

Fig. S1. The MBP only control for MBP pull-down assays. MBP protein does not bind to the tag-free *Synechocystis* Cas1, Cas2, or Cas4 proteins.



Fig. S2. The binding ability of Ca4-Cas1 complex to the splayed DNA. EMSA is performed using 5' Cy3-labelled splayed prepsapcer DNA (22-bp duplex with 7-nt overhangs). Different concentrations of Cas proteins (0, 0.2, 0.4, 1, and 4 μ M) were incubated with 0.2 μ M splayed prepsapcer DNA. Percent of the free DNA is calculated based on the gray scanning analysis.



Fig. S3. Crystal Structure of Se-Met Cas1 of *Synechocystis.* (a) Crystals of Se-Met Cas1. (b) Crystal structure of Se-Met Cas1. Two protomers form a homodimer via N-terminal domains.



Fig. S4. Alignment of Cas1 and Cas2 from *Synechocystis* sp. PCC6803, *E. faecalis* and *E. coli*. (a) Sequence alignment of Cas1 from various species. The secondary structure of *Synechocystis* sp. PCC6803 Cas1 is placed on top. Yellow boxes mark the homologous residues and red boxes mark the identical residues. (b) Superposition of Cas1 from various species. (c) Sequence alignment of Cas2 from various species. (d) Superposition of Cas2 from various species.



Fig. S5. Comparison of Cas1-Cas2-prespacer Architecture in Synechocystis sp.6803, *E. coli* and *E. faecalis.* The Cas1-Cas2-prespacer complexes of *Synechocystis* sp.6803 and *E. faecalis* have a similar Cas2 dimer orientation, which is different from that of *E. coli*.



Fig. S6. Gel-filtration Profiles of the Mutants in Lysine Clamp and Arginine Channel. Each mutant is indicated below the gel-filtration profiles.



Fig. S7. The Wedge Residues and the Grabber motif in Cas1 of *E. coli* (a, PDB ID: **5DS5) and** *E. faecalis* (b, PDB ID: **5XVN).** Y22 in *E. coli* and H11 *E. faecalis* function as a wedge to terminate the duplex region of the prespacer. The Grabber motif, Y86 and R84 in *E. coli* and D69 and R71 in *E. faecalis*, bends and guides the single stranded 3'-overhang to the active site of Cas1.



Fig. S8. Structure Reconstruction of the Cas4-Cas1 Complex by Negative Staining Electron Microscopy. (a) A representative raw negatively stained micrograph of Cas4-Cas1 complex. (b) The two-dimensional class averages of Cas4-Cas1 complex and the surface representation of the 3D reconstruction. (c) The Fourier shell correlation (FSC) curve of 3D reconstructed Cas4-Cas1 complex.

	Se-Met Cas1	Cas1-Cas2-22bp	Cas1-Cas2-26bp
Data collection			
Space group	P 6 ₅ 2 2	C 2 2 2 ₁	P 3 ₁
Cell dimensions			
<i>a</i> , <i>b</i> , <i>c</i> (Å)	119.7 119.7 401.7	125. 6 215.2 191.1	185.4 185.4 382.9
<i>α</i> , <i>β</i> , <i>γ</i> (°)	90.0 90.0 120.0	90.0 90.0 90.0	90.0 90.0 120.0
Resolution (Å)	50-2.89(2.95-2.89) *	50-3.70 (3.79-3.70) *	50-3.70 (3.82-3.70) *
$R_{\rm pim}$	2.8 (49.3)	7.7 (54.2)	10.4 (56.5)
Ι/σΙ	42.0 (2.0)	11.0 (2.0)	12.7 (2.1)
Completeness (%)	100.0 (100.0)	99.4 (99.4)	99.9(99.9)
Redundancy	73.4 (56.0)	12.5 (9.9)	10.5(10.4)
Refinement			
Resolution (Å)		33.81-3.72	47.56-3.70
No. reflections		25,222	156,966
$R_{\mathrm{work}}/R_{\mathrm{free}}(\%)$		27.36/32.34	29.57/34.31
No. atoms			
Protein		11,810	71,378
DNA		1,042	7,230
Water		2	0
B-factors			
Protein		40	138
DNA		71	212
R.m.s deviations			
Bond lengths (Å)		0.003	0.004
Bond angles (°)		0.670	0.949

Table S1. Data collection and refinement statistics.

*Highest resolution shell is shown in parenthesis.