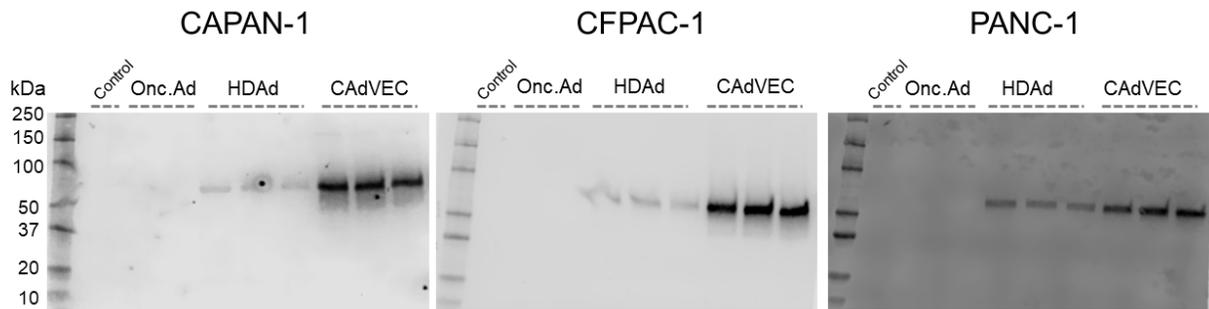


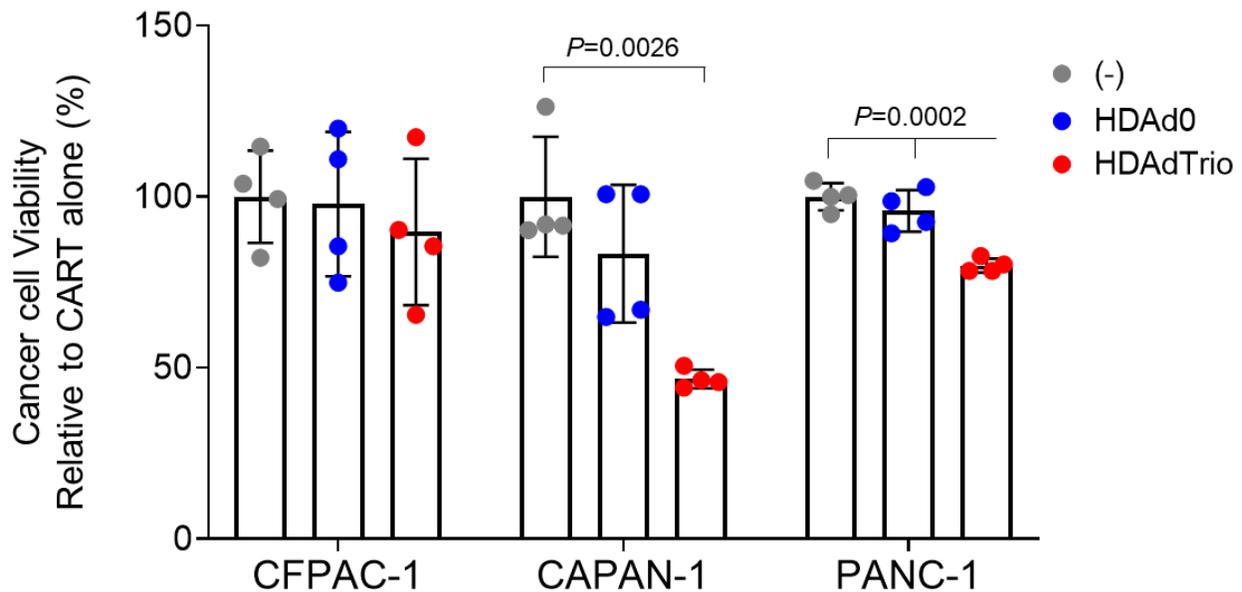
SUPPLEMENTARY INFORMATION

Oncolytic adeno-immunotherapy modulates the immune system enabling CAR T-cells to cure pancreatic tumors

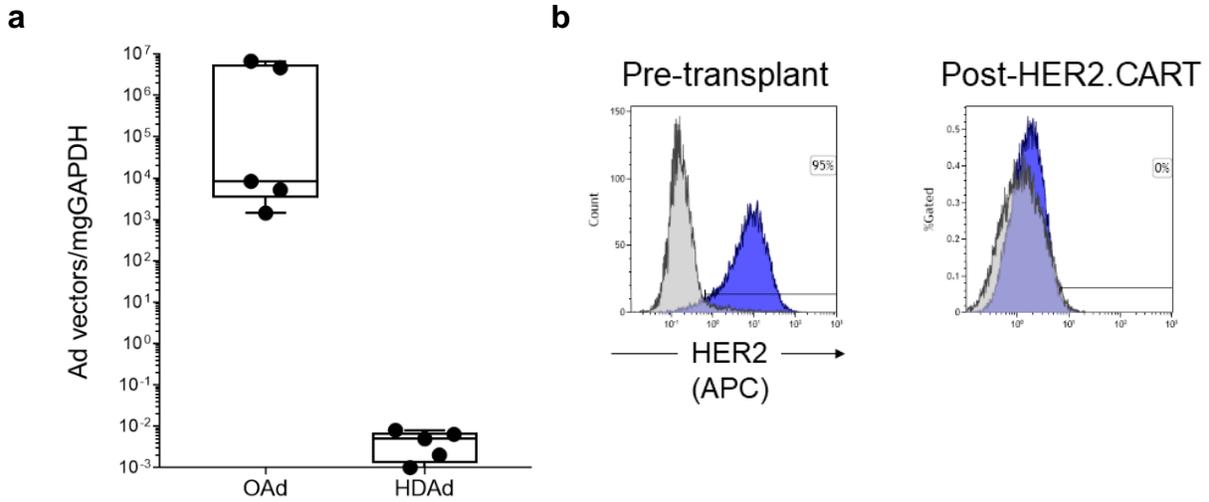
Rosewell Shaw et al.



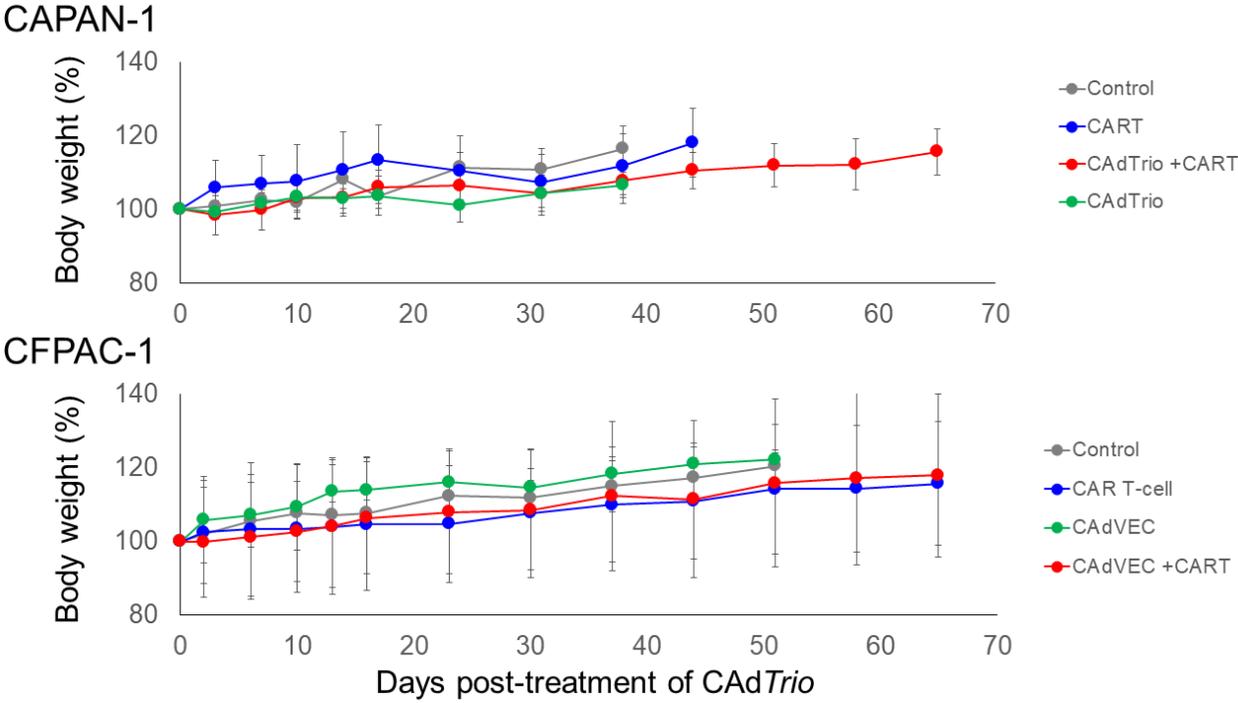
Supplementary Figure 1. Co-infection of OAd amplifies PD-L1 mini-antibody secretion. PANC-1, CAPAN-1 and CFPAC-1 were infected with total 10 vp/cell of HDAd $Trio$, OAd or CAAd $Trio$ (OAd:HDAd=1:1) (n=4 biologically independent samples). We sampled media 48 hours post-infection and quantified levels of PD-L1 mini-antibody by western blotting for PD-L1 mini-antibody.



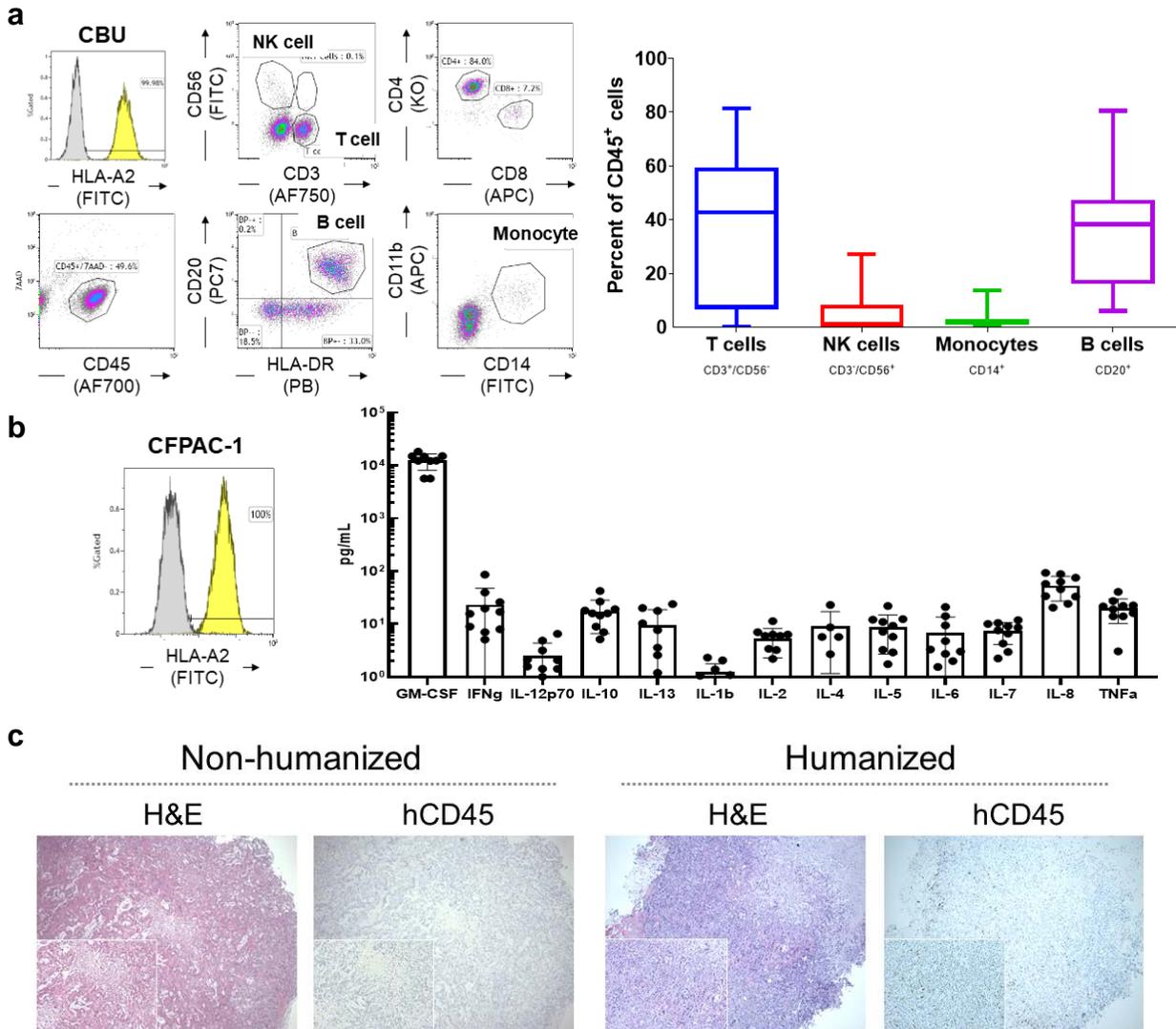
Supplementary Figure 2. Immunostimulatory molecules derived from *CAdTrio* enhance *HER2.CART* anti-tumor efficacy. PANC-1, CAPAN-1 and CFPAC-1 expressing *ffLuc* were infected with 100 vp/cell of HDAd0 (no transgene) or HDAdTrio. Cells were cultured with *HER2.CARTs* (E:T=1:40) at 24 hours post-infection. Cells were harvested 72 hours post-coculture, and viable cancer cells were analyzed by luciferase assay (n=4 independent biological samples). Data were normalized based on viability of *HER2.CART* alone. Data are presented as means \pm SD. P-values were determined by ordinary one-way ANOVA with Tukey multiple comparisons, $p=0.0026$ ($F_{2,9}=12.35$), $p=0.0002$ ($F_{2,9}=29.36$). Statistical significance set at $p < 0.05$, ns > 0.05 .



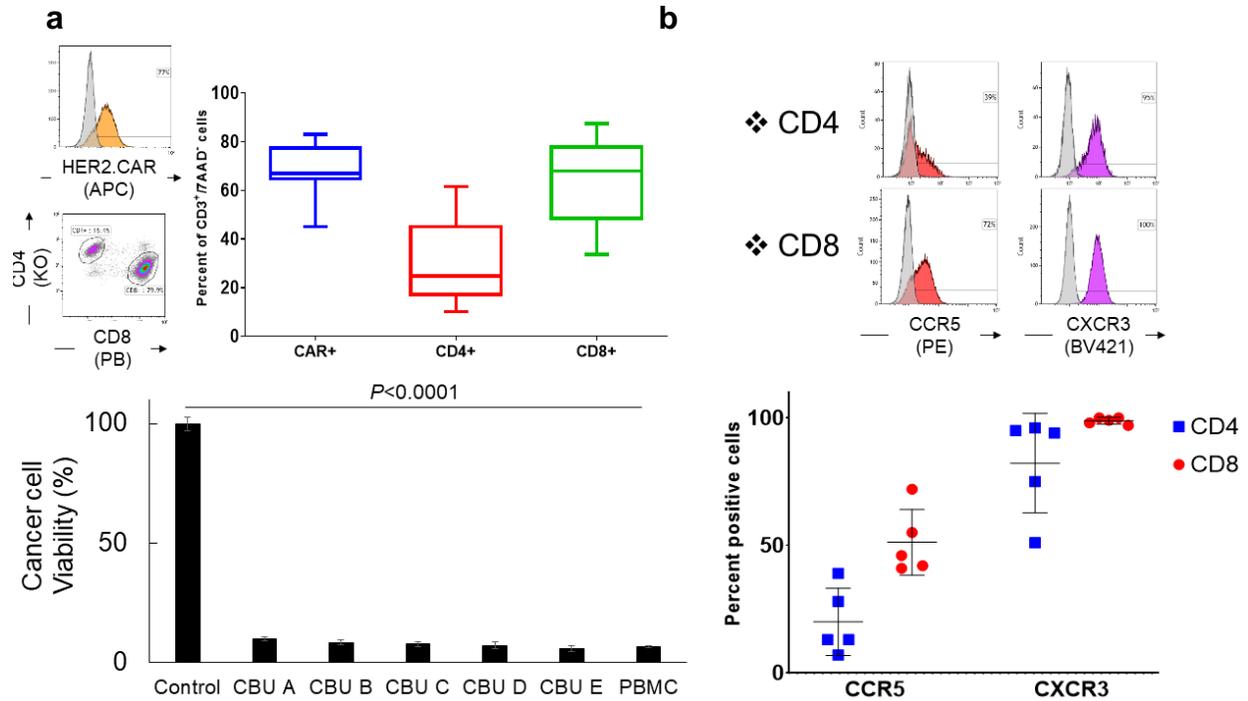
Supplementary Figure 3. Ad vectors persist at CAPAN-1 tumor site but do not control growth, and CAPAN-1 tumor causes antigen escape after HER2.CART treatment. (a) CAPAN-1 tumors treated with CAD $Trio$ alone were harvested when tumor volume reached $>1,500 \text{ mm}^3$. DNA was extracted from each tumor sample ($n=5$ animals), and the copy number of each Ad vector determined by quantitative PCR. Data was normalized with murine genomic GAPDH. Box plot elements: central line, median; box limit, upper and lower quartile; whisker, 1.5x inter-quartile range; points, outliers. **(b)** CAPAN-1 tumor treated with HER2.CART alone was harvested at the end of experiment (80 days post-infusion of HER2.CART), and HER2 expression of residual tumor was analyzed by flow cytometry.



Supplementary Figure 4. Combinatorial immunotherapy did not cause weight loss in xenograft mice. CAPAN-1 or CFPAC-1 cells were transplanted into the right flanks of NSG mice. A total of 1×10^7 vp of CAAdTrio (Onc:HD=1:1) were injected intra-tumorally. A total of 1×10^6 HER2.CAR T-cells were systemically administered 3 days post-injection of CAAdTrio (n=5 animals). Body weights were measured at different time points. Data are presented as means \pm SD.

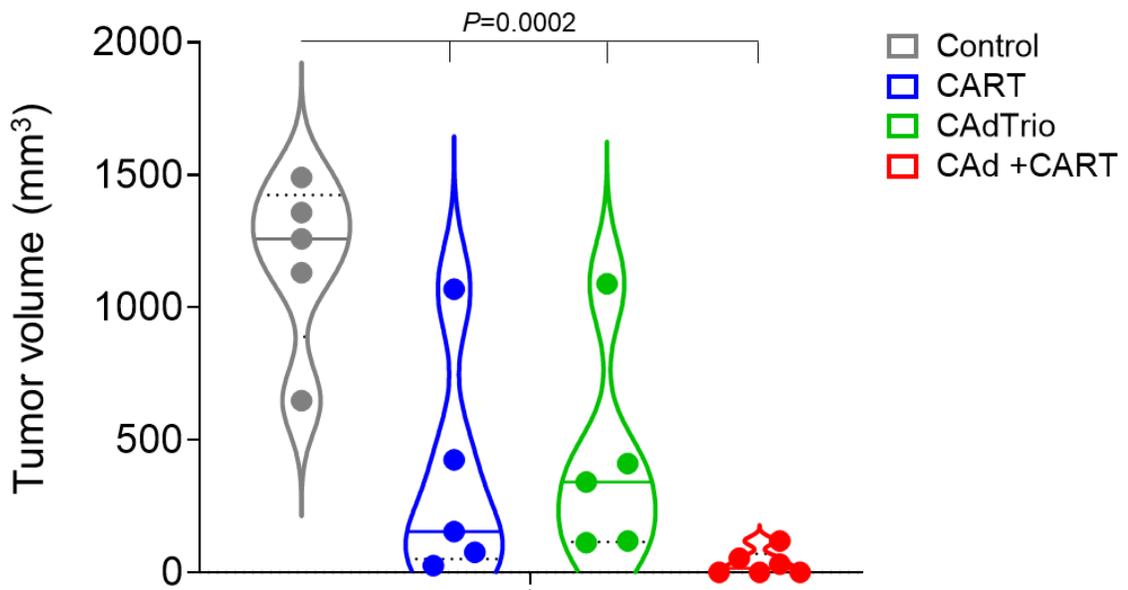


Supplementary Figure 5. Humanized mice reconstituted both innate and adaptive immune cells without inflammatory responses to transplanted CFPAC-1 cells. (a) Human immune subsets in PBMCs of humanized mice (n=30 animals) were characterized by flow cytometry 10 weeks post-transplant of HLA-A2⁺ CBU derived CD34⁺ cells. Box plot elements: central line, median; box limit, upper and lower quartile; whisker, 1.5x inter-quartile range; points, outliers. (b) HLA-A2 expression on CFPAC-1 was analyzed by flow cytometry. Plasma samples were collected from humanized mice transplanted with CFPAC-1 at 17 days post-transplant and circulating human Th1 and Th2 cytokines were measured. (c) CFPAC-1 cells were transplanted into the right flanks of non-humanized and humanized mice. Tumors were fixed at 21 days post-transplant, and sections were stained with anti-human CD45 IgG.

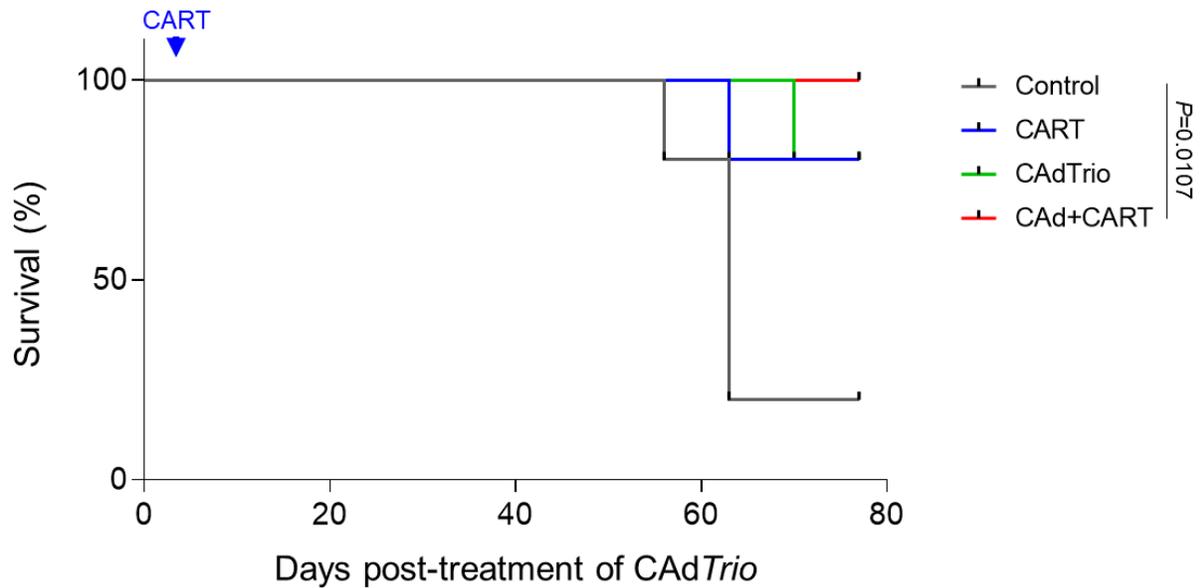


Supplementary Figure 6. Characterization of CBU derived HER2.CARTs. (a) HER2.CAR expression and T cell subsets of CBU derived HER2.CARTs (n=20 independent biological samples) were analyzed by flow cytometry. Box plot elements: central line, median; box limit, upper and lower quartile; whisker, 1.5x inter-quartile range; points, outliers. CFPAC-1 cells expressing *ffLuc* were cultured with 5 different CBU-derived HER2.CARTs or healthy donor PBMC-derived HER2.CARTs (E:T=1:10). Cells were harvested 72 hours post-coculture, and viable cancer cells were analyzed by luciferase assay (n=4 independent biological samples). Data are presented as means \pm SD. P-values were determined by ordinary one-way ANOVA with Tukey multiple comparisons, $p < 0.0001$ ($F_{1,084,3.252} = 2242$). Statistical significance set at $p < 0.05$, ns > 0.05 . (b) Chemokine receptors CCR5 and CXCR3 expressions were analyzed by flow cytometry on CBU derived HER2.CARTs (n=5).

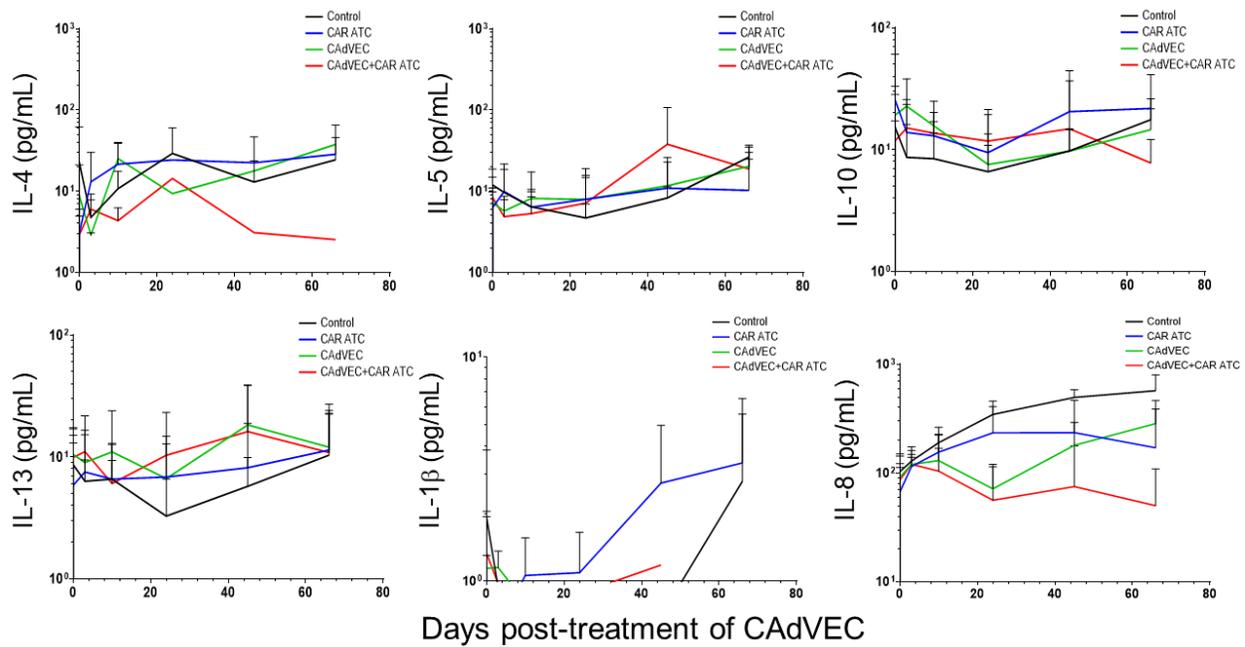
❖ 56 days post-injection of CAdTrio



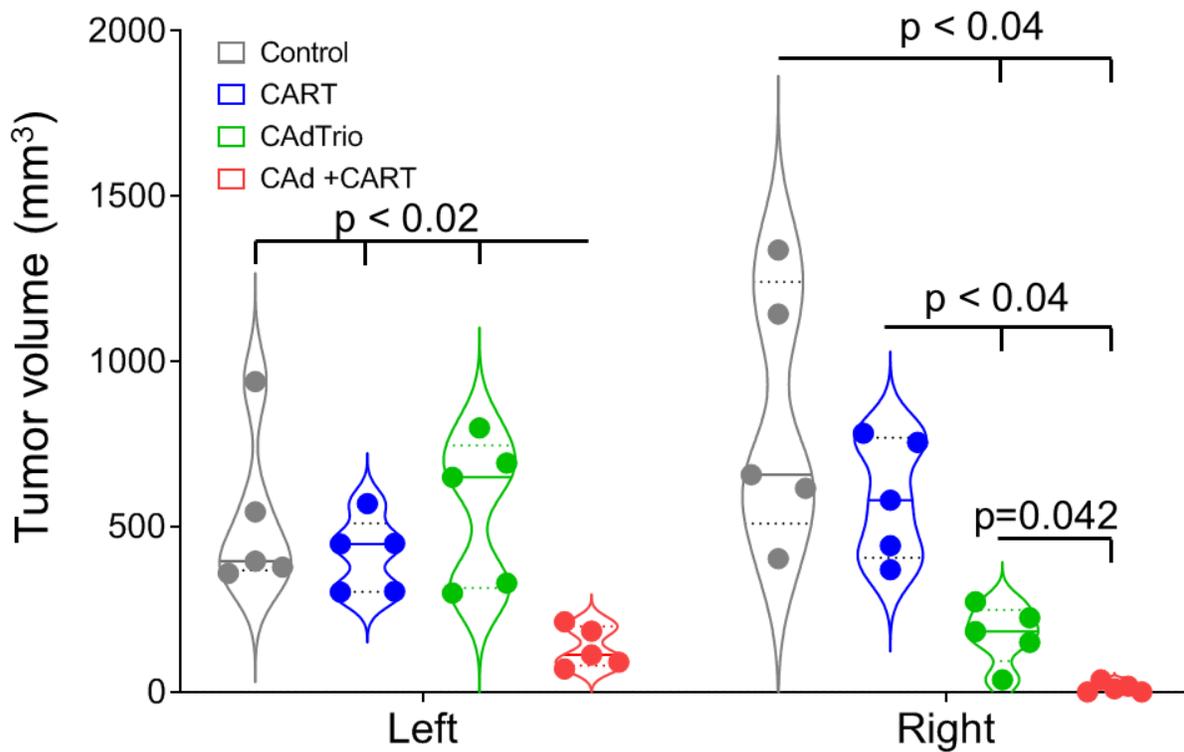
Supplementary Figure 7. Combination immunotherapy significantly controls CFPAC-1 tumor growth in humanized mice. CFPAC-1 cells were transplanted into the right flank of humanized mice (Control, CART alone, CAdTrio alone: n=5 animals, CAdTrio+CART: n=6 animals). A total of 1×10^7 vp of CAdTrio (Onc:HD=1:1) were injected into the tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdTrio. Tumor volumes shown here from day 56 post injection of CAd, individual data points are represented in violin plot with means \pm SD, $p=0.0002$. P-values were determined using ordinary one-way ANOVA with Tukey multiple comparisons, ($F(3,17)=11.66$). Statistical significance set at $p < 0.05$, ns > 0.05 .



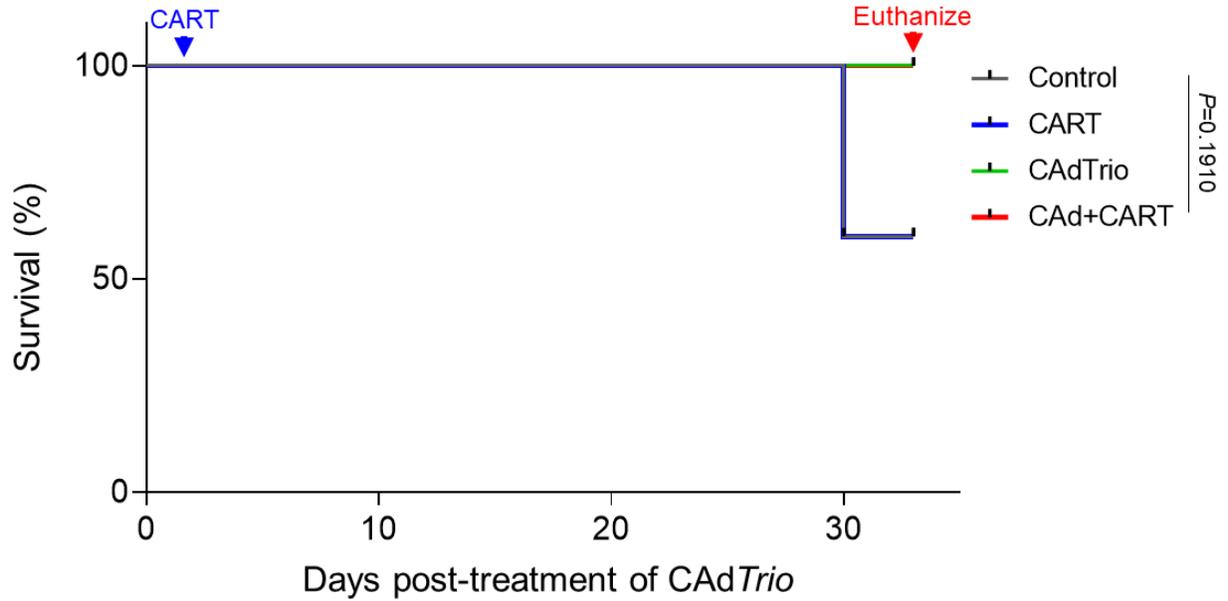
Supplementary Figure 8. Immunotherapy components control CFPAC-1 tumor growth in humanized mouse model. CFPAC-1 cells were transplanted into the right flank of humanized mice (Control, CART alone, CAdTrio alone: n=5 animals, CAdTrio+CART: n=6 animals). A total of 1×10^7 vp of CAdTrio (OAd:HD=1:1) were injected into the tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdTrio. Kaplan-Meier survival curve based on the timepoint at which tumor volumes necessitated euthanasia ($> 1,500 \text{ mm}^3$) after CAdTrio administration in mice, $p=0.0107$. P-values were determined using the Log-rank Mantel-Cox test (dF=3). Statistical significance set at $p < 0.05$, ns > 0.05 .



Supplementary Figure 9. Th2 cytokine expression was not induced by any treatment condition. CFPAC-1 cells were transplanted into the right flank of humanized mice (Control, CART alone, CAdTrio alone: n=5 animals, CAdTrio+CART: n=6 animals). A total of 1×10^7 vp of CAdTrio (Onc:HD=1:1) were injected into the tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdTrio. Serum samples were collected from mice at 0, 3, 10, 24, 45, and 66 days post-injection of CAdTrio, and human Th1 and Th2 cytokine levels in serum were measured by Multiplex. Data are presented as means \pm SD.

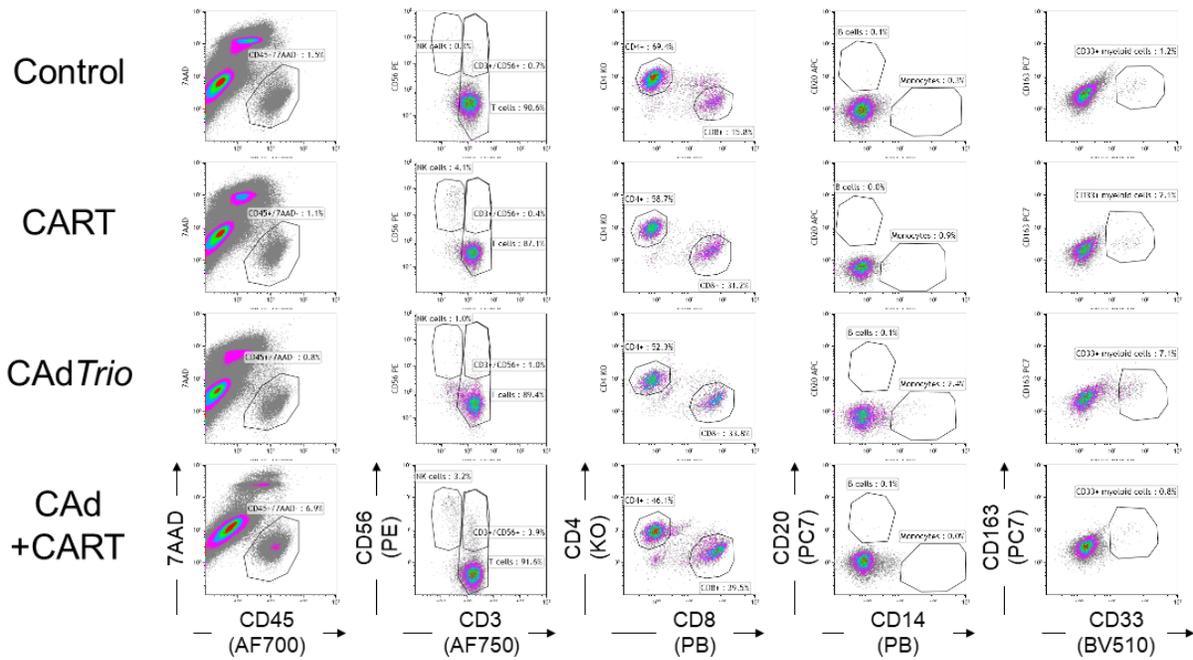


Supplementary Figure 10. Combination immunotherapy significantly controls CFPAC-1 tumor growth in humanized mice with multiple tumors. CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=5 animals). A total of 1×10^7 vp of *CAdTrio* (Onc:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of *CAdTrio*. Tumor volumes shown here from day 31 post injection of *CAd*, individual data points are represented in violin plot with means \pm SD. P-values were determined using ordinary one-way ANOVA with Tukey multiple comparisons, $p < 0.02$ ($F(1.977, 7.908)=7.426$), $p < 0.04$, $p=0.042$ ($F(1.511, 6.046)=18.35$). Statistical significance set at $p < 0.05$, ns > 0.05 .



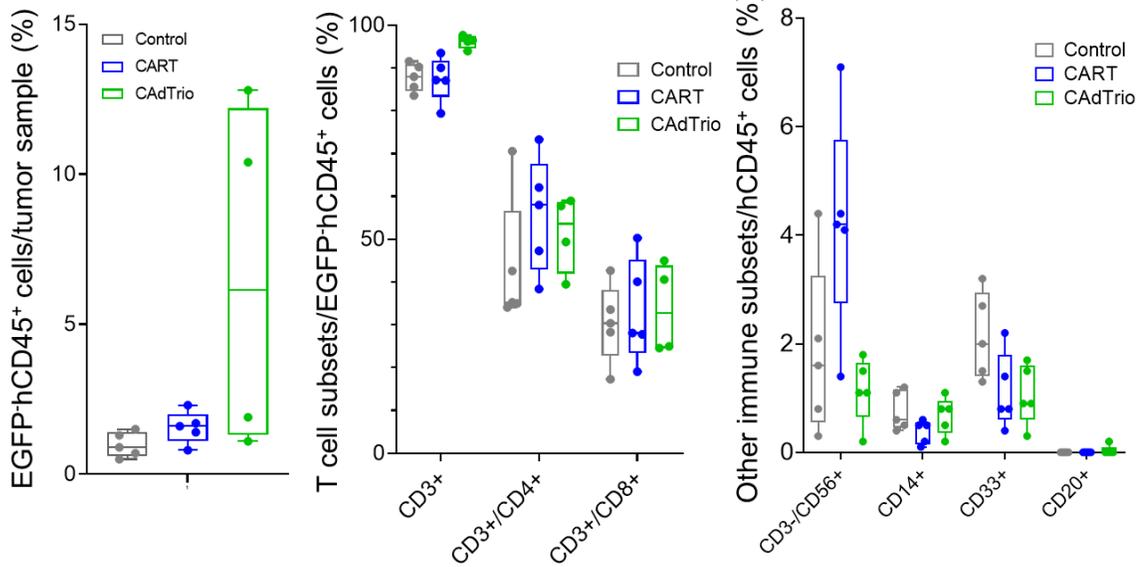
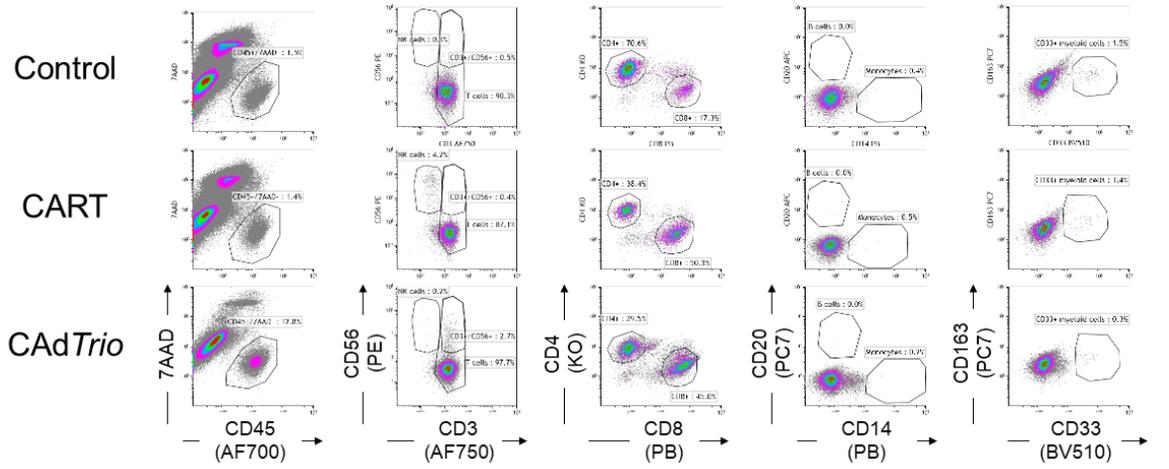
Supplementary Figure 11. Kaplan-Meier survival curve in humanized mice with multiple tumors. CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=5 animals). A total of 1×10^7 vp of CAdTrio (Onc:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdTrio. Kaplan-Meier survival curve based on the timepoint at which tumor volumes necessitated euthanasia ($> 1,500 \text{ mm}^3$) after CAdTrio administration in mice, $p=0.1910$. P-values were determined using the Log-rank Mantel-Cox test (dF=3). Statistical significance set at $p < 0.05$, ns > 0.05 .

❖ Left tumor



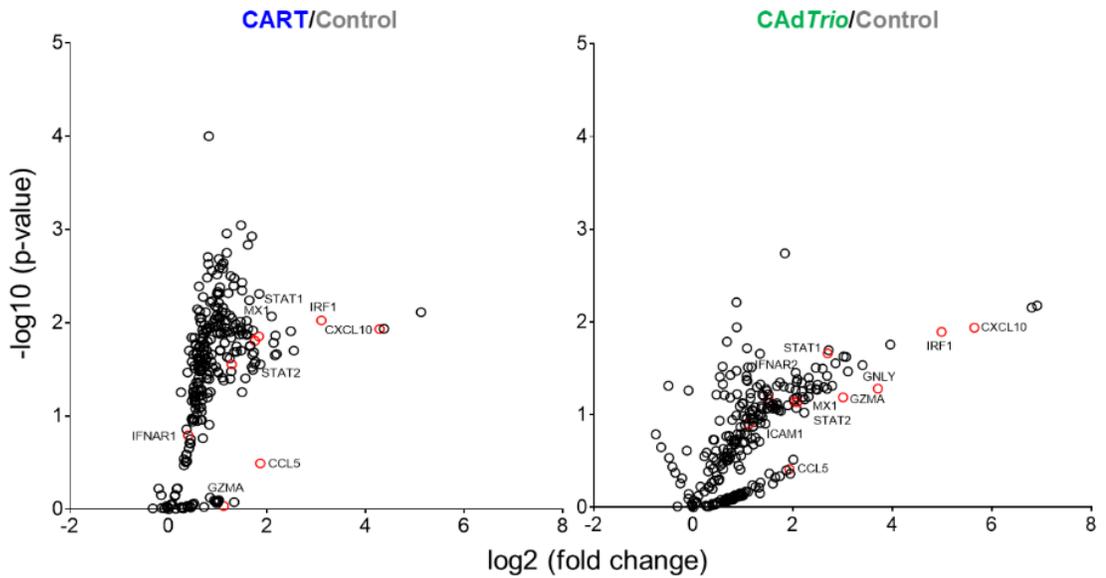
Supplementary Figure 12. Combination immunotherapy enhances immune infiltration to untreated tumor site in humanized mice with multiple PDAC tumors. CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=5 animals). A total of 1×10^7 vp of CAdTrio (Onc:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdTrio. CFPAC-1 tumors were harvested from humanized mice at 31 days post-injection of CAdTrio, and tumor infiltrating human immune cells were analyzed by flow cytometry. Shown here is representative data from one of five animals per group.

a ❖ Right tumor

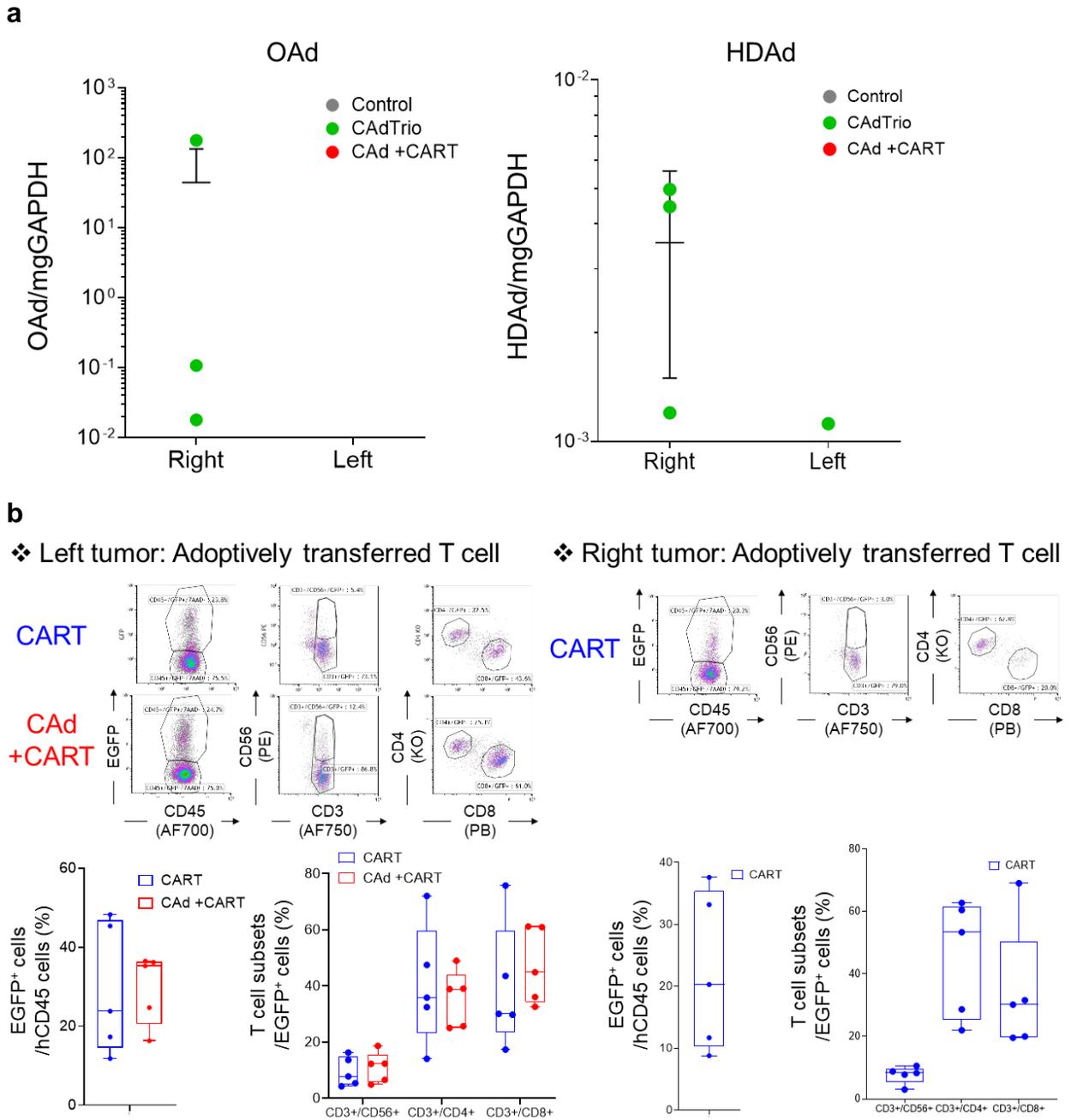


b

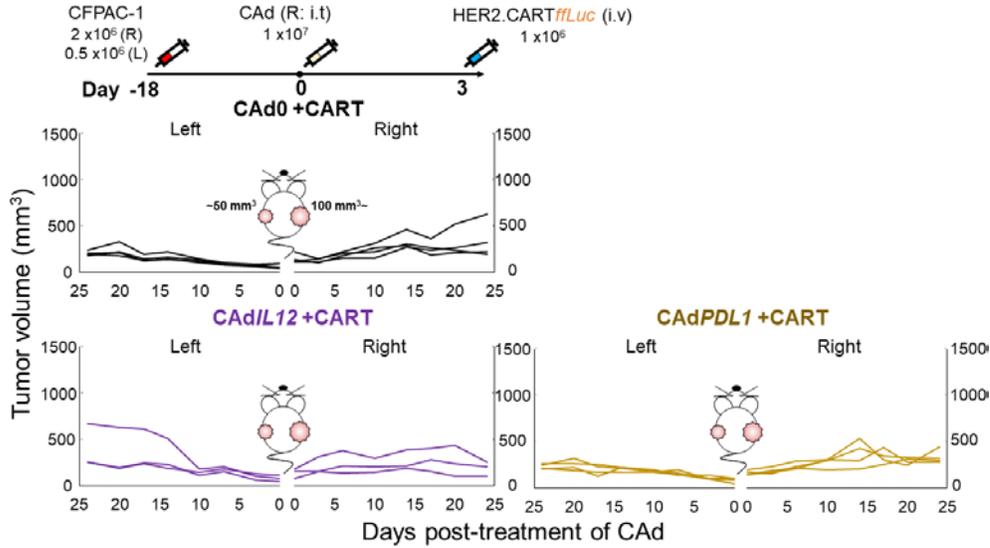
Right tumor



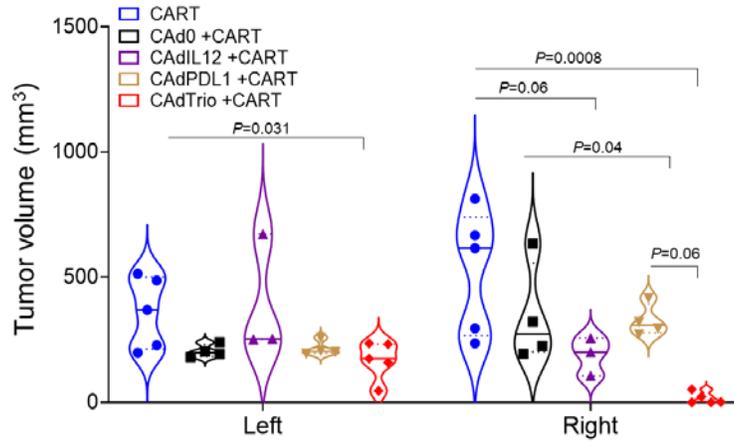
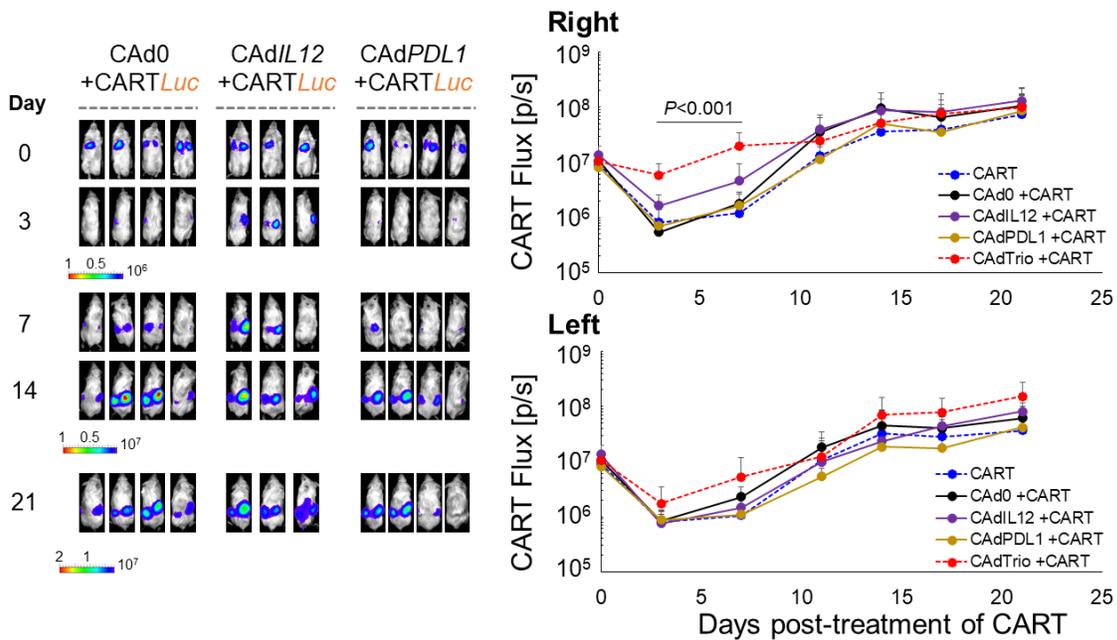
Supplementary Figure 13. Local combination immunotherapy treatment does not alter infiltration of immune cell subsets. CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=5 animals). A total of 1×10^7 vp of *CAdTrio* (Onc:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection. **(a)** CFPAC-1 tumors were harvested from humanized mice at 31 days post-injection of *CAdTrio*, and tumor infiltrating human immune cells were analyzed with flow cytometry and **(b)** total RNA was extracted from whole tumor. Gene expression was profiled with Nanostring. Genes showing more than 75% coefficient of variation (CV) compared to control tumors are shown. Box plot elements: central line, median; box limit, upper and lower quartile; whisker, 1.5x inter-quartile range; points, outliers.



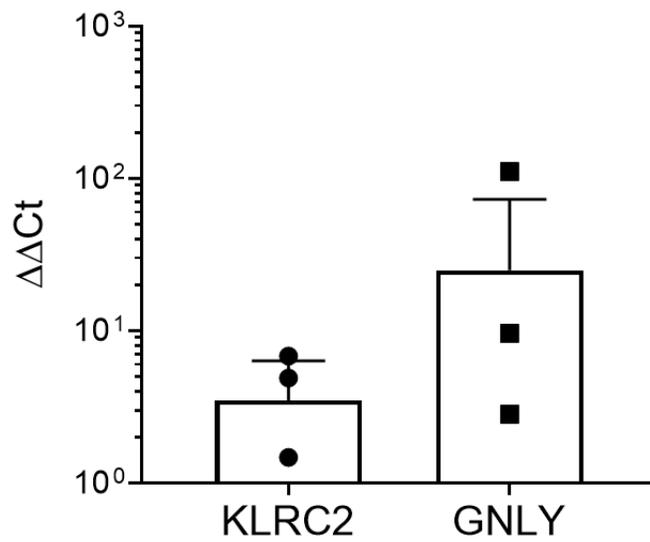
Supplementary Figure 14. Residual Ad vectors and HER2.CART at tumor sites. CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=5 animals). A total of 1×10^7 vp of *CAdTrio* (Onc:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffluc* were systemically administered 3 days post-injection of *CAdTrio*. Tumors were harvested 31 days post-injection of *CAdTrio*. **(a)** Total DNA was extracted from both right and left tumors separately, and residual Ad vectors were quantified and normalized with murine genomic GAPDH. **(b)** Adoptively transferred HER2.CARTs (EGFP⁺) at both tumor sites were separately analyzed by flow cytometry. Box plot elements: central line, median; box limit, upper and lower quartile; whisker, 1.5x inter-quartile range; points, outliers.

a

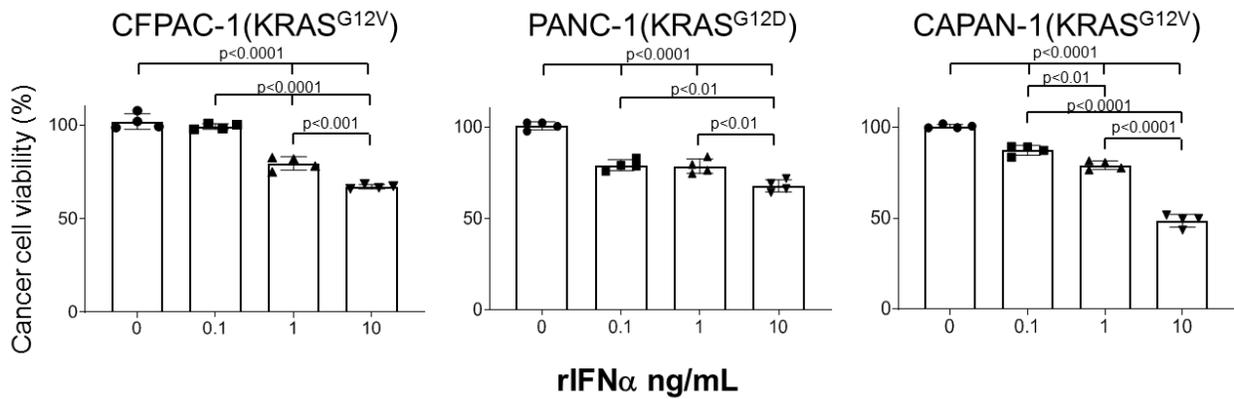
❖ 24 days post-injection of CAAd

**b**

Supplementary Figure 15. CAd components differently contribute to adoptively transferred HER2.CARTs in humanized mice with multiple PDAC tumors. (a) CFPAC-1 cells were transplanted into the right and left flanks of humanized mice (n=3 (CAdIL12) or 4 (CAd0, CAdPDL1) animals). A total of 1×10^7 vp of CAdS (OAd:HD=1:1) were injected into the right tumor. A total of 1×10^6 HER2.CARTs expressing *ffLuc* were systemically administered 3 days post-injection of CAdS. Tumor volumes were monitored at different time points. Tumor volumes shown here are from day 24 post injection of CAd, individual data points are represented in violin plot with means \pm SD. For comparison, tumor volumes from CART and CAdTrio+CART treated mice at day 24 post injection of CAd from **Figure 5** are shown. P-value was determined using unpaired two-tailed T test for left tumors. $p < 0.031$ ($t=2.597$, $df=8$). P-values were determined using ordinary one-way ANOVA with Tukey multiple comparisons for right tumors, $p=0.0008$ ($DF=16$), $p=0.06$ ($DF=16$), $p=0.04$ ($DF=16$), $p=0.06$ ($DF=16$). Statistical significance set at $p < 0.05$, $ns > 0.05$. **(b)** Bioluminescence of HER2.CARTs was monitored at different time points. For comparison, bioluminescence data of CART and CAdTrio+CART treated mice from **Figure 5** are shown here, denoted by dashed lines. Data are presented as means \pm SD. P-values were determined using ordinary one-way ANOVA with Tukey multiple comparisons, $p < 0.001$ ($F=8.641$). Statistical significance set at $p < 0.05$, $ns > 0.05$.



Supplementary Figure 16. *CAdTrio* induces *KLRC2* and *GNLY* expression at early time points. (a) CFPAC-1 cells were transplanted into the right flank of humanized mice, a total of 1×10^7 vp of *CAdTrio* (Onc:HD=1:1) were injected into the tumor. CFPAC-1 tumors were harvested from humanized mice (non-humanized mice: n=4 animals, humanized mice: n=5 animals) at 3 days post-injection of *CAdTrio*, and total RNA was extracted from whole tumors. *KLRC2* and *GNLY* genes were quantified and normalized with human β -Actin. Data are presented as means \pm SD.



Supplementary Figure 17. IFN α induces some PDAC cell death *in vitro*. PANC-1, CAPAN-1 and CFPAC-1 expressing ffLuc were cultured with increasing doses of rIFN α (n=4 independent biological samples). Cells were harvested 72 hours post-coculture, and viable cells were analyzed by luciferase assay. Data are presented as means \pm SD. P-values were determined by ordinary one-way ANOVA with Tukey multiple comparisons. CFPAC (F(3,12)=132.20), Panc1 (F(3,12)=74.83), CAPAN1 (F(3,12)=292.9). Statistical significance set at $p < 0.05$, ns > 0.05 .