

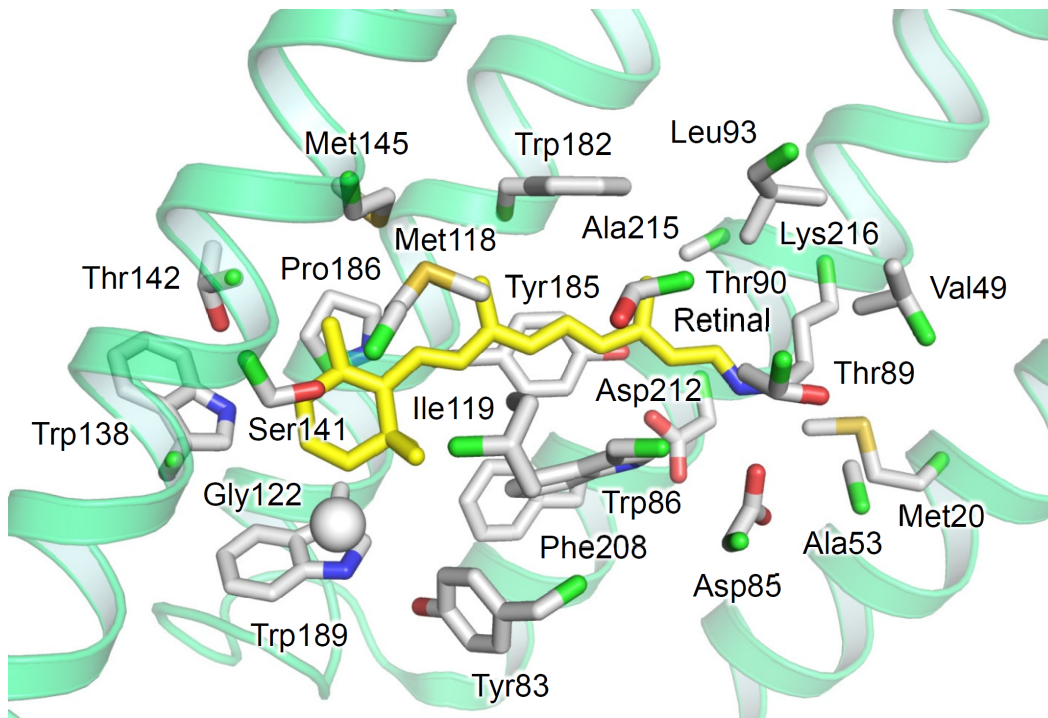
**Supplementary Information:**

**Exploration of natural red-shifted rhodopsins using a machine learning-  
based Bayesian experimental design**

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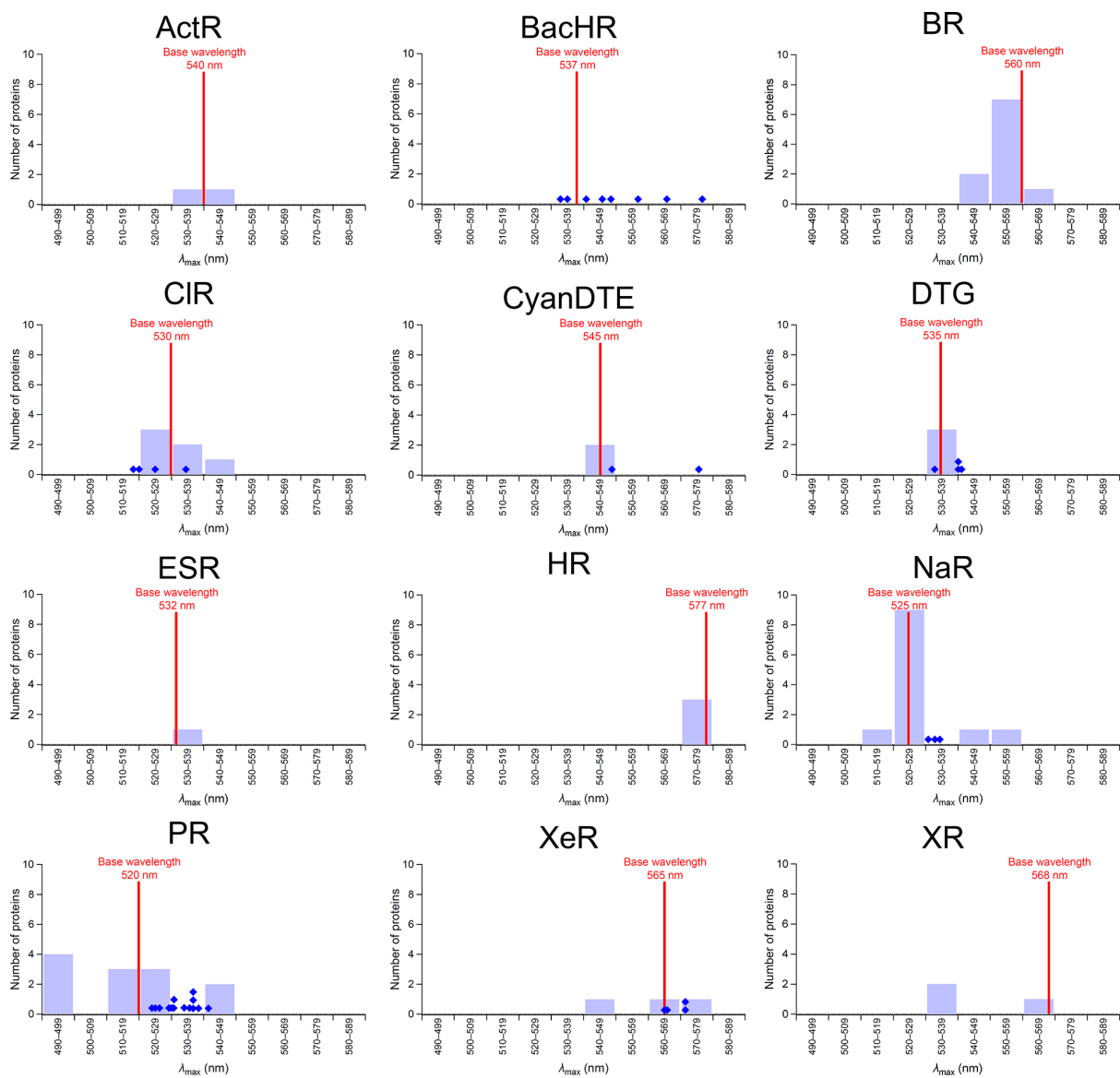
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<i>i</i>	1	2	3	4	5	6	7	8
Residue in BR	Met20	Val49	Ala53	Tyr83	Asp85	Trp86	Thr89	Thr90
<i>i</i>	9	10	11	12	13	14	15	16
Residue in BR	Leu93	Met118	Ile119	Gly122	Trp138	Ser141	Thr142	Met145
<i>i</i>	17	18	19	20	21	22	23	24
Residue in BR	Trp182	Tyr185	Pro186	Trp189	Phe208	Asp212	Ala215	Lys216

**Supplementary Figure 1. Amino acid residues around the retinal chromophore.** The structure of the 24 amino acid residues around the retinal used in the current ML model in the X-ray crystallographic structure of BR (PDB ID: 1IW6 (Matsui et al. *J. Mol. Biol.* (2002) 324, pp. 469–481)). The C $\alpha$  atom of Gly122 is shown as a white sphere. For clarity, the ribbon models of helices B, C, and E were omitted. The table lists the residue numbers and names of each residue in BR.



**Supplementary Figure 2. Distribution of  $\lambda_{\max}$  of wild-type microbial rhodopsins.**

Distribution of the number of microbial rhodopsins against their  $\lambda_{\max}$  (blue columns) for ion-pumping rhodopsin subfamilies. The base wavelengths of each subfamily and experimentally observed wavelength obtained in this study were indicated by red lines and blue diamonds, respectively. Since  $\lambda_{\max}$  of BacHR was not reported when we had constructed the training data, the distribution of the number of BacHRs against their  $\lambda_{\max}$  cannot be shown. Then,  $\lambda_{\max}$  of the first BacHR from cyanobacterium *Mastigocladopsis repens* was reported at 537 nm (Hasemi et al. *J. Biol. Chem.* (2016) 291, pp. 355–362)), and it was set to the base wavelength of BacHRs.