A Facile Colorimetric Method for the Quantification of Labile Iron Pool and Total Iron in Cells and Tissue Specimens

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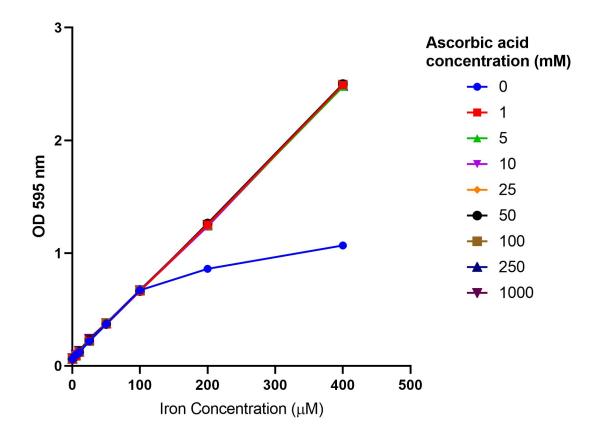
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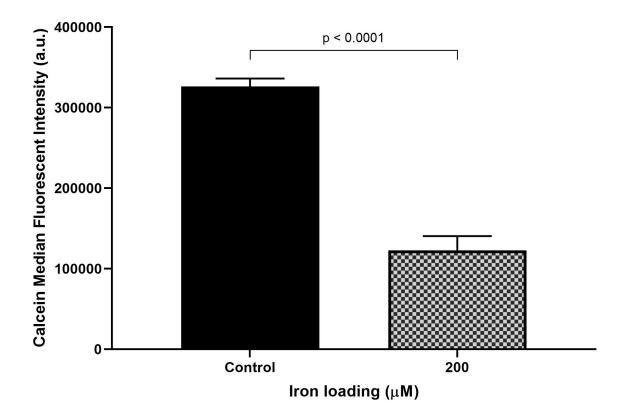
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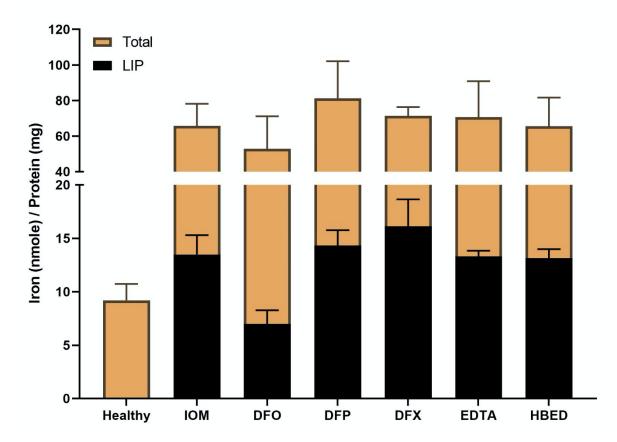
Supplementary Figures and Tables:



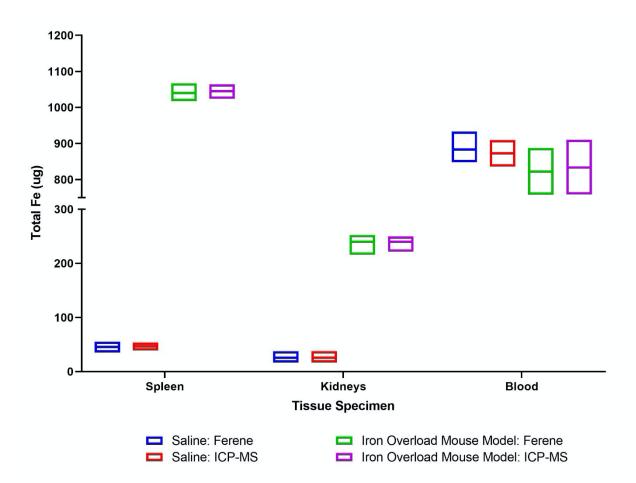
Supplementary Figure S1. The standard curve for the ferene assay with different concentrations of ascorbic acid in the working solution. While ascorbic acid concentrations is critical for the differentiation of labile iron and total iron measurements, the standard curve was prepared under different ascorbic acid concentrations (mM). The linear regression for these three iron standard curves are not different. All experiments were done in at least triplicates. Error bars show standard deviations. Statistical analysis were performed using GraphPad Prism.



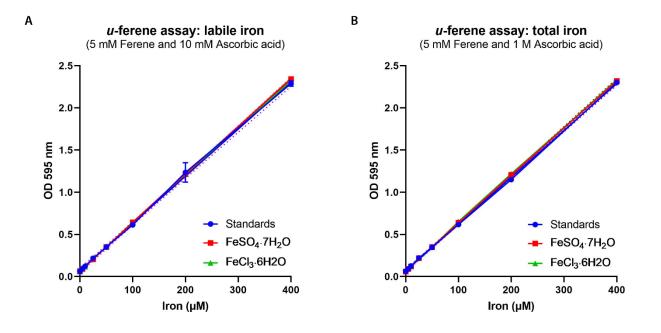
Supplementary Figure S2. The effect of iron loading on labile iron concentration. Calcein is a fluorophore which quenches in the presence of iron. This fluorophore was used to monitor the changes in the intracellular labile iron pool using flow cytometry. Unpaired t-test was performed using GraphPad Prism. All experiments were done in at least triplicates. Error bars show standard deviations. Statistical analysis were performed using GraphPad Prism.



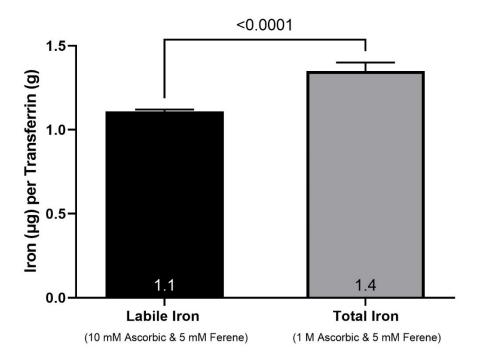
Supplementary Figure S3. Changes in the labile iron pool (LIP) in the presence of different chelators. LIP (using 10 mM ascorbic acid in the working solution) and total iron was quantified from iron overloaded HepG2 cell lysates; IOM – iron overload model with no chelator, DFO – deferoxamine, DFP – Deferiprone, DFX – Deferasirox, EDTA - Ethylenediaminetetraacetic acid, and HBED - N, N-bis(2-hydroxybenzyl)ethylenediamine-N,N-diacetic acid. Healthy cell lysates were used a control for changes in LIP and total iron. At 10 mM ascorbic acid concentration in working solution, only DFO treated lysates had a significant reduction in the LIP (p<0.0001). All experiments were done in at least triplicates. Error bars show standard deviations. Statistical analysis were performed using GraphPad Prism.



Supplementary Figure S4. Quantification of total iron in the remaining tissue specimens; organs (spleen and kidneys) and fluids (blood). Total iron was measured in both saline (red and blue) and iron-dextran iron overload mouse model groups (green and purple) comparing the ferene assay (blue and green) to ICP-MS (red and purple). Three biological replicates were measured per group. Statistical analysis were performed using GraphPad Prism.



Supplementary Figure S5. The standard curve for the ferene assay with different source of primary iron sources – ferric chloride and ferrous sulphate. Iron standards were prepared from either ferric chloride or ferrous sulphate, ranging from 0 to 1000 μ M. The standard curve was generated using the experimental conditions defined for labile iron measurements (5 mM ferene and 10 mM ascorbic acid) and total iron measurements (5 mM ferene and 10 mM ascorbic acid) and total iron measurements (5 mM ferene and 10 mM ascorbic acid) and total iron measurements (5 mM ferene and 10 mM ascorbic acid) and total iron measurements. Error bars show standard deviations. Simple linear regressions and their respective 95% confidence intervals were determined using GraphPad Prism. The slope and y-intercept determined from the linear regression were not significantly different amongst the different sources of iron, as determined by the overlap in the 95% confidence intervals.



Supplementary Figure S6. Labile and total iron were determined using the *u***-ferene assay in holo-transferrin.** All experiments were done in at least triplicates. Error bars show standard deviations. Unpaired t-test was performed to compare labile and total iron measurements. One sample t test was performed to compare total iron measurements with the iron content determined in the certificate of analysis from Sigma Aldrich. All statistical analyses were performed using GraphPad Prism.

Supplementary Table 1. A list of dilutions used for the analysis of total iron in biological samples

| biological sample | dilution for <i>u</i> - ferene | dilution for ICP-MS |
|----------------------|-----------------------------------|---------------------|
| cell lysates | 1 | 3 |
| plasma | 1 | 3 |
| liver | 40 | 5000 |
| spleen | 20 | 1000 |
| heart | 1 | 40 |
| kidneys | 8 | 250 |
| urine | 1 | 10 |
| feces | 80 | 4000 |
| blood | 10 | 1500 |

Supplementary Table 2. The template of running assay for the determination of iron content

| | | W | | | | | | monium | | |
|---------------------|--------|---|---|---|----|----|----|--------|-----|------|
| Analytes | | 0 | 1 | 5 | 10 | 25 | 50 | 100 | 250 | 1000 |
| | Fe | | | | | | | | | |
| | Fe-DFO | | | | | | | | | |
| | DFO | | | | | | | | | |
| iron | 0 | | | | | | | | | |
| | 5 | | | | | | | | | |
| | 10 | | | | | | | | | |
| sta | 25 | | | | | | | | | |
| i standards (µM) | 50 | | | | | | | | | |
| | 100 | | | | | | | | | |
| | 200 | | | | | | | | | |
| | 400 | | | | | | | | | |

Supplementary Table 3. The main components in the *u*-ferene assay's working solution

| working solution components | labile iron | total iron | | | | |
|---|-------------|------------|--|--|--|--|
| Ferene | 5 mM | 5 mM | | | | |
| Ascorbic acid | 10 mM | 1 M | | | | |
| Prepared in ammonium acetate buffer (pH 4.5, 2.5 M) | | | | | | |