KMKSHHLRWGL	299
24	RLQVLPNSPCV	8	GLPPSAARKVL	14	NLKLMIWIEM	60	KMHQHGQHRQL	214	PMEPSPCTGV	293
25	KLHPNTCAKGM	8	TLKSHHHWGQL	14	PLRPPSLPTKM	60	RLHGRPTRWNL	212	KMHQHGQHRQL	286
26	TLSPSTGTISM	8	RLIPWRTLLL	14	VLGWDEPGTV	60	LLPLAPRRGV	193	HLMGTRGHRCL	282
27	RLCARMWNQSV	8	RMPPTPAPKGM	14	EMLTGAVGTP	60	HLHSHARTPRM	190	ALREKPRLPRL	268
28	AMRPPSMATRL	8	HMPRDQNRVAL	14	PMIAIHPSSQL	59	KLGLLTMVGV	188	HLRLRHHHREL	267
29	ALPLRPARLSL	8	PLHGRQLRAL	14	TLQEQTARTRM	59	QLSGAHHHRAL	187	RMPRPPISPL	263
30	RLRWIMLRDAL	8	TLMKRPGLRGL	14	RMHARMTSKL	59	RLHHPHQLRPM	178	RLRPHMLPRL	260
31	CLESCLRAMQL	8	LLPPTPPHAV	14	AMPRAPRSSWV	59	HLMGTRGHRCL	175	HMARPGPHRHL	256
32	PLPLTQRTPAV	8	RLARSQQHSL	14	GLSGPCQPAL	58	RMPAQRVAQAM	170	HLHPHPPGAL	247
33	NLRQREGTRIL	7	RMDHMPWHVRL	14	RLQMSSRRHHV	58	KLNGQVSEML	169	HMHRPRPSGRL	241
34	RLPSLERWPRL	7	EMPMPAAQPL	14	RMTPLQAEAL	58	TLTPAQPPVLL	168	HMQRGTHTKV	229
35	GMWARSVRASL	7	HLSKPRRRHEL	14	TMPPGLWQAV	58	LMCTMAAASRL	168	HMPKPRTRVPL	208
36	ALRGVRRTRVL	7	ALCLASREVL	14	RLAPPRLPHIV	57	RLPTMGTPPL	167	HLHSHARTPRM	206
37	TLLVSTLRGCL	7	TMQHGAAQTAL	13	RLPAPHRPETV	57	MLKRPRTPRQV	165	MLFDGLPLLV	202
38	RLSQSHRPRPL	7	TMRTCTERTPM	13	RLMWAKQEMLL	57	KMKMKHHRPL	164	RLHVAPGQRRM	193
39	WMRWVREPVRM	7	EMAPHAMLERV	13	RLSRPPLPTGV	56	HLRLRHHHREL	162	SMPKRGTQVTM	186
40	QLSAPLAPRSL	7	NLTWLPATRL	13	RLPMHPPALAV	56	PLSAWTATASL	162	RMPPPRPLAL	184
41	LLRGQRLEHLL	7	RLHQPPRPLPL	13	RLPEPPGPAAV	56	LLQEEHRHPTV	162	RLHHRVVLPL	179
42	CLLTTPTPTAL	7	RLDKVPRPGPL	13	RMARHRPLQL	56	RMPRPPISPL	160	QMTARGRVHLL	177
43	QLTRWRHRSKGM	7	GLQRVTGHRPL	13	RMACGQHPGPL	56	RLHKSPPWRSV	160	RLPLPAPRLV	177
44	HLPWLPKHGRM	7	KLAPPNRPPL	13	KLFSGKSGQVL	55	RLPARCLGAV	160	RLPEPPGPAAV	172
45	PMAFRGWHTWM	7	HMRRPPVRL	13	RLPARPGLRV	55	RMPPLQSGCL	159	TLAPPAPRPL	169
46	TMHGPRSRRL	7	ELPGCAPQCLM	13	GMSMGPSQCR	55	RLRPGSNAVLM	153	TLPLHQPPKLV	169
47	SLAWRAGAMDM	7	PLGPPRLRPL	13	DMTSRLVRWAM	54	HMPRHRWAARV	153	HLARPPPARL	164
48	RLNPKGTSAM	7	SLPSAPRHGNL	13	ELRRAEELRNV	54	ALAPTRATRRV	151	RMRAPHPHTRL	164
49	RLSKSLRHTCM	7	PMPAPPLTLP	13	QLRLASTPRGL	54	DLLPQPWARGM	149	RMRFSHPHKQL	162
50	AMMQSLPAPGM	7	VLLPPLESPL	13	KMRRTGLLDV	54	PMVSGEHRGAV	149	AMARPPPARL	161

Summary of results from each round of affinity selection on a 11mer yeast HLA-A*02 library with TCR2. Numbers (#) indicate counts of the top-50 mimotopes. Top-two mimotopes from the round 4 screen were highlighted.

Table S6, related to Figure 4.

Sample ID	Gene ID	Unique mut ID	Var class	Var type	AA var	coverage	Ref depth	Alt depth	af	dbsnp	exac	counts_cohomic	filter
A6	ABCA13	chr7:48313276:C>G	exonic	nonsense	S1338X	74	64	10	0.144	FALSE	FALSE	0	PASS
A6	ABCC11	chr16:48201528:G>T	exonic	nonsynonymous_snv	T1312K	143	119	24	0.172	FALSE	FALSE	0	PASS
A6	ADAM7	chr8:24346811:G>A	exonic	nonsynonymous_snv	D411N	71	55	16	0.233	FALSE	FALSE	0	PASS
A6	ADGRB3	chr6:69653730:G>C	exonic	nonsynonymous_snv	V347L	74	64	10	0.145	FALSE	FALSE	0	PASS
A6	AKAP9	chr7:91707139:G>A	exonic	nonsynonymous_snv	E2291K	110	96	14	0.133	FALSE	FALSE	0	PASS
A6	ALS2	chr2:202622283:C>A	exonic	nonsynonymous_snv	S438I	92	76	16	0.18	FALSE	FALSE	0	PASS
A6	ANK2	chr4:114274311:G>C	exonic	nonsynonymous_snv	D1513H	85	68	17	0.206	FALSE	FALSE	0	PASS
A6	ANKFN1	chr17:54520251:G>T	exonic	nonsynonymous_snv	W355C	86	75	11	0.136	FALSE	FALSE	0	PASS
A6	ANKLE2	chr12:133327337:C>T	exonic	nonsynonymous_snv	E247K	265	200	65	0.246	FALSE	FALSE	0	PASS
A6	ANKS1B	chr12:99640454:T>G	exonic	nonsynonymous_snv	K649Q	83	72	11	0.14	FALSE	FALSE	0	PASS
A6	ARHGEF17	chr11:73021562:A>C	exonic	nonsynonymous_snv	T627P	360	290	70	0.196	FALSE	FALSE	0	PASS
A6	ARL2	chr11:64785853:G>A	exonic	nonsynonymous_snv	G28E	318	265	53	0.168	FALSE	FALSE	0	PASS
A6	ASB9	chrX:15266932:C>T	exonic	nonsynonymous_snv	G232S	204	167	37	0.184	FALSE	FALSE	0	PASS
A6	ASIC2	chr17:32483281:G>T	exonic	nonsynonymous_snv	L91I	513	426	87	0.17	FALSE	FALSE	0	PASS
A6	ATP13A5	chr3:193048947:T>C	exonic	nonsynonymous_snv	M476V	115	71	44	0.385	FALSE	FALSE	0	PASS
A6	ATP4B	chr13:114312422:C>T	exonic	nonsynonymous_snv	R13H	685	631	54	0.079	TRUE	TRUE	0	PASS
A6	ATP7A	chrX:77296175:G>A	exonic	nonsynonymous_snv	E1249K	235	211	24	0.104	FALSE	FALSE	0	PASS
A6	AUTS2	chr7:69364300:G>A	exonic	nonsynonymous_snv	R113H	80	52	28	0.353	TRUE	TRUE	3	PASS
A6	BAI3	chr6:69653730:G>C	exonic	nonsynonymous_snv	V347L	74	64	10	0.135	FALSE	FALSE	0	PASS
A6	BCOR	chrX:39933652:G>C	exonic	nonsynonymous_snv	P316R	654	593	61	0.094	FALSE	FALSE	0	PASS
A6	BLVR4	chr7:43830961:A>T	exonic	nonsynonymous_snv	Y83F	73	61	12	0.173	FALSE	FALSE	0	PASS
A6	C1orf173	chr1:75065441:C>T	exonic	nonsynonymous_snv	R555H	67	58	9	0.134	TRUE	TRUE	1	PASS
A6	C20orf141	chr20:2795930:C>A	exonic	nonsynonymous_snv	R34S	498	421	77	0.155	FALSE	FALSE	0	PASS
A6	C20orf203	chr20:31238353:C>A	exonic	nonsynonymous_snv	A156S	742	622	120	0.162	FALSE	FALSE	0	PASS
A6	CACNA1A	chr19:13409626:C>G	exonic	nonsynonymous_snv	E941Q	697	576	121	0.174	FALSE	FALSE	0	PASS
A6	CAST	chr5:96031574:C>G	exonic	nonsynonymous_snv	S58C	54	37	17	0.32	FALSE	FALSE	0	PASS
A6	CC2D2A	chr4:15504120:G>C	exonic	nonsynonymous_snv	E106Q	232	177	55	0.239	FALSE	FALSE	0	PASS
A6	CCDC159	chr19:11460855:T>A	exonic	nonsynonymous_snv	I69N	149	135	14	0.098	FALSE	FALSE	0	PASS
A6	CCDC22	chrX:49104195:C>A	exonic	nonsynonymous_snv	A353D	1037	946	91	0.088	FALSE	FALSE	0	PASS
A6	CCN1	chr10:97817990:G>A	exonic	nonsynonymous_snv	E371K	120	93	27	0.229	FALSE	FALSE	1	PASS
A6	CEP128	chr14:81251610:C>T	exonic	nonsynonymous_snv	E614K	109	75	34	0.315	FALSE	FALSE	0	PASS
A6	CEP95	chr17:62533725:T>C	exonic	nonsynonymous_snv	L765S	63	51	12	0.2	FALSE	FALSE	0	PASS
A6	CHIC1	chrX:72783256:G>C	exonic	nonsynonymous_snv	E46Q	338	270	68	0.203	FALSE	FALSE	0	PASS
A6	CLGN	chr4:141317335:T>C	exonic	nonsynonymous_snv	I303M	149	120	29	0.197	FALSE	FALSE	0	PASS
A6	CNBD1	chr8:88249177:T>C	exonic	nonsynonymous_snv	L203P	67	48	19	0.288	FALSE	FALSE	0	PASS
A6	COL22A1	chr8:139618637:G>C	exonic	nonsynonymous_snv	P1364R	69	55	14	0.211	FALSE	FALSE	0	PASS
A6	COL4A1	chr13:110835623:C>A	splicing	splice	G633V	151	109	42	0.278	FALSE	FALSE	0	PASS
A6	COL9A1	chr6:70965071:G>T	exonic	nonsynonymous_snv	P509H	61	55	6	0.109	FALSE	FALSE	0	PASS
A6	CTNNA2	chr2:80816447:C>A	exonic	nonsynonymous_snv	P676T	114	88	26	0.232	FALSE	FALSE	0	PASS
A6	DAW1	chr2:228767798:G>C	exonic	nonsynonymous_snv	Q192H	175	149	26	0.152	FALSE	FALSE	0	PASS
A6	DCAF8L1	chrX:27999212:T>A	exonic	nonsynonymous_snv	E80D	457	372	85	0.187	FALSE	FALSE	0	PASS
A6	DECR1	chr8:91049143:A>G	exonic	nonsynonymous_snv	K205E	58	49	9	0.166	FALSE	FALSE	0	PASS
A6	DIRAS3	chr1:68512702:C>G	exonic	nonsynonymous_snv	K93N	179	152	27	0.154	FALSE	FALSE	0	PASS
A6	DLC1	chr8:13356569:G>A	exonic	nonsynonymous_snv	R338C	89	79	10	0.12	TRUE	TRUE	2	PASS
A6	DNA2	chr10:70192032:G>T	exonic	nonsynonymous_snv	Q602K	89	78	11	0.131	FALSE	FALSE	0	PASS
A6	DNAH3	chr16:21069431:T>A	exonic	nonsynonymous_snv	Q1300H	53	46	7	0.145	FALSE	FALSE	0	PASS
A6	DNAH6	chr2:85035598:C>A	exonic	nonsynonymous_snv	L3891M	81	73	8	0.108	FALSE	FALSE	0	PASS
A6	DUSP27	chr1:167095602:G>A	exonic	nonsynonymous_snv	G412R	199	141	58	0.294	FALSE	FALSE	0	PASS
A6	DYNC111	chr7:95614255:G>C	exonic	nonsynonymous_snv	D254H	102	61	41	0.404	FALSE	FALSE	0	PASS
A6	EDNRB	chr13:78493532:C>A	exonic	nonsynonymous_snv	E73D	157	118	39	0.252	FALSE	FALSE	0	PASS
A6	EHMT1	chr9:140708956:A>T	exonic	nonsynonymous_snv	D1085V	346	276	70	0.203	FALSE	FALSE	0	PASS
A6	EMD	chrX:153609138:C>T	exonic	nonsynonymous_snv	S142F	295	235	60	0.205	FALSE	FALSE	0	PASS
A6	ERICH3	chr1:75065441:C>T	exonic	nonsynonymous_snv	R555H	67	58	9	0.1343	TRUE	TRUE	1	PASS
A6	FAM181B	chr11:82443751:G>A	exonic	nonsynonymous_snv	R341C	348	308	40	0.117	FALSE	FALSE	0	PASS
A6	FAT2	chr5:150886965:C>A	exonic	nonsynonymous_snv	R4089S	192	142	50	0.263	FALSE	FALSE	0	PASS
A6	FAT3	chr11:92086057:A>G	exonic	nonsynonymous_snv	N260S	130	118	12	0.097	FALSE	FALSE	0	PASS
A6	FBXO4	chr5:41927145:C>G	exonic	nonsynonymous_snv	L74V	130	116	14	0.113	FALSE	FALSE	0	PASS
A6	FGD1	chrX:54472725:G>T	exonic	nonsynonymous_snv	F901L	527	481	46	0.087	FALSE	FALSE	0	PASS
A6	FGFR2	chr10:123324084:G>A	exonic	nonsynonymous_snv	S129L	70	56	14	0.207	FALSE	FALSE	0	PASS
A6	FICD	chr12:108913075:C>A	exonic	nonsense	Y400X	249	207	42	0.171	FALSE	FALSE	0	PASS
A6	FIG4	chr6:110064446:C>T	exonic	nonsynonymous_snv	S337L	51	44	7	0.15	FALSE	FALSE	0	PASS
A6	FLT4	chr5:180041175:C>T	exonic	nonsynonymous_snv	R1075Q	682	543	139	0.204	FALSE	FALSE	2	PASS
A6	FPR1	chr19:52249356:G>T	exonic	nonsynonymous_snv	P298T	235	177	58	0.249	FALSE	FALSE	0	PASS
A6	FREM1	chr9:14859371:G>C	exonic	nonsynonymous_snv	F147L	184	156	28	0.156	TRUE	TRUE	0	PASS
A6	FRK	chr6:116325135:G>C	exonic	nonsense	S124X	69	57	12	0.181	FALSE	FALSE	0	PASS
A6	GALNT13	chr2:154996951:C>T	exonic	nonsense	Q82X	82	59	23	0.285	FALSE	FALSE	0	PASS
A6	GLB1L	chr2:220107905:C>G	exonic	nonsynonymous_snv	L68F	193	158	35	0.184	FALSE	FALSE	0	PASS
A6	GNA12	chr7:2883750:C>T	exonic	nonsynonymous_snv	E16K	40	24	16	0.405	FALSE	FALSE	0	PASS
A6	GOLGA6L2	chr15:23686407:C>A	exonic	nonsynonymous_snv	R405S	737	644	93	0.127	FALSE	FALSE	0	PASS

A6	RP11-321N4.5	chr6:86322638:C>A	splicing	splice	G85G	68	57	11	0.162	FALSE	FALSE	0	PASS
A6	RP11-399J13.3	chr11:64785853:G>A	exonic	nonsynonymous_snv	G28E	324	271	53	0.164	FALSE	FALSE	0	PASS
A6	RTC8	chr22:32792234:C>T	exonic	nonsynonymous_snv	A273T	77	65	12	0.164	FALSE	FALSE	0	PASS
A6	RTP2	chr3:187416515:T>C	exonic	nonsynonymous_snv	H150R	883	685	198	0.225	FALSE	FALSE	0	PASS
A6	SACS	chr13:23911873:C>T	exonic	nonsynonymous_snv	E2048K	107	92	15	0.146	FALSE	FALSE	1	PASS
A6	SC5D	chr11:121177887:T>C	exonic	nonsynonymous_snv	L189S	116	102	14	0.126	TRUE	TRUE	0	PASS
A6	SHISA8	chr22:42306502:C>A	exonic	nonsynonymous_snv	G241C	530	461	69	0.131	FALSE	FALSE	0	PASS
A6	SLC10A2	chr13:103703691:A>T	exonic	nonsynonymous_snv	L226Q	132	83	49	0.373	TRUE	TRUE	0	PASS
A6	SLC12A8	chr3:124854600:C>A	exonic	nonsense	E217X	192	143	49	0.257	TRUE	TRUE	0	PASS
A6	SLC14A1	chr18:43319130:G>T	exonic	nonsynonymous_snv	L222F	93	67	26	0.284	FALSE	FALSE	0	PASS
A6	SLC16A2	chrX:73740853:G>C	exonic	nonsynonymous_snv	M153I	188	141	47	0.252	FALSE	FALSE	0	PASS
A6	SLC2A13	chr12:40441855:C>G	exonic	nonsynonymous_snv	W238C	74	64	10	0.143	FALSE	FALSE	1	PASS
A6	SLC40A1	chr2:190428865:T>A	exonic	nonsynonymous_snv	T283S	198	179	19	0.099	FALSE	FALSE	0	PASS
A6	SLC4A10	chr2:162627561:G>C	exonic	nonsynonymous_snv	E54Q	35	29	6	0.188	FALSE	FALSE	0	PASS
A6	SLCSA7	chr2:108627285:G>C	exonic	nonsynonymous_snv	E571Q	61	49	12	0.206	FALSE	FALSE	0	PASS
A6	SORCS1	chr10:108536359:T>C	exonic	nonsynonymous_snv	Y273C	66	59	7	0.118	TRUE	TRUE	0	PASS
A6	SPANXN1	chrX:144337210:G>C	exonic	nonsynonymous_snv	R32T	131	107	24	0.187	FALSE	FALSE	0	PASS
A6	SPEN	chr1:16260921:C>T	exonic	nonsynonymous_snv	A2729V	209	174	35	0.17	TRUE	TRUE	0	PASS
A6	SRRM1	chr1:24981357:C>T	exonic	nonsynonymous_snv	S351L	89	81	8	0.098	FALSE	FALSE	2	PASS
A6	ST3GAL4	chr11:126277484:G>T	exonic	nonsynonymous_snv	R120L	398	356	42	0.106	FALSE	FALSE	0	PASS
A6	SUGP1	chr19:19389597:C>T	exonic	nonsynonymous_snv	E513K	215	175	40	0.189	FALSE	FALSE	0	PASS
A6	SYNCRIP	chr6:86322638:C>A	exonic	nonsense	G551X	68	57	11	0.17	FALSE	FALSE	0	PASS
A6	SYNE1	chr6:152675975:T>C	exonic	nonsynonymous_snv	Y3582C	126	111	15	0.125	FALSE	FALSE	0	PASS
A6	TACR3	chr4:104512737:C>A	exonic	nonsynonymous_snv	W331L	82	66	16	0.202	FALSE	FALSE	0	PASS
A6	TAS2R13	chr12:11061142:C>G	exonic	nonsynonymous_snv	W252C	100	68	32	0.323	FALSE	FALSE	0	PASS
A6	TBC1D23	chr3:100038044:C>T	exonic	nonsynonymous_snv	P607S	115	104	11	0.099	FALSE	FALSE	0	PASS
A6	TBX1	chr22:19753930:G>T	exonic	nonsynonymous_snv	R343L	297	254	43	0.147	FALSE	FALSE	0	PASS
A6	TECTA	chr11:121028698:C>T	exonic	nonsynonymous_snv	S1485F	359	288	71	0.199	FALSE	FALSE	1	PASS
A6	TMEM200C	chr18:5890672:T>A	exonic	nonsynonymous_snv	Y464F	428	274	154	0.361	FALSE	FALSE	0	PASS
A6	TMEM239	chr20:2795930:C>A	exonic	nonsynonymous_snv	R34S	502	424	78	0.156	FALSE	FALSE	0	PASS
A6	TP53	chr17:7577082:C>A	exonic	nonsense	E247X	109	90	19	0.18	FALSE	FALSE	37	PASS
A6	TPO	chr2:1499952:G>A	exonic	nonsynonymous_snv	R733K	542	493	49	0.091	FALSE	FALSE	0	PASS
A6	TRPM8	chr2:234839319:G>A	exonic	nonsynonymous_snv	V42M	44	32	12	0.281	FALSE	FALSE	0	PASS
A6	TRRAP	chr7:98602936:G>A	exonic	nonsynonymous_snv	R359K	450	378	72	0.16	FALSE	FALSE	0	PASS
A6	TTN	chr2:179456135:G>T	exonic	nonsynonymous_snv	P20106H	125	94	31	0.252	FALSE	FALSE	1	PASS
A6	TUBGCP6	chr22:50659585:G>A	exonic	nonsynonymous_snv	S1068L	696	590	106	0.153	FALSE	FALSE	0	PASS
A6	URAD	chr13:28552584:C>A	exonic	nonsense	E61X	148	121	27	0.186	FALSE	FALSE	0	PASS
A6	USH2A	chr1:216595521:G>T	exonic	nonsynonymous_snv	P53Q	141	114	27	0.194	FALSE	FALSE	0	PASS
A6	UTP20	chr12:101767513:G>C	exonic	nonsynonymous_snv	D2367H	138	124	14	0.106	TRUE	TRUE	0	PASS
A6	VAX1	chr10:118896040:G>T	exonic	nonsense	Y124X	465	377	88	0.19	TRUE	TRUE	0	PASS
A6	VMA21	chrX:150565812:C>T	exonic	nonsynonymous_snv	A11V	264	223	41	0.157	FALSE	FALSE	0	PASS
A6	VPS13C	chr15:62170909:C>T	exonic	nonsynonymous_snv	E3347K	40	27	13	0.332	TRUE	FALSE	0	PASS
A6	VWA5A	chr11:123988993:G>T	exonic	nonsynonymous_snv	G115V	128	97	31	0.246	TRUE	TRUE	0	PASS
A6	WDR24	chr16:735301:C>A	exonic	nonsynonymous_snv	V659F	884	695	189	0.214	FALSE	FALSE	0	PASS
A6	XIRP2	chr2:167992549:G>T	exonic	nonsynonymous_snv	S180I	64	53	11	0.181	FALSE	FALSE	0	PASS
A6	XKR7	chr20:30584398:C>A	exonic	nonsynonymous_snv	P293Q	660	564	96	0.146	FALSE	FALSE	0	PASS
A6	YJEFN3	chr19:19640173:G>T	exonic	nonsynonymous_snv	R20S	707	577	130	0.184	FALSE	FALSE	0	PASS
A6	ZIC1	chr3:147131169:A>T	exonic	nonsynonymous_snv	Q392L	297	234	63	0.213	FALSE	FALSE	0	PASS
A6	ZNF197	chr3:44671013:G>C	exonic	nonsynonymous_snv	D123H	139	106	33	0.237	FALSE	FALSE	0	PASS
A6	ZNF197;ZNF660-ZNF197	chr3:44671013:G>C	exonic	nonsynonymous_snv	D123H	138	105	33	0.242	FALSE	FALSE	0	PASS
A6	ZNF367	chr9:99180195:G>C	exonic	nonsynonymous_snv	I40M	298	241	57	0.193	FALSE	FALSE	0	PASS
A6	ZNF383	chr19:3726574:A>G	exonic	nonsynonymous_snv	M45V	162	89	73	0.451	TRUE	TRUE	0	PASS
A6	ZNF41	chrX:47307429:T>A	exonic	nonsynonymous_snv	E580D	99	85	14	0.148	FALSE	FALSE	0	PASS
A6	ZNF768	chr16:30536686:G>A	exonic	nonsynonymous_snv	R259W	327	286	41	0.127	FALSE	FALSE	1	PASS

Exome-seq result of the resected tumor from Stanford NSCLC patient A6.