



## Supporting Information

for *Adv. Sci.*, DOI: 10.1002/adv.202003091

Avidity-based selection of tissue-specific CAR-T cells  
from a combinatorial cellular library of CARs

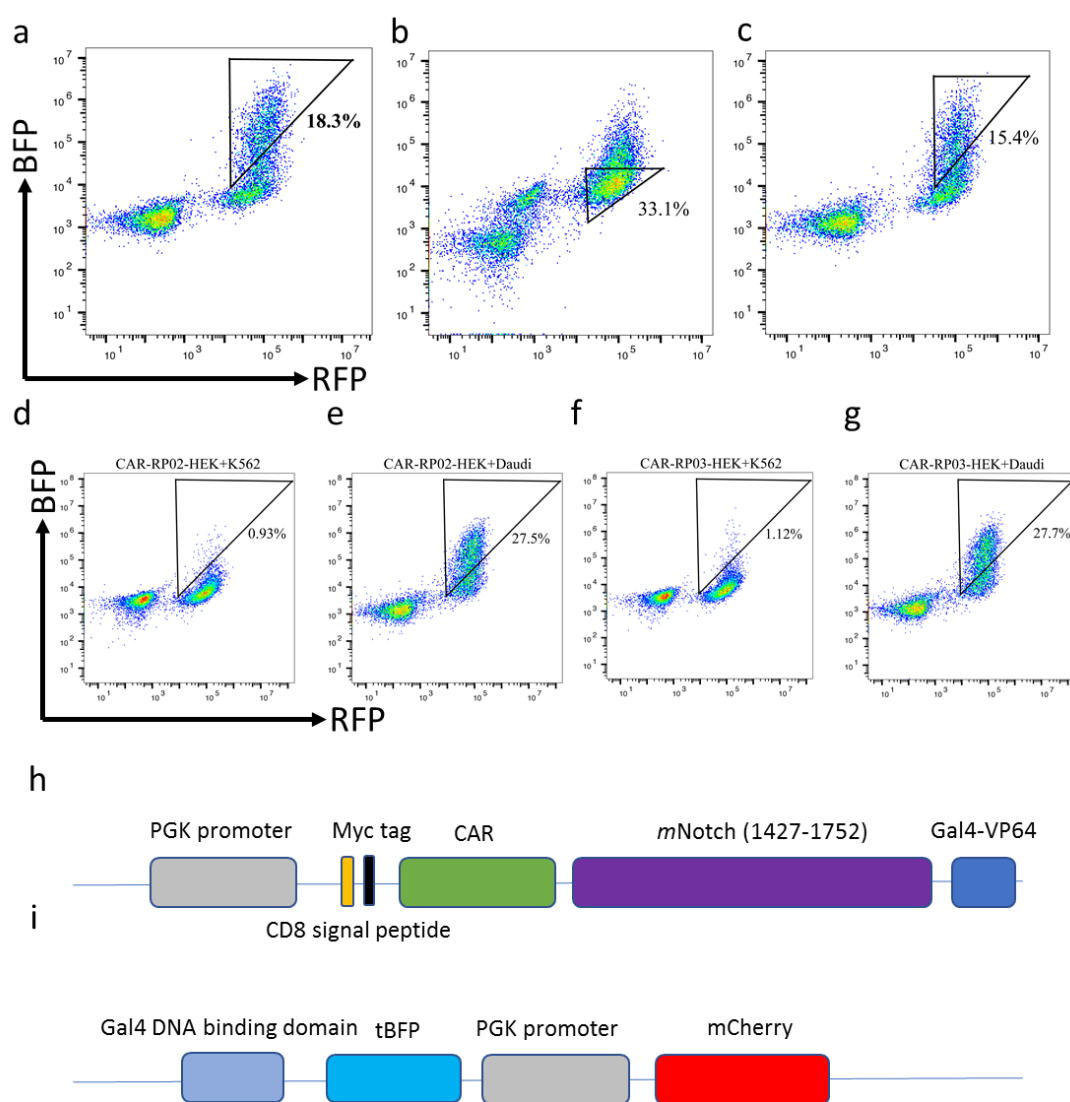
*Peixiang Ma<sup>#</sup>, Ping Ren<sup>#</sup>, Chuyue Zhang<sup>#</sup>, Jiaxing Tang<sup>#</sup>, Zheng Yu, Xuekai Zhu, Kun Fan, Guanglei Li, Wei Zhu, Wei Sang, Chenyu Min, Wenzhang Chen, Xingxu Huang, Guang Yang<sup>\*</sup>, Richard A Lerner<sup>\*</sup>*

## Supporting Information

### **Avidity-based selection of tissue-specific CAR-T cells from a combinatorial cellular library of CARs**

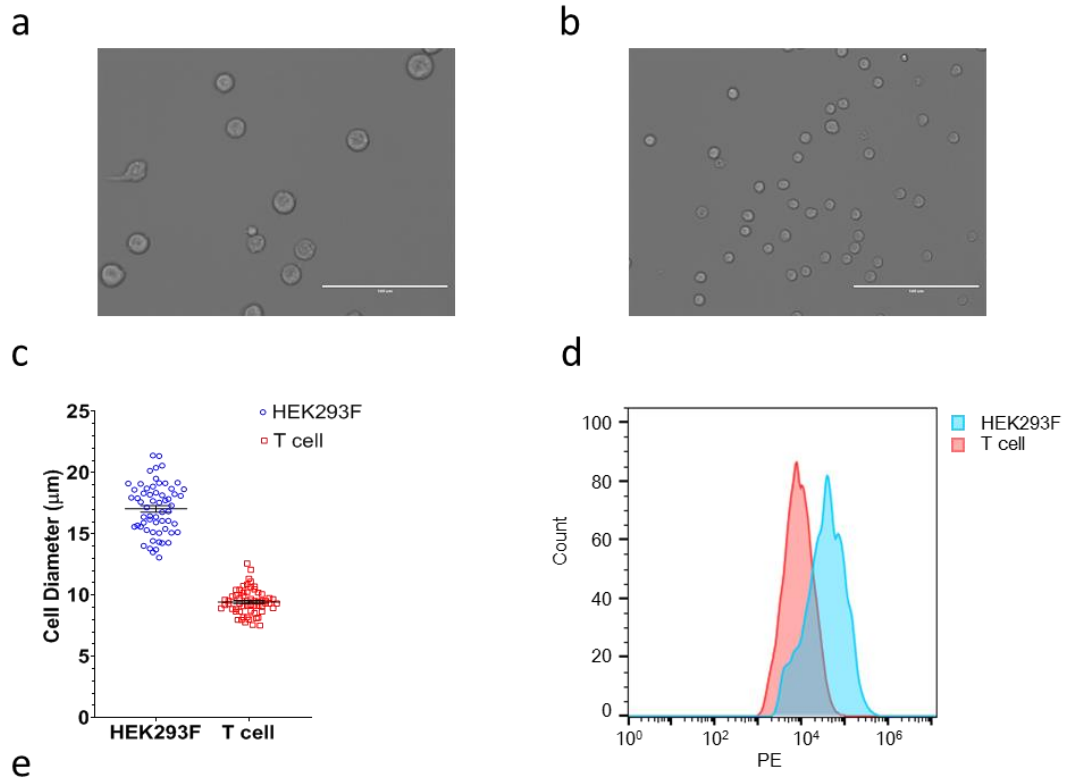
Peixiang Ma<sup>#</sup>, Ping Ren<sup>#</sup>, Chuyue Zhang<sup>#</sup>, Jiaying Tang<sup>#</sup>, Zheng Yu, Xuekai Zhu, Kun Fan, Guanglei Li, Wei Zhu, Wei Sang, Chenyu Min, Wenzhang Chen, Xingxu Huang, Guang Yang<sup>\*</sup>, Richard A Lerner<sup>\*</sup>.

## Supplementary Figures:



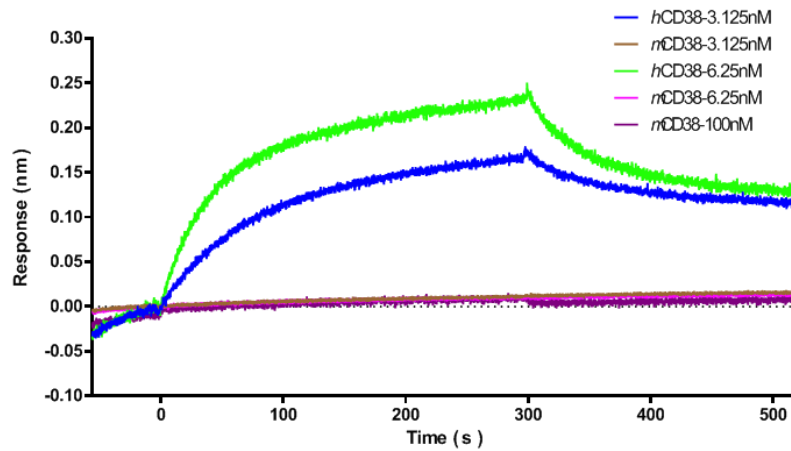
**Figure S1. Proof of concept of CAR selection using cell-cell interaction.** (a) The high-affinity scFV 028 was spiked into a single chain fragment variable (scFv) antibody library at a ratio of  $1:10^4$ . (b) In the positive selection group, the CAR-cell library was activated by K562-CD38 cells. (c) In the negative-positive selection group, initially healthy PBMCs with low expression of CD38 were used to activate the CAR-cell library. The activated cells were discarded as they were the result of activation by non-tumor cells. The remaining, non-activated cells were collected and exposed to K562-CD38 cells for activation. The positive cells were collected for sequencing. RP02-CAR synNotch was expressed in HEK-293 cells, then co-cultured with  $CD38^-$  (K562) cells (d) and  $CD38^+$  (Daudi) cells (e). RP03-CAR synNotch was

expressed in HEK-293 cells, then co-cultured with CD38<sup>-</sup> (K562) cells **(f)** and CD38<sup>+</sup> (Daudi) cells **(g)**. **(h)** The CAR-synNotch construct consists of an N-terminal extracellular domain containing a *hCD8α* signaling peptide, a myc peptide, and a CAR; a transmembrane domain consisting of *mNotch1* core (from 1427 to 1752 amino acids); and an intracellular domain consisting of Gal4-VP64. **(i)** The effector construct was designed to monitor chimeric Notch signaling, in which synNotch activation resulted in Gal4-VP64 release and, subsequently, BFP expression through interaction between the released Gal4-VP64 and the Gal4 DNA binding domain. A *mCherry* fluorescent protein was used in the construct as an internal transduction marker.



**Figure S2 | Comparison of CAR density on the cell surface of HEK293F and human T cells.** (a) Representative images of HEK293F cells grown in suspension. Scale bar is 100  $\mu\text{m}$ . (b) Representative images of human T cells grown in suspension. Scale bar is 100  $\mu\text{m}$ . (c) Diameter of HEK293F and primary T cells. The diameter of HEK293F is  $17.1 \pm 0.3$   $\mu\text{m}$  and primary T is  $9.4 \pm 0.1$   $\mu\text{m}$ . (n=60) (d) Cell surface synNotch on HEK293F and primary T cells was analyzed using PE-labelled rabbit anti- myc antibody. (e) Comparison of cell surface density of synNotch on HEK293F and primary T cells.

a



b

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hCD38-ECD    -VPRWRQQWSGPGTTKRFPEIVLARCVKYTEI-HPEMRHVDCQSVWDAFKGAFISKHPCN 58
mCD38-ECD    LRPRSLLVWTGEPPTTKHFSDFLGRCLIYTIQLRPEMRDQNCQEILSTFKGAFVSKNPCN 60
                **      *:*  ***:*  :  .*  **:  ***:*  :****.  :**  .  :*****:**:**

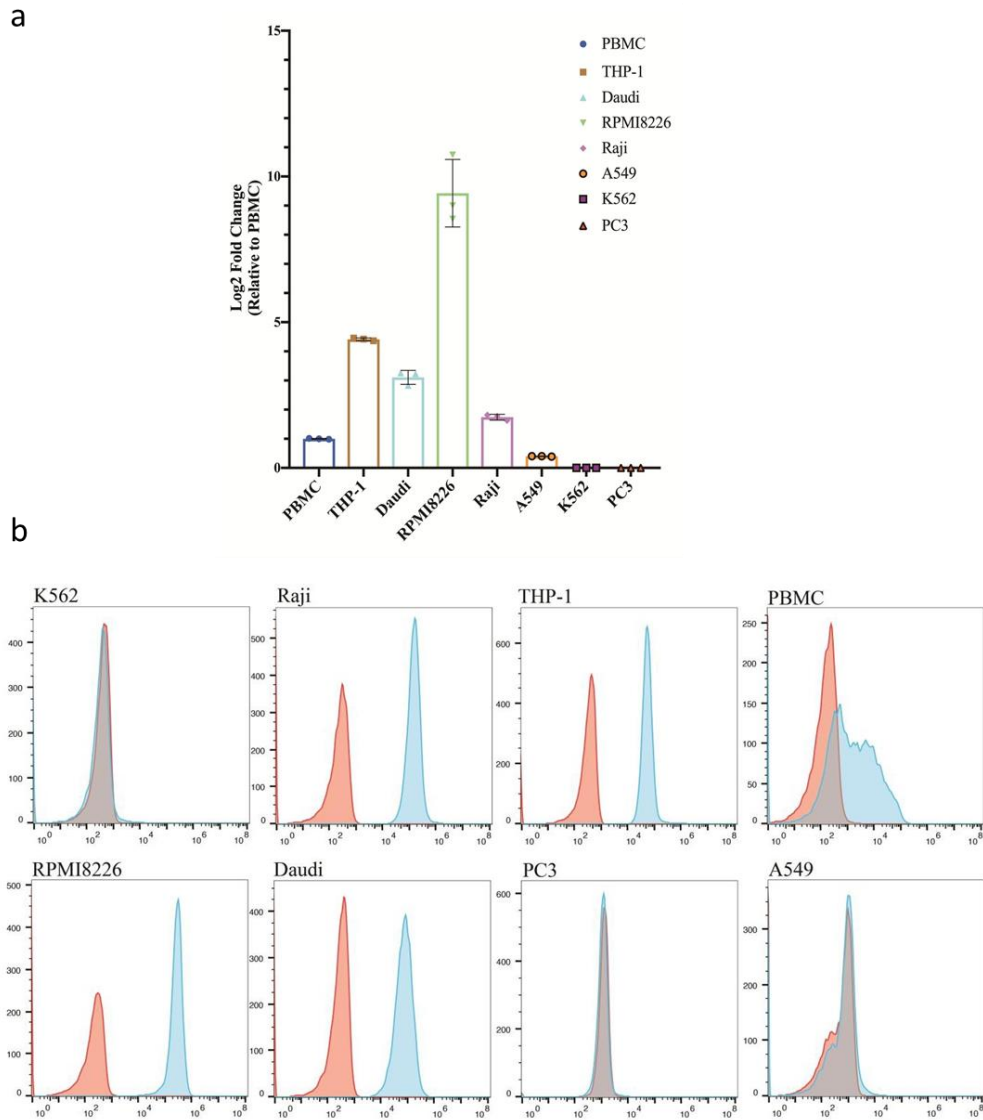
hCD38-ECD    I TEEDYQPLMKLGTQTVPCNKILLWSRIKD LAHQFTQVQRDMFTLEDTLGLYLADDLTWC 118
mCD38-ECD    I TREDYAPLVKLVLTQT IPCNKTLFWSKSKHLAHQYTWIQGKMFTLEDTLGYIADDLRWC 120
                **  ***  **:*  ***  :***  *  ***:  *  ***  *  :*  .  *****:****  **

hCD38-ECD    GEFNTSKINYQSCPWRKDCSNNPVSVFWKTVSRRFAEAAACDVVHVMNLNGSRSKIFDKNS 178
mCD38-ECD    GDPSTSDMNYVSCPWSENCNPNPIIVFWKVISQKFAEDACGVVQVMLNGSLREPFYKNS 180
                *  :  .**  .**  **  *  *  :*  ***  :***  .  *  :***  **  **  :***  *  :  *  ***

hCD38-ECD    TFGSVEVHNLQPEKVQTLAWVIHGGRDSRDLCDPTIKELESII SKRNIQFSCKNIYR 238
mCD38-ECD    TFGSVEVFLDPAKVKHLQA WVMHDI EGASSNACSSSSLNELKMI VQKRNMI FACVDNYR 240
                *****  .  *  :***  .  *  :***  *  .  *  :  *  .  :***  *  :  ***  *  :  *  :  **

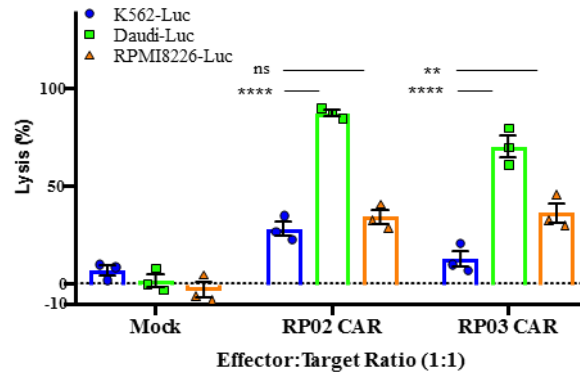
hCD38-ECD    PDKFLQCVKNPEDSSCTSEI 258
mCD38-ECD    PARFLQCVKNPEHPSCRLNT 260
                *  :*****  **  :
  
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**Figure S3 | Interactions between purified scFv antibodies and *h*CD38-ECD / *m*CD38-ECD proteins. (a) Comparison of binding affinities of RP02 and RP03 antibodies to *h*CD38-ECD / *m*CD38-ECD by surface plasmon resonance (SPR) measurement. (b) Sequence alignment between *h*CD38-ECD and *m*CD38-ECD.**



**Figure S4 | CD38 expression in different cells. (a)** Relative expression levels of CD38 in indicated cells were measured by real-time quantitative reverse transcription PCR (qRT-PCR). The results were normalized to GAPDH mRNA levels and represent the means  $\pm$  SEM. (n = 3) **(b)** Cell surface expression of CD38 was evaluated by FACS.

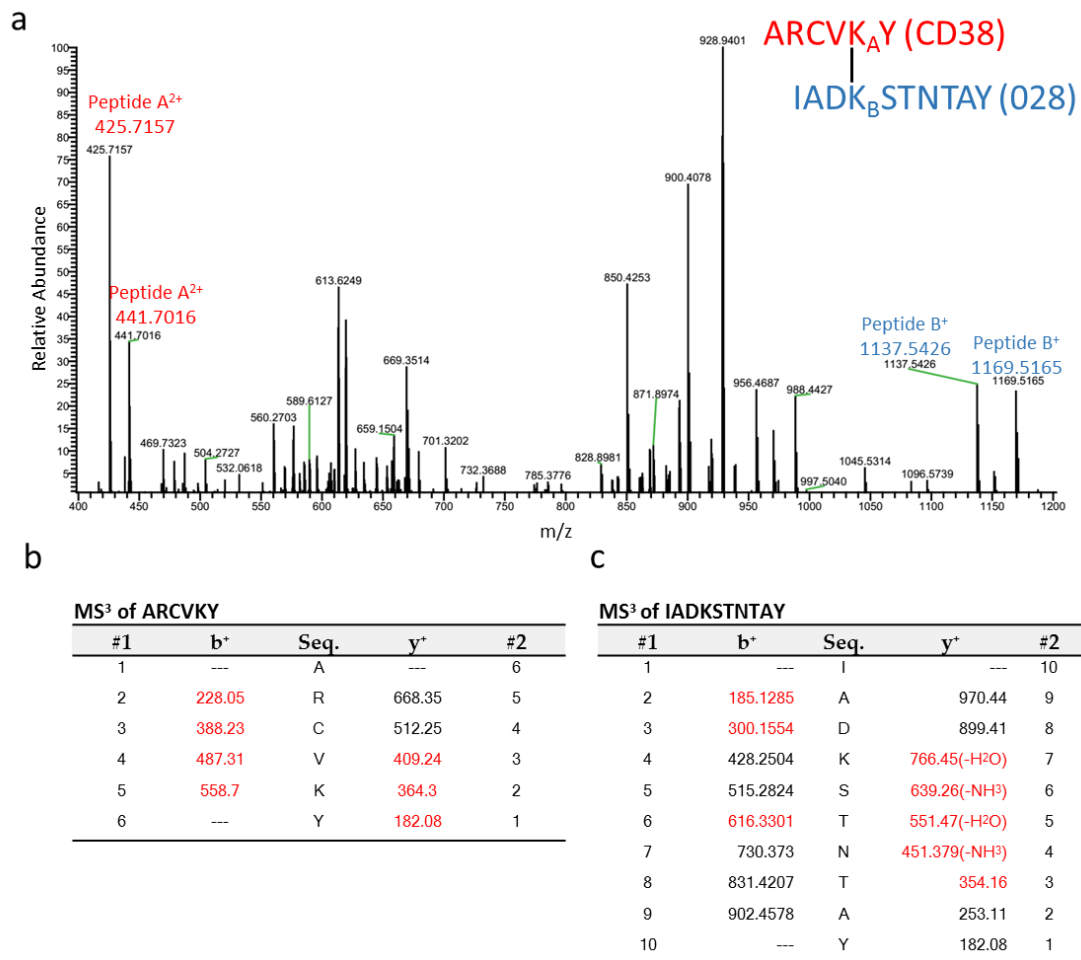
### Specific Killing of CD38<sup>+</sup> Cells by CD38 CAR-T Cells (24 h)



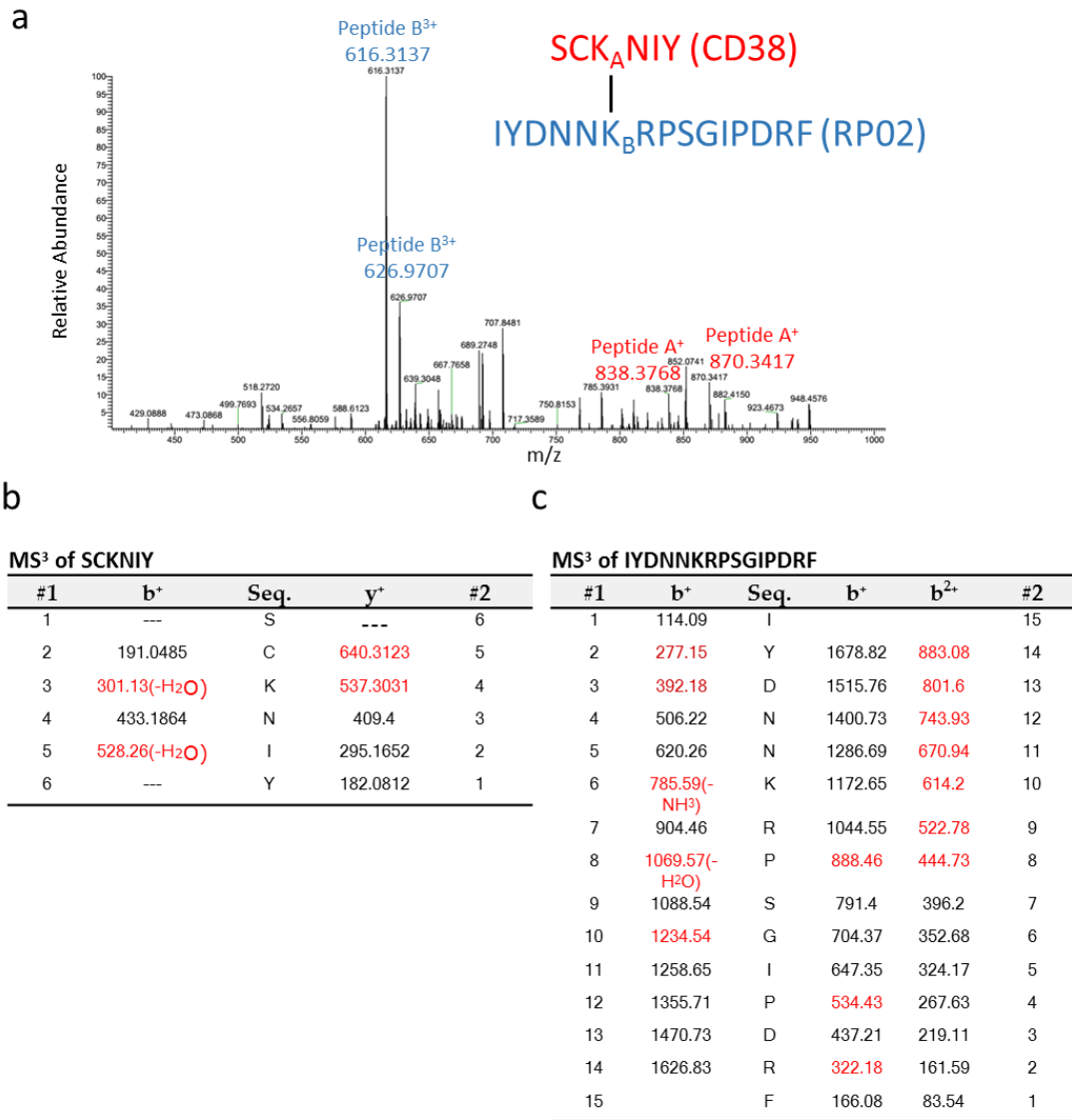
**Figure S5 | Evaluation of anti-tumor efficiency of RP02 CAR-T and RP03 CAR-T.**

In contrast to results seen with mock T cells, co-culture of CD38<sup>+</sup> target tumor cells (Daudi and RPMI8226) with RP02 or RP03 anti-CD38 CAR-T cells for 24 hours at an E:T ratio of 1:1 resulted in significant cytotoxicity. Data are represented as the mean  $\pm$  s.e.m. of  $n = 3$  technical replicates. Significance was considered as \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ ; \*\*\*\* $p < 0.0001$ .

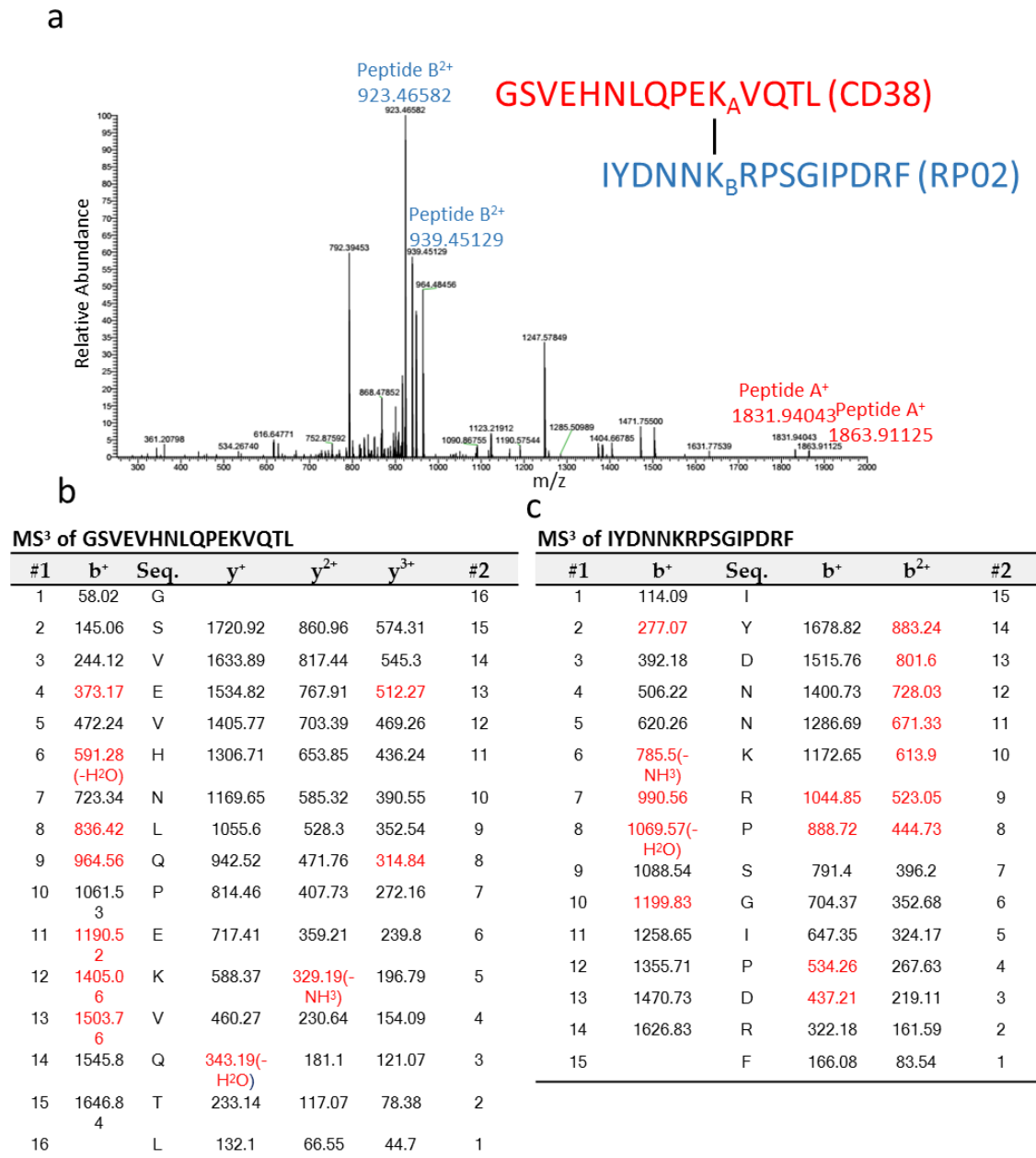




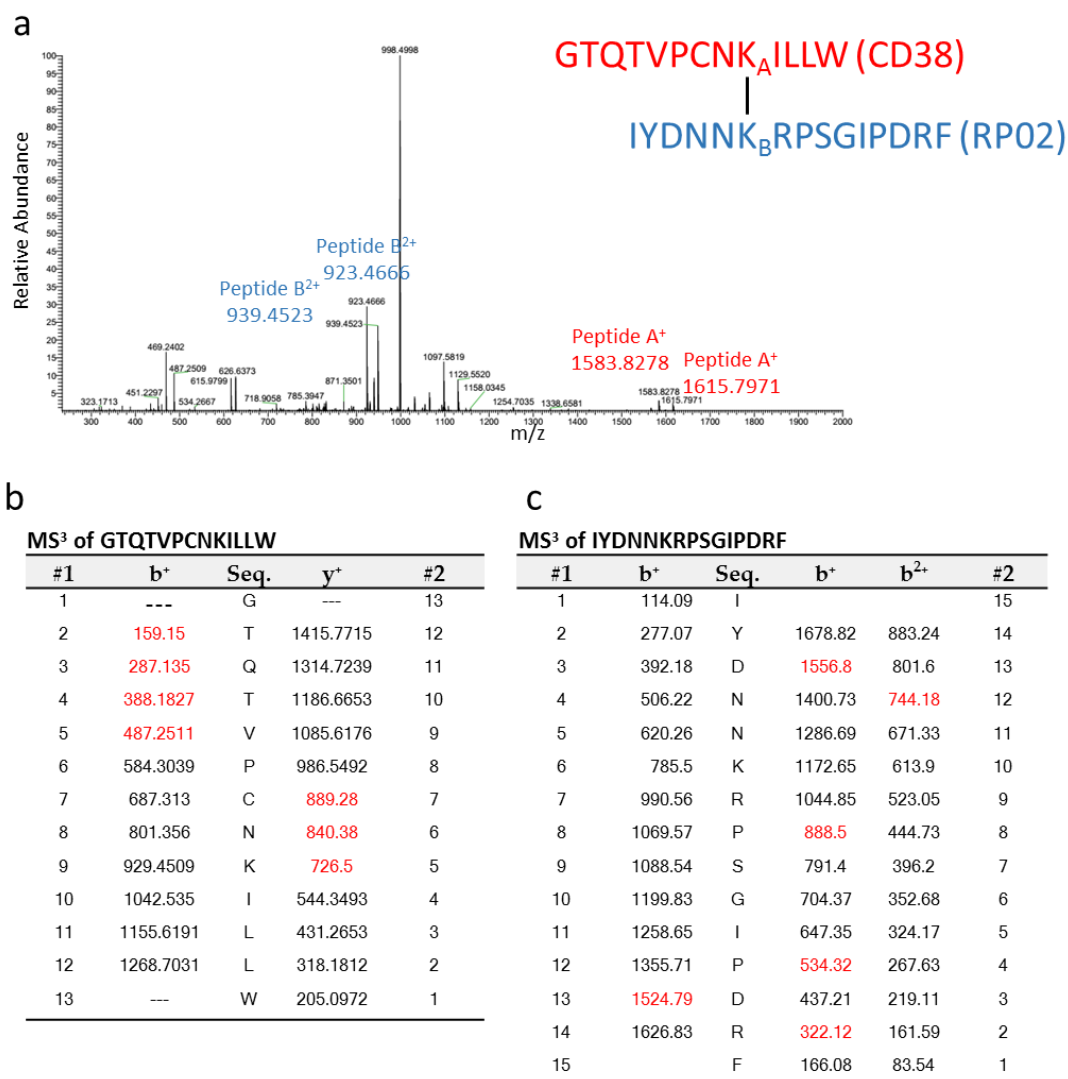
**Figure S6 | 028 scFv cross-linked with *h*CD38-ECD via IADKSTNTAY and ARCVKY peptides.** (a) Secondary MS showing cross-linked peptides ARCVKY-IADKSTNTAY. (b) Peptide mass fingerprint for ARCVKY. (c) Peptide mass fingerprint for IADKSTNTAY. The MS data were deposited in the proteomics identifications (PRIDE) database with accession number PXD019944.



**Figure S7 | RP02 scFv cross-linked with hCD38-ECD via IYDNNKRPSGIPDRF and SCKNIY peptides.** (a) Secondary MS showing cross-linked peptides SCKNIY-IYDNNKRPSGIPDRF. (b) Peptide mass fingerprint for SCKNIY. (c) Peptide mass fingerprint for IYDNNKRPSGIPDRF. The MS data were deposited in the proteomics identifications (PRIDE) database with accession number PXD019868.



**Figure S8 | RP02 scFv cross-linked with hCD38-ECD via IYDNNKRPSGIPDRF and GSVEVHNLQPEKVQTL peptides.** (a) Secondary MS showing cross-linked peptides GSVEVHNLQPEKVQTL-IYDNNKRPSGIPDRF. (b) Peptide mass fingerprint for GSVEVHNLQPEKVQTL. (c) Peptide mass fingerprint for IYDNNKRPSGIPDRF. The MS data were deposited in the proteomics identifications (PRIDE) database with accession number PXD019868.



**Figure S9 | RP02 scFv cross-linked with hCD38-ECD via IYDNNKRPSGIPDRF and GTQTVPCNKILLW peptides.** (a) Secondary MS showing cross-linked peptides GTQTVPCNKILLW-IYDNNKRPSGIPDRF. (b) Peptide mass fingerprint for GTQTVPCNKILLW. (c) Peptide mass fingerprint for IYDNNKRPSGIPDRF. The MS data were deposited in the proteomics identifications (PRIDE) database with accession number PXD019868.

	CDR3
028-V <sub>H</sub>	EPGERDPD-AVDI
RP02-V <sub>H</sub>	ED----YYYMDV
RP03-V <sub>H</sub>	DGLPRWNDIWFDL

	CDR3
028-V <sub>L</sub>	QQYNS--YPLT
RP02-V <sub>L</sub>	GTWDSSLSAGV
RP03-V <sub>L</sub>	TSFTT-FKTWV

**Figure S10 | Sequence alignment of variable domains.** The CDR3 sequences of 028, RP02 and RP03 scFvs were aligned. The sequences were numbered using the Kabat numbering scheme.