## **Supplementary Information**

## Presynaptic Kv3 channels are required for fast and slow endocytosis of synaptic vesicles

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Figure S1. Kv3.3 KO or Kv3.3<sub>G592R</sub> KI reduced the normalized Rate<sub>decay</sub> at hippocampal synapses, related to Figure 4.

(A) The normalized Rate<sub>decay</sub> (Nor. Rate<sub>decay</sub>) after 200 APs at 20 Hz in WT, Kv.3.3<sup>-/-</sup> (KO), or Kv3.3<sub>G592R</sub> boutons at 34-37°C. The normalized Rate<sub>decay</sub> was measured from F<sub>SypH</sub> with a different normalization method: ΔF, the F<sub>SypH</sub> increase induced by the 200-AP train, but not the F<sub>SypH</sub> baseline, was normalized as 100%. \*\*: p < 0.01 (comparison between KO and WT);</li>
\*: p < 0.05 (comparison between Kv3.3<sub>G592R</sub> and WT boutons); statistical test is t test. The experimental number is written inside the bar; each experiment contained 20-30 boutons; 1-3 experiments were taken from 1 culture; each culture was from 3-5 mice.

The normalization method used here removed the impact of  $\Delta F$  on the measurement of the Rate<sub>decay</sub>. The results shown in panel A are similar to those (See Figures 4E and 4F) without such normalization, indicating that the results drawn from Figures 4E and 4F are independent of the changes in  $\Delta F$ .

(B) Similar to panel A, except from WT boutons, WT boutons in the presence of 20 μM latruculin A (WT + Lat A), Kv.3.3<sup>-/-</sup> boutons (KO), or KO boutons in the presence of 20 μM latruculin A (KO + Lat A). \*\*: p < 0.01 (comparison between WT and WT+Lat A); N.S.: not significant, p > 0.05 (comparison between KO and KO + Lat A); statistical test is t test.

The results shown in panel B are similar to those (See Figures 4I and 4J) without such normalization, indicating that the results drawn from Figures 4I and 4J are independent of changes in  $\Delta F$ .



Figure S2. Peak SypH fluorescence increase induced by 200 APs at 20 Hz in various conditions, related to Figure 4.

- (A)Peak SypH fluorescence increase induced by 200 APs at 20 Hz ( $\Delta$ F, mean + s.e.m.) at hippocampal cultures in five conditions: 1) WT boutons, 2) Kv3.3<sup>-/-</sup> boutons (KO), 3) Kv3.3<sub>G592R</sub> boutons, 4) Kv.3.3<sup>-/-</sup> boutons overexpressed with WT Kv3.3 (KO + WT), 5) Kv.3.3<sup>-/-</sup> boutons overexpressed with W496F Kv3.3 mutant with no ionic conductance (KO + Mutant). \*\*: p < 0.01 (comparison between WT and KO); \*: p < 0.05 (comparison between WT and Kv3.3<sub>G592R</sub>); statistical test is t test. The experimental number is written inside the bar; each experiment contained 20-30 boutons; 1-3 experiments were taken from 1 culture; each culture was from 3-5 mice.
- (B) Similar to A, except from different conditions: 1) WT boutons, 2) WT boutons in the presence of 20 μM latrunculin A (WT + Lat A), 3) Kv.3.3<sup>-/-</sup> boutons (KO), and 4) KO boutons in the presence of 20 μM latrunculin A (KO + Lat A). \*\*: p < 0.01 (comparison between WT and WT + Lat A); N.S.: not significant, p > 0.05 (comparison between KO and KO + Lat A); statistical test is t test.



## Figure S3. W496F Kv3.3 mutation abolishes potassium currents in CHO cells, related to Figure 4.

- Left and Middle: representative traces of currents recorded in CHO cells expressing wild type Kv3.3 or W496F Kv3.3. Currents were evoked from a holding potential of -70 mV to test potentials between -80 mV and +70 mV in 10 mV increments.
- **Right:** bar graph shows group data for current density recorded at +70 mV. Current density was calculated by dividing the peak current by cell capacitance. Values are shown as mean  $\pm$  s.e.m. (n = 4, 4), and significance was tested using an unpaired two-tailed t- test, \*\*\*p=0.0002.



Figure S4. Kv3.1 knockout did not affect endocytosis at hippocampal synapses, related to Figure 4.

(A-B)  $F_{SypH}$  (mean + s.e.m., A) and Rate<sub>decay</sub> (mean + s.e.m., B) induced by 200 APs at 20 Hz in WT and Kv3.1<sup>-/-</sup> (Kv3.1 KO) boutons at 34-37°C.  $F_{SypH}$  (A) was normalized to baseline; s.e.m. (A) is plotted every 4 s. The experimental number is written inside the bar of panel B, which also applies to panel A. Rate<sub>decay</sub> in panel B is not significantly different (p > 0.05, t test).