

Figure S1. Effect of E484K substitution on C144/C051/C052 lineage neutralizing potency, related to Figure 1.

Neutralization of rVSV/SARS-CoV-2 2E1 or a plaque purified E484K mutant thereof, in 293T/ACE2cl.22 cells. Infected (%GFP+) cells relative to no antibody controls, mean and range of two independent experiments is plotted.



		11011011000000	-								
		(passage)	T478K	T478R	G485S	G485D	F486V	F486S	F486I	F486P	F486L
	10µg/ml	1D7 (p1)	0.018	0	0	0	0	0	0	0	0
0140	10µg/ml	1D7 (p2)	0.668	0.087	0.036	0.006	0.008	0.016	0	0	0.039
0143	10µg/ml	2E1 (p1)	0.037	0.052	0	0.028	0	0	0	0	0
	10µg/ml	2E1 (p2)	0.199	0.615	0	0.075	0	0	0	0	0.066
	10µg/ml	1D7 (p1)	0.264	0	0.022	0.015	0.016	0.011	0.032	0	0.045
0104	10µg/ml	1D7 (p2)	0.798	0.018	0.075	0	0.029	0.024	0.048	0	0.028
C164	10µg/ml	2E1 (p1)	0.282	0.331	0.032	0	0.026	0	0	0	0.022
	10µg/ml	2E1 (p2)	0.402	0.546	0.022	0	0.02	0	0	0	0
COFF	10µg/ml	1D7 (p1)	0.006	0	0.195	0.127	0.275	0.213	0.069	0.05	0
C055	10ua/ml	2E1 (p2)	0.028	0	0.221	0.189	0.365	0.135	0.019	0	0



Figure S2. Effects of somatic mutation on potency and viral escape in the C143/C164/C055 class 2 antibody lineage, related to Figure 1.

(A) Neutralization potency (IC₅₀) of C143, C164 and C055 measured using HIV-1-based SARS-CoV-2 variant pseudotypes and HT1080/ACE2cl.14 cells. The E484K substitution was constructed in an R683G (furin cleavage site mutant) background to increase infectivity. Mean of two independent experiments.

(B) Decimal fraction (color gradient; white = 0, red = 1) of Illumina sequence reads encoding the indicated RBD substitutions following rVSV/SARS-CoV-2 replication (1D7 and 2E1 virus

isolates) in the presence of the indicated amounts of antibodies for the indicated number of passages.

(C) C164 neutralization of rVSV/SARS-CoV-2 1D7, 2E1 or plaque purified mutants thereof, isolated following antibody selection, in 293T/ACE2cl.22 cells. Infected (%GFP+) cells relative to no antibody controls, mean and range of two independent experiments is plotted. (D) As in C for antibody C055.



Figure S3. Effect of viral substitutions on neutralization by C548 and C549, related to Figure 1

(A) C548 neutralization of HIV-1-based SARS-CoV-2 pseudotypes harboring the indicated substitutions that were identified by selection experiments. Infection (NanoLuc luciferase activity) is normalized to that obtained in the absence of antibody, mean and range of two independent experiments is plotted.

(B) As in A for antibody C549.



Figure S4. Effect of viral substitutions on neutralization by C098 and C099, related to Figure 2

(A) C098 neutralization of HIV-1-based SARS-CoV-2 pseudotypes harboring the indicated substitutions that were identified by selection experiments. Infection (NanoLuc luciferase activity) is normalized to that obtained in the absence of antibody, mean and range of two independent experiments is plotted.

(B) As in A for antibody C099.



Figure S5 Cryo-EM data processing and X-ray structures, related to Figures 6,7

(A-C) Representative micrograph, 2D class averages, FSC plots calculated using the goldstandard FSC criteria, and local resolution maps rendered in cryoSparc v2.15 for the cryo-EM structures of (A) C051-S, (B) C032-S, and (C) C548-S complexes.

(D-G) Cartoon representations of crystal structures of (D) C098, (E) C099, (F) C032, and (G) C080 Fabs.

(H) Cartoon representation of C098 Fab – SARS-CoV-2 RBD crystal structure.

(I) Cartoon representation of C099-CR3022 – SARS-CoV-2 RBD crystal structure.



Figure S6. Class 1 and 2 antibody sequence alignments and interactions with RBD, related to Figure 6.

(A) Sequence alignment between heavy and light chains of C098 and C099 relative to inferred germline sequences. Paratope residues highlighted in red.

(B) C098 epitope (light green surface on RBD with paratope sidechains from C098 highlighted as sticks).

(C) C099 epitope (light cyan surface on RBD with paratope sidechains from C099 highlighted as sticks). Somatic hypermutations found in C099 are highlighted with a red box.

(D) Overlay of V_H - V_L domains of class 1 Fabs bound to RBD (C098, green – this study; C099, blue – this study; C102, salmon – PDB 7K8M; CC12.3, cyan – PDB 6XC4; B38, gray – PDB 7BZ5; CV30, yellow – PDB 6XE1).

(E) Overlay of CDRH3 loops of class 1 Fabs described in panel D at the RBD interface.

(F) Sequence alignment between heavy and light chains of C144 and C051 relative to inferred germline sequences. Paratope residues highlighted in red.

(G) C144 epitope (light blue surface on RBD with paratope sidechains from C144 highlighted as sticks).

(H) C051 epitope (light orange surface on RBD with paratope sidechains from C051 highlighted as sticks).



Figure S7. Class 2 and 3 antibody sequence alignments and homology models, related to Figure 7.

(A) Sequence alignment between heavy and light chains of C548/C549 and C032/C080 antibody pairs relative to inferred germline sequences. Paratope residues highlighted in red.
(B) Homology model of C549-RBD complex. Antibody somatic mutations are shown as sticks. Residues changed by somatic hypermutation at the predicted RBD interface are indicated by an asterisk and enclosed in a red box.

(C) Predicted interactions between RBD (light gray) and C549 homology model LC residues (violet). C548 residues (light green) are shown.

(D) Homology model of the C080-RBD complex. Antibody somatic mutations are shown as sticks. Residues changed by somatic hypermutation at the predicted RBD interface are indicated by an asterisk and enclosed in a red box.

(E) Homology model of the C080-SARS-CoV RBD complex. Predicted RBD epitope and Fab paratope are shown as colored surface and sticks, respectively. Residues changed by somatic hypermutation at the predicted RBD interface are indicated by an asterisk and enclosed in a red box. Sequence differences in SARS-RBD relative to SARS-CoV-2 RBD are indicated with italics.

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				Heavy	Light				
	MADID	IGHV	IGHD	IGHJ	CDRH3	IGLV	IGLI	CDRL3	
	Germline	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	ARYYDFWSGYYFDY	IGLV2-14*01	IGLJ1*01	SSYTSSST-V	
C144	(1.3 months)	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	EGEVEG.NYSRDR	IGLV2-14*01	IGLJ1*01	R.	
C051	(6.2 months)	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	EGDVEG.H.SYSRDR	IGLV2-14*01	IGLJ1*01	NNN.R.	
C052	(6.2 months)	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	EGDVEGYSRDR	IGLV2-14*01	IGLJ1*01	NNN.R.	
C053	(6.2 months)	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	EGDVEGYSRDR	IGLV2-14*01	IGLJ1*01	AR.	
C054	(6.2 months)	IGHV3-53*01	IGHD3-3*01	IGHJ4*02	EGDVEGFS.LYSRDR	IGLV2-14*01	IGLJ1*01	FN.R.	
	Germline	IGHV1-69*01	IGHD3-10*01	IGHJ6*02	AYGYYYYYGMDV	IGLV9-49*01	IGLJ3*02	GADHGSGSNFV-V	
C548	(1.3 months)	IGHV1-69*01	IGHD3-10*01	IGHJ6*02	ARREPRD	IGLV9-49*01	IGLJ3*02	QG.	
C549	(6.2 months)	IGHV1-69*01	IGHD3-10*01	IGHJ6*02	ARREPPRDFF	IGLV9-49*01	IGLJ3*02	EGTG.	
	Germline	IGHV5-51*01	IGHD6-19*01	IGHJ2*01	ARAVDWYFDL	IGLV1-40*01	IGLJ1*01	QSYDSSLSYV	
C032	(1.3 months)	IGHV5-51*01	IGHD6-19*01	IGHJ2*01	<u>GV</u>	IGLV1-40*01	IGLJ1*01	AL	
C080	(6.2 months)	IGHV5-51*01	IGHD6-19*01	IGHJ2*01	GV	IGLV1-40*01	IGLJ1*01	SG.VDL	
	Germline	IGHV3-53*01	IGHD6-19*01	IGHJ3*02	ARYSSGDI	IGKV3-20*01	IGKJ1*01	QQYGSSP-T	
C098	(1.3 months)	IGHV3-53*01	IGHD6-19*01	IGHJ3*02	DLGT	IGKV3-20*01	IGKJ1*01	G.	
C099	(6.2 months)	IGHV3-53*01	IGHD6-19*01	IGHJ3*02	DLGT	IGKV3-20*01	IGKJ1*01	G.	
	Germline	IGHV3-66*01	IGHD4-23*01	IGHJ3*02	ARVAFDI	IGLV2-23*03	IGLJ3*02	CSYAGSSTFV	
C143	(1.3 months)	IGHV3-66*01	IGHD4-23*01	IGHJ3*02	DSSEVRDHPGHPGRSVG	IGLV2-23*03	IGLJ3*02	A	
C164	(1.3 months)	IGHV3-66*01	IGHD4-23*01	IGHJ3*02	DSSEVRDHPGHPGRSVG	IGLV2-23*02	IGLJ3*02	A	
C055	(6.2 months)	IGHV3-66*01	IGHD4-23*01	IGHJ3*02	DSSEVRDHPGHPGRSVG	IGLV2-23*01	IGLJ3*02	<mark>H</mark>	
	Germline	IGHV4-4*02	IGHD5-18*01	IGHJ4*02	ARDTAMYFDY	IGLV2-14*01	IGLJ3*02	SSYTSSSTL-	
C132	(1.3 months)	IGHV4-4*02	IGHD5-18*01	IGHJ4*02	GGGPE	IGLV2-14*01	IGLJ3*02	L	
C512	(6.2 months)	IGHV4-4*02	IGHD5-18*01	IGHJ4*02	.KGG.RGPES	IGLV2-14*01	IGLJ3*02	FAL	

Table S1 Clonally related antibody lineages in this study

Table S2. X-ray data collection and refinement statistics.

	C098 Fab SARS-CoV-2 RBD	C099 Fab CR3022 Fab SARS-CoV-2 RBD	C032 Fab	C080 Fab	C098 Fab	C099 Fab
PDB ID						
Data collection ^{a,b}						
Space group	C2	P41212	C2	P212121	P6522	P212121
Cell Dimenstions						
a, b, c (Å)	191.3, 87.8, 56.8	110, 110, 228.6	133.9, 61.7, 69.2	58.6, 67.4, 130.6	88.1, 88.1, 216.2	44.1, 92.6, 115.2
α, β, γ (°)	90, 99.8, 90	90, 90, 90	90, 97.3, 90	90, 90, 90	90, 90, 120	90, 90, 90
Resolution (Å)	40.7-2.0 (2.1-2.0)	39.6-2.6 (2.7-2.6)	36.0-2.1 (2.13-2.06)	36.6-1.9 (2.0-1.9)	43.2-1.4 (1.5-1.4)	43.0-1.3 (1.31-1.26)
R _{merge} (%)	9.7 (79.4)	25.2 (405)	7.7 (95.0)	16.73 (175.9)	7.9 (185.9)	9.6 (152.4)
R _{nim} (%)	6.5 (53.6)	7.4 (121.9)	5.2 (66.0)	8.1 (85.9)	2.0 (46.2)	4.3 (70.4)
CC ₁₂ (%)	99 5 (57 7)	99 5 (35 1)	99.2 (52.1)	99 2 (39 9)	99 9 (67 2)	99.7 (48.5)
< <u>I</u> / <u></u> <u></u> <u></u> <u></u> <u></u>	68(13)	89(11)	59(11)	79(20)	174(18)	83(10)
Completeness (%)	95 7 (97 0)	100 (100)	94 3 (95 5)	98.8 (97.4)	99.4 (99.1)	90 5 (97 7)
Redundancy	3.0 (3.0)	12.9 (12.6)	2.9 (2.8)	5.2 (5.2)	16.5 (16.8)	5.6 (5.5)
Wilson <i>B</i> -factor	30.4	58.6	40.9	24.2	18.9	13.2
Refinement and Validation						
Resolution (Å)	40.7-2.0 (2.1-2.0)	39.6-2.6 (2.66-2.6)	36.0-2.1 (2.13-2.06)	36.6-1.9 (2.0-1.9)	43.2-1.4 (1.5-1.4)	43.0-1.3 (1.31-1.26)
Unique Reflections	57,268 (5,762)	43,864 (2,600)	32,875 (3,295)	41.224 (4.044)	92.932 (9.042)	115.943 (12.409)
Number of atoms			-,	, (,)	, _,, , _ (, , ,)	,- (,,)
Protein	4750	8.109	3244	3266	3352	3266
Ligand	14	14	-	-	-	-
Waters	480	94	132	371	336	625
Rwork/Rfree (%)	17.9/20.8	18.9/23.7	20.0/22.7	18.6/22.3	18.6/20.5	18.0/20.1
R.m.s. deviations						
Bond lengths (Å)	0.009	0.009	0.008	0.012	0.01	0.011
Bond angles (°)	1.1	1.1	1.1	1.1	1.2	1.2
Poor rotamers (%)	0	0.82	0	0.55	1.0	1.10
Ramachandran plot						
Favored (%)	97.4	95.9	96.5	97.9	98.6	98.1
Allowed (%)	2.1	3.8	3.5	2.1	1.4	1.7
Disallowed (%)	0.5	0.3	0	0	0	0.2
Average B-factor (Å)	40.6	59.7	55.4	31.2	31.0	19.9

^aFor each structure reported, data were derived from a single crystal. ^bNumbers in parentheses correspond to the highest resolution shell

	C032	C548	
	SARS-CoV-2 S6P	SARS-CoV-2 S 6P	SARS-CoV-2 S 6P
PDB			
EMD			
Data collection conditions			
Microscope	Talos Arctica	Talos Artica	Talos Artica
Camera	Gatan K3 Summit	Gatan K3 Summit	Gatan K3 Summit
Magnification	45,000x	45,000x	45,000x
Voltage (kV)	200	200	200
Recording mode	counting	counting	counting
Dose rate (e ⁻ /pixel/s)	13.5	13.5	13.5
Electron dose $(e^{-/} Å^2)$	60	60	60
Defocus range (um)	0.7 - 2.0	0.7 - 2.0	0.7 - 2.0
Pixel size (\mathring{A})	0.8689	0.8689	0.8689
Micrographs collected	3 480	3 402	1959
Micrographs used	2 683	2 927	1687
Total extracted particles	844 544	390 630	554 852
Refined particles	192 286	134 506	94 255
Symmetry imposed	C1	C3	C3
Nominal Man Resolution (Å)	01	05	05
FSC 0.143 (unmasked/masked)	5 4/3 4	4 9/3 5	1 5/3 5
FSC 0.143 local (unmasked/masked)	6.7/4.4	5.8/4.1	N/A
Refinement and Validation			
Initial model used	6XKI	78.90	7K43
Number of atoms	OTILL	/10/0	/11/15
Protein	25 866	28 401	28136
Ligand	56	462	574
ManCC (global/local)	0 70/0 69	0.80/0.77	0.77/0.76
Man sharpening B-factor	69.3	64 5	110.1
R m s deviations	07.0	0.110	11001
Bond lengths (Å)	0.004	0.008	0.003
Bond angles (°)	0.64	1.1	0.52
MolProbity score	2.18	2.29	2.48
Clashscore (all atom)	16.3	16.4	18.26
Poor rotamers (%)	0	0.2	0.38
Ramachandran plot	~		
Favored (%)	92.4	93.8	97
Allowed (%)	7.5	6.9	2.7
Disallowed (%)	0	0.3	0.3