



## Supplementary Material

**Supplementary Figure 1.** Recombinant ATAT purifications. (A) Coomassie blue-stained SDS-PAGE of the 6xHis-tagged *Tc*ATAT purification from *E. coli* used for rabbit immunization and polyclonal antibodies production. MWM, molecular weight marker; (-), bacterial lysate uninduced with IPTG; FT, flowthrough obtained after incubating the induced bacterial lysate with the Ni<sup>+</sup>-agarose resin; W, wash; D1-D4, elution with NaH<sub>2</sub>PO<sub>4</sub> 100 mM, Tris 10 mM, Urea 8 M pH 5.9; E1-4, elution with NaH<sub>2</sub>PO<sub>4</sub> 100 mM, Tris 10 mM, Urea 8 M pH 4.5. (B) Coomassie blue-stained SDS-PAGE of the HA-tag *Tc*ATAT purification from *T. cruzi* epimastigotes using an anti-HA resin. This protein was used for the autoacetylation assay. MWM, molecular, weight marker; FT, flowthrough obtained after incubating the resin with the epimastigotes lysate; W, wash; 1-4, elution with HA peptide 0.1 mg/mL.

Tb427_030013700	1 MTH-	NVMCDDVL	PQLNL-PDGVTR	WNANLLEE	ERRLRNSDGH	ADRII
LmjF.25.1150	1 MRRR	PPQLTKTKLADEEV	PELTLVPDGVSR	NTGSDLDALLNA	IARRGGAEAAQ	QDLERKLC
TcCLB.467287.10	1 MSS-	TSQ-VALL	PKLSL-PDGVT <mark>V</mark>	NDGTALEY	ERRCNNVDEH	AVHLM
Tb427_030013700	46 LTIN	TLGKRSKEAQSLNT	ILTSV <mark>P</mark> RLRENRI	DARLYLLC <mark>HG</mark> GF	GVGILKIGVK	RLFV <mark>VP</mark> PS
LmjF.25.1150	61 RTID	ILGARSQQAQEINA	VLTSVARLREN <mark>S</mark>	IFRLYLLTONHF	GVGILKVGVK	KLFVTHP <mark>V</mark>
TcCLB.467287.10	45 QTIN	ILG <mark>I</mark> RSKEAQ <mark>C</mark> LNT	VLTSVARLRENR	DARVYLLCODGY	GVGILKMGVK	KLFVTHPS
Tb427_030013700	106 HAGL	MEIEPVCVLDFFVD	TS <mark>NQRQGYGKILI</mark>	FEHMLAFERLS-	PGDVAIDRPS	VKFLAFLR
LmjF.25.1150	121 TC <mark>GL</mark>	VEVDPLCVLDFYVD	E <mark>SCQRQGYGKML</mark>	YSHMLKAEHVSF	P <mark>EV</mark> LAIDRPS	N <mark>KL</mark> LGFLR
TcCLB.467287.10	105 YSSL	VEIDPLCVLDFFVD	TSFQR <mark>KGFGKT</mark> LI	FDAMLLN <mark>EGL</mark> N-	PGEVAIDRPS	VKFLAFL <mark>Q</mark>
Tb427_030013700	165 KHYG	LVEYTPQSNNFVVF	HKYFERHQQQRR(	SVGGSGRSC	·YQHQN	ETTTQ
LmjF.25.1150	181 KHYG	L <mark>AA</mark> YTPQ <mark>V</mark> NNFVVF	HSFFDHTTVSER(	GKLL	·YQHQN	
TcCLB.467287.10	164 KYYG	LVEYTPQSNNFVVF	HRYFDKWQPQR-(	GKGHW <mark>G</mark> GNAVPI	'RSLVRPQNGL	RVYPKYQS
Tb427_030013700	214 QLGT	QSGLLEDINQTHPA	PSYALRGVVMGH	ΓGPPLD <mark>L</mark> TNVTζ	200kpyh0pfa	TGRKTSYE
LmjF.25.1150	215	RAPS	P	Α		
TcCLB.467287.10	223 TTGP	NNNFE <mark>ED</mark> ATH <mark>R</mark> TPP	P	PP <mark>Ι</mark> PPPLV	yp <mark>0</mark> gsvts <b>p-</b> g	VGKKT <mark>A</mark> YE
Tb427_030013700	274 LQYE	RYLQSQNCRPTG	NAGYGGGNGPASS	SAEVRATNCQAF	RRTSPTRSGV	PYNIINGS
LmjF.25.1150	221		RASPFPSS	ANATVAIGGAKA	KNWT P	
TcCLB.467287.10	267 LQYE	EYLREQAYRRRQGG	DPRLQPVPNPVSS	SSEIVA <mark>ASC</mark> GAF	RR <mark>M</mark> SPTRSGV	QYNIISGT

**Supplementary Figure 2.** Multiple sequence alignment of *Tc*ATAT (TcCLB.467287.10) and its homologues in *Trypanosoma brucei* (Tb427\_030013700) and *Leishmania mayor* (LmjF.25.1150) using T-coffee server and colored with Boxshade. The acetyltransferase domain is boxed in green.



**Supplementary Figure 3.** Immunolocalization of ATAT-HA with rat monoclonal anti-HA antibodies in Dm28c p*Tc*INDEX-GW ATAT-HA epimastigotes uninduced (Without tet) and induced with 0.5  $\mu$ g/ml tetracycline 24 hours post-induction (h.p.i). Bar: 5  $\mu$ m. DAPI was used as nucleus and kinetoplast marker. The light blue arrow indicates the kinetoplast and the pink arrow indicates the nucleus.



**Supplementary Figure 4. (A)** Growth curve of Dm28c epimastigotes in the presence of increasing concentrations of Oryzalin (0-300 µM) (left panel) and number of parasites *versus* the log [Oryzalin]

at 72 h (right panel). The latter plot was fitted with the non-parametric regression log(inhibitor) vs. response -Variable slope (four parameters) in GraphPad Prism version 8.0 to obtain the IC<sub>50</sub> value and the R<sup>2</sup> of the fit. **(B)** DIC images of the morphological changes in Dm28c epimastigotes with different concentrations of Oryzalin at 72 hours. Bar = 10  $\mu$ m.



**Supplementary Figure 5. (A)** Immunolocalization of ATAT-HA with rat monoclonal anti-HA antibodies in isolated cytoskeletons of Dm28c pTcINDEX-GW ATAT-HA epimastigotes induced with 0.5  $\mu$ g/ml tetracycline for 24 h (**B**) Immunolocalization of acetylated a-tubulin with mouse monoclonal anti-acetylated a-tubulin (anti-AcTub) in isolated cytoskeletons of Dm28c pTcINDEXGW ATAT-HA epimastigotes cytoskeletons uninduced (-Tet) and induced with 0.5  $\mu$ g/ml tetracycline for 24 h (**+** Tet). (**C**) Immunolocalization of ATAT-HA with rat monoclonal anti-HA antibodies and mouse 5 polyclonal anti-paraflagellar rod 2 from *T. cruzi* (anti-PFR) in isolated cytoskeletons of Dm28c pTcINDEX-GW ATAT-HA epimastigotes induced with 0.5  $\mu$ g/ml tetracycline for 24 h.



**Supplementary Figure 6. (A)** Viability (% of live epimastigotes) of Dm28c p*Tc*INDEX-GW ATAT-HA epimastigotes uninduced (grey bar, -tet) and induced with 0.5  $\mu$ g/ml tetracycline (green bar, +tet) determined by counting live cells with a hematocytometer using Erythrosin B staining for 7 days. **(B)** Epimastigotes movements were examined using the computer-assisted semen analysis (CASA) system (Microptic, SCA evolution). The mean path velocity (VAP,  $\mu$ m/sec) was plotted at 24 and 48 h.p.i. without (grey bar, -tet) and with 0.5  $\mu$ g/ml tetracycline (green bar, +tet). Experiments were performed in triplicates.



**Supplementary Figure 7.** Nucleus/Kinetoplast content (N/K) of Dm28c p*Tc*INDEX-GW ATAT-HA epimastigotes cultures in the absence (-) or presence (+) of 0.5  $\mu$ g/ml tetracycline at different time points (24 and 48 hours). Data from three independent experiments were considered in the analysis (n = 300 cells for each column).