nature research

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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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| Fora | all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section. |
|------|---|
| n/a | Confirmed |
| | $oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| | 🗷 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| | The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section. |
| x | A description of all covariates tested |
| × | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| | For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i> |
| × | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| x | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| x | Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated |
| | Our web collection on <u>statistics for biologists</u> contains articles on many of the points above. |
| Sof | tware and code |

Policy information about <u>availability of computer code</u>

Data collection

Cell Profiler v3.1.9; TopSpin v3.5; Bio-Rad Image Lab v6.0.1; GROMACS v2018.2

Data analysis

ImageJ v1.52a; TopSpin v3.5; FoxTrot v2.0.2; VMD v1.9.3

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Provide your data availability statement here.

Life sciences study design

| All studies must d | isclose on these points even when the disclosure is negative. | | |
|--|---|--|--|
| Sample size | For Microtubule bench experiments, the length analyzed along the microtubule network was at least 0.5 mm for each condition. The number of clusters was determined and indicated. Paired two-sample Kolmogorov-Smirnov - and t-tests were used to test the significance. Four biological replicates were performed for each condition and values were then processed as described by Maucuer et al. J Cell Sci (2018). For Stress granule experiments, cell cytoplasm and nucleus were detected automatically by using Cell Profiler. Values were then analyzed in order to statistically distinguish the distribution of YB-1 or Lin28 protein in cells. The cell number chosen provides a sufficiently good statistical significance regarding the changes analyzed in this study. For Molecular Dynamics simulations, the sample size for prediction was based on 200 ns of NPT production run (except for Lin28:RRM1-TDP43:Lin28:RNA trimer complex where we ran only 10 ns of MD, which was sufficient to observe the destabilization of the complex) | | |
| Data exclusions | NO | | |
| Replication | Experiments were repeated in two or three independent replicates, as indicated in figure legends or in the online methods to reliably support conclusions stated in the manuscript. | | |
| Randomization | No data ramdomization was performed for any of experiments cited in this study. | | |
| Blinding | No blinding was applied in this study. | | |
| Reporting for specific materials, systems and methods We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response. | | | |
| Materials & e | xperimental systems Methods | | |
| n/a Involved in | ` | | |
| Antibodi | es | | |
| Eukaryot | ic cell lines | | |
| 🗶 🗌 Palaeont | ology and archaeology MRI-based neuroimaging | | |
| Animals and other organisms | | | |
| Human r | Human research participants | | |
| Clinical d | Clinical data | | |
| | | | |
| Dual use | research of concern | | |

Antibodies

Antibodies used Information about the commercial antibodies are provided in the manuscript. Oly YB-1 antibody are home made.

Validation The home made primary antibody, Anri-YB-1, has been tested by siRNA (Figure S7B)

Eukaryotic cell lines

| Policy information about <u>cell lines</u> | | | |
|---|--|--|--|
| Cell line source(s) | NCS-34, HeLa cells, HEK 293 (ATTC) | | |
| Authentication | None | | |
| Mycoplasma contamination | HeLa cells and HEK 293 celles were tested negative for mycoplasma contamination. | | |
| Commonly misidentified lines (See ICLAC register) | The cell line used in this study is not listed in the ICLAC database. | | |