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								ELISA	(OD 450	nm)
		CDR H3	Identity		CDR L3	Identity				
Antibody	V _H gene	(a.a.)	(%)	V⊾ gene	(a.a.)	(%)	Neut.	S2P	RBD	Ctrl
C1A-A1	IGHV1-69	15	99.31	IGKV3-11	11	99.64	-	2.46	0.26	0.00
C1A-A6	IGHV3-11	15	98.96	IGKV1-13	9	98.92	+	2.55	2.81	0.00
C1A-A11	IGHV3-30	20	99.31	IGKV1-33	9	99.28	+	2.87	2.85	00.0
C1A-B1	IGHV1-2*02	18	97.57	IGKV3-20	10	98.58	+	2.44	0.04	0.00
C1A-B3	IGHV3-53	14	97.89	IGKV1-9	9	98.92	++	2.66	2.94	0.00
C1A-B5	IGHV4-59*11	11	94.74	IGKV4-1*01	9	95.96	-	2.73	0.04	0.00
C1A-B6	IGHV1-24	14	98.61	IGLV2-8	10	99.31	-	3.08	0.00	0.00
C1A-B12	IGHV3-53	14	98.6	IGKV1-9	9	98.57	++	2.69	2.97	0.00
C1A-C1	IGHV3-23	14	99.31	IGKV1-39	10	100	++	2.95	2.97	0.00
C1A-C2	IGHV3-53	14	98.6	IGKV1-9	9	97.49	++	2.81	2.86	0.00
C1A-C4	IGHV3-53	14	98.25	IGKV1-9	9	97.13	++	2.89	2.85	0.00
C1A-C6	IGHV1-69	15	98.96	IGKV3-11	11	98.92	-	2.58	0.02	0.01
C1A-C7	IGHV4-39	12	99.31	IGLV2-14	10	94.79	-	1.04	2.08	1.37
C1A-C8	IGHV1-24	16	99.65	IGLV3-21	12	98.57	-	3.02	0.00	0.00
C1A-C9	IGHV1-69	14	97.92	IGKV1-39	8	98.92	+	2.99	2.93	0.00
C1A-C10	IGHV3 -23	17	98.61	IGLV2-23	12	99.65	+	2.75	0.04	0.00
C1A-D1	IGHV4-39	18	91.75	IGKV1-33	9	93.19	-	2.06	0.30	0.05
C1A-D9	IGHV3-30	20	98.61	IGKV2-30	10	100	-	2.66	1.78	0.02
C1A-D11	IGHV1-69	14	98.26	IGKV3-11	11	99.28	-	2.39	0.00	0.00
C1A-E1	IGHV3-9*01	13	89.58	IGKV1-39	10	91.04	-	2.70	2.82	0.00
C1A-E3	IGHV1-18	16	98.61	IGKV1-39	5	98.92	+	2.74	2.86	0.00
C1A-E4	IGHV3-30	14	97.92	IGKV3-20	9	100	-	2.43	0.03	0.00
C1A-E8	IGHV3-48	21	97.22	IGKV2-28	9	100	-	2.73	0.02	0.01
C1A-E9	IGHV3-30	14	100	IGKV3-20	9	100	-	1.81	0.00	0.00
C1A-F2	IGHV3-30-3	15	99.65	IGKV2-28	8	99.66	-	2.25	0.01	0.00
C1A-F3	IGHV4-61	13	98.63	IGKV1-39	10	99.28	-	2.40	0.07	0.01
C1A-F4	IGHV3-30-3	14	99.65	IGKV1-5	9	99.64	+	2.84	2.77	0.00
C1A-F7	IGHV3-15	11	91.84	IGKV4-1	9	93.6	-	2.42	0.01	0.00
C1A-F10	IGHV3-53	14	97.89	IGKV1-9	9	98.21	++	2.98	2.69	0.00
C1A-F11	IGHV1-69	21	89.24	IGKV1-33	8	94.98	-	2.39	2.37	0.00
C1A-F12	IGHV4-39	15	90.03	IGLV1-40	11	95.49	-	2.62	2.78	0.01
C1A-G4	IGHV3-23	17	99.65	IGLV2-23	11	99.65	-	2.57	0.00	0.00
C1A-G9	IGHV3-30-3	14	98.61	IGKV3-20	8	100	-	2.46	0.01	0.00
C1A-G11	IGHV1-69	12	97.92	IGKV1-5	9	99.28	-	2.75	0.01	0.00
C1A-G12	IGHV3-30-3	14	98.26	IGKV3-15	8	98.92	-	2.51	0.07	0.07
C1A-H5	IGHV3-53	14	97.54	IGKV1-9	9	97.49	++	2.69	2.67	0.00
C1A-H6	IGHV3-53	14	98.95	IGKV1-9	9	98.21	++	2.90	2.74	0.04
C1A-H10	IGHV3-21	18	99.31	IGKV3-15	10	100	-	2.64	0.03	0.02
C1A-H11	IGHV3-30	16	98.61	IGLV2-14	10	99.31	-	2.27	0.00	0.00
C1A-H12	IGHV4-39	15	98.28	IGKV3-15	9	99.28	-	2.66	0.01	0.01
C1B-A3	IGHV3-30-3	14	99.65	IGKV3-11	9	100	-	2.00	0.02	0.01
C1B-A5	IGHV3-30-3	14	99.65	IGKV3-20	9	98.94	-	2.20	0.01	0.01
C1B-A11	IGHV3-30-3	14	98.26	IGKV3-20	9	99.29	-	2.35	0.28	0.00

Table S1, Related to Table 1. Properties of monoclonal antibodies isolated from COVID-19 convalescent individual C1.

Antibodies highlighted in gray are somatic variants of the same antibody. CDR loop lengths are shown as numbers of amino acids (a.a.). ELISA values are colored in shades of blue according to their magnitude; darker shades are reflective of a stronger signal. S2P: prefusion stabilized version of the SARS-CoV-2 S ectodomain; RBD: receptor-binding domain; Ctrl: negative control protein Lujo virus GP1. Neut: neutralizing activity as shown in Figure S1G. ++ signifies >90% neutralization, and + signifies neutralization not meeting this threshold.

	C1A-B3/RBD ^a (PDB 7KFV)	C1A-B12/RBD ^a (PDB 7KFW)	C1A-C2/RBD ^a (PDB 7KFX)	C1A-F10/RBD ^a (PDB 7KFY)
Data collection				
Space group Cell dimensions	P212121	P212121	C2221	C2221
a, b, c (Å)	84.8, 113.3, 268.89	84.8, 113.3, 268.9	83.5, 149.2, 146.1	85.7, 146.8, 144.6
α,β,γ (°)	90.00, 90.00, 90.00	90.00, 90.00, 90.00	90.00, 90.00, 90.00	90.00, 90.00, 90.00
Resolution (Å)	200-2.77 (2.94- 2.77) ^b	200-2.10 (2.23- 2.10) ^b	200-2.22 (2.36- 2.22) ^b	200-2.16 (2.29- 2.16) ^b
R _{sym} R _{pim} L/ a	0.38 [´] (3.17) ^b 0.11 (1.03) ^{b,c} 5 4 (0 7) ^b	0.238 (2.01) ^b 0.095 (0.85) ^{b,c} 9 0 (1 4) ^b	0.136 (2.23) ^b 0.062 (1.01) ^{b,c} 6 9 (0.6) ^b	0.118 (1.13) ^b 0.076 (0.61) ^{b,c} 6 2 (0 7) ^b
Completeness (%)	98.1 (88.6) ^b 6 9 (6 5) ^b	98.9 (94.7) ^b 7 0 (6 4) ^b	99.1 (97.1) ^b	94.0 (89.4) ^b 2 1 (2 0) ^b
Redundancy	0.9 (0.3)	7.0 (0.4)	5.0 (5.0)	2.1 (2.0)
Refinement				
Resolution (Å)	133.92-2.79	134.44-2.10	74.60-2.23	74.04-2.16
No. reflections	63645	149225	44541	48373
R _{work} / R _{free}	0.19/0.23	0.18/0.22	0.19/0.23	0.19/0.22
No. atoms				
Protein	14414	14511	4861	4855
Ligand/ion	42	42	14	14
Water	400	1642	325	532
<i>B</i> -factors (Ų)				
Protein	88	47	63	52
Ligand/ion	96	99	103	101
Water	66	53	65	55
R.m.s. deviations				
Bond lengths (Å)	0.008	0.008	0.008	0.008
Bond angles (°)	0.980	0.970	1.000	1.000

Table S2, Related to Figure 1. Data collection and refinement statistics

^a Numbers of crystals for C1A-B3, C1A-B12, C1A-C2 and C1A-F10 data were 1 each. ^b Values in parentheses are for the highest-resolution shell. ^c Values from program *aimless*.

Table S3, Related to Figures 3 and 4. Human derived SARS-CoV-2 S sequences containing mutations of interest.

Virus name	PANGO lineage	Accession No.	S mutations of interest	Location	Collection date	Originating laboratory	Submitting laboratory	Authors
hCoV- 19/USA/FL- CDC-STM- 000013- F04/2021	B.1.1.7	EPI_ISL_884605	D614G, N501Y, Q493K, Y114del	Florida, United Sates	01/14/2021	Respiratory Viruses Branch, Centers for Disease Control and Prevention	Respiratory Viruses Branch, Centers for Disease Control and Prevention	Cook, P.W., Batra, D., Rambo- Martin, B.L., de Feo, E., Antico, J., Tran, C., Tolentino, M., Wickline, S., Gietzen, K., Sickler, B., Liu, J., Allen, E., Febbo, P., Galloway, S., Washington, N.L., White, S., Levan, G., Barret, K.S., Cirulli, E., Bolze, A., Ascencio, A., Rivera-Garcia, C., Cho, R., Nguyen, J., Wang, S., Ramirez, J., Cassens, T., Sandoval, E., Isaksson, M., Lee, W., Becker, D., Laurent, M., Lu, J., Paden, C.R., Tong, S., MacCannell, D.
nCoV- 19/Israel/CVL- 618ngs/2020	B.1.362	EPI_ISL_889024	Q493K	Israel	12/29/2020	Central Virology Laboratory	Israel National Consortium for SARS- CoV-2 sequencing	Neta Zuckerman, Errat Danan Bucris, Michal Mandelboim, Dana Bar-Ilan, Oran Erster, Tzvia Mann, Oran Erster, A. Zeevi, Assaf Rokney, Joseph Jaffe, Eva Nachum, Maya Davidovich Cohen, Ephraim Fass, Gal Zizelski Valenci, Mor Rubinstein, Efrat Rorman, Israel Nissan, Efrat Glick-Saar, Omri Nayshool, Gideon Rechavi, Ella Mendelson, Orna Mor
hCoV- 19/Switzerland/ BS- 42473265/2020	B.1.160	EPI_ISL_830854	D614G, Q493K	Basel, Switzerland	10/07/2020	University Hospital Basel, Clinical Virology	University Hospital Basel, Clinical Bacteriology	Tim Roloff, Madlen Stange, Helena MB Seth-Smith, Alfredo Mari, Karoline Leuzinger, Julia Bielicki, Manuel Battegay, Hans Hirsch, Adrian Egli
hCoV- 19/Switzerland/ BS- 42471446/2020	B.1.160	EPI_ISL_830843	D614G, Q493K	Basel, Switzerland	10/06/2020	University Hospital Basel, Clinical Virology	University Hospital Basel, Clinical Bacteriology	Tim Roloff, Madlen Stange, Helena MB Seth-Smith, Alfredo Mari, Karoline Leuzinger, Julia Bielicki, Manuel Battegay, Hans Hirsch, Adrian Egli
hCoV- 19/England/MIL K- 11C2FCD/2021	B.1.1.7	EPI_ISL_1006449	D614G, Q493R, N501Y, Y144del	United Kingdom, England	01/28/2021	Lighthouse Lab in Milton Keynes	Wellcome Sanger Institute for the COVID- 19 Genomics UK (COG- UK) Consortium	The Lighthouse Lab in Milton Keynes and Alex Alderton, Roberto Amato, Sonia Goncalves, Ewan Harrison, David K. Jackson, Ian Johnston, Dominic Kwiatkowski, Cordelia Langford, John Sillitoe on behalf of the Wellcome Sanger Institute COVID-19 Surveillance Team
hCoV- 19/USA/CA- CDC-STM- A100413/2021	B.1.1.7	EPI_ISL_850699	D614G, N501Y, Y489H, Y144del	California, United States	01/07/2021	Helix / Illumina	Genomics and Discovery, Respiratory Viruses Branch, Division of Viral Diseases, Centers for Disease Control and Prevention	Peter W. Cook, Dhwani Batra, Ben L. Rambo-Martin Eileen de Feo, Jan Antico, Christine Tran, Matthew Tolentino, Shannon Wickline, Kim Gietzen, Brad Sickler, Jingtao Liu, Eric Allen, Phil Febbo, Summer Galloway, Nicole L. Washington, Simon White, Geraint Levan, Kelly Schiabor Barrett, Elizabeth Cirulli, Alexandre Bolze, Ary Ascencio, Charlotte Rivera- Garcia, Ryan Cho, Jason Nguyen, Sherry Wang, Jimmy Ramirez, Tyler Cassens, Efren Sandoval, Magnus Isaksson, William Lee, David Becker, Marc Laurent, James Lu, Clinton R. Paden, Suxiang Tong, Duncan MacCannell
hCoV- 19/England/MIL K-99469D/2020	B.1.1.7	EPI_ISL_552392	D614G, Y489H	Ünited Kingdom, England	08/25/2020	Lighthouse Lab in Milton Keynes	Wellcome Sanger Institute for the COVID- 19 Genomics UK (COG- UK) consortium	The Lighthouse Lab in Milton Keynes and Alex Alderton, Roberto Amato, Sonia Goncalves, Ewan Harrison, David K. Jackson, Ian Johnston, Dominic Kwiatkowski, Cordelia Langford, John Sillitoe on behalf of the Wellcome Sanger Institute COVID-19 Surveillance Team (http://www.sanger.ac.uk/covid- team)
hCoV- 19/England/205 261299/2020	B.1.1.7	EPI_ISL_754289	D614G, N501Y, Y144del	United Kingdom, England	12/20/2020	Respiratory Virus Unit, National Infection	COVID-19 Genomics UK (COG-	PHE Covid Sequencing Team

						Service, Public Health England	UK) Consortium	
hCoV-19/South Africa/Tygerber g-461/2020	B.1.351	EPI_ISL_745186	D614G, E484K, K417N, N501Y, L242del, A243del, L244del	South Africa, Western Cape	12/07/2020	Wallaceden e Clinic wc WAL	National Health Laboratory Service (NHLS), Tygerberg	Susan Engelbrecht, Kayla Delaney, Bronwyn Kleinhans, Houriiyah Tegally, Eduan Wilkindon, Gert van Zyl, Wolfgang Preiser, Tulio de Oliveira
hCoV- 19/Brazil/AM- 20143138FN- R2/2020	P.1	EPI_ISL_811149	D614G, K417T, E484K, N501Y	Brazil, Amazonas, Manaus	12/30/2020	Laboratorio de Ecologia de Doencas Transmissiv eis na Amazonia, Instituto Leonidas e Maria Deane - Fiocruz Amazonia	Laboratorio de Ecologia de Doencas Transmissiv eis na Amazonia, Instituto Leonidas e Maria Deane - Fiocruz Amazoni	Valdinete Nascimento, Victor Souza, André Corado, Fernanda Nascimento, George Silva, Ágatha Costa, Debora Duarte, Luciana Gonçalves, Matilde Mejía, Karina Pessoa, Maria Júlia Brandão, Michele Jesus, Felipe Naveca
hCoV- 19/England/MIL K- F9DBDB/2021	B.1.429	EPI_ISL_852237	D614G, E484K, N501Y, S494P	United Kingdom, England	01/08/2021	Lighthouse Lab in Milton Keynes	Wellcome Sanger Institute for the COVID- 19 Genomics UK (COG- UK) Consortium	The Lighthouse Lab in Milton Keynes and Alex Alderton, Roberto Amato, Sonia Goncalves, Ewan Harrison, David K. Jackson, Ian Johnston, Dominic Kwiatkowski, Cordelia Langford, John Sillitoe on behalf of the Wellcome Sanger Institute COVID-19 Surveillance Team
hCoV- 19/Mozambique /INS- K008170/2021	B.1.351	EPI_ISL_964941	D614G, K417N, E484K, N501Y, L242del, A243del, L244del	Africa, Mozambique	01/14/2021	Instituto Nacional de Saude (INS), Mozambique	KRISP, KZN Research Innovation and Sequencing Platform	Nalia Ismael, Nadia Sitoe, Paulo Arnaldo, Nedio Mabunda, Giandhari J, Pillay S, Emmanuel S, Tegally H, Wilkinson E, de Oliveira T
hCoV- 19/Denmark/D CGC- 5481/2020	B.1.1.29 8	EPI_ISL_620806	D614G, N440D, Y453F	Denmark, Nordjylland	09/28/2020	Department of Virus and Microbiologi cal Special Diagnostics, Statens Serum Institut, Denmark	Albertsen lab, Department of Chemistry and Bioscience, Aalborg University, Denmark	Danish Covid-19 Genome Consortia
hCoV- 19/England/MIL K- 11C2FCD/2021	B.1.1.7	EPI_ISL_1006449	D614G, N501Y, Q493R , Y144del	United Kingdom, England	01/28/2021	Lighthouse Lab in Milton Keynes	Wellcome Sanger Institute for the COVID- 19 Genomics UK (COG- UK) Consortium	The Lighthouse Lab in Milton Keynes and Alex Alderton, Roberto Amato, Sonia Goncalves, Ewan Harrison, David K. Jackson, Ian Johnston, Dominic Kwiatkowski, Cordelia Langford, John Sillitoe on behalf of the Wellcome Sanger Institute COVID-19 Surveillance Team
hCoV- 19/England/LO ND- 12E726E/2021	B.1.1.7	EPI_ISL_997803	D614G, F486L, N501Y, Y144del	United Kingdom, England	02/10/2021	University College London, Great Ormond Street Hospital for Children Trust, Imperial College Healthcare NHS Trust	COVID-19 Genomics UK (COG- UK) Consortium	Sergi Castellano, Rachel Williams, Mark Kristiansen, Paola Resende Silva, Sunando Roy, Tony Brooks, Helena Tutill, Paola Niola, Patricia Dyal, Charlotte Williams, Leysa Forrest, Yasmin Panchbhaya, Jacqueline Findlay, Samuel Weeks, Julianne Brown, Kathryn Harris, Paul Randell, James Price, Alison Holmes, Judith Breuer

Not all S mutations found in the respective sequences are shown. RBD mutations of interest are shown in bold, and NTD deletions relevant to those shown in Figure S7L are shown in regular font. The Y453F_{RBD} mutation found in hCoV-19/Denmark/DCGC-5481/2020 is shown because it is a REGN10933 resistance mutation detected in vitro (Baum et al., 2020) and has also been associated with mink-derived SARS-CoV-2 sequences. We gratefully acknowledge the listed authors from the originating laboratories responsible for obtaining the specimens and the submitting laboratories where genetic sequence data were generated and shared through the GISAID Initiative, on which this research is based.

Table S4, Related to Figures 5 and 6. Summary of the observed effects of S mutations on the activity of monoclonal antibodies and polyclonal serum IgG.

S mutations	Effect on monoc	lonal antibodies	Effect on	Additional notes		
	Resistance	Sensitive	polyclonal			
			serum IgG			
N439K	REGN10987 (with	All C1A-V⊦3-53	Modest to	Described as a circulating variant with		
	fourteenfold	antibodies, B38,	no effect in	maintained fitness (Thomson et al., 2021)		
	resistance)	CC12.1,	4/4 donors			
		REGN10933				
Q493K	C1A-V _H 3-53	C1A-V _H 3-53	Resistance	Also described through in vitro resistance		
	antibodies with low	antibodies with	in 1/4	mapping efforts (Baum et al., 2020; Weisblum et		
	RBD affinity, B38,	high RBD affinity,	donors	al., 2020)		
	REGN10933	CC12.1,		Observed in other recent human-derived		
		REGN10987		SARS-CoV-2 S sequences (Table S3)		
Q493R	C1A-V _H 3-53	C1A-V _H 3-53	Partial	Also described through in vitro resistance		
	antibodies with low	antibodies with	resistance	mapping efforts (Baum et al., 2020; Weisblum et		
	RBD affinity, B38,	high RBD affinity,	in 1/4	al., 2020)		
	REGN10933	CC12.1,	donors	Observed in other recent human-derived		
		REGN10987		SARS-CoV-2 S sequences (Table S3)		
Day 152*	All C1A-V _H 3-53	REGN10987	Resistance	Seven of the eight RBD mutations it contains		
	antibodies, B38,		in 4/4	have been observed in other human-derived		
	CC12.1,		donors	SARS-CoV-2 S sequences. The only exception		
	REGN10933		tested	is F486I, although it is similar to F486L, which		
				has been observed (Figure 3B; Table S3)		
Day 146*	All C1A-V _H 3-53	REGN10987 (with	Resistance	All of the RBD mutations it contains have been		
	antibodies, B38,	fourfold	in 4/4	observed at least individually in other human-		
	CC12.1,	resistance)	donors	derived SARS-CoV-2 S sequences (Figure 3B;		
	REGN10933			Table S3)		