Image-guided Raman spectroscopy probe-tracking for tumour margin delineation

Conor C. Horgan,^{a,b,c} Mads S. Bergholt,^{a,b,c,#} May Zaw Thin,^d Anika Nagelkerke,^{a,b,c,†} Robert Kennedy,^e Tammy L. Kalber,^d Daniel J. Stuckey,^d Molly M. Stevens^{a,b,c,*}

^aDepartment of Materials, Imperial College London, London, UK.

^bDepartment of Bioengineering, Imperial College London, London, UK.

^cInstitute of Biomedical Engineering, Imperial College London, London, UK.

^dCentre for Advanced Biomedical Imaging, University College London, London, UK.

^eGuy's and St Thomas' NHS Foundation Trust, Oral/Head and Neck Pathology, King's College London, London, UK.

[#]Current address: Centre for Craniofacial and Regenerative Biology, King's College London, London, UK. [†]Current address: Groningen Research Institute of Pharmacy, Pharmaceutical Analysis, University of Groningen, The Netherlands.

Supplementary Information



Fig. S1 Schematic of image-guided Raman spectroscopic system software flow; (I) Video is initialised with diagnostic probe and interrogation area within FOV, (II) User interaction enables coloured-marker-based image segmentation which is combined with probe kinematic information for real-time probe tracking, (III) Once selected, the software begins near real-time data acquisition, continually tracking the diagnostic probe and recording spectral diagnostic information until the user starts a diagnostic acquisition, (IV) When the user starts a diagnostic acquisition, the coordinates of the probe are recorded and a detailed spectral signal acquired. This spectral signal is then diagnosed as either positive (V, i) or negative (V, ii) using a pre-developed spectroscopic diagnostic model. The diagnosis is then overlaid onto the imaging information at the coordinates where the measurement was acquired. Positive diagnosis coordinates are connected to form a boundary that delineates the tumour margin.



Fig. S2 (a-d) Image processing for coloured marker-based tracking takes (a) an input image frame and converts it from an RGB image to (b) an HSV image before (c) HSV-based image segmentation for identification and isolation of the coloured fiducial markers, which (d) is then combined with a priori knowledge of the probe geometry to calculate the pose of the spectroscopic probe and the location of the probe tip (black circle).



Fig. S3 (a-d) Sequential coloured marker-based tracking video frames of the spectroscopic probe during a video sequence, with identified fiducial markers and probe tip, demonstrating robust tracking performance following partial occlusion of the spectroscopic probe.



Fig. S4 (a) Video frame of mock Raman spectroscopic margin delineation video sequence of fresh chicken tissue used for characterisation of probe tracking error with overlaid ground truth and algorithm-determined probe-tip motion for entire video sequence. (b) Analysis of the processing time for the core spectroscopic margin delineation algorithm during (i) baseline loop iteration with 0.1 second integration time (prior to spectroscopic diagnostic acquisitions), (ii) initial spectroscopic diagnostic acquisition with 1 second integration time, (iii) subsequent spectroscopic diagnostic acquisitions (iv) baseline loop iterations with 0.1 second integration time, (iii) subsequent spectroscopic diagnostic acquisitions) (n = 3). (c-d) Tracking error for probe tip and fiducial markers in (c) X and (d) Y for the mock spectroscopic margin delineation video sequence.



Fig. S5 (a-c) Video frames from the image-guided Raman spectroscopic system showing the delineated region of chicken fat tissue with safety margins value of (a) 0, (b) 1.5 mm, and (c) 3 mm. (d-f) Sequential video frames from the image-guided Raman spectroscopic system indicating iterative suggested measurement locations (yellow squares) dependent on the spatial distribution of existing positive diagnostic acquisitions. (g-i) Video frames from the image-guided Raman spectroscopic system showing the delineated region of chicken fat tissue with diagnostic thresholds of (g) 0%, (h) 85%, and (i) 100% where green squares indicate locations of negative (non-cancerous) acquisitions and red squares indicate positive (cancerous) acquisitions. (j-k) Video frames from the image-guided Raman spectroscopic system showing the resulting AR display of spatial spectroscopic diagnostic coordinates when video stabilisation is (j) enabled and (k) disabled.



Fig. S6 (a) Video frame from the image-guided Raman spectroscopic system showing chicken muscle and chicken fat tissue. (b) Mean Raman spectra of chicken muscle tissue and chicken fat tissue (n = 25). (c) PLS-DA latent variable 1 (LV1) scores for chicken muscle tissue and chicken fat tissue Raman spectra. (d) PLS-DA latent variable 1.



Fig. S7 (a,c,e) Video frames from our image-guided Raman spectroscopic system showing the delineated margin of chicken fat tissue following 22-26 diagnostic spectral acquisitions with safety margin sizes of 0, 1.5 mm, and 3 mm for (a) specimen 1, (c) specimen 2, and (e) specimen 3. (b,d,f) Corresponding margin delineation accuracies indicating true positive (green), false negative (red), and false positive (yellow) diagnostic regions for (b) specimen 1, (d) specimen 2, and (f) specimen 3.



Fig. S8 Mean normalised true positive, false negative, and false positive margin delineated areas for the *ex vivo* chicken tissue specimens.



Fig. S9 (a-c) PLS-DA latent variables (a) 1, (b) 2, and (c) 3 for the PLS-DA Raman spectroscopic diagnostic model of human tissue biopsies.



Fig. S10 Magnified H&E stained section of SCC biopsy specimen (scale bar = 2 mm).



Fig. S11 (a-b) PLS-DA latent variables (a) 1 and (b) 2 for the PLS-DA Raman spectroscopic diagnostic model of SW1222 colorectal xenograft tumours in nu/nu mice.



Fig. S12 (a) Photographs of 0 μ M, 2 μ M, 4 μ M, and 20 μ M PPIX optical tissue phantoms inserted into chicken muscle tissue. (b-c) Fluorescence imaging of 0 μ M, 2 μ M, 4 μ M, and 20 μ M PPIX optical tissue phantoms in *ex vivo* chicken tissue. (d) Raw and (e) processed Raman spectra of the PPIX optical tissue phantoms and *ex vivo* chicken muscle tissue (n = 30). (f) PCA of the PPIX optical tissue phantoms and *ex vivo* chicken tissue Raman spectra.



Fig. S13 (a) Photograph of the 0 μ M PPIX optical tissue phantom inserted into chicken tissue used to generate the PLS-DA model. (b) Mean Raman spectra of chicken tissue and optical tissue phantom (n = 30). (c) PLS-DA latent variable 1 (LV1) scores for the chicken tissue and the optical tissue phantom Raman spectra. (d) PLS-DA latent variable 1.



Fig. S14 (**a**,**c**,**e**) Exemplar margin delineation of (**a**) 4 μ M PPIX, (**c**) 2 μ M PPIX, and (**e**) 0 μ M PPIX optical tissue phantoms using fluorescence-guided Raman spectroscopic margin delineation (with safety margins of 0, 3.5, and 7 mm) and fluorescence imaging. (**b**,**d**,**f**) Corresponding true positive, false negative, and false positive areas for fluorescence-guided Raman spectroscopic margin delineation (with safety margins of 0, 3.5, and 7 mm) and fluorescence imaging.



Fig. S15 (a-b) Mean phantom margin delineation accuracy for the 0 μ M, 2 μ M, and 4 μ M PPIX optical tissue phantoms (n = 3) using (a) fluorescence-guided Raman spectroscopic margin delineation and (b) fluorescence imaging. (c) Comparison of the mean margin delineation accuracy of fluorescence-guided Raman spectroscopic margin delineation and fluorescence imaging across the fluorescent optical tissue phantoms (2 μ M and 4 μ M PPIX).