

1. Supplementary Tables

Supplementary Table 1: DAKD and its analogues used in this work with different isotope labeling schemes or chemical modifications. Peptides for ssNMR experiments were uniformly ^{13}C , ^{15}N -labelled on specific residue sites.

Peptide	Labeling scheme	Abbreviation
DAKD	$(^{13}\text{C}, ^{15}\text{N})\text{K-RPPGFSPF}^{(a)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -K ₁ DAKD
	K- $(^{13}\text{C}, ^{15}\text{N})\text{R-PPGF-}(^{13}\text{C}, ^{15}\text{N})\text{S-PF}^{(b)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -R ₂ S ₇ DAKD
	KR- $(^{13}\text{C}, ^{15}\text{N})\text{P-PGFSPF}^{(a)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -P ₃ DAKD
	KRP- $(^{13}\text{C}, ^{15}\text{N})\text{P-GFSPF}^{(a)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -P ₄ DAKD
	KRPP- $(^{13}\text{C}, ^{15}\text{N})\text{G-}(^{13}\text{C}, ^{15}\text{N})\text{F-SPF}^{(a)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -G ₅ F ₆ DAKD
	KRPPGFS- $(^{13}\text{C}, ^{15}\text{N})\text{P-}(^{13}\text{C}, ^{15}\text{N})\text{F}^{(b)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -S ₇ P ₈ DAKD
DALK	KRPPGFS- $(^{13}\text{C}, ^{15}\text{N})\text{P-}(^{13}\text{C}, ^{15}\text{N})\text{L}^{(a)}$	U- $[^{13}\text{C}, ^{15}\text{N}]$ -P ₈ L ₉ DALK
DAKD(N-Methyl F6)	KRPPG-(N-Methyl)F-SPF ^(a)	DAKD(N-Methyl F6)

(a) Thermo Fisher Scientific, Ulm, Germany, (b) Eurogentec, Cologne, Germany

Supplementary Table 2: Chemical Shift assignment of DAKD bound to B₁R*

Residue	Atoms								
	N	C'	C α	C β	C γ	C δ	C ϵ	C ζ	N(sidechain)
K1	40.2	173.8	55.9	33.4	-	29.5	41.8		32.5 (N ζ)
R2	121.5	174.8	55.0	30.8	27.5	43.3		158.7	83.5 (N ϵ) 73.6(N η)
P3	133.9	175.7	61.7	31.2	28.5	50.6			
P4	131.7	177.4	63.5	32.5	28.4	50.4			
G5	111.9	174.7	45.2						
F6	118.7	178.5	59.0	40.1	129.1	131.7	131.7	134.1	
S7	121.8	174.4	55.4	62.6					
P8	135.1	175.9	62.7	32.7	28.5	50.6			
F9	124.4	181.0	57.3	39.4	128.5	131.6	131.6	133.8	

* ^{13}C chemical shifts (in ppm) are referenced to TSP.

* ^{15}N chemical shifts (in ppm) are referenced to liquid NH_3 .

Supplementary Table 3: Chemical Shift assignment of DAKD without B₁R* (solid-state NMR)

Residue	Atoms								
	N	C'	C α	C β	C γ	C δ	C ϵ	C ζ	N(sidechain)
K1	40.7	173.4	55.5	33.6	22.5	29.4	42.2		33.4 (N ζ)
R2	121.1	175.1	55.0	30.3	26.8	43.2		158.8	83.2 (N ϵ) 72.6(N η)
P3	133.2	174.8	61.8	31.1	27.9	50.8			
P4	130.7	178.3	63.3	32.7	28.3	50.6			
G5	111.3	174.7	45.1						
F6	118.5	178.6	58.7	40.0	129.3	132.2	132.2	134.7	
S7	121.1	174.0	56.1	62.7					
P8	135.8	176.0	62.6	32.8	28.5	51.0			
F9	126.5	181.6	58.6	39.8	129.1	130.8	132.3	132.9	

* ¹³C chemical shifts (in ppm) are referenced to TSP.

* ¹⁵N chemical shifts (in ppm) are referenced to liquid NH₃.

Supplementary Table 4: Chemical Shift Differences of bound DAKD – free DAKD (solid-state NMR)

Residue	Atoms								
	N	C'	C α	C β	C γ	C δ	C ϵ	C ζ	N(sidechain)
K1	-0.5	+0.4	+0.4	-0.2		+0.1	-0.4		-0.9 (N ζ)
R2	+0.4	-0.3	0.0	+0.5	+0.7	+0.1		-0.1	+0.3(N ϵ) +1.0(N η)
P3	+0.7	+0.9	-0.1	+0.1	+0.6	-0.2			
P4	+1.0	-0.9	+0.2	-0.2	+0.1	-0.2			
G5	+0.6	0.0	+0.1						
F6	+0.2	-0.1	+0.3	+0.1	-0.2	-0.5	-0.5	-0.6	
S7	+0.7	+0.4	-0.7	-0.1					
P8	-0.7	-0.1	+0.1	-0.1	0.0	-0.4			
F9	-2.1	-0.6	-1.3	-0.4	-0.6	+0.8	-0.7	+0.9	

* ¹³C chemical shifts (in ppm) are referenced to TSP.

* ¹⁵N chemical shifts (in ppm) are referenced to liquid NH₃.

Supplementary Table 5: Chemical Shift assignment of DAKD without B₁R* in solution without detergent at 298K *298K, H₂O/D₂O, 50mM MES, pH5.6, 100mM NaCl

Residue	Atoms												
	N	NH	C'	C α	C β	C γ	C δ	C ϵ	H α	H β	H γ	H δ	H ϵ
K1	-	-	172.3	55.5	33.2	23.8	29.1	41.9	3.94	1.79	1.34	1.60	2.90
R2	125.1	8.66	173.3	54.2	30.0	26.7	43.3		4.50	1.62	1.54	2.98	7.01
P3	-		174.4	-	30.7	27.3	50.6		-	1.79/2.28	1.92	2.74/3.39	
P4	-		177.4	63.3	31.9	27.4	50.4		4.30	1.80/2.19	1.94	3.56/3.71	
G5	109.1	8.35	173.7	45.0					3.74/3.83				
F6	120.2	7.89	175.3	57.9	39.8	-	-	-	4.45	2.94	-	7.11	7.22
S7	119.8	8.04	172.3	55.5	63.7				4.54	3.61			
P8	-		175.3	63.4	31.7	26.7	50.6		4.20	1.71/2.04	1.60/1.80	3.46	
F9	123.5	7.33	180.0	58.7	40.2	-	-	-	4.32	2.84/3.04	-	7.13	7.24

* ¹³C chemical shifts (in ppm) are referenced to TSP.* ¹⁵N chemical shifts (in ppm) are referenced to liquid NH₃.

Supplementary Table 6: Solution NMR and refinement statistics for free DAKD in solution without detergents

	ARIA	CYANA (TALOS+)	CYANA (TALOS-N)	Forward cluster A
NMR distance and dihedral constraints				
Distance constraints				
Total NOE	73	-	-	-
Intra-residue	48	-	-	-
Inter-residue				
Sequential ($ i - j = 1$)	24	-	-	-
Medium-range ($ i - j < 4$)	1	-	-	-
Long-range ($ i - j > 5$)	-	-	-	-
Intermolecular	-	-	-	-
Hydrogen bonds	0	-	-	-
Total dihedral angle restraints	14	11	11	11
ϕ	7	4	4	4
ψ	7	7	7	7
Structure statistics				
Violations (mean and s.d.)				
Distance constraints (Å)	0.039 ± 0.002	-	-	-
Dihedral angle constraints (°)	0.83 ± 0.19	2.64 ± 0.88	2.56 ± 0.87	0.20 ± 0.07
Max. dihedral angle violation (°)	5.42	11.90	15.35	2.22
Max. distance constraint violation (Å)	0.312	-	-	-
Number of CYANA restraints violations*	-	3	2	0
CYANA target function (10^{-2})	-	1.59	1.88	0.75
Deviations from idealized geometry				
Bond lengths (Å)	0.008	0.001	0.001	0.001
Bond angles (°)	0.6	0.3	0.3	0.3
Impropers (°)	1.05	0.64	0.64	0.64
Average pairwise r.m.s. deviation** (Å)				
Heavy	2.80 ± 0.73 (20)	1.63 ± 0.24 (10)	1.57 ± 0.23 (10)	1.68 ± 0.24 (10)
Backbone	1.29 ± 0.47 (20)	0.57 ± 0.13 (10)	0.53 ± 0.12 (10)	0.58 ± 0.13 (10)
Ramachandran distribution				
Most favored (%)	98.3	100	100	100
Additional allowed (%)	1.7	0	0	0
Generally allowed (%)	0	0	0	0
Disallowed region (%)	0	0	0	0

* Violations are counted if at least 3 out of 10 structural models show violation

** The number of structural models for computing pairwise r.m.s. deviation was listed in ().

Supplementary Table 7: ssNMR and refinement statistics for DAKD bound to B1R

	CYANA (TALOS+)	CYANA (TALOS-N)	CYANA (PREDITOR)	Forward cluster A
NMR distance and dihedral constraints				
Distance constraints				
Total NOE	-	-	-	-
Intra-residue	-	-	-	-
Inter-residue				
Sequential ($ i - j = 1$)	-	-	-	-
Medium-range ($ i - j < 4$)	-	-	-	-
Long-range ($ i - j > 5$)	-	-	-	-
Intermolecular	-	-	-	-
Hydrogen bonds	-	-	-	-
Total dihedral angle restraints	11	11	10	13
ϕ	4	4	4	5
ψ	7	7	6	8
Structure statistics				
Violations (mean and s.d.)				
Distance constraints (Å)	-	-	-	-
Dihedral angle constraints (°)	0.43 ± 0.14	1.00 ± 0.43	0.87 ± 0.29	0.78 ± 0.26
Max. dihedral angle violation (°)	4.78	10.85	8.72	5.48
Max. distance constraint violation (Å)	-	-	-	-
Number of CYANA restraints violations*	0	2	2	1
CYANA target function (10^{-3})	7.61	14.7	7.11	8.53
Deviations from idealized geometry				
Bond lengths (Å)	0.001	0.001	0.001	0.001
Bond angles (°)	0.3	0.3	0.3	0.3
Impropers (°)	0.64	0.64	0.64	0.64
Average pairwise r.m.s. deviation** (Å)				
Heavy	1.67 ± 0.18	1.46 ± 0.31	2.07 ± 0.31	1.49 ± 0.34
Backbone	0.50 ± 0.09	0.36 ± 0.10	0.82 ± 0.23	0.39 ± 0.12
Ramachandran distribution				
Most favored (%)	100	97	60	100
Additional allowed (%)	0	3	40	0
Generally allowed (%)	0	0	0	0
Disallowed region (%)	0	0	0	0

* Violations are counted if at least 3 out of 10 structural models show violation

** Pairwise r.m.s. deviations were calculated on 10 structural models.

Supplementary Table 8: SSNMR and TALOS+/CYANA refinement statistics for DAKD and BK in different states

	DAKD (+ B1R)	DAKD (- B1R)	BK (+ B2R)	BK (- B2R)
NMR distance and dihedral constraints				
Distance constraints				
Total NOE	-	-	-	-
Intra-residue	-	-	-	-
Inter-residue				
Sequential ($ i - j = 1$)	-	-	-	-
Medium-range ($ i - j < 4$)	-	-	-	-
Long-range ($ i - j > 5$)	-	-	-	-
Intermolecular	-	-	-	-
Hydrogen bonds	-	-	-	-
Total dihedral angle restraints	11	11	10	10
ϕ	4	4	4	4
ψ	7	7	6	6
Structure statistics				
Violations (mean and s.d.)				
Distance constraints (Å)	-	-	-	-
Dihedral angle constraints (°)	0.43 ± 0.14	1.99 ± 0.67	2.32 ± 0.77	0.64 ± 0.21
Max. dihedral angle violation (°)	4.78	10.59	23.21	6.36
Max. distance constraint violation (Å)	-	-	-	-
Number of CYANA restraints violations*	0	2	1	1
CYANA target function (10^{-3})	7.61	13.8	21.1	5.07
Deviations from idealized geometry				
Bond lengths (Å)	0.001	0.001	0.001	0.001
Bond angles (°)	0.3	0.3	0.3	0.3
Impropers (°)	0.64	0.65	0.62	0.63
Average pairwise r.m.s. deviation** (Å)				
Heavy	1.67 ± 0.18	1.73 ± 0.25	1.63 ± 0.22	2.29 ± 0.42
Backbone	0.50 ± 0.09	0.43 ± 0.12	0.28 ± 0.07	0.58 ± 0.12
Ramachandran distribution				
Most favored (%)	100	100	77	97
Additional allowed (%)	0	0	23	3
Generally allowed (%)	0	0	0	0
Disallowed region (%)	0	0	0	0

* Violations are counted if at least 3 out of 10 structural models show violation

** Pairwise r.m.s. deviations were calculated on 10 structural models.

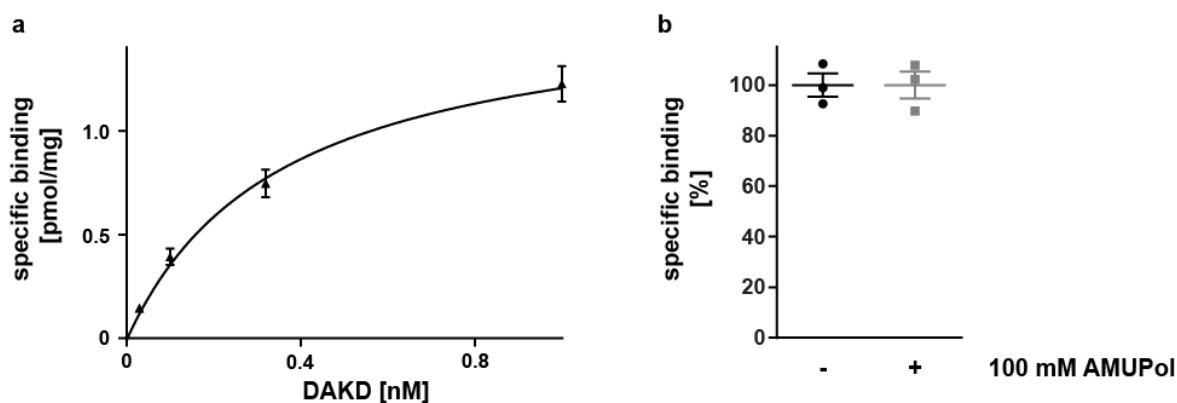
Supplementary Table 9: Summary of GPCR template X-ray structures used for B₁R/B₂R comparative modeling.

Template Name	Species	Sequence identity to B ₁ R (%)	Sequence identity to B ₂ R (%)	PDB code	Resolution (Å)	Reference
angiotensin II type 1 receptor	human	33	34	4yay	2.9	1
C-C chemokine receptor type 5	human	28	24	4mbs	2.7	2
κ-opioid receptor	human	24	27	4djh	2.9	3
orexin receptor type 2	human	20	21	4s0v	2.5	4
P ₂ Y ₁ receptor	human	22	25	4xnv	2.2	5
δ-opioid receptor	human	26	26	4n6h	1.8	6
μ-opioid receptor	human	24	26	4dkl	2.8	7
lysophosphatidic acid receptor 1	human	21	18	4z35	2.9	8
nociceptin receptor	human	24	27	4ea3	3.0	9
rhodopsin	bovine	21	23	2x72	3.0	10
5-hydroxytryptamine receptor 2B	human	19	21	4ib4	2.7	11
β ₁ -adrenergic receptor	turkey	19	19	2y03	2.9	12
proteinase-activated receptor 1	human	20	24	3vw7	2.2	13
muscarinic acetylcholine receptor M2	human	19	20	3uon	3.0	14
histamine H1 receptor	human	22	21	3rze	3.1	15
muscarinic acetylcholine receptor M3	rat	18	21	4u15	2.8	16
β ₂ -adrenergic receptor	human	19	20	3sn6	3.2	17
β ₂ -adrenergic receptor	human	19	21	2rh1	2.4	18
adenosine receptor A2a	human	21	22	3qak	2.7	19
adenosine receptor A2a	human	21	22	3eml	2.6	20
sphingosine 1-phosphate receptor 1	human	21	21	3v2y	2.8	21
dopamine D3 receptor	human	20	25	3pbl	2.9	22
5-hydroxytryptamine receptor 1B	human	19	21	4iar	2.7	23
free fatty acid receptor 1	human	19	15	4phu	2.3	24

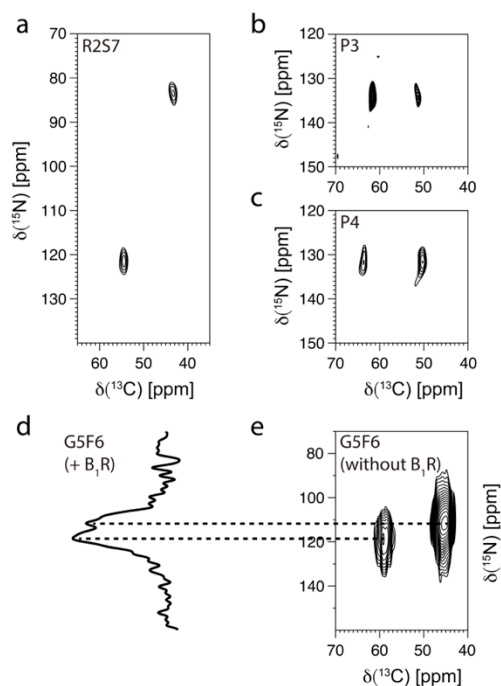
Supplementary Table 11: Comparison of peptide N-terminal binding region of B₁R from different mammals. The conserved binding residues are highlighted in yellow. The positions at which where changes in charge occur are labeled in red. Residue and BW numbering are based on human B₁R. All the non-rodent sequences as well as the common rat, mouse and Chinese hamster sequences were obtained from Uniprot. Sequences of other rodents were fished using Blast. The alignment were performed by Multalin server²⁵ using default parameters and the figure was prepared from the output of ESPript 3 (<http://nar.oxfordjournals.org/content/42/W1/W320.long>).

		270	280	290
		<i>E6.58</i>	<i>R6.62</i>	<i>E7.28 D7.29 D7.32</i>
Primates	Homo sapiens (human)	HFFAFLEFLFQVQAVRGC	FWEDFDL	DLGLQLA
	Pan troglodytes (chimpanzee)	HFFAFLEFLFQVQAVRGC	FWEDFDL	DLGLQLA
	Chlorocebus aethiops (African green monkey)	HFFAFLEFLFQVQAIRGCF	WEDFDL	DLGLQLA
	Papio anubis (olive baboon)	HFFAFLEFLFQVQAVRGC	FWEDFDL	DLGLQLA
Lagomorpha	Oryctolagus cuniculus (European rabbit)	HFFAFLECLWQVHAIGGCF	WEEFTD	DLGLQLS
Scandentia	Tupaia minor (pygmy tree shrew)	HCFAFLEFLFQVRAVRGC	FWEDFDL	DLGLQLA
Carnivora	Canis lupus familiaris (dog)	HFFAFLEFLTQVQVVRGC	FWENFKD	DLGLQYA
Rodents	Muridae Rattus norvegicus (brown rat)	HFFAFLDFLVQVRVIQDC	SWKEITD	DLGLQLA
	Muridae Mus musculus (house mouse)	HFFAFLDFLVQVRVIQDC	SWKEITD	DLGLQLA
	Cricetidae Cricetulus griseus (Chinese hamster)	HFFAFLDFLVQVKAIQHCS	WKEFTD	DLGLQLA
	Cricetidae Mesocricetus auratus (golden hamster)	HFFAFLDFLVQVKAIQDC	SWKEFTD	DLGLQLA
	Cricetidae Peromyscus maniculatus bairdii (deer mouse)	HFFAFLDFLVQVKAIQSC	AWKEFTD	DLGLQLA
	Cricetidae Microtus ochrogaster (prairie vole)	HFFAFLDFLVQVKVVQDC	SWKEFTD	DLGLQLA
	Cricetidae Neotoma lepida (desert woodrat)	HFFAFLDFLVQVKVIQDC	SWKEFTD	DLGLQLA
	Dipodidae Jaculus jaculus (lesser Egyptian jerboa)	HFFAFLDFLVQVRAIQDC	FWKEVTD	DLGLQLA
	Spalacidae Nannospalax galili (Northern Israeli blind subterranean mole rat)	HFFAFLDFLVQVRAIQSC	FWKEITD	DLGLQLA
	Bathyergidae Heterocephalus glaber (naked mole rat)	HFFAFLEFLFHVQAVQGC	FWEEVTD	DLGLQLA
	Bathyergidae Fukomys damarensis (Damara mole rat)	HFFAFLEFLFHVQAIQGC	CCWEEITD	DLGLQLA

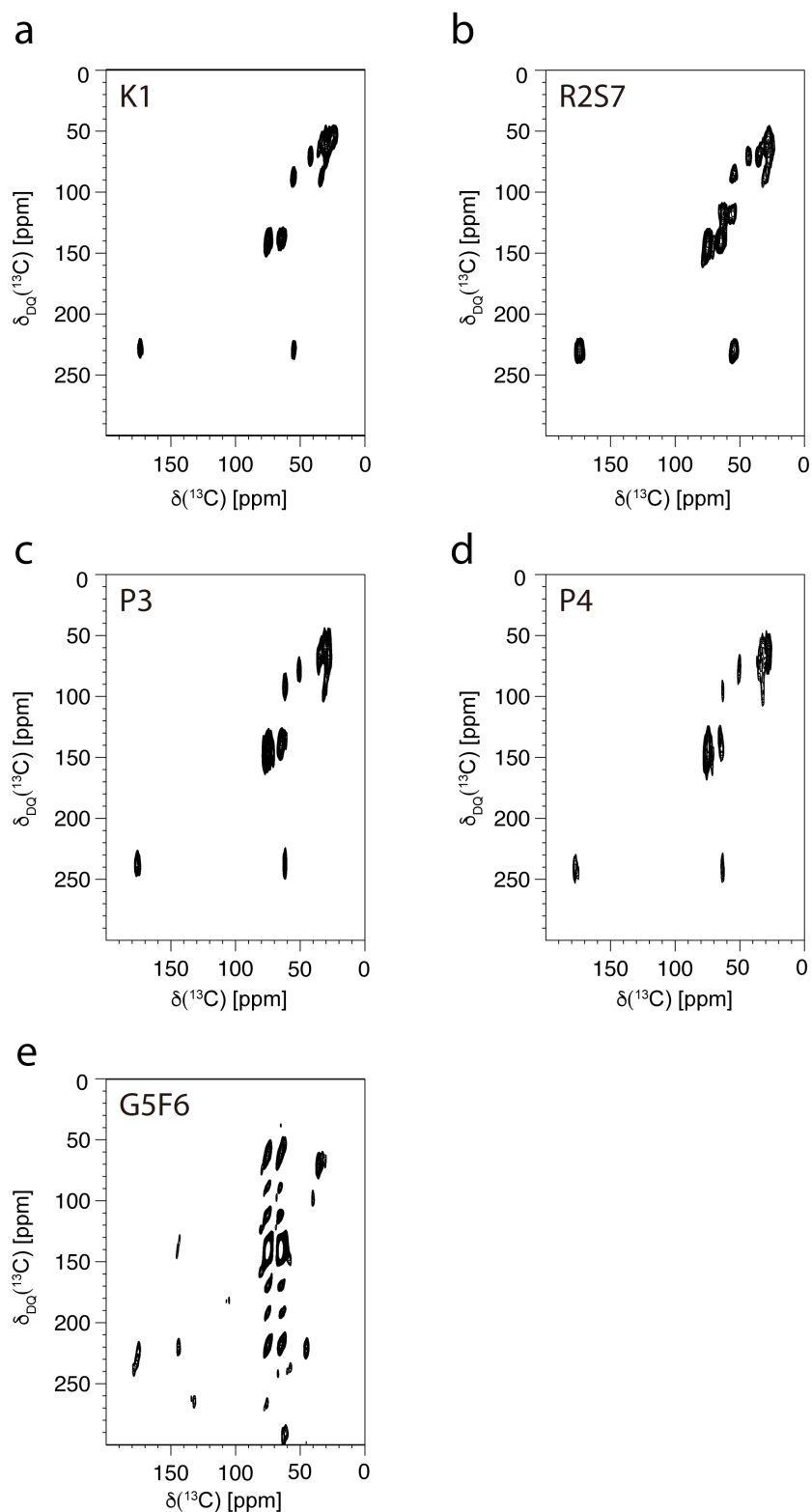
2. Supplementary Figures



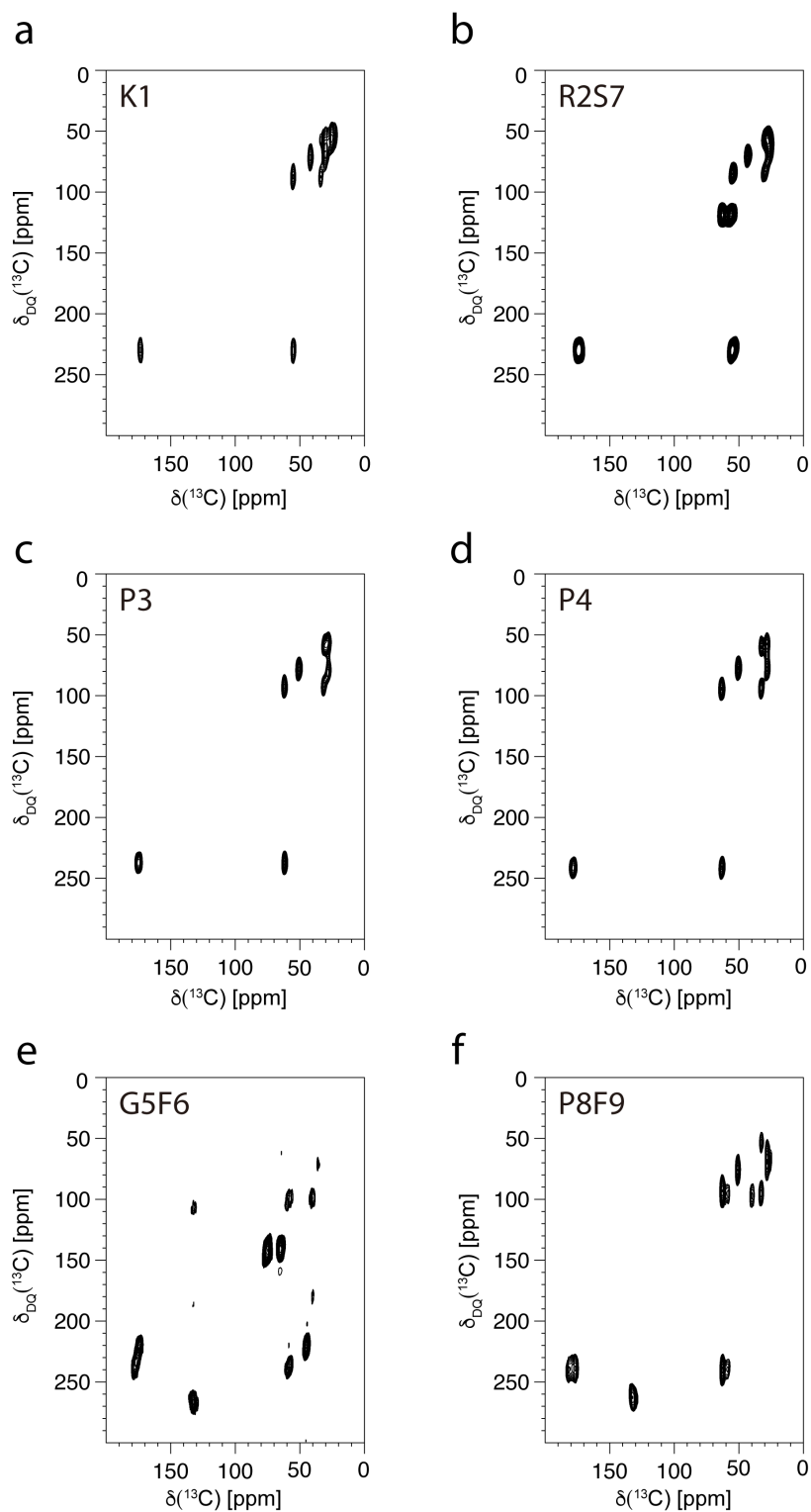
Supplementary Figure 1: DAKD binds with high affinity to detergent-solubilized B₁R (a) and total DAKD binding is not affected by the presence of the bi-nitroxide radical AMUPol (b). (a) Binding of DAKD to the B₁R solubilized in a mixed micelle of DDM and CHS can be saturated and takes place with high affinity (0.35 ± 0.08 nM), indicating that the receptor remains functional after solubilization from the membrane. (b) Total binding of DAKD to B₁R is not influenced by the presence of AMUPol confirming that radical addition does not disrupt the receptor-ligand complex before freezing the sample in the spectrometer. For both assays, B₁R was solubilized for 3 h in 1% DDM, 0.1% CHS in 50 mM HEPES-NaOH pH 7.6, 150 mM NaCl, 5% glycerol. Non-solubilized material was pelleted by ultracentrifugation. The receptor was immobilized on StrepTactin (StrepTactin SuperFlow Plus, Qiagen) beads and incubated with increasing concentrations of radiolabeled DAKD (Perkin Elmer) to determine the dissociation constant or with 10 nM radiolabeled DAKD in the presence of 100 mM AMUPol to assess the influence of the radical on total binding. Non-specific binding was assessed in the presence of 10 μ M unlabeled DAKD. Bound from unbound ligand was separated by transferring StrepTactin-immobilized receptors onto GF/B filters and washing with 50 mM HEPES-NaOH pH 7.6, 150 mM NaCl, 5% glycerol, 0.05% DDM, 0.005% CHS. Remaining radioactivity was measured by liquid scintillation counting. The binding assays were carried out in triplicate (n=3).



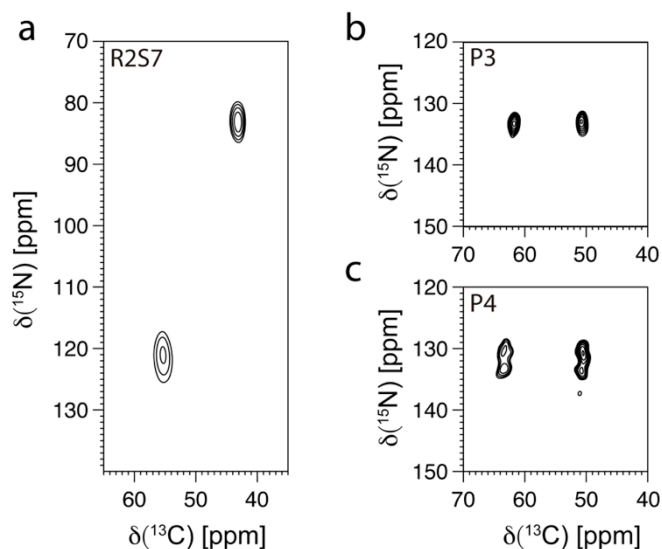
Supplementary Figure 2: Subsections of ^{15}N - ^{13}C TEDOR spectra of differently labeled DAKD bound to B_1R . (a) $\text{U-}[^{13}\text{C}, ^{15}\text{N}]\text{-R}_2\text{S}_7$ DAKD, (b) $\text{U-}[^{13}\text{C}, ^{15}\text{N}]\text{-P}_3$ DAKD, (c) $\text{U-}[^{13}\text{C}, ^{15}\text{N}]\text{-P}_4$ DAKD. Further subsections of $\text{U-}[^{13}\text{C}, ^{15}\text{N}]\text{-R}_2\text{S}_7$ DAKD are shown in Supplementary Figure 7. Panel (d) presents CAN 1D ^{15}N spectra of $\text{U-}[^{13}\text{C}, ^{15}\text{N}]\text{-G}_5\text{F}_6$ DAKD bound to B_1R . Panel (e) shows NCA spectra of this peptide without receptor. The ^{15}N chemical shifts of G5 and F6 can be easily assigned based on their characteristic values and high similarity to the values as indicated by the dashed horizontal lines.



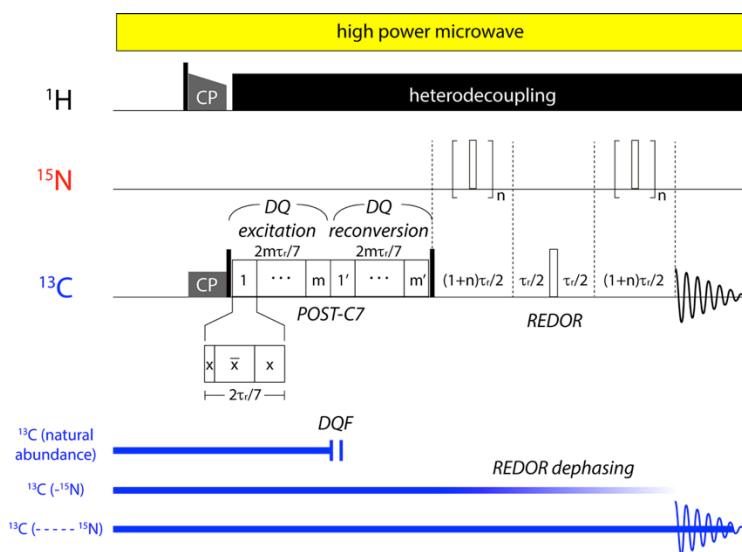
Supplementary Figure 3: DQ-SQ ^{13}C - ^{13}C 2D spectra of DAKD bound to B_1R . (a) U- ^{13}C , ^{15}N -K₁ DAKD, (b) U- ^{13}C , ^{15}N -R₂S₇ DAKD, (c) U- ^{13}C , ^{15}N -P₃ DAKD, (d) U- ^{13}C , ^{15}N -P₄ DAKD, (e) U- ^{13}C , ^{15}N -G₅F₆ DAKD. The “ridge” in spectrum (e) is caused by the natural abundance ^{13}C - ^{13}C pair in glycerol, which was significantly reduced by the use of ^{13}C -depleted glycerol in later measurements on other samples.



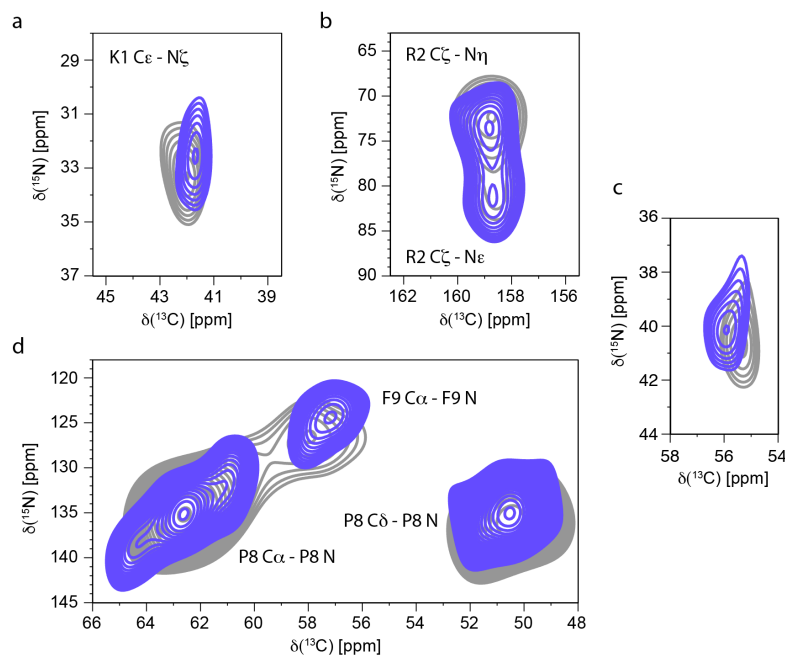
Supplementary Figure 4: DQ-SQ ^{13}C - ^{13}C 2D spectra of DAHD without B₁R. (a) U- ^{13}C , ^{15}N]-K₁ DAHD, (b) U- ^{13}C , ^{15}N]-R₂S₇ DAHD, (c) U- ^{13}C , ^{15}N]-P₃ DAHD, (d) U- ^{13}C , ^{15}N]-P₄ DAHD, (e) U- ^{13}C , ^{15}N]-G₅F₆ DAHD, (f) U- ^{13}C , ^{15}N]-P₈F₉ DAHD.



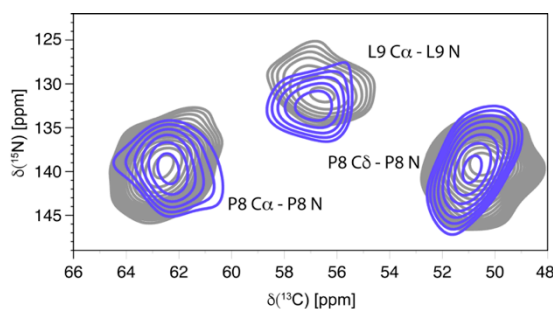
Supplementary Figure 5: ^{15}N - ^{13}C 2D TEDOR spectra of DAKD peptide without receptor. The peptides were labeled using different labeling schemes (Supplementary Table 1). (a) U- ^{13}C , ^{15}N -R₂S₇ DAKD, (b) U- ^{13}C , ^{15}N -P₃ DAKD, (d) U- ^{13}C , ^{15}N -P₄ DAKD.



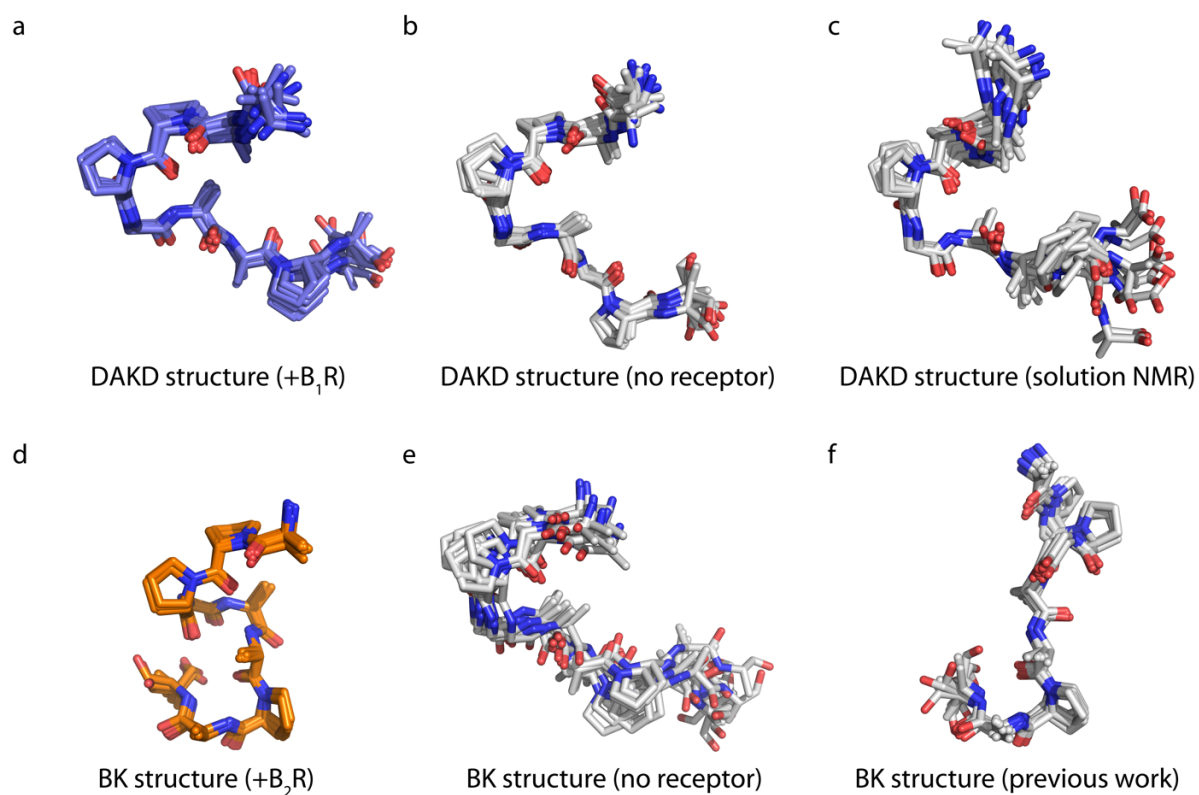
Supplementary Figure 6: DQ and REDOR doubly-filtered ^{13}C 1D pulse sequence used for acquiring the spectrum in Fig. 2f. The double-quantum-filter (DQF) removes efficiently natural abundance ^{13}C signals. Subsequently, ^{13}C magnetization of spins experiencing large ^{13}C - ^{15}N dipolar couplings (e.g. from ^{13}C - ^{15}N single bond distances) get dephased during a rotational-echo double-resonance spectroscopy (REDOR) recoupling period. The remaining signals mainly come from the ^{13}C labeled sites, which are further away from ^{15}N spins (i.e. those which are not directly bonded).



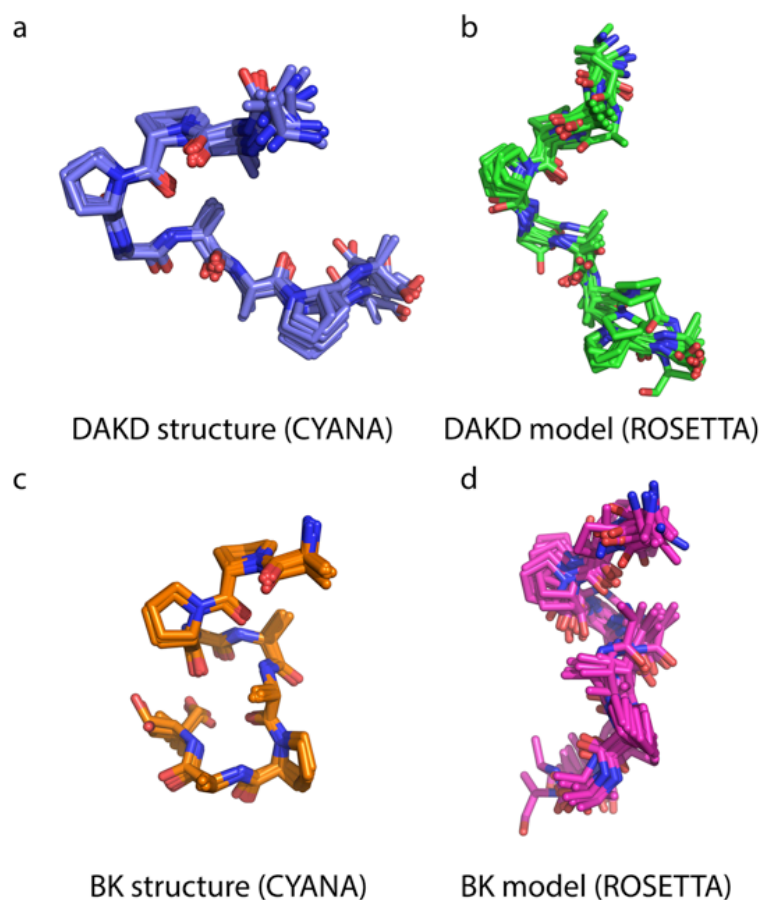
Supplementary Figure 7: Comparison of ^{15}N - ^{13}C TEDOR 2D spectra of DAKD free (grey) and bound (blue) to B_1R . N- and C-terminal chemical shift changes: Lys and Arg side chain regions of ^{15}N - ^{13}C TEDOR spectra of U- ^{13}C , ^{15}N -K₁ DAKD (a) and U- ^{13}C , ^{15}N -R₂S₇ DAKD (b). Chemical shift perturbations of the C ϵ -N ζ cross-peak of K1^{DAKD} and the C ζ -N ϵ cross-peak of R2^{DAKD} indicate the involvement of these side chains in binding. (c) Comparison of ^{15}N - ^{13}C TEDOR 2D spectra of U- ^{13}C , ^{15}N -K₁ DAKD free and bound to B_1R . Small chemical shift changes on both the ^{13}C and ^{15}N dimensions are observed for amine ^{15}N / $^{13}\text{C}\alpha$ cross peak of K1^{DAKD} upon binding to B_1R . (d) The ^{15}N - ^{13}C TEDOR spectrum of U- ^{13}C , ^{15}N -P₈F₉ DAKD reveals a chemical shift change for the C α -N cross-peak of F9^{DAKD} indicating a backbone conformational change upon binding.



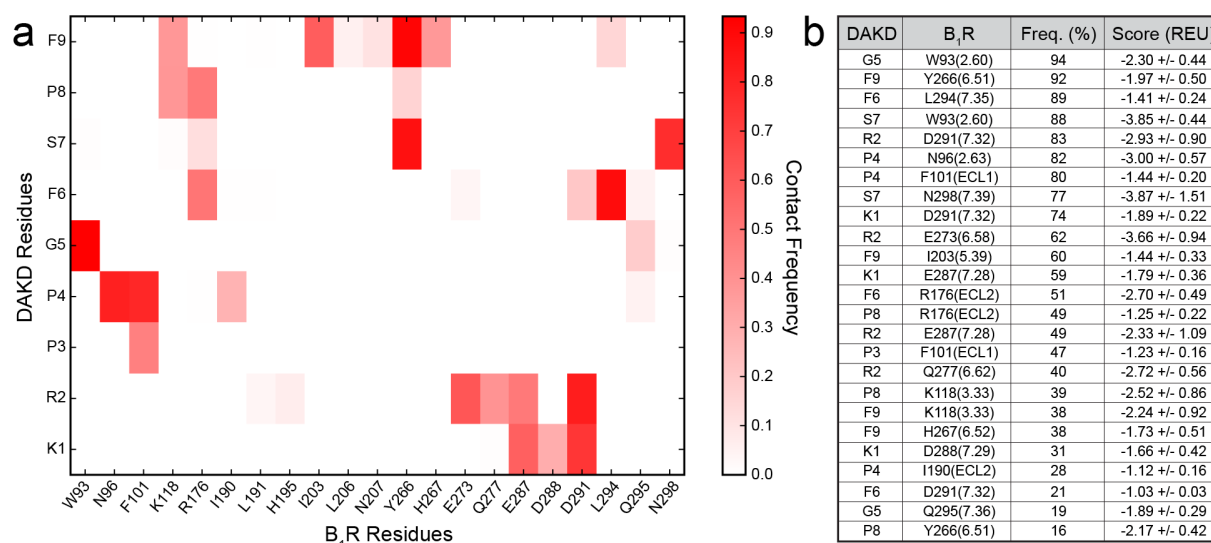
Supplementary Figure 8: ^{15}N - ^{13}C TEDOR 2D spectra show significant backbone chemical shift changes of antagonist peptide U- ^{13}C , ^{15}N -P₈L₉ DALK upon binding to human B_1R . The spectrum of DALK bound to B_1R is shown in blue and the spectrum of DALK without receptor is shown in grey.



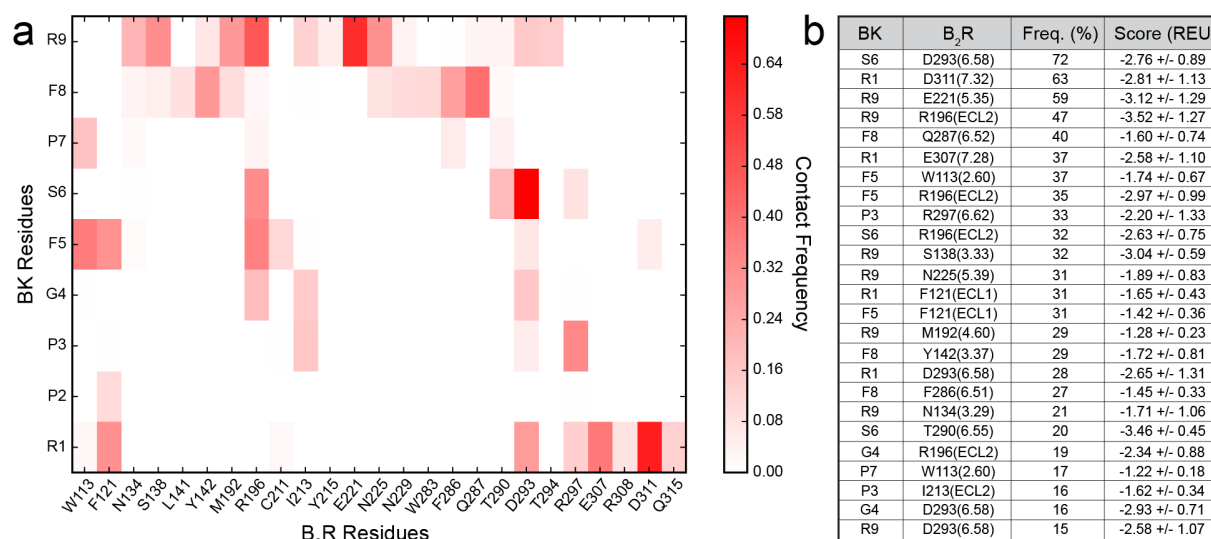
Supplementary Figure 9: Comparison of DAKD and BK NMR structures. DAKD bound to B₁R (a) and in frozen solution as determined from chemical shift by DNP-enhanced MAS-NMR (b). DAKD in solution determined by liquid-state NMR from chemical shifts and distance restraints (c). BK bound to B₂R re-calculated from TALOS+ from previously published chemical shifts²⁶ (d) and without receptor in frozen solution (e). Previously published structure of receptor-bound BK (f). Only backbone and C β atoms are shown. Each bundle contains 10 structures/models. The C-terminal residues of DAKD experience a subtle structural rearrangement upon binding to B₁R. In contrast, the C-terminal part of BK shows large conformational changes between receptor-bound and free states. The structure refinement statistics is summarized in Supplementary Tables 6-8.



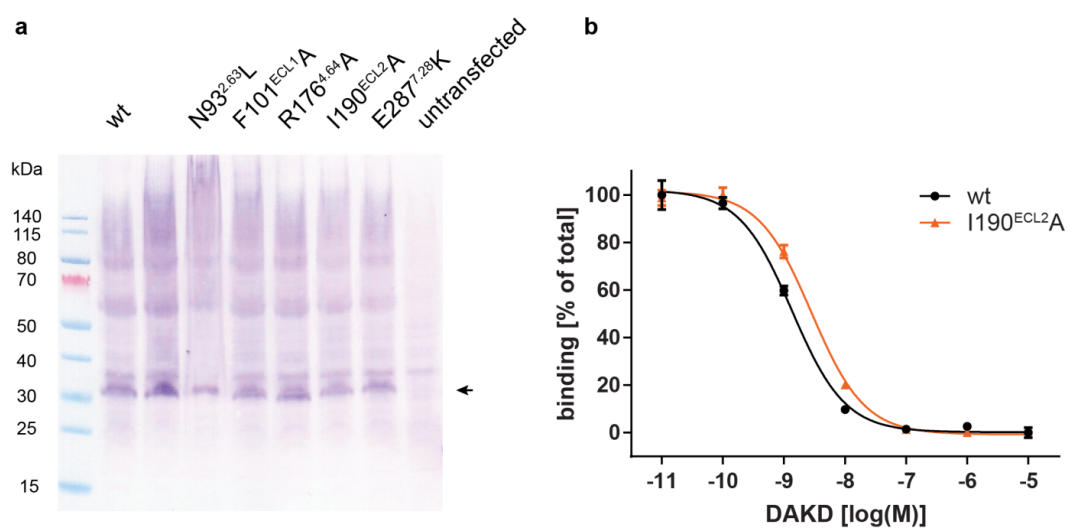
Supplementary Figure 10: Comparison of NMR structures (a,c) and Rosetta models (b,d) of DAKD (a,b) and BK (c,d) bound to human B₁R and human B₂R respectively. Only backbone part and C β atoms of are shown. Each bundle contains 10 structures/models. The characteristic V-shape folding of DAKD (a) and S-shape folding of BK (c) were reproduced by Rosetta modeling (b,d). The backbone heavy atom RMSD between the final set of 10 Rosetta models and the NMR structure of BK is 2.1 Å on average. In case of DAKD, the average backbone RMSD between the Rosetta created ensemble and the NMR structural model obtained from the forward-predicted cluster A is 2.2 Å. The RMSD between the Rosetta created ensemble and the structure obtained by back-calculating from TALOS+ torsion angle restraints is 2.3 Å. Thus, the Rosetta models clearly confirm the overall structural motifs of both peptide ligands as inferred from the NMR chemical shift data. Some small torsion angle differences were observed between NMR and Rosetta models, which can be attributed to the receptor background in which the Rosetta modeling was performed. For DAKD, torsion angle differences for the first two N-terminal residues were observed, which could arise from some structural variability of ECL3 and thus different ligand interaction modes in our B₁R models. In case of BK, different values for the ϕ angle of F8^{BK} and ψ angle of P7^{BK} were obtained, reflecting a slightly more extended conformation of the C-terminal tail. Our B₂R binding model shows interactions of BK with TMH3 at the lower end of the binding pocket, which requires the C-terminus to stretch, leading to the described structural differences.



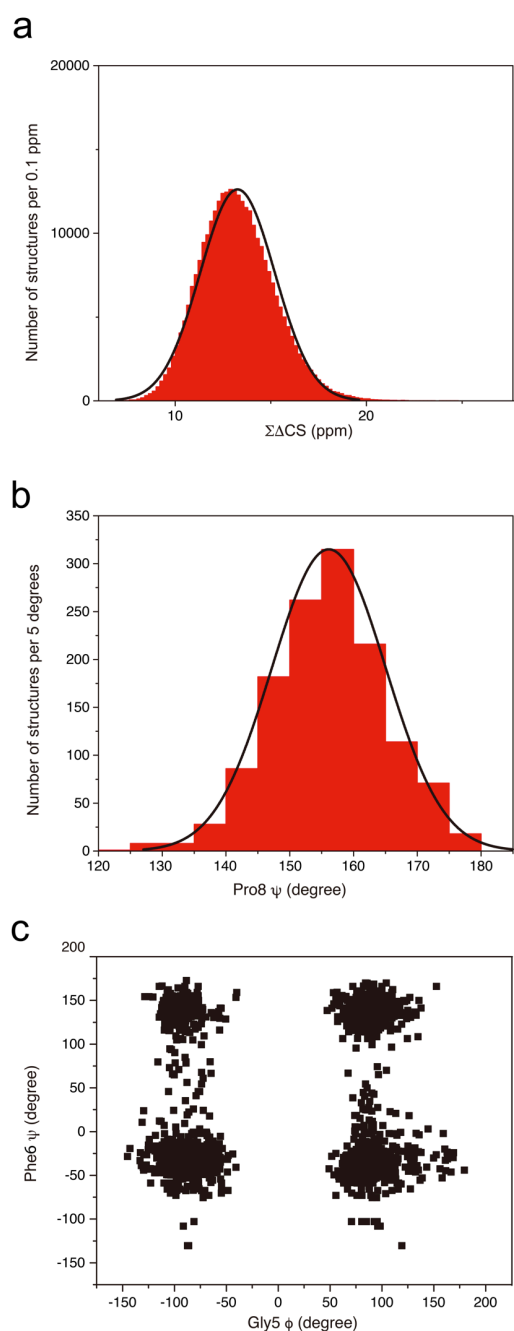
Supplementary Figure 11: Analysis of the B₁R-DAKD receptor-peptide interface. (a) Heatmap displaying the frequency of DAKD-B₁R contacts with a Rosetta score better than -1.0 REU as observed across the 1000 top docking models. (b) Summary of the 25 most frequent residue pairs in DAKD-B₁R models.



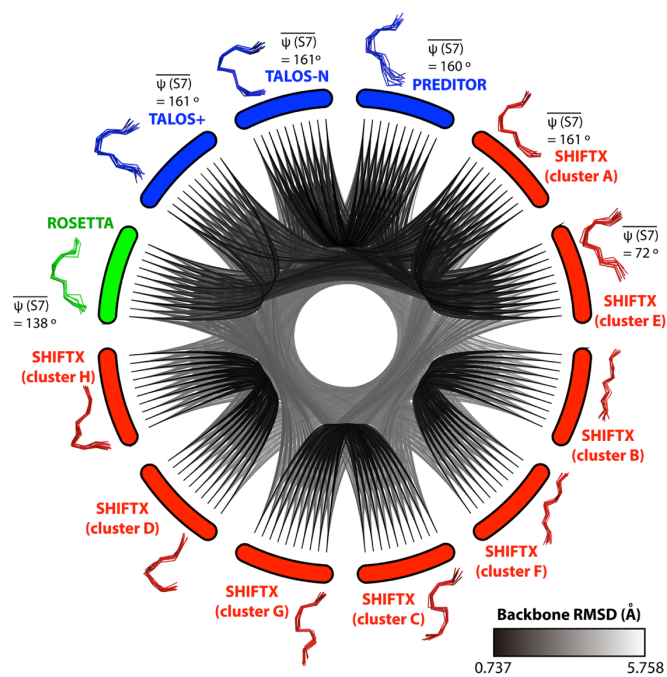
Supplementary Figure 12: Analysis of the B₂R-BK receptor-peptide interface. (a) Heatmap displaying the frequency of BK-B₂R contacts with a Rosetta score better than -1.0 REU as observed across the 1000 top docking models. (b) Summary of the 25 most frequent residue pairs in BK-B₂R models. The N-terminal residue R1^{BK} is close to D293^{6.58}, E307^{7.28} and D311^{7.32} (Figures 5b,d). Its positively charged amine-terminus and side chain make multiple interactions with these residues. We found the R1^{BK} side chain not in a single position but instead to sample several orientations and make electrostatic interactions with D293^{6.58} and D311^{7.32} as well as H-bond contacts with Q315^{7.36}. At its C-terminal end, BK is fixed by an H-bond between the R9^{BK} carboxyl group and residue S138^{3.33}, which is structurally similar to the DAKD-B₁R complex where F9^{DAKD} interacts with K118^{3.33} (Figures 5b,f). The side chain of R9^{BK} was found to participate in frequent interactions with E221^{5.35} and N225^{5.39} at the extracellular end of TMH V (Figure 5f). The BK C-terminus is further stabilized by B₂R residue F286^{6.51}, which has a strong effect on ligand binding²⁷. We observed this residue to pack against P7^{BK} and F8^{BK} and, together with L141^{3.36}, Y142^{3.37} and T290^{6.56}, form a hydrophobic environment for F8^{BK} (Figure 5f). A contribution of Y142^{3.37} and T290^{6.56} to ligand binding can be inferred from the available mutational data, too. Furthermore, our B₂R binding model predicts several interactions that have not been described before, such as hydrophobic contacts between F5^{BK} and the aromatic side chains of W113^{2.60} and F121 in ECL1 as well as several H-bonds between the central portion of the BK backbone and R196 in ECL2. Interestingly, we have identified and experimentally validated the corresponding residues in B₁R (W93^{2.60}, F101 and R176) as important DAKD interactions sites, which suggests that this group of conserved receptor residues is important for binding the middle region of both peptide ligands. For example, R176 in B₁R and R196 in B₂R, play a similar role as H-bond donor. However, due to its more extended, open conformation DAKD forms a dense H-bond network with R176 and additional receptor residues, which is not observed to this high extent for BK.



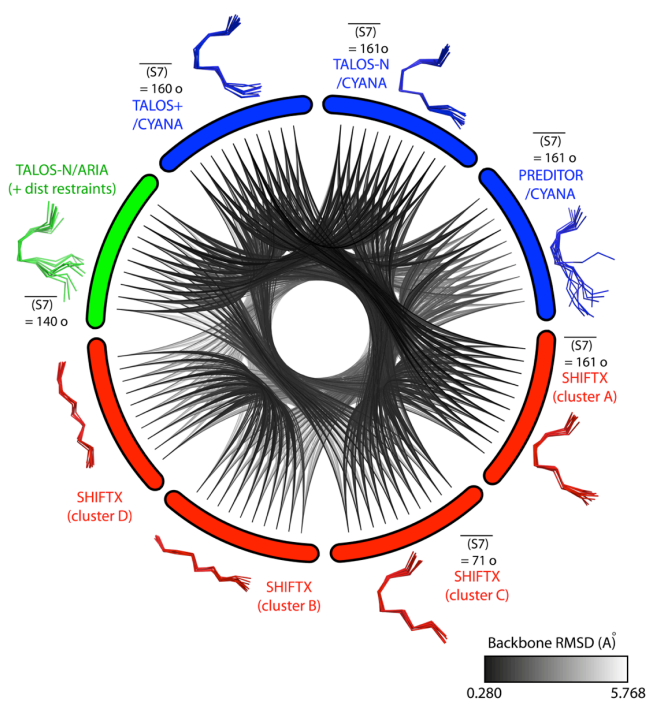
Supplementary Figure 13: Expression of B₁R mutants (a) and competition binding experiments of wild type and I190^{ECL2}A B₁R (b). The expression level of B₁R mutants was assessed by Western blot using an M2 FLAG antibody against the N-terminal FLAG tag. All mutants express comparably to wild type (wt). The lane between wt and N93^{2,63}L contains an unrelated sample. (b) Competition binding analysis of wild type and I190^{ECL2}A B₁R expressed in HEK cells. Replacing I190^{ECL2} with an alanine residue results in a slight decrease of the IC₅₀ of DAKD (I190^{ECL2}A 2.8 ± 0.2 nM, wild type (wt) 1.4 ± 0.7 nM; n=2). See Figure 4b for comparative binding experiments.



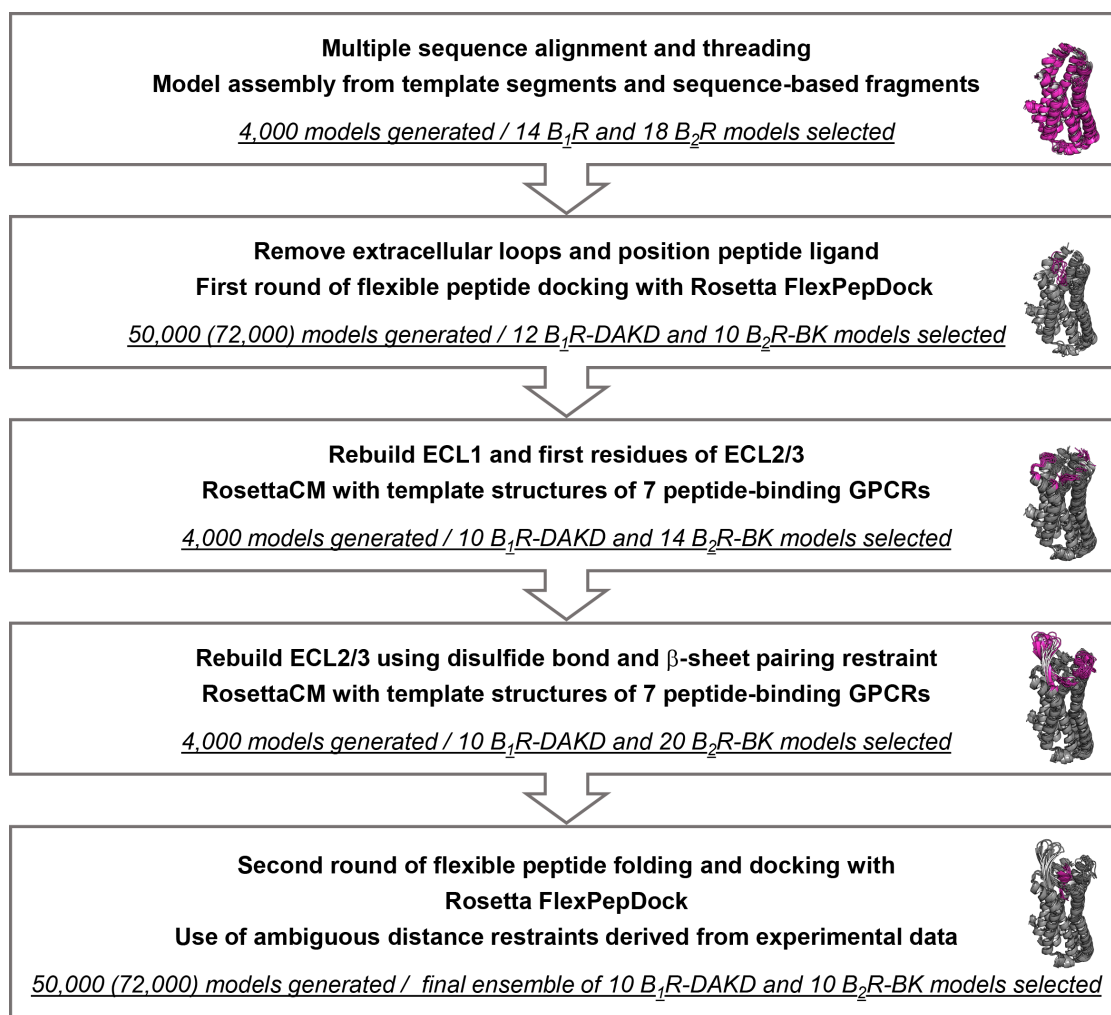
Supplementary Figure 15: Characteristics of Flexible-Meccano/SHIFTX-based forward chemical shift fitting approach on B₁R-bound DAKD. (a) Overall Gaussian-like distribution of conformations along $\Sigma\Delta\text{CS}$ in Flexible-Meccano/SHIFTX approach. (b) An example showing the Gaussian-like torsion angle distribution in the top 1490 structures selected by the parameter *cutoff* (see online methods). (c) An example showing independent torsion angle ambiguity in torsion angle restraints obtained from top 1490 structures in the Flexible Meccano/SHIFTX-based forward fitting.



Supplementary Figure 16: RMSD analysis of DAKD structures/models generated using different approaches. The string plot is generated using CIRCOS-0.69.3²⁸.

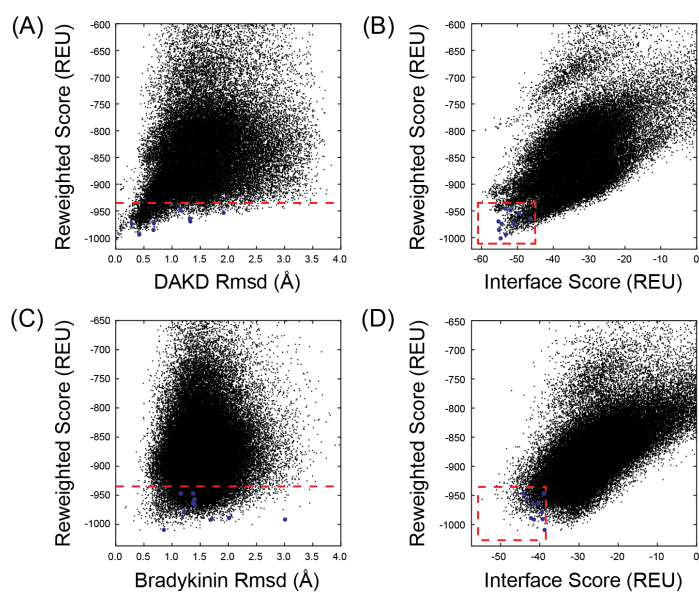


Supplementary Figure 17: RMSD analysis of non-bound DAKD structures/models generated using different approaches. The string plot is generated using CIRCOS-0.69.3²⁸.

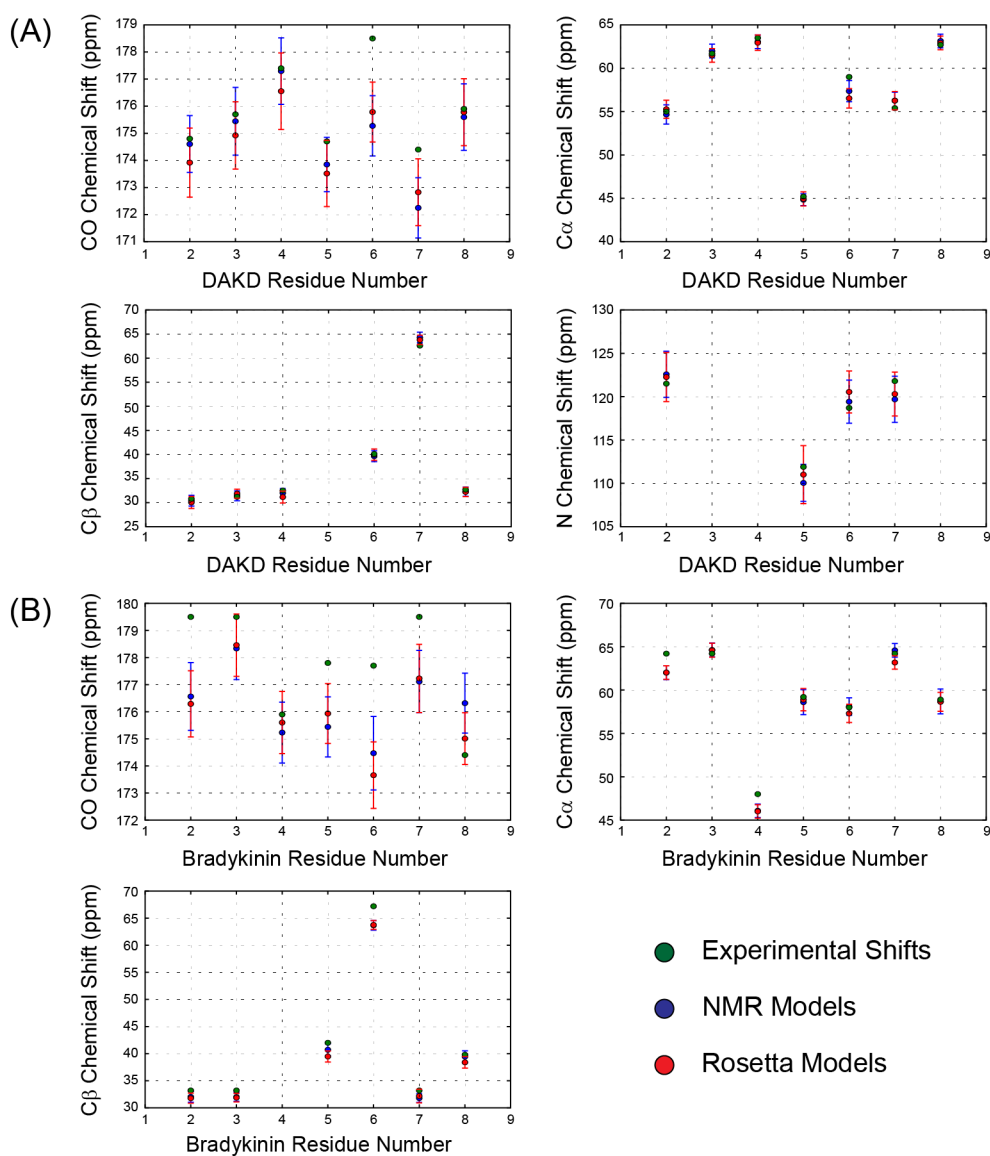


Supplementary Figure 18: Workflow of B₁R-DAKD and B₂R-bradykinin modeling and docking.

Ensembles of B₁R and B₂R models in complex with their endogenous ligands DAKD and bradykinin were constructed by several rounds of comparative modeling, loop building and ligand docking. Comparative modeling of the transmembrane helix region was performed using 24 GPCR template structures, whereas for loop modeling only seven peptide-binding GPCRs with higher sequence similarity to B₁R and B₂R were used as templates. Peptide docking was guided by loose distance restraints derived from mutational data previously reported for B₁R and B₂R. Representative models for each modeling stage are depicted in the flowchart. For selecting the final cluster, a chemical shift filtering step was applied (see online methods).



Supplementary Figure 19: Rosetta reweighted score vs. peptide backbone RMSD and interface score plots for docking of DAKD to B₁R (A and B) and bradykinin to B₂R (C and D). The ten largest clusters (shown as blue circles) were created from the best 1,000 models by reweighted and interface score (area within the red rectangle). The reweighted score combine the Rosetta total, peptide and interface energy.



Supplementary Figure 20: Comparison of experimental and SPARTA+ back-calculated chemical shifts for NMR and Rosetta structural models of (A) DAKD and (B) bradykinin. Calculated chemical shifts represent the average of the final NMR structural models, as obtained by the forward prediction method (DAKD, cluster A) or by the TALOS+ method (BK) and the 10 final Rosetta models. Error bars depict the prediction uncertainty of SPARTA+. Chemical shift values of the first and last residue were not calculated because the prediction requires torsion angle information from two direct neighbor residues. The SPARTA+ calculated chemical shifts for the presented Rosetta ensemble of DAKD and BK gave an average RMSD of 1.33 ± 0.07 ppm and 1.99 ± 0.10 ppm, respectively, which compares to 1.34 ± 0.14 ppm and 1.82 ± 0.06 ppm for the respective NMR models. Using SHIFTX2, a comparable good agreement was obtained and RMSD values were 1.56 ± 0.10 ppm and 1.98 ± 0.06 ppm for Rosetta models and 1.65 ± 0.15 ppm and 1.89 ± 0.06 ppm for NMR models of DAKD and BK, respectively.

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