

BMJ Open

BMJ Open is committed to open peer review. As part of this commitment we make the peer review history of every article we publish publicly available.

When an article is published we post the peer reviewers' comments and the authors' responses online. We also post the versions of the paper that were used during peer review. These are the versions that the peer review comments apply to.

The versions of the paper that follow are the versions that were submitted during the peer review process. They are not the versions of record or the final published versions. They should not be cited or distributed as the published version of this manuscript.

BMJ Open is an open access journal and the full, final, typeset and author-corrected version of record of the manuscript is available on our site with no access controls, subscription charges or pay-per-view fees (<http://bmjopen.bmj.com>).

If you have any questions on BMJ Open's open peer review process please email info.bmjopen@bmj.com

BMJ Open

The MUKnine OPTIMUM protocol: A screening study to identify high risk multiple myeloma patients suitable for novel treatment approaches combined with a phase II study evaluating optimised combination of biological therapy in newly diagnosed high risk multiple myeloma and plasma cell leukaemia

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2020-046225
Article Type:	Protocol
Date Submitted by the Author:	27-Oct-2020
Complete List of Authors:	Brown, Sarah; University of Leeds, Clinical Trials Research Unit Sherratt, Debbie; University of Leeds, Clinical Trials Research Unit Hinsley, Samantha; University of Leeds Clinical Trials Research Unit Flanagan, Louise; University of Leeds Clinical Trials Research Unit Roberts, Sadie; University of Leeds Clinical Trials Research Unit Walker, Katrina; University of Leeds Clinical Trials Research Unit Hall, Andrew; University of Leeds Clinical Trials Research Unit Pratt, Guy; Queen Elizabeth Hospital Messiou, Christina; Institute of Cancer Research Sutton Jenner, Matthew ; Southampton General Hospital Kaiser, Martin; Institute of Cancer Research,
Keywords:	Myeloma < HAEMATOLOGY, Clinical trials < THERAPEUTICS, STATISTICS & RESEARCH METHODS

SCHOLARONE™
Manuscripts



I, the Submitting Author has the right to grant and does grant on behalf of all authors of the Work (as defined in the below author licence), an exclusive licence and/or a non-exclusive licence for contributions from authors who are: i) UK Crown employees; ii) where BMJ has agreed a CC-BY licence shall apply, and/or iii) in accordance with the terms applicable for US Federal Government officers or employees acting as part of their official duties; on a worldwide, perpetual, irrevocable, royalty-free basis to BMJ Publishing Group Ltd ("BMJ") its licensees and where the relevant Journal is co-owned by BMJ to the co-owners of the Journal, to publish the Work in this journal and any other BMJ products and to exploit all rights, as set out in our [licence](#).

The Submitting Author accepts and understands that any supply made under these terms is made by BMJ to the Submitting Author unless you are acting as an employee on behalf of your employer or a postgraduate student of an affiliated institution which is paying any applicable article publishing charge ("APC") for Open Access articles. Where the Submitting Author wishes to make the Work available on an Open Access basis (and intends to pay the relevant APC), the terms of reuse of such Open Access shall be governed by a Creative Commons licence – details of these licences and which [Creative Commons](#) licence will apply to this Work are set out in our licence referred to above.

Other than as permitted in any relevant BMJ Author's Self Archiving Policies, I confirm this Work has not been accepted for publication elsewhere, is not being considered for publication elsewhere and does not duplicate material already published. I confirm all authors consent to publication of this Work and authorise the granting of this licence.

1
2
3
4 The MUKnine OPTIMUM protocol: A screening study to identify high risk multiple myeloma
5 patients suitable for novel treatment approaches combined with a phase II study evaluating
6 optimised combination of biological therapy in newly diagnosed high risk multiple myeloma
7 and plasma cell leukaemia.
8
9

10 Sarah Brown^{1*}, Debbie Sherratt^{1*}, Samantha Hinsley¹, Louise Flanagan¹, Sadie Reed¹, Katrina
11 Walker¹, Andrew Hall¹, Guy Pratt⁴, Christina Messiou³, Matthew Jenner^{2**} and Martin Kaiser^{3**},
12 on behalf of the Myeloma UK Early Phase Clinical Trial Network
13
14

15 Corresponding authors:

16 Sarah Brown and Martin Kaiser
17
18
19
20

21 Author Details

22
23 1 Clinical Trials Research Unit, Leeds Institute of Clinical Trials Research, University of
24 Leeds, Leeds UK. Emails: d.sherratt@leeds.ac.uk; Samantha.Hinsley@glasgow.ac.uk;
25 I.m.flanagan@leeds.ac.uk; S.N.Roberts@leeds.ac.uk; K.M.Walker@leeds.ac.uk;
26 A.Hall2@leeds.ac.uk; s.brown@leeds.ac.uk
27

28 2 Southampton General Hospital, Southampton UK. Email: matthew.jenner@uhs.nhs.uk
29

30 3 Institute of Cancer Research, London UK. Email: Martin.kaiser@icr.ac.uk
31

32 4 Queen Elizabeth Hospital, Birmingham UK. Email: guy.pratt@uhb.nhs.uk
33

34 *Sarah Brown & Debbie Sherrat contributed equally as joint first authors

35 **Matthew Jenner & Martin Kaiser contributed equally as joint last authors
36
37
38

39 **Word count: 3993**
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract

Introduction: Multiple myeloma (MM) is a plasma cell tumour with over 5800 new cases each year in the UK. The introduction of biological therapies has improved outcomes for the majority of MM patients, but in approximately 20% of patients the tumour is characterised by genetic changes which confer a significantly poorer prognosis, generally termed as high risk (HR) MM. It is important to diagnose these genetic changes early and identify more effective first-line treatment options for these patients.

Methods and analysis: The Myeloma UK *nine* OPTIMUM trial (MUK*nine*) is designed to evaluate novel treatment strategies for patients with HRMM. Patients with suspected or newly diagnosed MM, fit for intensive therapy, are offered participation in a central tumour genetic screening protocol (MUK*nine a*). Patients identified as molecularly HR are invited into the phase II, single-arm, multi-centre trial (MUK*nine b*) investigating an intensive treatment schedule comprising bortezomib (Velcade), lenalidomide (Revlimid), daratumumab (Darzalex), low-dose cyclophosphamide & dexamethasone, with single high dose melphalan and autologous stem cell transplantation followed by combination consolidation and maintenance therapy. The trial uses a Bayesian decision rule to determine if this treatment strategy is sufficiently active for further study.

Patients identified as not having HR disease will receive standard local treatment and followed up in a cohort study.

Exploratory studies include longitudinal whole body diffusion weighted MRI for imaging Minimal Residual Disease (MRD) testing.

Ethics and dissemination: Ethics approval obtained. Results of study and substudies will be submitted for publication in a peer-reviewed journal.

Trial Registration: ISRCTN16847817, May 2017

Key Words: Newly diagnosed multiple myeloma, daratumumab, phase II, Bayesian, minimal residual disease

Article summary

Strengths and limitations of the study

- This is the first time in the UK that newly diagnosed multiple myeloma patients may be entered into a clinical trial prospectively according to their genetic risk profile
- A flexible multiple outcome, multi-stage Bayesian design is used to enable early stopping for lack of efficacy
- No concurrent control arm is included due to the availability of near concurrent historical control data from the Myeloma XI trial

For peer review only

Introduction

Multiple myeloma (MM) is a clonal disorder of plasma cells which accumulate in the bone marrow leading to cytopenias, bone resorption, renal impairment, infection and the production of a monoclonal protein¹. MM represents 1.5% of all malignant diseases, with an incidence of 9/100,000 per year accounting for around 5800 new cases each year in the UK (3000 deaths per year)². Median age at diagnosis is 69 years but 37% of patients are diagnosed before the age of 65 (including 15% <55)³. Median overall survival (OS) of younger patients is approximately 10 years⁴⁻¹¹. However approximately 20% of patients have a significantly worse prognosis, with estimated survival of <3 years and are characterised as having high risk (HR) disease^{7 12 13}. A number of genetic lesions and gene expression profiles (GEP) have been identified as associated with HR disease⁷, and molecular risk models based on these markers can be used to predict HR disease in a clinical setting. Further research is ongoing to identify additional HR markers and to better understand the mechanisms driving this tumour biology.

Unfortunately, patients with HR disease have, in terms of absolute outcome, benefitted less from the introduction of novel therapies than standard risk (SR) patients¹⁴⁻²⁰. It is important to define the optimal way to treat this group of patients given the number of available novel agents with favourable toxicity profiles allowing the use of combination therapy, consolidation and maintenance therapy. Here we describe the protocol for the MUKnine trial, a phase II study evaluating optimised combination of biological therapy in patients with newly diagnosed HRMM and plasma cell leukaemia (PCL), incorporating a screening and observational study for patients with standard risk disease. The trial has completed recruitment and is currently in follow-up.

Defining high risk disease

In a recent meta-analysis of 1,905 trial patients from the MRC Myeloma IX and NCRI Myeloma XI trials, recurrent chromosomal translocations t(4;14), t(14;16), t(14;20) and copy number aberrations (CNA) gain(1q) or del(17p) were independently associated with shorter PFS and OS. Presence of two or more such HR lesions, also termed double-hit⁷, was associated with particularly adverse outcome and increased specificity of outcome prediction considering individual lesions in isolation. The co-segregation model is exclusively based on molecular features of the tumour cell and contrasts to risk predictors which require inclusion of clinical risk markers (renal function, age, performance status) or their proxies, such as the international staging system (ISS)¹². For participants fit to receive intensive therapy, HR can thus be specifically defined by presence of two or more cytogenetically adverse lesions [t(4;14), t(14;16), t(14;20), del(1p32) gain(1q) or del(17p)].

The prognostic relevance of GEP risk signatures, in particular EMC-92, from which the SKY92 MMProfiler diagnostic assay was developed, has been demonstrated in the Myeloma IX trial dataset²¹. A recent analysis including Myeloma IX and Myeloma XI trial patients demonstrated independent association of GEP SKY92 high risk and genetic HR markers with adverse outcome in MM^{11 13 21-24}. Results suggest that both tests assay different clinically relevant qualities of HR biology. Combining GEP and double-hit genetic risk information identifies about 20-30% of patients with markedly short PFS and OS.

The exact impact of single nucleotide variants on MM risk status is still under investigation. However, very recent evidence, published after design of MUKnine, seems to confirm that structural aberrations such as translocations and CNA are the dominant markers of HRMM,

1
2
3 although detail on their assessment varies²⁵⁻²⁷. The observation of poor prognosis associated
4 with HR disease defined by such molecular criteria is consistent with clinical studies carried
5 out by other trial groups^{5-11 21 22 24 28 29}. Clearly a focused approach to improve the treatment
6 and outcome of this poor performing subgroup of MM patients is essential.
7
8
9

10 Treatment

11
12 Recent data has demonstrated efficacy of the combination of multiple novel agents in HR
13 disease³⁰. Until the molecular mechanisms contributing to HR biology can be directly targeted,
14 combinations of multiple novel agents and ongoing therapy to induce and maintain remission
15 are the most efficacious therapeutic principles³¹.
16

17 Maximising exposure to novel agents as an alternative to multi-drug cytotoxic alkylating
18 chemotherapy is hypothesised to benefit HR patients. Ongoing use of a combination of
19 biologic agents with favourable toxicity profiles can potentially minimise the chance of relapse
20 due to sustained multi-angled pressure on the MM re-populating cell pool.
21

22
23 Long-term exposure to thalidomide does not benefit HR patients^{32 33}. However, lenalidomide
24 maintenance in newly diagnosed HR patients (t(4;14) or del 17p) does have a PFS and OS
25 benefit³⁴. There is a substantial body of evidence suggesting that HR patients benefit from
26 long-term exposure to proteasome inhibition such as bortezomib³⁵⁻³⁹.
27

28 The combination of bortezomib and lenalidomide as induction and consolidation therapy is
29 safe and deliverable with a number of studies using this approach⁴⁰. Adding
30 cyclophosphamide to this triplicate approach is safe, nevertheless the lenalidomide,
31 cyclophosphamide, bortezomib and dexamethasone (RCVD) combination failed to show any
32 additional benefit to RVD (lenalidomide, bortezomib and dexamethasone) in the EVOLUTION
33 study⁴¹. However, this study evaluated all genetic risk groups and it is hypothesised that the
34 addition of low dose alkylating therapy may present an additional benefit in a HR population
35 with highly proliferative subclones.
36
37

38 Daratumumab is a monoclonal antibody that targets the CD38 molecule and has multiple
39 mechanisms of action against MM cells. It has demonstrated activity in MM as a single agent
40 and in combinations with lenalidomide and dexamethasone where it enhances the potency of
41 other drugs such as lenalidomide offering an interesting alternative to chemotherapy in MM⁴².
42 The addition of daratumumab to standard of care regimens improved outcome and combining
43 with lenalidomide or bortezomib appears to improve the poor outcomes associated with HR
44 disease^{43 44}.
45
46

47 Whilst tandem ASCT may offer prolongation of response in comparison with single procedures
48 the comparative studies reported at time of design of MUK^{nine} were undertaken in an era in
49 which novel agents were not routinely incorporated in clinical practice⁴⁵. Recent exploratory
50 analyses have suggested the potential advantage of tandem ASCT for patients with high risk
51 disease⁴⁶. Depth of response is associated with duration of response and therefore optimising
52 the induction, consolidation and maintenance approach with a single ASCT is an alternative
53 way to achieve MRD negative disease state. Melphalan has been combined with bortezomib
54 in phase II studies demonstrating safety and improvement in complete response rates
55 compared with conventional high dose melphalan conditioning⁴⁷. Although a recent report
56 stated no PFS benefit of a Velcade-augmented ASCT in a randomised trial, results for an ultra-
57 high risk group such as double-hit MM are unknown⁴⁸. The highly proliferative behaviour of
58
59
60

double-hit disease and GEP high risk provides rationale for a bridging treatment for the three months recovery period post-ASCT.

Rapid tumour evolution and associated early relapse are key characteristics of HRMM, even in patients who have achieved deep remission after ASCT⁴⁹. Maintaining multi-agent treatment intensity around and long-term after ASCT to limit size of the clonal pool as well as molecular avenues for tumour escape seems currently one of the most promising treatment strategies for HRMM, with the aim of achieving sustained deep responses in at least some patients⁵⁰. Longitudinal minimal residual disease monitoring can predict remission status with higher sensitivity than standard biochemical/protein analyses and could be of use in identifying HRMM patients benefitting most from treatment early. As bone marrow biopsy based MRD assessment may be biased due to spatial disease heterogeneity, sensitive whole body imaging can be performed in parallel to capture residual disease in other bone marrow or soft tissue areas. Whole body diffusion weighted MRI is a particularly sensitive imaging modality for MM, and standardised image acquisition and interpretation guidelines make implementation in multi-centre clinical trials feasible^{51 52}.

In line with this, the MUKnine OPTIMUM trial has been designed to evaluate the following treatment regimen in patients with HRMM, the full schedule is given in Table 1:

- **CVRDd (induction) – Cyclophosphamide, bortezomib (Velcade), lenalidomide (Revlimid), daratumumab (Darzalex), dexamethasone**
Based on the EVOLUTION trial⁴¹. Daratumumab doses are used in ongoing clinical trials⁵³.
- **Melphalan - Bortezomib ASCT**
Melphalan 200 mg/m² is standard practice in Europe for salvage treatment ^{54 55}. The addition of bortezomib in phase II studies demonstrated safety and improvement in complete response (CR) rates compared with conventional high dose melphalan conditioning ⁴⁷. Velcade weekly monotherapy during the clinical recovery period from ASCT limits very early disease relapse in the HR population.
- **VRDd (consolidation 1) – Bortezomib, lenalidomide, daratumumab, dexamethasone**
Doses for VRd combination are based on IFM 2008-01⁴⁰ and IFM 2009-02/DFCI. Daratumumab doses are used in current clinical trials⁵³.
- **VRD (consolidation 2) – Bortezomib, lenalidomide, daratumumab**
The dose of VRD during consolidation 2 is used to minimise effects of long term corticosteroid use and risks of long term neuropathy with weekly bortezomib with no break in treatment. Utilising existing daratumumab dosing schedules it is anticipated this will be a tolerable longer term combination.
- **RD (maintenance) – Lenalidomide, daratumumab**
The dose of Lenalidomide is based on two pivotal studies^{34 56} and is the current dose used in the Myeloma XI trial²⁰. Daratumumab doses are used in current clinical trials⁵³.

Current protocols: MUKnine a, v2.0, 25/07/2018. MUKnine b, v4.0, 14/05/2020.

Methods and analysis

Aims

- To assess whether future trials in this setting are feasible and to determine risk status for participants with MM in order to deliver novel therapy to those deemed HR
- To determine whether it is possible to improve the outcome of HR patients by utilising multiple biological agents during induction, ASCT, consolidation and maintenance, and to provide evidence for the future evaluation of these high-cost interventions.

Primary Objectives

- Assess whether molecular risk-defining investigations can be turned around within 8 weeks
- Determine whether the combination of three novel agents bortezomib, lenalidomide & daratumumab in combination with low-dose cyclophosphamide & dexamethasone is sufficiently active in terms of PFS in a HR population to take forward to a phase III trial

Secondary Objectives

Secondary objectives include evaluating safety and toxicity profiles of trial treatment, evaluating additional measures of treatment activity and assessing quality of life. In patients not identified as having HR disease, secondary objectives are to summarise treatment pathways and clinical outcomes in this setting.

Exploratory Objectives

To explore novel molecular biomarkers associated with treatment activity, and evaluate germline variability/mutations, genomic instability and clonal evolution.

An exploratory imaging sub-study is included to explore the association of imaging MRD status with clinical outcomes and to assess patterns of disease distribution by whole body DW-MRI.

Trial design

The MUKnine OPTIMUM trial is comprised of two components, MUKnine *a* and MUKnine *b*, as outlined in Figure 1. MUKnine *a* is a genetic screening component, where patients with suspected symptomatic MM will be screened to determine their risk status. Patients identified as not having HR disease will receive treatment as standard of care and will have data collected on their treatment and survival. Patients who are identified as having HR disease or PCL are invited to take part in the second component, MUKnine *b*, a single arm phase II, multi-centre trial. MUKnine *b* incorporates interim assessments for futility using a Bayesian strategy for monitoring multiple outcomes proposed by Thall, Simon and Estey^{57 58} and extended by Thall and Sung⁵⁹. The trial is single arm to ensure a feasible sample size given the availability of molecularly matched individual participant data from currently running trials (Myeloma XI/XI+). This provides a body of almost concurrent control data available for the purpose of exploratory statistical comparison.

Diffusion-weighted whole body MRI (DW-MRI) is a functional method capable of detecting small-volume disease activity in MM^{60 61}, being used in standard practice at several academic

1
2
3 UK hospitals already, demonstrating excellent performance in guiding therapy on a day-to-
4 day basis. An exploratory sub-study is incorporated in MUK*nine* using DW-MRI for disease
5 distribution assessment and imaging MRD in combination with cellular (bone marrow) MRD.
6
7

8 9 Sample size

10 Recent data from the Myeloma XI trial demonstrate a median PFS for patients with HR disease
11 in the intensive pathway of 19.7 months (598 patients¹²). With a median PFS of 19-20 months
12 in the control arm, we require 92-94 patients to observe a 25% difference in median PFS
13 (corresponding to a difference of 4.8-5.0 months) in the 85% credible interval. Allowing for
14 slight changes in the actual count data, we require 95 HR patients to be registered.
15
16

17 A sample size re-estimation using individual patient data from Myeloma XI/XI+, when
18 available, allows the number of HR patients required to detect a 25% difference in median
19 PFS to be increased to 105. In order to include 105 HR patients, approximately 620 patients
20 with MM would need to be registered at diagnosis, assuming approximately 10-15% failed
21 diagnostic tests, and approximately 20% patients identified as HR.
22
23

24 The trial design includes interim analyses after every cohort of 10 MUK*nine b* participants
25 have been followed-up to 120 days post-ASCT. Until recruitment is complete, the trial could
26 be terminated early for futility on the basis of MRD status and PFS at 100 days post-ASCT.
27
28

29 Consent, eligibility, screening and registration

30
31 Participants are recruited from UK NHS hospitals. Hospital sites delivering the HR treatment
32 are approved sites within the Myeloma UK Early Phase Clinical Trials Network⁶² and patients
33 recruited from sites outside of the Network sites are referred to receive treatment. The imaging
34 sub-study is undertaken at select sites with appropriate radiology capacity. Assenting patients
35 will provide written informed consent and be registered.
36
37

38 Patients presenting who are likely to have symptomatic MM (identified by pre-tests performed
39 as standard) are approached prior to having a bone marrow biopsy for diagnosis or
40 confirmation of MM. A full list of inclusion and exclusion criteria are in Table 2.
41

42 Patients are provided with information about the trial and if agreeable are consented for the
43 bone marrow biopsy to allow samples to be sent to central laboratories and for screening. This
44 consent allows follow up data to be collected under the MUK*nine a* protocol if the patient is
45 found not to have HR disease. Patients are registered to the trial via a web-based system
46 (provided by University of Leeds) prior to any trial-specific assessments being conducted.
47 Participants can also optionally consent to the imaging sub-study. Participants retain the right
48 to withdraw at any time without giving reasons and without their further treatment being
49 prejudiced.
50
51

52 Bone marrow and blood samples are taken as per standard care and sent to the Institute of
53 Cancer Research, London (ICR) by next day postal delivery for genetic molecular risk profiling.
54

55 HR status is determined by the presence of one or more of the following, based on the
56 International Myeloma Working Group (IMWG) guidelines⁶³, the Myeloma IX trial and the
57 EMC92 GEP model^{3 5 10 21 64}:
58

- 59 • 2 or more adverse lesions [t(4;14), t(14;16), t(14;20), gain(1q), del(17p), del(1p)]
- 60 • GEP – high risk score as per EMC92/SKY92 GEP model

- PCL, defined as the presence of more than $2 \times 10^9/L$ peripheral blood plasma cells or a plasmacytosis accounting for >20% of the differential white cell count

Patients identified as having HR disease are provided with a patient information sheet detailing the HR treatment schedule in MUKnine b and consented if willing to participate. A further registration documents all patients going on to HR treatment. If the patient does not wish to receive HR treatment they continue with standard treatment and data collected through the MUKnine a protocol.

For all patients at screening, bone marrow samples are sent to Haematological Malignancy Diagnostic Service (HMDS), Leeds, for MRD monitoring. Blood and urine samples are sent to Clinical Immunology Service, University of Birmingham for disease response assessments. A cell-free DNA peripheral blood sample is sent to the ICR.

Interventions

Upon first consent, treatment with standard local treatment may commence for up to 2 cycles (up to 8 weeks) whilst central molecular risk profiling is performed. Treatment may be with cyclophosphamide, thalidomide, dexamethasone (CTD), cyclophosphamide, lenalidomide, dexamethasone (CRD), bortezomib, thalidomide, dexamethasone (VTD) or cyclophosphamide, bortezomib, dexamethasone (CVD) to further take part in the MUKnine trial. This allows participants to start treatment for MM while awaiting results from risk-defining genetic investigations.

MUKnine a: Participants not identified as having HR disease continue to receive standard treatment or treatment as directed by their clinician and are followed up regularly, with information on their treatment pathway and outcomes collected.

MUKnine b: Participants identified as having HR disease and who consent to take part in the HR treatment schedule receive treatment as in Table 1. Eligibility criteria to continue treatment through each stage of ASCT, consolidation part 1 and 2, and maintenance, are detailed in Table 2.

Each individual drug in the schedule may be dose reduced if toxicity is experienced, as deemed necessary by the treating physician and in line with standard reductions used for these treatments (Table 3). Dose reductions can be made for grade 1 toxicity (e.g. neuropathy) to maximise long-term tolerability and treatment effect in this patient group. Dose reductions from pre-trial treatment may be continued at induction treatment.

Trial Assessments

During treatment

MUKnine a: For non-HR participants a summary of treatment received in each phase of treatment is collected. Central samples are collected at the end of any line of standard treatment for response assessment. For patients participating in the imaging study a DW-MRI scan is performed at 100-120 days and 21 months post-ASCT, along with bone marrow, peripheral blood and urine samples for disease assessment.

1
2
3 MUKnine b: For HR participants, trial assessments are performed in line with the schedule of
4 assessments in Table 4. Data are collected at each cycle of treatment and at the end of each
5 phase of treatment.
6

7 Central laboratory investigations include:

- 8
- 9 • Bone marrow aspirate and peripheral blood for molecular profiling:
 - 10 ○ MLPA or equivalent platform for copy number aberrations [del(17p), gain(1q),
11 del(1p)]²⁸
 - 12 ○ RQ-PCR translocation assay or equivalent tool for prediction of HR
13 translocations [t(4;14), t(14;16) and t(14;20)]⁶⁵
 - 14 ○ Gene expression profiling based on Affymetrix HG-U133 Plus 2.0 or equivalent
15 platform with risk profile determined as per EMC92 model²³
 - 16 ○ Exploratory molecular analyses to identify potentially targetable mutations
 - 17 ■ Whole exome or whole genome next-generation sequencing
 - 18 ■ Gene expression profiling (GEP)
 - 19 ■ Epigenetic analyses
 - 20 ■ Germline variant analysis
 - 21 • Bone marrow aspirate for MRD analyses
 - 22 • Peripheral blood for disease assessment
 - 23 ○ Disease parameters, e.g. paraprotein, for serum response assessments
 - 24 ○ Beta-2-microglobulin
 - 25 ○ Albumin
- 26
27
28
29

30 Quality of Life questionnaires, EQ-5D, QLQ-C30 and QLQ-MY20, are collected from all
31 participants at baseline, and for participants who go on to HR treatment these are completed
32 at:
33

- 34 ○ End of induction treatment
 - 35 ○ 100 days post-ASCT then 3-monthly thereafter until disease progression.
- 36
37

38 *Follow-up*

39 Upon completion of treatment, patients are followed-up at 3 months, and then six-monthly
40 during standard of care visits, until second disease progression, death or withdrawal
41
42
43

44 *Imaging assessments*

45 All patients participating in the DW-MRI sub-study have whole body DW-MRI scan performed
46 at baseline, 100-120 days post-ASCT and at end of consolidation part 2.
47
48
49

50 Outcomes

51 *Primary endpoint*

52 MUKnine a:

53 The proportion of patients with molecular risk-defining investigations performed within 8
54 weeks.
55

56 MUKnine b:
57
58
59
60

1
2
3 The primary endpoints to determine whether to terminate the trial early for futility are

4 Minimal residual disease at 100 days post-ASCT

5 Progression-free survival at 100 days post-ASCT

6
7
8 The primary endpoint to assess efficacy of HR treatment if the trial is not stopped early for
9 futility is PFS at 18 months post-registration to screening.

11 12 13 *Secondary Endpoints*

14
15 MUKnine a: recruitment rates; PFS; OS; Second PFS (PFS2); treatment received;
16 overall response;

17
18 MUKnine b:

19
20 Safety and toxicity (adverse reactions (ARs), serious adverse events (SAEs), serious
21 adverse reactions (SARs) and suspected unexpected serious adverse reactions (SUSARs)
22 graded by common terminology criteria for adverse events (CTCAE) v5.0)

23
24 MRD at the end of induction therapy, and post- consolidation part 2

25
26 OS

27
28 Maximum and overall response at the end of induction therapy, 100 days post-ASCT
29 and postconsolidation part 2

30
31 Time to progression and time to maximum response

32
33 PFS2

34
35 Overall treatment benefit and clinician assessment of treatment benefit at the end of
36 induction therapy and 100 days post-ASCT

37
38 Quality of life as assessed by the EQ-5D, EORTC QLQ-C30 and EORTC QLQ-MY20

39
40 Treatment compliance

41 42 *Exploratory Endpoints*

43
44 Genomic instability, mutation rates and clonal evolution

45 46 47 *Imaging sub-study*

48
49 PFS; OS; Response; Patterns of disease distribution and discreet “3D phenotypes”

50 51 52 Statistical analysis

53
54 The MUKnine b trial is designed using a Bayesian approach to enable assessment of multiple
55 outcomes and incorporating multiple interim analyses.

56
57 The experimental treatment will be evaluated on an ongoing basis based on assessment of
58 MRD status and PFS. Interim assessments are made after cohorts of 10 participants have
59 been followed up to 100-120 days post-ASCT, and data reviewed by an independent Data
60

1
2
3 Monitoring and Ethics Committee (DMEC). The trial may be terminated early for futility on the
4 basis of MRD status and PFS at 100-120 days post-ASCT, using initial pre-defined stopping
5 boundaries based on Myeloma IX data. Following updated prior information becoming
6 available from Myeloma XI/XI+, these stopping boundaries were re-calculated to provide
7 updated decision criteria.
8

9
10 If the trial is not terminated early, up to 105 newly diagnosed patients with molecular HR
11 disease will be registered to treatment. With the availability of molecularly matched individual
12 participant data from currently running trials (Myeloma XI/XI+) a body of almost concurrent
13 control data is available to use for the purpose of exploratory statistical comparison.
14

15 The experimental treatment arm will be compared to control in terms of PFS at 18 months
16 post-registration to screening, expressed as a binary outcome, within the Bayesian framework.
17 Further analyses of PFS at 18 months will be performed outside of the Bayesian framework
18 using Kaplan-Meier estimation.
19

20 MUK*nine a* endpoints, and secondary and exploratory endpoints will be analysed using
21 summary statistics alongside confidence intervals where appropriate. All analyses are fully
22 detailed in a statistical analysis plan prior to being undertaken. Full statistical analysis for
23 MUK*nine* is also discussed in the MUK*nine* statistical methods paper (in preparation).
24
25

26 Trial conduct

27
28 Data are collected via electronic case report forms. Site monitoring of source data is performed
29 by CTRU following the trial monitoring plan. The trial is conducted in accordance with the
30 principles of Good Clinical Practice (GCP) and in line with the relevant Research Governance
31 Framework within the UK through adherence with University of Leeds CTRU standard
32 operating procedures. An independent DMEC reviews safety data on a regular basis to identify
33 any safety concerns or trends. An independent Trial Steering Committee periodically reviews
34 safety data and discusses recommendations made by the DMEC.
35
36
37
38

39 Patient and Public Involvement

40 Patients were involved in review and development of trial design, protocol and patient
41 information sheet.
42
43

44 **Ethics and dissemination**

45
46 The trial has national research ethics approval from the NHS National Research Ethics
47 Service, London South East Research Ethics Committee (Ref: 17/LO/0022, 17/LO/0023). All
48 patients provide written informed consent prior to take part in the trial at the hospital site where
49 they are recruited.
50
51

52 A manuscript with results of the MUK*nine b* study will be published in a peer-reviewed journal.
53 Separate manuscripts will be written for results of MUK*nine a* and each of the exploratory
54 objectives; these will also be submitted for publication in peer-reviewed journals. Upon
55 publication of the final long-term results of the study, requests for use of data may be made to
56 the CTRU and will be reviewed by the Trial Management Group.
57
58
59
60

Discussion

This is the first time in the UK genetic risk has been used prospectively in MM to identify participants to be treated in an academically-led clinical trial and select treatment based solely on this. It is hoped this trial will bring improved survival and longer term disease control for patients with HRMM in the future by providing an intensive treatment regimen specifically targeted at this difficult to treat disease sub-group. In addition, the trial will provide important evidence regarding feasibility of multi-centre molecular-risk stratified trials in MM at the point of diagnosis, using central molecular tumour investigations.

Intensive treatment in HR patients has been used outside the UK with some promising results but access to drugs in the UK has been challenging. This trial is designed to work within the UK NHS system and provide the best treatment for HR patients. The availability of novel targeted molecular therapies helps in treating the highly heterogeneous disease of MM. Ultimately data generated through this trial aims to support the case for access to combination therapies of expensive agents to patient subgroups with a high unmet need such as HR disease.

Abbreviations

ADCC	antibody dependant cytotoxicity
ADCP	antibody dependant cell phagocytosis
ALT	Alanine transaminase
ASCT	Autologous stem cell transplant
AST	Aspartate transaminase
CRD	Cyclophosphamide, Lenalidomide (Revlimid®), Dexamethasone
CTCAE	Common Terminology Criteria for Adverse Events
CTD	Cyclophosphamide, thalidomide, dexamethasone
DMEC	Data Monitoring and Ethics Committee
DW-MRI	Diffuse weighted magnetic resonance imaging
ECG	Electrocardiogram
ECOG	Eastern Cooperative Oncology Group performance status
¹⁸ F-FDG PET-CT	¹⁸ F-fluorodeoxyglucose - Positron emission tomography - computerised tomography
GEP	Gene expression profiling
HDM-V	High dose Melphalan with Velcade (bortezomib)
HR	High risk
IMiD	Immunomodulatory drugs
IMWG	International Myeloma Working Group
KCRD	Carfilzomib, Cyclophosphamide, Lenalidomide, Dexamethasone
MM	Multiple Myeloma
MRD	Minimal residual disease
MRI	Magnetic resonance imaging
OS	Overall survival
PCL	Plasma Cell Leukaemia
PFS	Progression-free survival
PFS2	Second progression-free survival
RCVD	Lenalidomide, Cyclophosphamide, Bortezomib, Dexamethasone
RVD	Lenalidomide, Bortezomib, Dexamethasone
RD	Lenalidomide, Dexamethasone
VRD	Bortezomib, Lenalidomide, Daratumumab
VRDd	Bortezomib, Lenalidomide, Daratumumab, dexamethasone

Competing interests

SH, DS, LF, SR, AH, KW and SB declare they have no competing interests. MK has served as a consultant and received honoraria from Celgene, Takeda, Amgen, Chugai, BMS, AbbVie and Janssen, and has received research funding from Celgene. GP served as a consultant and received honoraria from Janssen Oncology, Celgene, Amgen, Takeda, Gilead Sciences and Binding Site. MJ has served as a consultant and received honoraria from Celgene, Janssen, Takeda, Amgen, AbbVie and Novartis.

Funding

This trial is funded by Janssen, Celgene and Myeloma UK. Award/Grant number is not applicable.

Author Contributions

MJ, MK, GP, SB, LF, SH and DS designed the trial. SH, AH, KW and SB developed the statistical analysis plan and are responsible for the ongoing statistical monitoring, analysis and interpretation of data. DS, SH, MK and MJ wrote the manuscript. MJ, MK and GP perform the research and collect data. DS, LF and SR perform trial and data management. All authors reviewed and approved the final manuscript.

Acknowledgments

Janssen and Celgene provided funding to Myeloma UK and University of Leeds who in turn fund the MUK*nine* trial. Janssen and Celgene approved the design of the study but have no input in the collection, analysis or interpretation of data as this is a fully academically sponsored trial. Contact details of the sponsor, the University of Leeds, are accessible via the trial registration. Through the Myeloma UK Clinical Trials Network, Myeloma UK were involved in the study design and are actively involved in the collection and interpretation of data, as well as in the review of manuscripts arising from the study publishing trial outcomes.

For peer review only

Figure 1. MUKnine OPTIMUM trial design

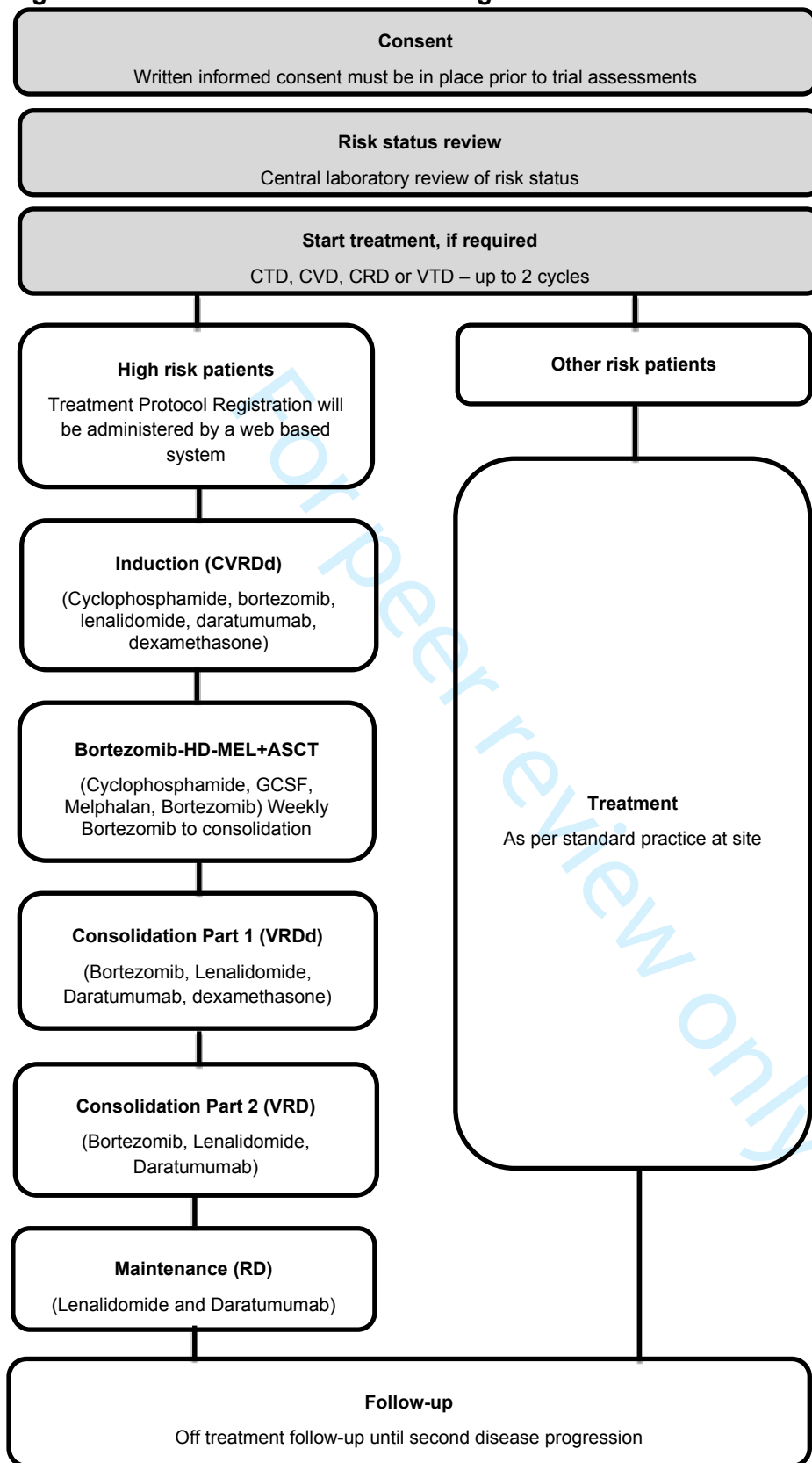


Table 1: Treatment Schedule

Induction Treatment			
Regimen: CVRDd to maximum response (or a maximum of 6 cycles of bortezomib)			Cycle duration: 21 days
Drug	Dose	Route	Days
Cyclophosphamide	500 mg	PO	1 and 8
Bortezomib	1.3 mg/m ²	SC	1, 4, 8, 11
Lenalidomide	25 mg	PO	1 – 14
Daratumumab	16 mg/kg (actual body weight)	IV	1, 8, 15** (Cycles 1 and 2)
			1 only (Cycle 3 onwards)
Dexamethasone*	20 mg – 40 mg	PO/IV	1, 4, 8, 11
Autologous Stem Cell Transplant			
Cyclophosphamide and GCSF mobilisation is recommended			
Regimen: Bortezomib HD-MEL+ASCT			
Drug	Dose	Route	Days
Melphalan	200 mg/m ²	IV	-1
Bortezomib	1.3mg/m ²	SC	-1, (8-18 hours post Melphalan)
Autologous stem cell return		IV	0
Bortezomib	1.3mg/m ²	SC	+5, +14, then weekly to consolidation 1
Consolidation Treatment 1			
To begin between 100 -120 days post ASCT			
Regimen: VRDd x 6 cycles*			Cycle duration: 28 days
Drug	Dose	Route	Days
Bortezomib	1.3mg/m ²	SC	1, 8,15, 22
Lenalidomide	25mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1
Dexamethasone*	20mg-40mg	PO/IV	1, 8,15**, 22
Consolidation Treatment 2			
Regimen: VRD x 12 cycles*			Cycle duration: 28 days
Drug	Dose	Route	Days
Bortezomib	1.3mg/m ²	SC	1, 8,15
Lenalidomide	25mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1

Maintenance Treatment			
Regimen: RD until progression		Cycle duration: 28 days	
Drug	Dose	Route	Days
Lenalidomide	10mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1

* On days where participants receive dexamethasone 40mg at site (i.e. pre Daratumumab infusion), dexamethasone must not be self-administered at home too.

** On day 15 participants will receive pre-med as per SPC (e.g. methylprednisolone)

Table 2: Eligibility Criteria for trial entry and continuing treatment through each stage

Inclusion Criteria	Exclusion Criteria
<p>Screening</p> <ul style="list-style-type: none"> Undergoing bone marrow investigation due to suspected symptomatic multiple myeloma or plasma cell leukaemia (PCL) or Participants with biopsy-confirmed symptomatic multiple myeloma, willing to undergo a further study bone marrow biopsy for molecular profiling. Participants previously screened but found not to have symptomatic multiple myeloma but now have suspected symptomatic multiple myeloma may be re-screened Aged 18 years or over Fit for intensive chemotherapy and autologous stem cell transplant (at clinician's discretion) Eastern Cooperative Oncology Group (ECOG) score ≤ 2 	<ul style="list-style-type: none"> Confirmed solitary bone/solitary extramedullary plasmacytoma. Primary diagnosis of Waldenstrom's Disease. Monoclonal gammopathy of undetermined significance or smouldering multiple myeloma unless progression to symptomatic multiple myeloma is highly suspected or confirmed Received therapy for multiple myeloma Prior or concurrent invasive malignancies Any uncontrolled or severe cardiovascular or pulmonary disease Grade 2 or greater peripheral neuropathy (per NCI-CTCAEv4.0) Known/underlying medical conditions that, in the investigator's opinion, would make the administration of the study drug hazardous Any clinically significant cardiac disease Known chronic obstructive pulmonary disease (COPD) Known to be seropositive for history of human immunodeficiency virus (HIV) or known to have active hepatitis B or hepatitis C. Any known allergies, hypersensitivity, or intolerance to corticosteroids, monoclonal antibodies or human proteins, or their excipients or known sensitivity to mammalian-derived products. Clinically significant allergies or intolerance to cyclophosphamide, lenalidomide, bortezomib, daratumumab or dexamethasone. Previous treatment with daratumumab or any other anti-CD38 therapies. Participants with contraindication to thromboprophylaxis. Participants with POEMS syndrome Any concurrent medical or psychiatric condition or disease

	<ul style="list-style-type: none"> • Known or suspected of not being able to comply with the study protocol • Participant is a woman who is pregnant, or breast-feeding, or planning to become pregnant while enrolled in this trial or within at least 6 months after the last dose of trial treatment. Or, participant is a man who plans to father a child while taking part in this trial or within at least 6 months after the last dose of trial treatment. • Major surgery within 2 weeks before treatment protocol registration or has not fully recovered from surgery, or has surgery planned during the time the participant is expected to participate in the study. Kyphoplasty or vertebroplasty is not considered major surgery. • Received an investigational drug (including investigational vaccines) or used an invasive investigational medical device within 4 weeks before treatment protocol registration or is currently enrolled in an interventional investigational study. <p><u>Imaging sub-study</u></p> <p>Only those taking part in the imaging sub study have these exclusions:</p> <ul style="list-style-type: none"> • MRI incompatible metal implant • Claustrophobia
<p>Treatment</p> <ul style="list-style-type: none"> • Confirmation of High Risk status from ICR. Participants with confirmed plasma cell leukaemia with >20% circulating plasma cells do not need confirmation of HR status from ICR to proceed to treatment. • Confirmation of receipt of baseline bone marrow at HMDS and, blood and urine samples at the University of Birmingham • Previously untreated participants, although participants may have received up to 2 cycles of CTD, CVD, CRD or VTD pre-trial induction chemotherapy while awaiting the results of the laboratory analysis. • Measurable disease before starting standard treatment <ul style="list-style-type: none"> ◦ Paraprotein \geq 5g/L or \geq 0.5 g/L for IgD subtypes OR Serum free kappa or lambda light chains \geq 100 mg/L with abnormal ratio (for light chain only myeloma) OR Urinary Bence Jones protein \geq 200 mg/24h. • Non-measurable participants providing they accept a 3 monthly bone marrow during induction and a 6 monthly bone marrow assessment during consolidation and maintenance. • Fit for intensive chemotherapy and autologous stem cell transplant (at clinician's discretion). • Eastern Cooperative Oncology Group (ECOG) Performance Status \leq2. 	<ul style="list-style-type: none"> • Solitary bone/solitary extramedullary plasmacytoma. • Primary diagnosis of amyloidosis, monoclonal gammopathy of undetermined significance or smouldering multiple myeloma or Waldenstrom's Disease. • Prior or concurrent invasive malignancies • Known/underlying medical conditions that, in the investigator's opinion, would make the administration of the study drug hazardous • Any clinically significant cardiac disease • Known chronic obstructive pulmonary disease (COPD) • Known to be seropositive for history of human immunodeficiency virus (HIV) or known to have active hepatitis B or hepatitis C. • Any known allergies, hypersensitivity, or intolerance to corticosteroids, monoclonal antibodies or human proteins, or their excipients or known sensitivity to mammalian-derived products. • Clinically significant allergies or known intolerance to cyclophosphamide, lenalidomide, bortezomib, daratumumab or dexamethasone. • Previous treatment with daratumumab or any other anti-CD38 therapies. • Participants with contraindication to thromboprophylaxis.

<ul style="list-style-type: none"> • The Celgene Pregnancy Prevention Plan must be followed and participants must agree to comply with this: <ul style="list-style-type: none"> • Females of childbearing potential (FCBP) must agree to utilise two reliable forms of contraception simultaneously or practice complete abstinence for at least for 28 days prior to starting trial treatment, during the trial and for at least 28 days after trial treatment discontinuation, and even in case of dose interruption, and must agree to regular pregnancy testing during this timeframe. • Males must agree to use a latex condom during any sexual contact with FCBP during the trial, including during dose interruptions and for 28 days following discontinuation from this trial even if he has undergone a successful vasectomy • Males must also agree to refrain from donating semen or sperm while on trial treatment including during any dose interruptions and for at least 6 months after discontinuation from this trial • All participants must agree to refrain from donating blood while on trial drug including during dose interruptions and for 28 days after discontinuation from this trial. • Laboratory Results <ul style="list-style-type: none"> • Calculated creatinine clearance $\geq 30\text{mL/min}$ (using Cockcroft-Gault formula). • ALT or AST ≤ 2.5 times upper limit of normal (ULN). • Bilirubin $\leq 2.0 \times \text{ULN}$, except in participants with congenital bilirubinemia, such as Gilbert syndrome (direct bilirubin ≤ 2.0 times ULN) • Platelet count $\geq 75 \times 10^9/\text{L}$. ($\geq 50 \times 10^9/\text{L}$ if multiple myeloma involvement in the bone marrow is $>50\%$). Platelet support is permitted. • Absolute neutrophil count (ANC) $\geq 1.0 \times 10^9/\text{L}$. Growth factor support is permitted. • Haemoglobin $\geq 80 \text{ g/L}$. (Participants may be receiving red blood cell (RBC) transfusions in accordance with institutional guidelines. • Corrected serum calcium $\leq 3.5 \text{ mmol/L}$ 	<ul style="list-style-type: none"> • Participants with POEMS syndrome • Any concurrent medical or psychiatric condition or disease • Known or suspected of not being able to comply with the study protocol • Participant is a woman who is pregnant, or breast-feeding, or planning to become pregnant while enrolled in this trial or within at least 6 months after the last dose of trial treatment. Or, participant is a man who plans to father a child while taking part in this trial or within at least 6 months after the last dose of trial treatment. • Major surgery within 2 weeks before treatment protocol registration or has not fully recovered from surgery, or has surgery planned during the time the participant is expected to participate in the study. Kyphoplasty or vertebroplasty is not considered major surgery. • Received an investigational drug (including investigational vaccines) or used an invasive investigational medical device within 4 weeks before treatment protocol registration or is currently enrolled in an interventional investigational study. <p><u>Imaging sub-study</u></p> <p>Only those taking part in the imaging sub study have these exclusions:</p> <ul style="list-style-type: none"> • MRI incompatible metal implant • Claustrophobia • Not received a DW-MRI at baseline
<p>Autologous Stem Cell Transplant</p> <ul style="list-style-type: none"> • Minimum stem cell harvest of 2×10^6 CD34+ cells/kg body weight. • Received a minimum of 4, unless CR has been achieved with a lesser number, or a maximum of 6 Induction (CVRDd) cycles (including standard treatment). • Achieved a response of stable disease (SD_ or better). • Dose modifications of any or all individual drugs within induction is permitted including complete stop of no more than one agent due to toxicity as 	<ul style="list-style-type: none"> • Participants that have progressive disease.

long as the required number of cycles have been received	
<p>Consolidation Part 1</p> <ul style="list-style-type: none"> • Undergone autologous transplant with HDM-V conditioning (Participants must have received a minimum of 100 mg/m² Melphalan in order to proceed with consolidation). • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modifications because of toxicity including complete stop of weekly bortezomib is permitted 	<ul style="list-style-type: none"> • Participants that have progressive disease.
<p>Consolidation Part 2</p> <ul style="list-style-type: none"> • Received 6 cycles of Consolidation Part 1 (VRDd) • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modification of any or all of the individual drugs in consolidation part 1 is permitted including complete stop of no more than one agent because of toxicity as long as the required number of cycles have been received. 	<ul style="list-style-type: none"> • Participants that have progressive disease.
<p>Maintenance</p> <ul style="list-style-type: none"> • Received 12 cycles of Consolidation Part 2 (VRD). • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modification of any or all of the individual drugs in consolidation part 2 is permitted including complete stop of no more than one agent because of toxicity as long as the required number of cycles have been received. 	<ul style="list-style-type: none"> • Participants that have progressive disease.

Table 3: Dose Modifications

Cyclophosphamide					
Modifications are at the discretion of the investigator					
<i>Renal Impairment</i> - A dose reduction of 50% for creatinine clearance of 10ml/min is recommended					
<i>Hepatic Impairment</i> - A dose reduction to 350mg is recommended with a serum bilirubin of >2.5 times the upper limit of normal					
Bortezomib					
Induction dose reductions					
Regimen: First dose reduction CVRDd				Cycle duration: 21 days	
Drug	Dose	Route	Days		
Cyclophosphamide	500 mg	PO	1 and 8		
Bortezomib	1.3 mg/m ²	SC	1, 8, 15		
Lenalidomide	25 mg	PO	1 – 14		
Daratumumab	16 mg/kg (actual body weight)	IV	1, 8, 15 (Cycles 1 and 2)		
			1 only (Cycle 3 onwards)		
Dexamethasone*	20 mg – 40 mg	PO/IV	1, 8, 15		
Post induction dose reductions					
Bortezomib schedule	Dose levels				
	0	-1	-2	-3	-4
Twice weekly schedules	1.3 mg/m ² d 1, 4, 8, 11	1.3 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 8, 15	1.3 mg/m ² d 1, 15	Stop
Once weekly schedules	1.3 mg/m ² d 1, 8, 15, (22)	1.0 mg/m ² d 1, 8, 15 (22)	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
Consolidation 1	1.3 mg/m ² d 1, 8, 15, 22	1.0 mg/m ² d 1, 8, 15, 22	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
Consolidation 2	1.3 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
<i>Neuropathy</i> - CTCAE Grade 1 with pain or grade 2- withhold bortezomib until returns to baseline. Dose reduce 1 level; CTCAE Grade 2 with pain or grade 3- withhold bortezomib until returns to baseline. Dose reduce 2 levels; CTCAE Grade 4 – discontinue treatment					
<i>Renal impairment</i> – dose reduce at the discretion of the clinician					
<i>Hepatic impairment</i> – moderate or severe impairment (>1.5 – 3x ULN) should start on a reduced dose of 0.7mg/m ² during the first cycle of treatment and dose escalate to 1.0 mg/m ² or dose reduce to 0.5mg/m ² may be considered					
<i>Grade 3 Non haematological toxicity</i> – withhold until symptoms of toxicity resolve and reduce one dose.					
<i>Grade 4 haematological toxicity</i> – withhold until symptoms of toxicity resolve and reduce one dose. Support may be given.					

Lenalidomide schedule	Dose levels					
	0	-1	-2	-3	-4	-5
	25mg	20mg	15mg	10mg	5mg	2.5 mg
<p><i>Thrombocytopenia</i> - $<25 \times 10^9/L$ stop lenalidomide for the remainder of the cycle. Return to $\geq 50 \times 10^9/L$ decrease by 1 dose level to resume the next cycle.</p> <p><i>Neutropenia</i> – first fall to $<0.5 \times 10^9/L$ omit lenalidomide until a return to $\geq 0.5 \times 10^9/L$ when neutropenia is the only toxicity. Resume lenalidomide at one dose lower. For each subsequent drop to $\geq 0.5 \times 10^9/L$ omit lenalidomide, resume lenalidomide decreased by 1 dose level at the next cycle.</p> <p><i>Renal impairment</i> – 30- 50 mL/min 10mg daily; < 30 mL/min, not requiring dialysis 7.5mg daily or 15mg every other day; < 30 mL/min, requiring dialysis 5mg daily administered following dialysis</p> <p><i>Other non haematological toxicities:</i> CTCAE grade 3 & 4 related to lenalidomide should be stopped and started 1 dose lower when toxicity has resolved to grade 2 at clinicians discretion. Rash – interrupt or discontinue for grade 2 or 3. Grade 4 rash discontinue including angioedema, exfoliative or bullous rash or Steven Johnson syndrome or toxic epidermal necrosis.</p>						
Daratumumab schedule	Frequency	Dose held	Dosing restart			
Induction cycles 1 and 2	Weekly	>3 days	Next planned weekly dose			
Induction cycles 3 – 6	Monthly	>1 week	Next planned weekly dose			
Consolidation 1, Consolidation 2, Maintenance	Monthly	>2 weeks	Next planned weekly dose			
<p>Follow the daratumumab SmPC. The daratumumab infusion must be withheld to allow for recovery from toxicity ONLY where any of the following criteria are met and the event cannot be ascribed to lenalidomide or cyclophosphamide.</p> <ul style="list-style-type: none"> • Grade 4 thrombocytopenia or Grade 3 thrombocytopenia with bleeding. • Grade 4 neutropenia, if this is the second occurrence despite growth factor support. • Febrile neutropenia of any grade. • Neutropenia with infection, of any grade. • Grade 3 or higher non-haematological toxicities with the following exceptions: <ul style="list-style-type: none"> ○ Grade 3 nausea that responds to antiemetic treatment within 8 days. ○ Grade 3 vomiting that responds to antiemetic treatment within 8 days. ○ Grade 3 diarrhoea that responds to anti-diarrhoeal treatment within 8 days. ○ Grade 3 fatigue that was present at baseline or that lasts for <8 days after the previous administration of daratumumab. ○ Grade 3 asthenia that was present at baseline or that lasts for <8 days after the previous administration of daratumumab. 						
Dexamethasone						
<p>Occasionally patients will not be able to tolerate because of corticosteroid effects. Dose reductions from 40 to 20mg daily. Further dose reductions to 10mg daily is acceptable followed by the omission of dexamethasone</p> <p>If the bortezomib schedule changes, dexamethasone should change in line with it.</p>						
Melphalan						
<p>Dose may be adjusted based on performance status and clinical judgment in discussion with the Chief Investigator</p> <p>GFR measured by Cockcroft & Gault formula or EDTA - >50ml/min 200mg/m²; 30-50ml/min 140mg/m²; <30ml/min 100mg/m²</p>						

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For peer review only

Table 4: Trial Assessments

Investigations	All Patients	Non HR Patients			HR Patients							
	Screening participants - All	Prior to any new line of treatment	Post any line of treatment	First and second disease progression	Before starting MUKnine treatment ¹	Prior to each cycle of induction treatment CVRDd ²	End of induction treatment	Autologous stem cell transplant ⁵	100-120 days post transplant	Prior to each cycle of consolidation part 1 (VRDd), consolidation part 2 (VRD) and maintenance treatment (RD)	End of consolidation part 1 (VRDd), consolidation part 2 (VRD) and maintenance treatment (RD)	First and second disease progression
Consent	X				X							
Medical history	X				X							
Symptom-directed physical exam (including weight, ECOG)	X	X	X		X	X	X	X	X	X	X	
Haematology & biochemistry test	X	X	X		X	X	X	X	X	X	X	
Disease assessment ³	X	X	X	X	X	X	X	X	X	X	X	X
DW-MRI Imaging ⁷	X								X		X (Part 2 only)	
ECG					X							
Pregnancy testing as required					X	X	X		X	X	X	
Participant questionnaires	X						X		X	X ⁶	X	
Details of treatment			X			X	X	X	X	X	X	
Clinical assessment of treatment benefit							X		X		X	
Central Laboratory Samples												
Bone marrow aspirate	X			X			X		X		X	X
Peripheral blood	X	X		X		X ⁴	X		X	X ^{4,6}	X	X
Urine sample	X	X		X		X ⁴	X		X	X ^{4,6}	X	X

1 Treatment must start within 14 days of registration to MUKnine treatment

2 All assessments must be performed within 72 hours prior to day 1 of each cycle of treatment

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

3 Response assessments must be made in line with the IMWG criteria

4 Cycle 1 day 1 only

5 Autologous stem cell transplant will be performed as per local practice with local monitoring of adverse events and haematology tests. Participants will be given weekly
6 bortezomib until 100-120 days post transplant, the assessments will be performed monthly during this time for the trial.

7 3 monthly during treatment

8 if site and participant taking part in the imaging sub-study

For peer review only

References

1. Kyle RA, Rajkumar SV. Criteria for diagnosis, staging, risk stratification and response assessment of multiple myeloma. *Leukemia* 2009;23(1):3-9. doi: leu2008291 [pii];10.1038/leu.2008.291 [doi]
2. Myeloma Statistics Cancer Research UK. <https://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/myeloma#heading-Two> 2015
3. Ries LAG YJ, Keel GE, Eisner MP, Lin YD, Horner M-J SEER Survival Monograph: Cancer Survival Among Adults U.S. SEER Program, 1988-2001, Patient and Tumor Characteristics.
4. Avet-Loiseau H. Ultra high-risk myeloma. *Hematology*;2010:489-93.
5. Avet-Loiseau H, Attal M, Campion L, et al. Long-term analysis of the IFM 99 trials for myeloma: cytogenetic abnormalities [t(4;14), del(17p), 1q gains] play a major role in defining long-term survival. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2012;30(16):1949-52. doi: 10.1200/jco.2011.36.5726 [published Online First: 2012/05/02]
6. Avet-Loiseau H, Durie BG, Cavo M, et al. Combining fluorescent in situ hybridization data with ISS staging improves risk assessment in myeloma: an International Myeloma Working Group collaborative project. *Leukemia* 2013;27(3):711-7. doi: 10.1038/leu.2012.282
7. Boyd KD, Ross FM, Chiecchio L, et al. A novel prognostic model in myeloma based on co-segregating adverse FISH lesions and the ISS: analysis of patients treated in the MRC Myeloma IX trial. *Leukemia* 2012;26(2):349-55. doi: 10.1038/leu.2011.204 [published Online First: 2011/08/13]
8. Chng WJ, Kuehl WM, Bergsagel PL, et al. Translocation t(4;14) retains prognostic significance even in the setting of high-risk molecular signature. *Leukemia* 2008;22(2):459-61. doi: 10.1038/sj.leu.2404934 [published Online First: 2007/09/07]
9. Morgan GJ, Davies FE, Gregory WM, et al. First-line treatment with zoledronic acid as compared with clodronic acid in multiple myeloma (MRC Myeloma IX): a randomised controlled trial. *Lancet* 2010;376(9757):1989-99. doi: 10.1016/s0140-6736(10)62051-x [published Online First: 2010/12/07]
10. Morgan GJ, Davies FE, Gregory WM, et al. Cyclophosphamide, thalidomide, and dexamethasone (CTD) as initial therapy for patients with multiple myeloma unsuitable for autologous transplantation. *Blood* 2011;118(5):1231-8. doi: 10.1182/blood-2011-02-338665
11. Shaughnessy JD, Jr., Zhan F, Burington BE, et al. A validated gene expression model of high-risk multiple myeloma is defined by deregulated expression of genes mapping to chromosome 1. *Blood* 2007;109(6):2276-84. doi: 10.1182/blood-2006-07-038430 [published Online First: 2006/11/16]
12. Shah V, Sherborne AL, Walker BA, et al. Prediction of outcome in newly diagnosed myeloma: a meta-analysis of the molecular profiles of 1905 trial patients. *Leukemia* 2017 doi: 10.1038/leu.2017.179

13. Shah V, Sherborne AL, Johnson DC, et al. Predicting ultrahigh risk multiple myeloma by molecular profiling: an analysis of newly diagnosed transplant eligible myeloma XI trial patients. *Leukemia* 2020 doi: 10.1038/s41375-020-0750-z
14. Dimopoulos MA, Oriol A, Nahi H, et al. Daratumumab, Lenalidomide, and Dexamethasone for Multiple Myeloma. *The New England journal of medicine* 2016;375(14):1319-31. doi: 10.1056/NEJMoa1607751 [published Online First: 2016/10/06]
15. Palumbo A, Chanan-Khan A, Weisel K, et al. Daratumumab, Bortezomib, and Dexamethasone for Multiple Myeloma. *The New England journal of medicine* 2016;375(8):754-66. doi: 10.1056/NEJMoa1606038 [published Online First: 2016/08/25]
16. Dimopoulos MA, Moreau P, Palumbo A, et al. Carfilzomib and dexamethasone versus bortezomib and dexamethasone for patients with relapsed or refractory multiple myeloma (ENDEAVOR): a randomised, phase 3, open-label, multicentre study. *The lancet oncology* 2016;17(1):27-38. doi: 10.1016/s1470-2045(15)00464-7 [published Online First: 2015/12/17]
17. Stewart AK. Carfilzomib for the treatment of patients with relapsed and/or refractory multiple myeloma. *Future Oncology*;11(15):2121-36.
18. Shah V, Sherborne AL, Walker BA, et al. Prediction of outcome in newly diagnosed myeloma: a meta-analysis of the molecular profiles of 1905 trial patients. *Leukemia* 2018;32(1):102-10. doi: 10.1038/leu.2017.179 [published Online First: 2017/06/07]
19. Chng WJ, Dispenzieri A, Chim CS, et al. IMWG consensus on risk stratification in multiple myeloma. *Leukemia* 2014;28(2):269-77. doi: 10.1038/leu.2013.247 [published Online First: 2013/08/27]
20. Jackson GH, Davies FE, Pawlyn C, et al. Lenalidomide maintenance versus observation for patients with newly diagnosed multiple myeloma (Myeloma XI): a multicentre, open-label, randomised, phase 3 trial. *The lancet oncology* 2019;20(1):57-73. doi: 10.1016/s1470-2045(18)30687-9 [published Online First: 2018/12/19]
21. Kuiper R, Broyl A, de Knecht Y, et al. A gene expression signature for high-risk multiple myeloma. *Leukemia* 2012;26(11):2406-13. doi: 10.1038/leu.2012.127
22. Decaux O, Lode L, Magrangeas F, et al. Prediction of survival in multiple myeloma based on gene expression profiles reveals cell cycle and chromosomal instability signatures in high-risk patients and hyperdiploid signatures in low-risk patients: a study of the Intergroupe Francophone du Myelome. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2008;26(29):4798-805. doi: 10.1200/jco.2007.13.8545 [published Online First: 2008/07/02]
23. Dickens NJ, Walker BA, Leone PE, et al. Homozygous deletion mapping in myeloma samples identifies genes and an expression signature relevant to pathogenesis and outcome. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2010;16(6):1856-64. doi: 10.1158/1078-0432.ccr-09-2831 [published Online First: 2010/03/11]

- 1
2
3 24. Mulligan G, Mitsiades C, Bryant B, et al. Gene expression profiling and
4 correlation with outcome in clinical trials of the proteasome inhibitor
5 bortezomib. *Blood* 2007;109(8):3177-88. doi: 10.1182/blood-2006-09-044974
6 [published Online First: 2006/12/23]
7
- 8 25. Walker BA, Boyle EM, Wardell CP, et al. Mutational Spectrum, Copy Number
9 Changes, and Outcome: Results of a Sequencing Study of Patients With
10 Newly Diagnosed Myeloma. *Journal of clinical oncology : official journal of the*
11 *American Society of Clinical Oncology* 2015;33(33):3911-20. doi:
12 10.1200/jco.2014.59.1503 [published Online First: 2015/08/19]
13
- 14 26. Samur MK, Samur AA, Fulciniti M, et al. Genome-Wide Somatic Alterations in
15 Multiple Myeloma Reveal a Superior Outcome Group. 2020;38(27):3107-18.
16 doi: 10.1200/jco.20.00461
17
- 18 27. Perrot A, Lauwers-Cances V, Tournay E, et al. Development and Validation of a
19 Cytogenetic Prognostic Index Predicting Survival in Multiple Myeloma.
20 2019;37(19):1657-65. doi: 10.1200/jco.18.00776
21
- 22 28. Kumar SK, Uno H, Jacobus SJ, et al. Impact of gene expression profiling-based
23 risk stratification in patients with myeloma receiving initial therapy with
24 lenalidomide and dexamethasone. *Blood* 2011;118(16):4359-62. doi:
25 10.1182/blood-2011-03-342089 [published Online First: 2011/08/24]
26
- 27 29. Meissner T, Seckinger A, Reme T, et al. Gene expression profiling in multiple
28 myeloma--reporting of entities, risk, and targets in clinical routine. *Clinical*
29 *cancer research : an official journal of the American Association for Cancer*
30 *Research* 2011;17(23):7240-7. doi: 10.1158/1078-0432.CCR-11-1628
31
- 32 30. Stewart AK, Rajkumar SV, Dimopoulos MA, et al. Carfilzomib, lenalidomide, and
33 dexamethasone for relapsed multiple myeloma. *New England Journal of*
34 *Medicine* 2015;372(2):142-52.
35
- 36 31. Palumbo A, Gay F, Cavallo F, et al. Continuous Therapy Versus Fixed Duration
37 of Therapy in Patients With Newly Diagnosed Multiple Myeloma. *Journal of*
38 *Clinical Oncology*;33(30):3459-66.
39
- 40 32. Brioli A, Kaiser MF, Pawlyn C, et al. Biologically defined risk groups can be used
41 to define the impact of thalidomide maintenance therapy in newly diagnosed
42 multiple myeloma. *Leuk Lymphoma* 2013;54(9):1975-81. doi:
43 10.3109/10428194.2012.760736 [published Online First: 2012/12/29]
44
- 45 33. Morgan GJ, Gregory WM, Davies FE, et al. The role of maintenance thalidomide
46 therapy in multiple myeloma: MRC Myeloma IX results and meta-analysis.
47 *Blood* 2012;119(1):7-15. doi: 10.1182/blood-2011-06-357038 [published
48 Online First: 2011/10/25]
49
- 50 34. Attal M, Lauwers-Cances V, Marit G, et al. Lenalidomide maintenance after
51 stem-cell transplantation for multiple myeloma. *The New England journal of*
52 *medicine* 2012;366(19):1782-91. doi: 10.1056/NEJMoa1114138 [published
53 Online First: 2012/05/11]
54
- 55 35. Barlogie B, Anaissie E, van Rhee F, et al. Incorporating bortezomib into upfront
56 treatment for multiple myeloma: early results of total therapy 3. *Br J Haematol*
57 2007;138(2):176-85. doi: 10.1111/j.1365-2141.2007.06639.x [published
58 Online First: 2007/06/27]
59
60

- 1
2
3 36. Bergsagel PL, Mateos MV, Gutierrez NC, et al. Improving overall survival and
4 overcoming adverse prognosis in the treatment of cytogenetically high-risk
5 multiple myeloma. *Blood* 2013;121(6):884-92. doi: 10.1182/blood-2012-05-
6 432203 [published Online First: 2012/11/21]
- 7
8 37. Harousseau JL, Attal M, Avet-Loiseau H, et al. Bortezomib plus dexamethasone
9 is superior to vincristine plus doxorubicin plus dexamethasone as induction
10 treatment prior to autologous stem-cell transplantation in newly diagnosed
11 multiple myeloma: results of the IFM 2005-01 phase III trial. *Journal of clinical
12 oncology : official journal of the American Society of Clinical Oncology*
13 2010;28(30):4621-9. doi: 10.1200/jco.2009.27.9158 [published Online First:
14 2010/09/09]
- 15
16 38. Neben K, Lokhorst HM, Jauch A, et al. Administration of bortezomib before and
17 after autologous stem cell transplantation improves outcome in multiple
18 myeloma patients with deletion 17p. *Blood* 2012;119(4):940-8. doi:
19 10.1182/blood-2011-09-379164
- 20
21 39. Shaughnessy JD, Zhou Y, Haessler J, et al. TP53 deletion is not an adverse
22 feature in multiple myeloma treated with total therapy 3. *Br J Haematol*
23 2009;147(3):347-51. doi: 10.1111/j.1365-2141.2009.07864.x [published
24 Online First: 2009/08/26]
- 25
26 40. Roussel M, Lauwers-Cances V, Robillard N, et al. Front-line transplantation
27 program with lenalidomide, bortezomib, and dexamethasone combination as
28 induction and consolidation followed by lenalidomide maintenance in patients
29 with multiple myeloma: a phase II study by the Intergroupe Francophone du
30 Myelome. *Journal of clinical oncology : official journal of the American Society
31 of Clinical Oncology* 2014;32(25):2712-7. doi: 10.1200/jco.2013.54.8164
32 [published Online First: 2014/07/16]
- 33
34 41. Kumar S, Flinn I, Richardson PG, et al. Randomized, multicenter, phase 2 study
35 (EVOLUTION) of combinations of bortezomib, dexamethasone,
36 cyclophosphamide, and lenalidomide in previously untreated multiple
37 myeloma. *Blood* 2012;119(19):4375-82. doi: 10.1182/blood-2011-11-395749
- 38
39 42. Plesner T, Arkenau HT, Gimsing P, et al. Phase 1/2 study of daratumumab,
40 lenalidomide, and dexamethasone for relapsed multiple myeloma. *Blood* 2016
41 doi: 10.1182/blood-2016-07-726729 [published Online First: 2016/08/18]
- 42
43 43. Katja C. Weisel JSM, Gordon Cook, Merav Leiba, Kenshi Suzuki, Shaji Kumar,
44 Michele Cavo, Herve Avet-Loiseau, Hang Quach, Vania Hungria, Suzanne
45 Lentzsch, Roman Hajek, Pieter Sonneveld, Kaida Wu, Xiang Qin, Christopher
46 Chiu, David Soong, Ming Qi, Jordan Mark Schechter, Meletios A. Dimopoulos.
47 Efficacy of daratumumab in combination with lenalidomide plus
48 dexamethasone (DRd) or bortezomib plus dexamethasone (DVd) in relapsed
49 or refractory multiple myeloma (RRMM) based on cytogenetic risk status.
50 2017 ASCO Annual Meeting 2017; J Clin Oncol 35, 2017 (suppl; abstr 8006)
- 51
52 44. Farooqui AA, Tariq M, Nabeel S, et al. Daratumumab-based three drug regimens
53 for high-risk multiple myeloma: A meta-analysis. 2020;38(15_suppl):e20549-
54 e49. doi: 10.1200/JCO.2020.38.15_suppl.e20549
- 55
56 45. Cavo M, Tosi P, Zamagni E, et al. Prospective, randomized study of single
57 compared with double autologous stem-cell transplantation for multiple
58
59
60

- myeloma: Bologna 96 clinical study. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2007;25(17):2434-41. doi: 10.1200/jco.2006.10.2509 [published Online First: 2007/05/09]
46. Cavo M, Gay F, Beksac M, et al. Autologous haematopoietic stem-cell transplantation versus bortezomib–melphalan–prednisone, with or without bortezomib–lenalidomide–dexamethasone consolidation therapy, and lenalidomide maintenance for newly diagnosed multiple myeloma (EMN02/HO95): a multicentre, randomised, open-label, phase 3 study. *The Lancet Haematology* 2020;7(6):e456-e68. doi: [https://doi.org/10.1016/S2352-3026\(20\)30099-5](https://doi.org/10.1016/S2352-3026(20)30099-5)
47. Barlogie B, Tricot GJ, van Rhee F, et al. Long-term outcome results of the first tandem autotransplant trial for multiple myeloma. *Br J Haematol* 2006;135(2):158-64. doi: 10.1111/j.1365-2141.2006.06271.x [published Online First: 2006/08/31]
48. Roussel M, Moreau P, Huynh A, et al. Bortezomib and high-dose melphalan as conditioning regimen before autologous stem cell transplantation in patients with de novo multiple myeloma: a phase 2 study of the Intergroupe Francophone du Myelome (IFM). *Blood* 2010;115(1):32-7. doi: 10.1182/blood-2009-06-229658 [published Online First: 2009/11/04]
49. Rawstron AC, Child JA, de Tute RM, et al. Minimal residual disease assessed by multiparameter flow cytometry in multiple myeloma: impact on outcome in the Medical Research Council Myeloma IX Study. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2013;31(20):2540-7. doi: 10.1200/JCO.2012.46.2119
50. Goicoechea I, Puig N, Cedena M-T, et al. Deep MRD profiling defines outcome and unveils different modes of treatment resistance in standard and high risk myeloma. *Blood* 2020 doi: 10.1182/blood.2020006731
51. Croft J, Riddell A, Koh D-M, et al. Inter-observer agreement of baseline whole body MRI in multiple myeloma. *Cancer Imaging* 2020;20(1):48. doi: 10.1186/s40644-020-00328-9
52. Messiou C, Hillengass J, Delorme S, et al. Guidelines for Acquisition, Interpretation, and Reporting of Whole-Body MRI in Myeloma: Myeloma Response Assessment and Diagnosis System (MY-RADS). 2019;291(1):5-13. doi: 10.1148/radiol.2019181949
53. ClinicalTrials.gov website.
54. Harousseau JL, Avet-Loiseau H, Attal M, et al. Achievement of at least very good partial response is a simple and robust prognostic factor in patients with multiple myeloma treated with high-dose therapy: long-term analysis of the IFM 99-02 and 99-04 Trials. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2009;27(34):5720-6. doi: 10.1200/jco.2008.21.1060 [published Online First: 2009/10/15]
55. Moreau P, Hullin C, Garban F, et al. Tandem autologous stem cell transplantation in high-risk de novo multiple myeloma: final results of the prospective and randomized IFM 99-04 protocol. *Blood* 2006;107(1):397-403. doi: 10.1182/blood-2005-06-2573 [published Online First: 2005/09/08]

- 1
2
3 56. McCarthy PL, Owzar K, Hofmeister CC, et al. Lenalidomide after stem-cell
4 transplantation for multiple myeloma. *The New England journal of medicine*
5 2012;366(19):1770-81. doi: 10.1056/NEJMoa1114083
6
- 7 57. Thall PF, Simon RM, Estey EH. Bayesian sequential monitoring designs for
8 single-arm clinical trials with multiple outcomes. *Stat Med* 1995;14(4):357-79.
9 [published Online First: 1995/02/28]
10
- 11 58. Thall PF, Simon RM, Estey EH. New statistical strategy for monitoring safety and
12 efficacy in single-arm clinical trials. *Journal of clinical oncology : official journal*
13 *of the American Society of Clinical Oncology* 1996;14(1):296-303. [published
14 Online First: 1996/01/01]
15
- 16 59. Thall PF, Sung HG. Some extensions and applications of a Bayesian strategy for
17 monitoring multiple outcomes in clinical trials. *Stat Med* 1998;17(14):1563-80.
18 [published Online First: 1998/08/12]
19
- 20 60. Wale A, Pawlyn C, Kaiser M, et al. Frequency, distribution and clinical
21 management of incidental findings and extramedullary plasmacytomas in
22 whole body diffusion weighted magnetic resonance imaging in patients with
23 multiple myeloma. *Haematologica* 2016;101(4):e142-4. doi:
24 10.3324/haematol.2015.139816 [published Online First: 2016/01/29]
25
- 26 61. Messiou C, Kaiser M. Whole body diffusion weighted MRI--a new view of
27 myeloma. *Br J Haematol* 2015;171(1):29-37. doi: 10.1111/bjh.13509
28 [published Online First: 2015/05/28]
29
- 30 62. Brown SR, Sherratt D, Booth G, et al. Experiences of establishing an academic
31 early phase clinical trials unit. *Clinical trials (London, England)*
32 2017:1740774517710250. doi: 10.1177/1740774517710250 [published
33 Online First: 2017/05/24]
34
- 35 63. Rajkumar SV, Harousseau JL, Durie B, et al. Consensus recommendations for
36 the uniform reporting of clinical trials: report of the International Myeloma
37 Workshop Consensus Panel 1. *Blood* 2011;117(18):4691-95. doi: blood-2010-
38 10-299487 [pii];10.1182/blood-2010-10-299487 [doi]
39
- 40 64. Office for National Statistics. Office for National Statistics: Cancer Statistics
41 Registrations, England (Series MB1) , No. 42, 2011 2013 [
42
- 43 65. Kaiser MF, Walker BA, Hockley SL, et al. A TC classification-based predictor for
44 multiple myeloma using multiplexed real-time quantitative PCR. *Leukemia*
45 2013;27(8):1754-7. doi: 10.1038/leu.2013.12 [published Online First:
46 2013/01/16]
47
48
49
50
51
52
53
54
55
56
57
58
59
60



STANDARD PROTOCOL ITEMS: RECOMMENDATIONS FOR INTERVENTIONAL TRIALS

SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	1
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	2
	2b	All items from the World Health Organization Trial Registration Data Set	1, 2, 4-9, 11-15, Tables 1 and 2
Protocol version	3	Date and version identifier	6
Funding	4	Sources and types of financial, material, and other support	14, 15
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	1,14
	5b	Name and contact information for the trial sponsor	15
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	14, 15
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	12, 13, 14

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

Introduction

Background and rationale

- 6a Description of research question and justification for undertaking the trial, including summary of relevant studies (published and unpublished) examining benefits and harms for each intervention
- 6b Explanation for choice of comparators

4-7
3, 8, 12

Objectives

- 7 Specific objectives or hypotheses

7, 8

Trial design

- 8 Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group), allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory)

2, 8

Methods: Participants, interventions, and outcomes

Study setting

- 9 Description of study settings (eg, community clinic, academic hospital) and list of countries where data will be collected. Reference to where list of study sites can be obtained

8, 9

Eligibility criteria

- 10 Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and individuals who will perform the interventions (eg, surgeons, psychotherapists)

Table 2

Interventions

- 11a Interventions for each group with sufficient detail to allow replication, including how and when they will be administered
- 11b Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose change in response to harms, participant request, or improving/worsening disease)
- 11c Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence (eg, drug tablet return, laboratory tests)
- 11d Relevant concomitant care and interventions that are permitted or prohibited during the trial

Table 1
9, 10 and Table 3
Detailed in main protocol
Table 2, Table 3

1	Outcomes	12	Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended	11, 12, further detail in MUK nine statistical methods paper and statistical analysis plan
2				
3				
4				
5				
6				
7				
8				
9				
10	Participant timeline	13	Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure)	9-11, Figure 1, Table 4
11				
12				
13	Sample size	14	Estimated number of participants needed to achieve study objectives and how it was determined, including clinical and statistical assumptions supporting any sample size calculations	8, 9
14				
15				
16	Recruitment	15	Strategies for achieving adequate participant enrolment to reach target sample size	Detailed in main protocol
17				
18				
19				

Methods: Assignment of interventions (for controlled trials)

Allocation:

24	Sequence generation	16a	Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any factors for stratification. To reduce predictability of a random sequence, details of any planned restriction (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants or assign interventions	NA, not randomised
25				
26				
27				
28				
29				
30	Allocation concealment mechanism	16b	Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned	NA, not randomised
31				
32				
33				
34	Implementation	16c	Who will generate the allocation sequence, who will enrol participants, and who will assign participants to interventions	NA, not randomised
35				
36				
37				
38	Blinding (masking)	17a	Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome assessors, data analysts), and how	NA, not randomised or blinded
39				
40				
41				
42				

1 17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant’s NA, not
 2 allocated intervention during the trial randomised or
 3 blinded
 4
 5

6 **Methods: Data collection, management, and analysis**

7
 8 Data collection 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related 9-13, Table 4
 9 methods processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of
 10 study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known.
 11 Reference to where data collection forms can be found, if not in the protocol
 12

13
 14 18b Plans to promote participant retention and complete follow-up, including list of any outcome data to be Detailed in main
 15 collected for participants who discontinue or deviate from intervention protocols protocol
 16

17 Data management 19 Plans for data entry, coding, security, and storage, including any related processes to promote data quality 13, 14
 18 (eg, double data entry; range checks for data values). Reference to where details of data management
 19 procedures can be found, if not in the protocol
 20

21 Statistical methods 20a Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the Detailed in MUK
 22 statistical analysis plan can be found, if not in the protocol nine statistical
 23 methods paper
 24 and statistical
 25 analysis plan
 26
 27

28 20b Methods for any additional analyses (eg, subgroup and adjusted analyses) Detailed in MUK
 29 nine statistical
 30 methods paper
 31 and statistical
 32 analysis plan
 33
 34
 35
 36
 37
 38
 39
 40
 41
 42
 43
 44
 45
 46

1	20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation)	Detailed in MUK nine statistical methods paper and statistical analysis plan
2			
3			
4			
5			
6			
7			
8			

Methods: Monitoring

11	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed	12, 13
12				
13				
14				
15				
16				
17				
18		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	9, 12
19				
20				
21	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse events and other unintended effects of trial interventions or trial conduct	10, 11, 27. Further detail in main protocol
22				
23				
24				
25	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	13
26				
27				
28				

Ethics and dissemination

31	Research ethics approval	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	2, 13
32				
33				
34	Protocol amendments	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals, regulators)	Detailed in main protocol
35				
36				
37				
38	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and how (see Item 32)	8-10, 13. Further detail in main protocol
39				
40				
41				
42				

1		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary studies, if applicable	9, 10. Further detail in main protocol
2				
3				
4				
5	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained in order to protect confidentiality before, during, and after the trial	Detailed in main protocol
6				
7				
8	Declaration of interests	28	Financial and other competing interests for principal investigators for the overall trial and each study site	14
9				
10				
11	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that limit such access for investigators	13, 14. Further detail in main protocol
12				
13				
14				
15				
16	Ancillary and post-trial care	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial participation	Detailed in main protocol
17				
18				
19	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals, the public, and other relevant groups (eg, via publication, reporting in results databases, or other data sharing arrangements), including any publication restrictions	2, 13
20				
21				
22				
23				
24		31b	Authorship eligibility guidelines and any intended use of professional writers	Detailed in main protocol
25				
26				
27		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	Detailed in main protocol
28				
29				
30				
31	Appendices			
32				
33	Informed consent materials	32	Model consent form and other related documentation given to participants and authorised surrogates	Detailed in main protocol
34				
35				
36	Biological specimens	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular analysis in the current trial and for future use in ancillary studies, if applicable	9, 10, Table 4. Further detail in main protocol
37				
38				
39				

1 *It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items.
2 Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons
3 [“Attribution-NonCommercial-NoDerivs 3.0 Unported”](#) license.
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

For peer review only

BMJ Open

The MUKnine OPTIMUM protocol: A screening study to identify high risk multiple myeloma patients suitable for novel treatment approaches combined with a phase II study evaluating optimised combination of biological therapy in newly diagnosed high risk multiple myeloma and plasma cell leukaemia

Journal:	<i>BMJ Open</i>
Manuscript ID	bmjopen-2020-046225.R1
Article Type:	Protocol
Date Submitted by the Author:	21-Jan-2021
Complete List of Authors:	Brown, Sarah; University of Leeds, Clinical Trials Research Unit Sherratt, Debbie; University of Leeds, Clinical Trials Research Unit Hinsley, Samantha; University of Leeds Clinical Trials Research Unit Flanagan, Louise; University of Leeds Clinical Trials Research Unit Roberts, Sadie; University of Leeds Clinical Trials Research Unit Walker, Katrina; University of Leeds Clinical Trials Research Unit Hall, Andrew; University of Leeds Clinical Trials Research Unit Pratt, Guy; Queen Elizabeth Hospital Messiou, Christina; Institute of Cancer Research Sutton Jenner, Matthew ; Southampton General Hospital Kaiser, Martin; Institute of Cancer Research,
Primary Subject Heading:	Haematology (incl blood transfusion)
Secondary Subject Heading:	Oncology
Keywords:	Myeloma < HAEMATOLOGY, Clinical trials < THERAPEUTICS, STATISTICS & RESEARCH METHODS

SCHOLARONE™
Manuscripts



I, the Submitting Author has the right to grant and does grant on behalf of all authors of the Work (as defined in the below author licence), an exclusive licence and/or a non-exclusive licence for contributions from authors who are: i) UK Crown employees; ii) where BMJ has agreed a CC-BY licence shall apply, and/or iii) in accordance with the terms applicable for US Federal Government officers or employees acting as part of their official duties; on a worldwide, perpetual, irrevocable, royalty-free basis to BMJ Publishing Group Ltd ("BMJ") its licensees and where the relevant Journal is co-owned by BMJ to the co-owners of the Journal, to publish the Work in this journal and any other BMJ products and to exploit all rights, as set out in our [licence](#).

The Submitting Author accepts and understands that any supply made under these terms is made by BMJ to the Submitting Author unless you are acting as an employee on behalf of your employer or a postgraduate student of an affiliated institution which is paying any applicable article publishing charge ("APC") for Open Access articles. Where the Submitting Author wishes to make the Work available on an Open Access basis (and intends to pay the relevant APC), the terms of reuse of such Open Access shall be governed by a Creative Commons licence – details of these licences and which [Creative Commons](#) licence will apply to this Work are set out in our licence referred to above.

Other than as permitted in any relevant BMJ Author's Self Archiving Policies, I confirm this Work has not been accepted for publication elsewhere, is not being considered for publication elsewhere and does not duplicate material already published. I confirm all authors consent to publication of this Work and authorise the granting of this licence.

1
2
3
4 The MUKnine OPTIMUM protocol: A screening study to identify high risk multiple myeloma
5 patients suitable for novel treatment approaches combined with a phase II study evaluating
6 optimised combination of biological therapy in newly diagnosed high risk multiple myeloma
7 and plasma cell leukaemia.
8
9

10 Sarah Brown^{1*}, Debbie Sherratt^{1*}, Samantha Hinsley¹, Louise Flanagan¹, Sadie Reed¹, Katrina
11 Walker¹, Andrew Hall¹, Guy Pratt⁴, Christina Messiou³, Matthew Jenner^{2**} and Martin Kaiser^{3**},
12 on behalf of the Myeloma UK Early Phase Clinical Trial Network
13
14

15 Corresponding authors:

16 Sarah Brown and Martin Kaiser
17
18
19
20

21 Author Details

22
23 1 Clinical Trials Research Unit, Leeds Institute of Clinical Trials Research, University of
24 Leeds, Leeds UK. Emails: d.sherratt@leeds.ac.uk; Samantha.Hinsley@glasgow.ac.uk;
25 I.m.flanagan@leeds.ac.uk; S.N.Roberts@leeds.ac.uk; K.M.Walker@leeds.ac.uk;
26 A.Hall2@leeds.ac.uk; s.brown@leeds.ac.uk
27

28 2 Southampton General Hospital, Southampton UK. Email: matthew.jenner@uhs.nhs.uk
29

30 3 Institute of Cancer Research, London UK. Email: Martin.kaiser@icr.ac.uk
31

32 4 Queen Elizabeth Hospital, Birmingham UK. Email: guy.pratt@uhb.nhs.uk
33

34 *Sarah Brown & Debbie Sherrat contributed equally as joint first authors

35 **Matthew Jenner & Martin Kaiser contributed equally as joint last authors
36
37
38

39 **Word count: 4333**
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abstract

Introduction: Multiple myeloma (MM) is a plasma cell tumour with over 5800 new cases each year in the UK. The introduction of biological therapies has improved outcomes for the majority of MM patients, but in approximately 20% of patients the tumour is characterised by genetic changes which confer a significantly poorer prognosis, generally termed high risk (HR) MM. It is important to diagnose these genetic changes early and identify more effective first-line treatment options for these patients.

Methods and analysis: The Myeloma UK *nine* OPTIMUM trial (MUK*nine*) evaluates novel treatment strategies for patients with HRMM. Patients with suspected or newly diagnosed MM, fit for intensive therapy, are offered participation in a tumour genetic screening protocol (MUK*nine a*), with primary endpoint proportion of patients with molecular screening performed within 8 weeks. Patients identified as molecularly HR are invited into the phase II, single-arm, multi-centre trial (MUK*nine b*) investigating an intensive treatment schedule comprising bortezomib, lenalidomide, daratumumab, low-dose cyclophosphamide & dexamethasone, with single high-dose melphalan and autologous stem cell transplantation (ASCT) followed by combination consolidation and maintenance therapy. MUK*nine b* primary endpoints are Minimal Residual Disease (MRD) at day 100 post-ASCT and progression-free survival. Secondary endpoints include response, safety and quality of life. The trial uses a Bayesian decision rule to determine if this treatment strategy is sufficiently active for further study.

Patients identified as not having HR disease receive standard treatment and followed up in a cohort study.

Exploratory studies include longitudinal whole body diffusion-weighted MRI for imaging MRD testing.

Ethics and dissemination: Ethics approval London South East Research Ethics Committee (Ref: 17/LO/0022, 17/LO/0023). Results of studies will be submitted for publication in a peer-reviewed journal.

Trial Registration: ISRCTN16847817, May 2017

Key Words: Newly diagnosed multiple myeloma, daratumumab, phase II, Bayesian, minimal residual disease

Article summary

Strengths and limitations of the study

- This is the first time in the UK that newly diagnosed multiple myeloma patients may be entered into a clinical trial prospectively according to their genetic risk profile
- A flexible multiple outcome, multi-stage Bayesian design is used to enable early stopping for lack of efficacy
- No concurrent control arm is included due to the availability of near concurrent historical control data from the Myeloma XI trial

For peer review only

Introduction

Multiple myeloma (MM) is a clonal disorder of plasma cells which accumulate in the bone marrow leading to cytopenias, bone resorption, renal impairment, infection and the production of a monoclonal protein¹. MM represents 1.5% of all malignant diseases, with an incidence of 9/100,000 per year accounting for around 5800 new cases each year in the UK (3000 deaths per year)². Median age at diagnosis is 69 years but 37% of patients are diagnosed before the age of 65 (including 15% <55)³. Median overall survival (OS) of younger patients is approximately 10 years⁴⁻¹¹. However approximately 20% of patients have a significantly worse prognosis, with estimated survival of <3 years and are characterised as having high risk (HR) disease^{7 12 13}. A number of genetic lesions and gene expression profiles (GEP) have been identified as associated with HR disease⁷, and molecular risk models based on these markers can be used to predict HR disease in a clinical setting. Further research is ongoing to identify additional HR markers and to better understand the mechanisms driving this tumour biology.

Unfortunately, patients with HR disease have, in terms of absolute outcome, benefitted less from the introduction of novel therapies than standard risk (SR) patients¹⁴⁻²⁰. It is important to define the optimal way to treat this group of patients given the number of available novel agents with favourable toxicity profiles allowing the use of combination therapy, consolidation and maintenance therapy. Here we describe the protocol for the MUKnine trial, a phase II study evaluating optimised combination of biological therapy in patients with newly diagnosed HRMM and plasma cell leukaemia (PCL), incorporating a screening and observational study for patients with standard risk disease. The trial has completed recruitment and is currently in follow-up.

Defining high risk disease

In a recent meta-analysis of 1,905 trial patients from the MRC Myeloma IX and NCRI Myeloma XI trials, recurrent chromosomal translocations t(4;14), t(14;16), t(14;20) and copy number aberrations (CNA) gain(1q) or del(17p) were independently associated with shorter PFS and OS. Presence of two or more such HR lesions, also termed double-hit⁷, was associated with particularly adverse outcome and increased specificity of outcome prediction considering individual lesions in isolation. The co-segregation model is exclusively based on molecular features of the tumour cell and contrasts to risk predictors which require inclusion of clinical risk markers (renal function, age, performance status) or their proxies, such as the international staging system (ISS)¹². For participants fit to receive intensive therapy, HR can thus be specifically defined by presence of two or more cytogenetically adverse lesions [t(4;14), t(14;16), t(14;20), del(1p32) gain(1q) or del(17p)].

The prognostic relevance of GEP risk signatures, in particular EMC-92, from which the SKY92 MMProfiler diagnostic assay was developed, has been demonstrated in the Myeloma IX trial dataset²¹. A recent analysis including Myeloma IX and Myeloma XI trial patients demonstrated independent association of GEP SKY92 high risk and genetic HR markers with adverse outcome in MM^{11 13 21-24}. Results suggest that both tests assay different clinically relevant qualities of HR biology. Combining GEP and double-hit genetic risk information identifies about 20-30% of patients with markedly short PFS and OS.

The exact impact of single nucleotide variants on MM risk status is still under investigation. However, very recent evidence, published after design of MUKnine, seems to confirm that structural aberrations such as translocations and CNA are the dominant markers of HRMM,

1
2
3 although detail on their assessment varies²⁵⁻²⁷. The observation of poor prognosis associated
4 with HR disease defined by such molecular criteria is consistent with clinical studies carried
5 out by other trial groups^{5-11 21 22 24 28 29}. Clearly a focused approach to improve the treatment
6 and outcome of this poor performing subgroup of MM patients is essential.
7
8
9

10 Treatment

11
12 Recent data has demonstrated efficacy of the combination of multiple novel agents in HR
13 disease³⁰. Until the molecular mechanisms contributing to HR biology can be directly targeted,
14 combinations of multiple novel agents and ongoing therapy to induce and maintain remission
15 are the most efficacious therapeutic principles³¹.
16

17 Maximising exposure to novel agents as an alternative to multi-drug cytotoxic alkylating
18 chemotherapy is hypothesised to benefit HR patients. Ongoing use of a combination of
19 biologic agents with favourable toxicity profiles can potentially minimise the chance of relapse
20 due to sustained multi-angled pressure on the MM re-populating cell pool.
21

22
23 Long-term exposure to thalidomide does not benefit HR patients^{32 33}. However, lenalidomide
24 maintenance in newly diagnosed HR patients (t(4;14) or del 17p) does have a PFS and OS
25 benefit³⁴. There is a substantial body of evidence suggesting that HR patients benefit from
26 long-term exposure to proteasome inhibition such as bortezomib³⁵⁻³⁹.
27

28 The combination of bortezomib and lenalidomide as induction and consolidation therapy is
29 safe and deliverable with a number of studies using this approach⁴⁰. Adding
30 cyclophosphamide to this triplicate approach is safe, nevertheless the lenalidomide,
31 cyclophosphamide, bortezomib and dexamethasone (RCVD) combination failed to show any
32 additional benefit to RVD (lenalidomide, bortezomib and dexamethasone) in the EVOLUTION
33 study⁴¹. However, this study evaluated all genetic risk groups and it is hypothesised that the
34 addition of low dose alkylating therapy may present an additional benefit in a HR population
35 with highly proliferative subclones.
36
37

38 Daratumumab is a monoclonal antibody that targets the CD38 molecule and has multiple
39 mechanisms of action against MM cells. It has demonstrated activity in MM as a single agent
40 and in combinations with lenalidomide and dexamethasone where it enhances the potency of
41 other drugs such as lenalidomide offering an interesting alternative to chemotherapy in MM⁴².
42 The addition of daratumumab to standard of care regimens improved outcome and combining
43 with lenalidomide or bortezomib appears to improve the poor outcomes associated with HR
44 disease^{43 44}.
45
46

47 Whilst tandem ASCT may offer prolongation of response in comparison with single procedures
48 the comparative studies reported at time of design of MUK^{nine} were undertaken in an era in
49 which novel agents were not routinely incorporated in clinical practice⁴⁵. Recent exploratory
50 analyses have suggested the potential advantage of tandem ASCT for patients with high risk
51 disease⁴⁶. Depth of response is associated with duration of response and therefore optimising
52 the induction, consolidation and maintenance approach with a single ASCT is an alternative
53 way to achieve MRD negative disease state. Melphalan has been combined with bortezomib
54 in phase II studies demonstrating safety and improvement in complete response rates
55 compared with conventional high dose melphalan conditioning⁴⁷. Although a recent report
56 stated no PFS benefit of a Velcade-augmented ASCT in a randomised trial, results for an ultra-
57 high risk group such as double-hit MM are unknown⁴⁸. The highly proliferative behaviour of
58
59
60

double-hit disease and GEP high risk provides rationale for a bridging treatment for the three months recovery period post-ASCT.

Rapid tumour evolution and associated early relapse are key characteristics of HRMM, even in patients who have achieved deep remission after ASCT⁴⁹. Maintaining multi-agent treatment intensity around and long-term after ASCT to limit size of the clonal pool as well as molecular avenues for tumour escape seems currently one of the most promising treatment strategies for HRMM, with the aim of achieving sustained deep responses in at least some patients⁵⁰. Longitudinal minimal residual disease monitoring can predict remission status with higher sensitivity than standard biochemical/protein analyses and could be of use in identifying HRMM patients benefitting most from treatment early. As bone marrow biopsy based MRD assessment may be biased due to spatial disease heterogeneity, sensitive whole body imaging can be performed in parallel to capture residual disease in other bone marrow or soft tissue areas. Whole body diffusion weighted MRI is a particularly sensitive imaging modality for MM, and standardised image acquisition and interpretation guidelines make implementation in multi-centre clinical trials feasible^{51 52}.

In line with this, the MUKnine OPTIMUM trial has been designed to evaluate the following treatment regimen in patients with HRMM, the full schedule is given in Table 1:

- **CVRDd (induction) – Cyclophosphamide, bortezomib (Velcade), lenalidomide (Revlimid), daratumumab (Darzalex), dexamethasone**
Based on the EVOLUTION trial⁴¹. Daratumumab doses are used in ongoing clinical trials⁵³.
- **Melphalan - Bortezomib ASCT**
Melphalan 200 mg/m² is standard practice in Europe for induction consolidation treatment^{54 55}. The addition of bortezomib in phase II studies demonstrated safety and improvement in complete response (CR) rates compared with conventional high dose melphalan conditioning⁴⁷. Velcade weekly monotherapy during the clinical recovery period from ASCT limits very early disease relapse in the HR population.
- **VRDd (consolidation 1) – Bortezomib, lenalidomide, daratumumab, dexamethasone**
Doses for VRd combination are based on IFM 2008-01⁴⁰ and IFM 2009-02/DFCI. Daratumumab doses are used in current clinical trials⁵³.
- **VRD (consolidation 2) – Bortezomib, lenalidomide, daratumumab**
The dose of VRD during consolidation 2 is used to minimise effects of long term corticosteroid use and risks of long term neuropathy with weekly bortezomib with no break in treatment. Utilising existing daratumumab dosing schedules it is anticipated this will be a tolerable longer term combination.
- **RD (maintenance) – Lenalidomide, daratumumab**
The dose of Lenalidomide is based on two pivotal studies^{34 56} and is the current dose used in the Myeloma XI trial²⁰. Daratumumab doses are used in current clinical trials⁵³.

Current protocols: MUKnine a, v2.0, 25/07/2018. MUKnine b, v4.0, 14/05/2020.

Methods and analysis

Aims

- To assess whether future trials in this setting are feasible and to determine risk status for participants with MM in order to deliver novel therapy to those deemed HR
- To determine whether it is possible to improve the outcome of HR patients by utilising multiple biological agents during induction, ASCT, consolidation and maintenance, and to provide evidence for the future evaluation of these high-cost interventions.

Primary Objectives

- Assess whether molecular risk-defining investigations can be turned around within 8 weeks
- Determine whether the combination of three novel agents bortezomib, lenalidomide & daratumumab in combination with low-dose cyclophosphamide & dexamethasone is sufficiently active in terms of PFS in a HR population to take forward to a phase III trial

Secondary Objectives

Secondary objectives include evaluating safety and toxicity profiles of trial treatment, evaluating additional measures of treatment activity and assessing quality of life. In patients not identified as having HR disease, secondary objectives are to summarise treatment pathways and clinical outcomes in this setting.

Exploratory Objectives

To explore novel molecular biomarkers associated with treatment activity, and evaluate germline variability/mutations, genomic instability and clonal evolution.

An exploratory imaging sub-study is included to explore the association of imaging MRD status with clinical outcomes and to assess patterns of disease distribution by whole body DW-MRI.

Trial design

The MUKnine OPTIMUM trial is comprised of two components, MUKnine *a* and MUKnine *b*, as outlined in Figure 1. MUKnine *a* is a genetic screening component, where patients with suspected symptomatic MM will be screened to determine their risk status. Patients identified as not having HR disease will receive treatment as standard of care and will have data collected on their treatment and survival. Patients who are identified as having HR disease or PCL are invited to take part in the second component, MUKnine *b*, a single arm phase II, multi-centre trial. MUKnine *b* incorporates interim assessments for futility using a Bayesian strategy for monitoring multiple outcomes proposed by Thall, Simon and Estey^{57 58} and extended by Thall and Sung⁵⁹. The trial is single arm to ensure a feasible sample size given the availability of molecularly matched individual participant data from currently running trials (Myeloma XI/XI+). This provides a body of almost concurrent control data available for the purpose of exploratory statistical comparison.

Diffusion-weighted whole body MRI (DW-MRI) is a functional method capable of detecting small-volume disease activity in MM^{60 61}, being used in standard practice at several academic

1
2
3 UK hospitals already, demonstrating excellent performance in guiding therapy on a day-to-
4 day basis. An exploratory sub-study is incorporated in MUK*nine* using DW-MRI for disease
5 distribution assessment and imaging MRD in combination with cellular (bone marrow) MRD.
6
7

8 9 Sample size

10 Recent data from the Myeloma XI trial demonstrate a median PFS for patients with HR disease
11 in the intensive pathway of 19.7 months (598 patients¹²). With a median PFS of 19-20 months
12 in the control arm, we require 92-94 patients to observe a 25% difference in median PFS
13 (corresponding to a difference of 4.8-5.0 months) in the 85% credible interval. Allowing for
14 slight changes in the actual count data, we require 95 HR patients to be registered.
15
16

17 A sample size re-estimation using individual patient data from Myeloma XI/XI+, when
18 available, allows the number of HR patients required to detect a 25% difference in median
19 PFS to be increased to 105. In order to include 105 HR patients, approximately 620 patients
20 with MM would need to be registered at diagnosis, assuming approximately 10-15% failed
21 diagnostic tests, and approximately 20% patients identified as HR.
22
23

24 The trial design includes interim analyses after every cohort of 10 MUK*nine b* participants
25 have been followed-up to 120 days post-ASCT. Until recruitment is complete, the trial could
26 be terminated early for futility on the basis of MRD status and PFS at 100 days post-ASCT.
27
28

29 Consent, eligibility, screening and registration

30
31 Participants are recruited from UK NHS hospitals. Hospital sites delivering the HR treatment
32 are approved sites within the Myeloma UK Early Phase Clinical Trials Network⁶² and patients
33 recruited from sites outside of the Network sites are referred to receive treatment, to ensure
34 sufficient patient reach to achieve target sample size. The imaging sub-study is undertaken at
35 select sites with appropriate radiology capacity. Assenting patients will provide written
36 informed consent and be registered.
37
38

39 Patients presenting who are likely to have symptomatic MM (identified by pre-tests performed
40 as standard) are approached prior to having a bone marrow biopsy for diagnosis or
41 confirmation of MM. A full list of inclusion and exclusion criteria are in Table 2. No age cut-off
42 is incorporated for transplant eligibility, as per Myeloma XI/XI+ and standard practice.
43
44

45 Patients are provided with information about the trial and if agreeable are consented for the
46 bone marrow biopsy to allow samples to be sent to central laboratories and for screening. This
47 consent allows follow up data to be collected under the MUK*nine a* protocol if the patient is
48 found not to have HR disease. Patients are registered to the trial via a web-based system
49 (provided by University of Leeds) prior to any trial-specific assessments being conducted.
50 Participants can also optionally consent to the imaging sub-study. Participants retain the right
51 to withdraw at any time without giving reasons and without their further treatment being
52 prejudiced.
53

54 Bone marrow and blood samples are taken as per standard care and sent to the Institute of
55 Cancer Research, London (ICR) by next day postal delivery for genetic molecular risk profiling.
56

57 HR status is determined by the presence of one or more of the following, based on the
58 International Myeloma Working Group (IMWG) guidelines⁶³, the Myeloma IX trial and the
59 EMC92 GEP model^{3 5 10 21 64}:
60

- 2 or more adverse lesions [t(4;14), t(14;16), t(14;20), gain(1q), del(17p), del(1p)]
- GEP – high risk score as per EMC92/SKY92 GEP model
- PCL, defined as the presence of more than $2 \times 10^9/L$ peripheral blood plasma cells or a plasmacytosis accounting for >20% of the differential white cell count

Patients identified as having HR disease are provided with a patient information sheet detailing the HR treatment schedule in MUKnine b and consented if willing to participate. A further registration documents all patients going on to HR treatment. If the patient does not wish to receive HR treatment they continue with standard treatment and data collected through the MUKnine a protocol.

For all patients at screening, bone marrow samples are sent to Haematological Malignancy Diagnostic Service (HMDS), Leeds, for MRD monitoring. Blood and urine samples are sent to Clinical Immunology Service, University of Birmingham for disease response assessments. A cell-free DNA peripheral blood sample is sent to the ICR.

Interventions

Upon first consent, treatment with standard local treatment may commence for up to 2 cycles (up to 8 weeks) whilst central molecular risk profiling is performed. Treatment may be with cyclophosphamide, thalidomide, dexamethasone (CTD), cyclophosphamide, lenalidomide, dexamethasone (CRD), bortezomib, thalidomide, dexamethasone (VTD) or cyclophosphamide, bortezomib, dexamethasone (CVD) to further take part in the MUKnine trial. This allows participants to start treatment for MM while awaiting results from risk-defining genetic investigations.

MUKnine a: Participants not identified as having HR disease continue to receive standard treatment or treatment as directed by their clinician and are followed up regularly, with information on their treatment pathway and outcomes collected.

MUKnine b: Participants identified as having HR disease and who consent to take part in the HR treatment schedule receive treatment as in Table 1. Eligibility criteria to continue treatment through each stage of ASCT, consolidation part 1 and 2, and maintenance, are detailed in Table 2.

Each individual drug in the schedule may be dose reduced if toxicity is experienced, as deemed necessary by the treating physician and in line with standard reductions used for these treatments (Table 3). Dose reductions can be made for grade 1 toxicity (e.g. neuropathy) to maximise long-term tolerability and treatment effect in this patient group. Dose reductions from pre-trial treatment may be continued at induction treatment. The majority of treatment is delivered in hospital, therefore adherence is as per protocol. Patients are reminded of treatment scheduling for oral medication at each cycle prescription.

Trial Assessments

During treatment

MUKnine a: For non-HR participants a summary of treatment received in each phase of treatment is collected. Central samples are collected at the end of any line of standard treatment for response assessment. For patients participating in the imaging study a DW-MRI

scan is performed at 100-120 days and 21 months post-ASCT, along with bone marrow, peripheral blood and urine samples for disease assessment.

MUKnine b: For HR participants, trial assessments are performed in line with the schedule of assessments in Table 4. Data are collected at each cycle of treatment and at the end of each phase of treatment, thus limiting loss to follow-up. All adverse events will be collected for all participants from the first IMP dose until 90 days after the date of the last dose of study drugs.

Central laboratory investigations include:

- Bone marrow aspirate and peripheral blood for molecular profiling:
 - MLPA or equivalent platform for copy number aberrations [del(17p), gain(1q), del(1p)]²⁸
 - RQ-PCR translocation assay or equivalent tool for prediction of HR translocations [t(4;14), t(14;16) and t(14;20)]⁶⁵
 - Gene expression profiling based on Affymetrix HG-U133 Plus 2.0 or equivalent platform with risk profile determined as per EMC92 model²³
 - Exploratory molecular analyses to identify potentially targetable mutations
 - Whole exome or whole genome next-generation sequencing
 - Gene expression profiling (GEP)
 - Epigenetic analyses
 - Germline variant analysis
- Bone marrow aspirate for MRD analyses
- Peripheral blood for disease assessment
 - Disease parameters, e.g. paraprotein, for serum response assessments
 - Beta-2-microglobulin
 - Albumin

Quality of Life questionnaires, EQ-5D, QLQ-C30 and QLQ-MY20, are collected from all participants at baseline, and for participants who go on to HR treatment these are completed at:

- End of induction treatment
- 100 days post-ASCT then 3-monthly thereafter until disease progression.

Follow-up

Upon completion of treatment, patients are followed-up at 3 months, and then six-monthly during standard of care visits, until second disease progression, death or withdrawal. Assessment via standard of care visits promotes participant retention and complete follow-up.

Imaging assessments

All patients participating in the DW-MRI sub-study have whole body DW-MRI scan performed at baseline, 100-120 days post-ASCT and at end of consolidation part 2.

Outcomes

Primary endpoint

MUKnine a:

The proportion of patients with molecular risk-defining investigations performed within 8 weeks.

MUKnine b:

The primary endpoints to determine whether to terminate the trial early for futility are

Minimal residual disease at 100 days post-ASCT

Progression-free survival at 100 days post-ASCT

The primary endpoint to assess efficacy of HR treatment if the trial is not stopped early for futility is PFS at 18 months post-registration to screening.

Secondary Endpoints

MUKnine a: recruitment rates; PFS; OS; Second PFS (PFS2); treatment received; overall response;

MUKnine b:

Safety and toxicity (adverse reactions (ARs), serious adverse events (SAEs), serious adverse reactions (SARs) and suspected unexpected serious adverse reactions (SUSARs) graded by common terminology criteria for adverse events (CTCAE) v5.0)

MRD at the end of induction therapy, and post- consolidation part 2

OS

Maximum and overall response at the end of induction therapy, 100 days post-ASCT and postconsolidation part 2

Time to progression and time to maximum response

PFS2

Overall treatment benefit and clinician assessment of treatment benefit at the end of induction therapy and 100 days post-ASCT

Quality of life as assessed by the EQ-5D, EORTC QLQ-C30 and EORTC QLQ-MY20

Treatment compliance

Exploratory Endpoints

Genomic instability, mutation rates and clonal evolution

Imaging sub-study

PFS; OS; Response; Patterns of disease distribution and discreet “3D phenotypes”

Statistical analysis

The *MUKnine b* trial is designed using a Bayesian approach to enable assessment of multiple outcomes and incorporating multiple interim analyses.

1
2
3 The experimental treatment will be evaluated on an ongoing basis based on assessment of
4 MRD status and PFS. Interim assessments are made after cohorts of 10 participants have
5 been followed up to 100-120 days post-ASCT, and data reviewed by an independent Data
6 Monitoring and Ethics Committee (DMEC). The trial may be terminated early for futility on the
7 basis of MRD status and PFS at 100-120 days post-ASCT, using initial pre-defined stopping
8 boundaries based on Myeloma IX data. Following updated prior information becoming
9 available from Myeloma XI/XI+, these stopping boundaries were re-calculated to provide
10 updated decision criteria.
11
12

13 If the trial is not terminated early, up to 105 newly diagnosed patients with molecular HR
14 disease will be registered to treatment. With the availability of molecularly matched individual
15 participant data from currently running trials (Myeloma XI/XI+) a body of almost concurrent
16 control data is available to use for the purpose of exploratory statistical comparison.
17
18

19 The experimental treatment arm will be compared to control in terms of PFS at 18 months
20 post-registration to screening, expressed as a binary outcome, within the Bayesian framework.
21 Further analyses of PFS at 18 months will be performed outside of the Bayesian framework
22 using Kaplan-Meier estimation.
23

24 MUK*nine* a endpoints, and secondary and exploratory endpoints will be analysed using
25 summary statistics alongside confidence intervals where appropriate. All analyses are fully
26 detailed in a statistical analysis plan prior to being undertaken. Full statistical analysis for
27 MUK*nine* is provided in Supplementary File 1, and discussed in the MUK*nine* statistical
28 methods paper (in preparation).
29
30

31 Trial conduct

32
33 Data are collected via electronic case report forms. Site monitoring of source data is performed
34 by CTRU following the trial monitoring plan. The trial is conducted in accordance with the
35 principles of Good Clinical Practice (GCP) and in line with the relevant Research Governance
36 Framework within the UK through adherence with University of Leeds CTRU standard
37 operating procedures. All information collected during the course of the trial will be kept strictly
38 confidential. Information will be held securely on paper and electronically at the CTRU. An
39 independent DMEC reviews safety data on a regular basis to identify any safety concerns or
40 trends. An independent Trial Steering Committee periodically reviews safety data and
41 discusses recommendations made by the DMEC.
42
43
44
45

46 Statement of indemnity

47
48 This trial is sponsored by The University of Leeds and the University of Leeds will be liable for
49 negligent harm caused by the design of the trial. The NHS has a duty of care to participants
50 treated, whether or not the participant is taking part in a clinical trial, and the NHS remains
51 liable for clinical negligence and other negligent harm to participants under this duty of care.
52

53 As this is a clinician-led trial, there are no arrangements for no-fault compensation. As this is
54 a clinician-led trial, there are no arrangements for no-fault compensation; however, usual
55 product liability will be covered by the manufacturer under the Consumer Protection Act 1987.
56
57
58

59 Patient and Public Involvement

1
2
3 Patients were involved in review and development of trial design, protocol and patient
4 information sheet (model consent form provided in Supplementary File 2).
5
6

7 **Ethics and dissemination**

8
9 The trial has national research ethics approval from the NHS National Research Ethics
10 Service, London South East Research Ethics Committee (Ref: 17/LO/0022, 17/LO/0023). All
11 patients provide written informed consent prior to take part in the trial at the hospital site where
12 they are recruited. Any required protocol amendments will be submitted to ethics and MHRA
13 (as appropriate), and will be implemented at the relevant sites once approved. Information on
14 amendments will be reported to the DMEC and TSC.
15
16

17
18 A manuscript with results of the MUK*nine b* study will be published in a peer-reviewed journal.
19 Separate manuscripts will be written for results of MUK*nine a* and each of the exploratory
20 objectives; these will also be submitted for publication in peer-reviewed journals. Credit for the
21 main results will be given to all those who have collaborated in the trial, through authorship
22 and contributorship. Uniform requirements for authorship for manuscripts submitted to medical
23 journals will guide authorship decisions. Professional writers are not intended to be used.
24 Upon publication of the final long-term results of the study, requests for use of data may be
25 made to the CTRU and will be reviewed by the Trial Management Group.
26
27
28
29

30 **Discussion**

31 This is the first time in the UK genetic risk has been used prospectively in MM to identify
32 participants to be treated in an academically-led clinical trial and select treatment based solely
33 on this. It is hoped this trial will bring improved survival and longer term disease control for
34 patients with HRMM in the future by providing an intensive treatment regimen specifically
35 targeted at this difficult to treat disease sub-group. In addition, the trial will provide important
36 evidence regarding feasibility of multi-centre molecular-risk stratified trials in MM at the point
37 of diagnosis, using central molecular tumour investigations.
38
39

40 Intensive treatment in HR patients has been used outside the UK with some promising results
41 but access to drugs in the UK has been challenging. This trial is designed to work within the
42 UK NHS system and provide the best treatment for HR patients. The availability of novel
43 targeted molecular therapies helps in treating the highly heterogeneous disease of MM.
44 Ultimately data generated through this trial aims to support the case for access to combination
45 therapies of expensive agents to patient subgroups with a high unmet need such as HR
46 disease.
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Abbreviations

ADCC	antibody dependant cytotoxicity
ADCP	antibody dependant cell phagocytosis
ALT	Alanine transaminase
ASCT	Autologous stem cell transplant
AST	Aspartate transaminase
CRD	Cyclophosphamide, Lenalidomide (Revlimid®), Dexamethasone
CTCAE	Common Terminology Criteria for Adverse Events
CTD	Cyclophosphamide, thalidomide, dexamethasone
DMEC	Data Monitoring and Ethics Committee
DW-MRI	Diffuse weighted magnetic resonance imaging
ECG	Electrocardiogram
ECOG	Eastern Cooperative Oncology Group performance status
¹⁸ F-FDG PET-CT	¹⁸ F-fluorodeoxyglucose - Positron emission tomography - computerised tomography
GEP	Gene expression profiling
HDM-V	High dose Melphalan with Velcade (bortezomib)
HR	High risk
IMiD	Immunomodulatory drugs
IMWG	International Myeloma Working Group
KCRD	Carfilzomib, Cyclophosphamide, Lenalidomide, Dexamethasone
MM	Multiple Myeloma
MRD	Minimal residual disease
MRI	Magnetic resonance imaging
OS	Overall survival
PCL	Plasma Cell Leukaemia
PFS	Progression-free survival
PFS2	Second progression-free survival
RCVD	Lenalidomide, Cyclophosphamide, Bortezomib, Dexamethasone
RVD	Lenalidomide, Bortezomib, Dexamethasone
RD	Lenalidomide, Dexamethasone
VRD	Bortezomib, Lenalidomide, Daratumumab
VRDd	Bortezomib, Lenalidomide, Daratumumab, dexamethasone

Competing interests

SH, DS, LF, SR, AH, KW and SB declare they have no competing interests. MK has served as a consultant and received honoraria from Celgene, Takeda, Amgen, Chugai, BMS, AbbVie and Janssen, and has received research funding from Celgene. GP served as a consultant and received honoraria from Janssen Oncology, Celgene, Amgen, Takeda, Gilead Sciences and Binding Site. MJ has served as a consultant and received honoraria from Celgene, Janssen, Takeda, Amgen, AbbVie and Novartis.

Funding

This trial is funded by Janssen, Celgene and Myeloma UK. Award/Grant number is not applicable.

Author Contributions

Sarah Brown & Debbie Sherrat contributed equally as joint first authors. Matthew Jenner & Martin Kaiser contributed equally as joint last authors

MJ, MK, GP, SB, LF, SH and DS designed the trial. SH, AH, KW and SB developed the statistical analysis plan and are responsible for the ongoing statistical monitoring, analysis and interpretation of data. DS, SH, MK and MJ wrote the manuscript. MJ, MK, CM and GP perform

1
2
3 the research and collect data. DS, LF and SR perform trial and data management. All authors
4 reviewed and approved the final manuscript.
5
6
7

8 **Acknowledgments**

9
10 Janssen and Celgene provided funding to Myeloma UK and University of Leeds who in turn
11 fund the MUK*nine* trial. Janssen and Celgene approved the design of the study but have no
12 input in the collection, analysis or interpretation of data as this is a fully academically
13 sponsored trial. Contact details of the sponsor, the University of Leeds, are accessible via the
14 trial registration. Through the Myeloma UK Clinical Trials Network, Myeloma UK were involved
15 in the study design and are actively involved in the collection and interpretation of data, as
16 well as in the review of manuscripts arising from the study publishing trial outcomes.
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Table 1: Treatment Schedule

Induction Treatment			
Regimen: CVRDd to maximum response (or a maximum of 6 cycles of bortezomib)			Cycle duration: 21 days
Drug	Dose	Route	Days
Cyclophosphamide	500 mg	PO	1 and 8
Bortezomib	1.3 mg/m ²	SC	1, 4, 8, 11
Lenalidomide	25 mg	PO	1 – 14
Daratumumab	16 mg/kg (actual body weight)	IV	1, 8, 15** (Cycles 1 and 2)
			1 only (Cycle 3 onwards)
Dexamethasone*	20 mg – 40 mg	PO/IV	1, 4, 8, 11
Autologous Stem Cell Transplant			
Cyclophosphamide and GCSF mobilisation is recommended			
Regimen: Bortezomib HD-MEL+ASCT			
Drug	Dose	Route	Days
Melphalan	200 mg/m ²	IV	-1
Bortezomib	1.3mg/m ²	SC	-1, (8-18 hours post Melphalan)
Autologous stem cell return		IV	0
Bortezomib	1.3mg/m ²	SC	+5, +14, then weekly to consolidation 1
Consolidation Treatment 1			
To begin between 100 -120 days post ASCT			
Regimen: VRDd x 6 cycles*			Cycle duration: 28 days
Drug	Dose	Route	Days
Bortezomib	1.3mg/m ²	SC	1, 8,15, 22
Lenalidomide	25mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1
Dexamethasone*	20mg-40mg	PO/IV	1, 8,15**, 22
Consolidation Treatment 2			
Regimen: VRD x 12 cycles*			Cycle duration: 28 days
Drug	Dose	Route	Days
Bortezomib	1.3mg/m ²	SC	1, 8,15
Lenalidomide	25mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1

Maintenance Treatment			
Regimen: RD until progression		Cycle duration: 28 days	
Drug	Dose	Route	Days
Lenalidomide	10mg	PO	1 - 21
Daratumumab	16mg/kg (actual body weight)	IV	1

* On days where participants receive dexamethasone 40mg at site (i.e. pre Daratumumab infusion), dexamethasone must not be self-administered at home too.

** On day 15 participants will receive pre-med as per SPC (e.g. methylprednisolone)

Table 2: Eligibility Criteria for trial entry and continuing treatment through each stage

Inclusion Criteria	Exclusion Criteria
<p>Screening</p> <ul style="list-style-type: none"> Undergoing bone marrow investigation due to suspected symptomatic multiple myeloma or plasma cell leukaemia (PCL) or Participants with biopsy-confirmed symptomatic multiple myeloma, willing to undergo a further study bone marrow biopsy for molecular profiling. Participants previously screened but found not to have symptomatic multiple myeloma but now have suspected symptomatic multiple myeloma may be re-screened Aged 18 years or over Fit for intensive chemotherapy and autologous stem cell transplant (at clinician's discretion) Eastern Cooperative Oncology Group (ECOG) score ≤ 2 	<ul style="list-style-type: none"> Confirmed solitary bone/solitary extramedullary plasmacytoma. Primary diagnosis of Waldenstrom's Disease. Monoclonal gammopathy of undetermined significance or smouldering multiple myeloma unless progression to symptomatic multiple myeloma is highly suspected or confirmed Received therapy for multiple myeloma Prior or concurrent invasive malignancies Any uncontrolled or severe cardiovascular or pulmonary disease Grade 2 or greater peripheral neuropathy (per NCI-CTCAEv4.0) Known/underlying medical conditions that, in the investigator's opinion, would make the administration of the study drug hazardous Any clinically significant cardiac disease Known chronic obstructive pulmonary disease (COPD) Known to be seropositive for history of human immunodeficiency virus (HIV) or known to have active hepatitis B or hepatitis C. Any known allergies, hypersensitivity, or intolerance to corticosteroids, monoclonal antibodies or human proteins, or their excipients or known sensitivity to mammalian-derived products. Clinically significant allergies or intolerance to cyclophosphamide, lenalidomide, bortezomib, daratumumab or dexamethasone. Previous treatment with daratumumab or any other anti-CD38 therapies. Participants with contraindication to thromboprophylaxis. Participants with POEMS syndrome Any concurrent medical or psychiatric condition or disease

	<ul style="list-style-type: none"> • Known or suspected of not being able to comply with the study protocol • Participant is a woman who is pregnant, or breast-feeding, or planning to become pregnant while enrolled in this trial or within at least 6 months after the last dose of trial treatment. Or, participant is a man who plans to father a child while taking part in this trial or within at least 6 months after the last dose of trial treatment. • Major surgery within 2 weeks before treatment protocol registration or has not fully recovered from surgery, or has surgery planned during the time the participant is expected to participate in the study. Kyphoplasty or vertebroplasty is not considered major surgery. • Received an investigational drug (including investigational vaccines) or used an invasive investigational medical device within 4 weeks before treatment protocol registration or is currently enrolled in an interventional investigational study. <p><u>Imaging sub-study</u></p> <p>Only those taking part in the imaging sub study have these exclusions:</p> <ul style="list-style-type: none"> • MRI incompatible metal implant • Claustrophobia
<p>Treatment</p> <ul style="list-style-type: none"> • Confirmation of High Risk status from ICR. Participants with confirmed plasma cell leukaemia with >20% circulating plasma cells do not need confirmation of HR status from ICR to proceed to treatment. • Confirmation of receipt of baseline bone marrow at HMDS and, blood and urine samples at the University of Birmingham • Previously untreated participants, although participants may have received up to 2 cycles of CTD, CVD, CRD or VTD pre-trial induction chemotherapy while awaiting the results of the laboratory analysis. • Measurable disease before starting standard treatment <ul style="list-style-type: none"> ◦ Paraprotein $\geq 5\text{g/L}$ or $\geq 0.5\text{ g/L}$ for IgD subtypes OR Serum free kappa or lambda light chains $\geq 100\text{ mg/L}$ with abnormal ratio (for light chain only myeloma) OR Urinary Bence Jones protein $\geq 200\text{ mg/24h}$. • Non-measurable participants providing they accept a 3 monthly bone marrow during induction and a 6 monthly bone marrow assessment during consolidation and maintenance. • Fit for intensive chemotherapy and autologous stem cell transplant (at clinician's discretion). • Eastern Cooperative Oncology Group (ECOG) Performance Status ≤ 2. 	<ul style="list-style-type: none"> • Solitary bone/solitary extramedullary plasmacytoma. • Primary diagnosis of amyloidosis, monoclonal gammopathy of undetermined significance or smouldering multiple myeloma or Waldenstrom's Disease. • Prior or concurrent invasive malignancies • Known/underlying medical conditions that, in the investigator's opinion, would make the administration of the study drug hazardous • Any clinically significant cardiac disease • Known chronic obstructive pulmonary disease (COPD) • Known to be seropositive for history of human immunodeficiency virus (HIV) or known to have active hepatitis B or hepatitis C. • Any known allergies, hypersensitivity, or intolerance to corticosteroids, monoclonal antibodies or human proteins, or their excipients or known sensitivity to mammalian-derived products. • Clinically significant allergies or known intolerance to cyclophosphamide, lenalidomide, bortezomib, daratumumab or dexamethasone. • Previous treatment with daratumumab or any other anti-CD38 therapies. • Participants with contraindication to thromboprophylaxis.

<ul style="list-style-type: none"> • The Celgene Pregnancy Prevention Plan must be followed and participants must agree to comply with this: <ul style="list-style-type: none"> • Females of childbearing potential (FCBP) must agree to utilise two reliable forms of contraception simultaneously or practice complete abstinence for at least for 28 days prior to starting trial treatment, during the trial and for at least 28 days after trial treatment discontinuation, and even in case of dose interruption, and must agree to regular pregnancy testing during this timeframe. • Males must agree to use a latex condom during any sexual contact with FCBP during the trial, including during dose interruptions and for 28 days following discontinuation from this trial even if he has undergone a successful vasectomy • Males must also agree to refrain from donating semen or sperm while on trial treatment including during any dose interruptions and for at least 6 months after discontinuation from this trial • All participants must agree to refrain from donating blood while on trial drug including during dose interruptions and for 28 days after discontinuation from this trial. • Laboratory Results <ul style="list-style-type: none"> • Calculated creatinine clearance $\geq 30\text{mL/min}$ (using Cockcroft-Gault formula). • ALT or AST ≤ 2.5 times upper limit of normal (ULN). • Bilirubin $\leq 2.0 \times \text{ULN}$, except in participants with congenital bilirubinemia, such as Gilbert syndrome (direct bilirubin ≤ 2.0 times ULN) • Platelet count $\geq 75 \times 10^9/\text{L}$. ($\geq 50 \times 10^9/\text{L}$ if multiple myeloma involvement in the bone marrow is $>50\%$). Platelet support is permitted. • Absolute neutrophil count (ANC) $\geq 1.0 \times 10^9/\text{L}$. Growth factor support is permitted. • Haemoglobin $\geq 80 \text{ g/L}$. (Participants may be receiving red blood cell (RBC) transfusions in accordance with institutional guidelines. • Corrected serum calcium $\leq 3.5 \text{ mmol/L}$ 	<ul style="list-style-type: none"> • Participants with POEMS syndrome • Any concurrent medical or psychiatric condition or disease • Known or suspected of not being able to comply with the study protocol • Participant is a woman who is pregnant, or breast-feeding, or planning to become pregnant while enrolled in this trial or within at least 6 months after the last dose of trial treatment. Or, participant is a man who plans to father a child while taking part in this trial or within at least 6 months after the last dose of trial treatment. • Major surgery within 2 weeks before treatment protocol registration or has not fully recovered from surgery, or has surgery planned during the time the participant is expected to participate in the study. Kyphoplasty or vertebroplasty is not considered major surgery. • Received an investigational drug (including investigational vaccines) or used an invasive investigational medical device within 4 weeks before treatment protocol registration or is currently enrolled in an interventional investigational study. <p><u>Imaging sub-study</u></p> <p>Only those taking part in the imaging sub study have these exclusions:</p> <ul style="list-style-type: none"> • MRI incompatible metal implant • Claustrophobia • Not received a DW-MRI at baseline
<p>Autologous Stem Cell Transplant</p> <ul style="list-style-type: none"> • Minimum stem cell harvest of 2×10^6 CD34+ cells/kg body weight. • Received a minimum of 4, unless CR has been achieved with a lesser number, or a maximum of 6 Induction (CVRDd) cycles (including standard treatment). • Achieved a response of stable disease (SD_ or better). • Dose modifications of any or all individual drugs within induction is permitted including complete stop of no more than one agent due to toxicity as 	<ul style="list-style-type: none"> • Participants that have progressive disease.

long as the required number of cycles have been received	
<p>Consolidation Part 1</p> <ul style="list-style-type: none"> • Undergone autologous transplant with HDM-V conditioning (Participants must have received a minimum of 100 mg/m² Melphalan in order to proceed with consolidation). • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modifications because of toxicity including complete stop of weekly bortezomib is permitted 	<ul style="list-style-type: none"> • Participants that have progressive disease.
<p>Consolidation Part 2</p> <ul style="list-style-type: none"> • Received 6 cycles of Consolidation Part 1 (VRDd) • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modification of any or all of the individual drugs in consolidation part 1 is permitted including complete stop of no more than one agent because of toxicity as long as the required number of cycles have been received. 	<ul style="list-style-type: none"> • Participants that have progressive disease.
<p>Maintenance</p> <ul style="list-style-type: none"> • Received 12 cycles of Consolidation Part 2 (VRD). • Neutrophils $\geq 1.0 \times 10^9/L$. Growth factor support is permitted. • Platelet count $\geq 75 \times 10^9/L$. Platelet support is permitted. • Dose modification of any or all of the individual drugs in consolidation part 2 is permitted including complete stop of no more than one agent because of toxicity as long as the required number of cycles have been received. 	<ul style="list-style-type: none"> • Participants that have progressive disease.

Table 3: Dose Modifications

Cyclophosphamide					
Modifications are at the discretion of the investigator					
<i>Renal Impairment</i> - A dose reduction of 50% for creatinine clearance of 10ml/min is recommended					
<i>Hepatic Impairment</i> - A dose reduction to 350mg is recommended with a serum bilirubin of >2.5 times the upper limit of normal					
Bortezomib					
Induction dose reductions					
Regimen: First dose reduction CVRDd				Cycle duration: 21 days	
Drug	Dose	Route	Days		
Cyclophosphamide	500 mg	PO	1 and 8		
Bortezomib	1.3 mg/m ²	SC	1, 8, 15		
Lenalidomide	25 mg	PO	1 – 14		
Daratumumab	16 mg/kg (actual body weight)	IV	1, 8, 15 (Cycles 1 and 2)		
			1 only (Cycle 3 onwards)		
Dexamethasone*	20 mg – 40 mg	PO/IV	1, 8, 15		
Post induction dose reductions					
Bortezomib schedule	Dose levels				
	0	-1	-2	-3	-4
Twice weekly schedules	1.3 mg/m ² d 1, 4, 8, 11	1.3 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 8, 15	1.3 mg/m ² d 1, 15	Stop
Once weekly schedules	1.3 mg/m ² d 1, 8, 15, (22)	1.0 mg/m ² d 1, 8, 15 (22)	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
Consolidation 1	1.3 mg/m ² d 1, 8, 15, 22	1.0 mg/m ² d 1, 8, 15, 22	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
Consolidation 2	1.3 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 8, 15	1.0 mg/m ² d 1, 15	0.7 mg/m ² d 1, 15,	Stop
<i>Neuropathy</i> - CTCAE Grade 1 with pain or grade 2- withhold bortezomib until returns to baseline. Dose reduce 1 level; CTCAE Grade 2 with pain or grade 3- withhold bortezomib until returns to baseline. Dose reduce 2 levels; CTCAE Grade 4 – discontinue treatment					
<i>Renal impairment</i> – dose reduce at the discretion of the clinician					
<i>Hepatic impairment</i> – moderate or severe impairment (>1.5 – 3x ULN) should start on a reduced dose of 0.7mg/m ² during the first cycle of treatment and dose escalate to 1.0 mg/m ² or dose reduce to 0.5mg/m ² may be considered					
<i>Grade 3 Non haematological toxicity</i> – withhold until symptoms of toxicity resolve and reduce one dose.					
<i>Grade 4 haematological toxicity</i> – withhold until symptoms of toxicity resolve and reduce one dose. Support may be given.					

Lenalidomide schedule	Dose levels					
	0	-1	-2	-3	-4	-5
	25mg	20mg	15mg	10mg	5mg	2.5 mg
<p><i>Thrombocytopenia</i> - $<25 \times 10^9/L$ stop lenalidomide for the remainder of the cycle. Return to $\geq 50 \times 10^9/L$ decrease by 1 dose level to resume the next cycle.</p> <p><i>Neutropenia</i> – first fall to $<0.5 \times 10^9/L$ omit lenalidomide until a return to $\geq 0.5 \times 10^9/L$ when neutropenia is the only toxicity. Resume lenalidomide at one dose lower. For each subsequent drop to $\geq 0.5 \times 10^9/L$ omit lenalidomide, resume lenalidomide decreased by 1 dose level at the next cycle.</p> <p><i>Renal impairment</i> – 30- 50 mL/min 10mg daily; < 30 mL/min, not requiring dialysis 7.5mg daily or 15mg every other day; < 30 mL/min, requiring dialysis 5mg daily administered following dialysis</p> <p><i>Other non haematological toxicities:</i> CTCAE grade 3 & 4 related to lenalidomide should be stopped and started 1 dose lower when toxicity has resolved to grade 2 at clinicians discretion. Rash – interrupt or discontinue for grade 2 or 3. Grade 4 rash discontinue including angioedema, exfoliative or bullous rash or Steven Johnson syndrome or toxic epidermal necrosis.</p>						
Daratumumab schedule	Frequency	Dose held	Dosing restart			
Induction cycles 1 and 2	Weekly	>3 days	Next planned weekly dose			
Induction cycles 3 – 6	Monthly	>1 week	Next planned weekly dose			
Consolidation 1, Consolidation 2, Maintenance	Monthly	>2 weeks	Next planned weekly dose			
<p>Follow the daratumumab SmPC. The daratumumab infusion must be withheld to allow for recovery from toxicity ONLY where any of the following criteria are met and the event cannot be ascribed to lenalidomide or cyclophosphamide.</p> <ul style="list-style-type: none"> • Grade 4 thrombocytopenia or Grade 3 thrombocytopenia with bleeding. • Grade 4 neutropenia, if this is the second occurrence despite growth factor support. • Febrile neutropenia of any grade. • Neutropenia with infection, of any grade. • Grade 3 or higher non-haematological toxicities with the following exceptions: <ul style="list-style-type: none"> ○ Grade 3 nausea that responds to antiemetic treatment within 8 days. ○ Grade 3 vomiting that responds to antiemetic treatment within 8 days. ○ Grade 3 diarrhoea that responds to anti-diarrhoeal treatment within 8 days. ○ Grade 3 fatigue that was present at baseline or that lasts for <8 days after the previous administration of daratumumab. ○ Grade 3 asthenia that was present at baseline or that lasts for <8 days after the previous administration of daratumumab. 						
Dexamethasone						
<p>Occasionally patients will not be able to tolerate because of corticosteroid effects. Dose reductions from 40 to 20mg daily. Further dose reductions to 10mg daily is acceptable followed by the omission of dexamethasone</p> <p>If the bortezomib schedule changes, dexamethasone should change in line with it.</p>						
Melphalan						
<p>Dose may be adjusted based on performance status and clinical judgment in discussion with the Chief Investigator</p> <p>GFR measured by Cockcroft & Gault formula or EDTA - >50ml/min 200mg/m²; 30-50ml/min 140mg/m²; <30ml/min 100mg/m²</p>						

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For peer review only

Table 4: Trial Assessments

Investigations	All Patients	Non HR Patients			HR Patients							
	All Screening participants	Prior to any new line of treatment	Post any line of treatment	First and second disease progression	Before starting MUKnine treatment ¹	Prior to each cycle of induction treatment CVRDd ²	End of induction treatment	Autologous stem cell transplant ⁵	100-120 days post transplant	Prior to each cycle of consolidation part 1 (VRDd), consolidation part 2 (VRD) and maintenance treatment (RD)	End of consolidation part 1 (VRDd), consolidation part 2 (VRD) and maintenance treatment (RD)	First and second disease progression
Consent	X				X							
Medical history	X				X							
Symptom-directed physical exam (including weight, ECOG)	X	X	X		X	X	X	X	X	X	X	
Haematology & biochemistry test	X	X	X		X	X	X	X	X	X	X	
Disease assessment ³	X	X	X	X	X	X	X	X	X	X	X	X
DW-MRI Imaging ⁷	X								X		X (Part 2 only)	
ECG					X							
Pregnancy testing as required					X	X	X		X	X	X	
Participant questionnaires	X						X		X	X ⁶	X	
Details of treatment			X			X	X	X	X	X	X	
Clinical assessment of treatment benefit							X		X		X	
Central Laboratory Samples												
Bone marrow aspirate	X			X			X		X		X	X
Peripheral blood	X	X		X		X ⁴	X		X	X ^{4,6}	X	X
Urine sample	X	X		X		X ⁴	X		X	X ^{4,6}	X	X

1 Treatment must start within 14 days of registration to MUKnine treatment

2 All assessments must be performed within 72 hours prior to day 1 of each cycle of treatment

1
2
3 3 Response assessments must be made in line with the IMWG criteria

4 4 Cycle 1 day 1 only

5
6 5 Autologous stem cell transplant will be performed as per local practice with local monitoring of adverse events and haematology tests. Participants will be given weekly
7 bortezomib until 100-120 days post transplant, the assessments will be performed monthly during this time for the trial.

8 6 3 monthly during treatment

9
10 7 if site and participant taking part in the imaging sub-study

11
12
13 Figure 1: MUKnine OPTIMUM trial design

14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

For peer review only

References

1. Kyle RA, Rajkumar SV. Criteria for diagnosis, staging, risk stratification and response assessment of multiple myeloma. *Leukemia* 2009;23(1):3-9. doi: leu2008291 [pii];10.1038/leu.2008.291 [doi]
2. Myeloma Statistics Cancer Research UK. <https://www.cancerresearchuk.org/health-professional/cancer-statistics/statistics-by-cancer-type/myeloma#heading-Two> 2015
3. Ries LAG YJ, Keel GE, Eisner MP, Lin YD, Horner M-J SEER Survival Monograph: Cancer Survival Among Adults U.S. SEER Program, 1988-2001, Patient and Tumor Characteristics.
4. Avet-Loiseau H. Ultra high-risk myeloma. *Hematology*;2010:489-93.
5. Avet-Loiseau H, Attal M, Campion L, et al. Long-term analysis of the IFM 99 trials for myeloma: cytogenetic abnormalities [t(4;14), del(17p), 1q gains] play a major role in defining long-term survival. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2012;30(16):1949-52. doi: 10.1200/jco.2011.36.5726 [published Online First: 2012/05/02]
6. Avet-Loiseau H, Durie BG, Cavo M, et al. Combining fluorescent in situ hybridization data with ISS staging improves risk assessment in myeloma: an International Myeloma Working Group collaborative project. *Leukemia* 2013;27(3):711-7. doi: 10.1038/leu.2012.282
7. Boyd KD, Ross FM, Chiecchio L, et al. A novel prognostic model in myeloma based on co-segregating adverse FISH lesions and the ISS: analysis of patients treated in the MRC Myeloma IX trial. *Leukemia* 2012;26(2):349-55. doi: 10.1038/leu.2011.204 [published Online First: 2011/08/13]
8. Chng WJ, Kuehl WM, Bergsagel PL, et al. Translocation t(4;14) retains prognostic significance even in the setting of high-risk molecular signature. *Leukemia* 2008;22(2):459-61. doi: 10.1038/sj.leu.2404934 [published Online First: 2007/09/07]
9. Morgan GJ, Davies FE, Gregory WM, et al. First-line treatment with zoledronic acid as compared with clodronic acid in multiple myeloma (MRC Myeloma IX): a randomised controlled trial. *Lancet* 2010;376(9757):1989-99. doi: 10.1016/s0140-6736(10)62051-x [published Online First: 2010/12/07]
10. Morgan GJ, Davies FE, Gregory WM, et al. Cyclophosphamide, thalidomide, and dexamethasone (CTD) as initial therapy for patients with multiple myeloma unsuitable for autologous transplantation. *Blood* 2011;118(5):1231-8. doi: 10.1182/blood-2011-02-338665
11. Shaughnessy JD, Jr., Zhan F, Burington BE, et al. A validated gene expression model of high-risk multiple myeloma is defined by deregulated expression of genes mapping to chromosome 1. *Blood* 2007;109(6):2276-84. doi: 10.1182/blood-2006-07-038430 [published Online First: 2006/11/16]
12. Shah V, Sherborne AL, Walker BA, et al. Prediction of outcome in newly diagnosed myeloma: a meta-analysis of the molecular profiles of 1905 trial patients. *Leukemia* 2017 doi: 10.1038/leu.2017.179

13. Shah V, Sherborne AL, Johnson DC, et al. Predicting ultrahigh risk multiple myeloma by molecular profiling: an analysis of newly diagnosed transplant eligible myeloma XI trial patients. *Leukemia* 2020 doi: 10.1038/s41375-020-0750-z
14. Dimopoulos MA, Oriol A, Nahi H, et al. Daratumumab, Lenalidomide, and Dexamethasone for Multiple Myeloma. *The New England journal of medicine* 2016;375(14):1319-31. doi: 10.1056/NEJMoa1607751 [published Online First: 2016/10/06]
15. Palumbo A, Chanan-Khan A, Weisel K, et al. Daratumumab, Bortezomib, and Dexamethasone for Multiple Myeloma. *The New England journal of medicine* 2016;375(8):754-66. doi: 10.1056/NEJMoa1606038 [published Online First: 2016/08/25]
16. Dimopoulos MA, Moreau P, Palumbo A, et al. Carfilzomib and dexamethasone versus bortezomib and dexamethasone for patients with relapsed or refractory multiple myeloma (ENDEAVOR): a randomised, phase 3, open-label, multicentre study. *The lancet oncology* 2016;17(1):27-38. doi: 10.1016/s1470-2045(15)00464-7 [published Online First: 2015/12/17]
17. Stewart AK. Carfilzomib for the treatment of patients with relapsed and/or refractory multiple myeloma. *Future Oncology*;11(15):2121-36.
18. Shah V, Sherborne AL, Walker BA, et al. Prediction of outcome in newly diagnosed myeloma: a meta-analysis of the molecular profiles of 1905 trial patients. *Leukemia* 2018;32(1):102-10. doi: 10.1038/leu.2017.179 [published Online First: 2017/06/07]
19. Chng WJ, Dispenzieri A, Chim CS, et al. IMWG consensus on risk stratification in multiple myeloma. *Leukemia* 2014;28(2):269-77. doi: 10.1038/leu.2013.247 [published Online First: 2013/08/27]
20. Jackson GH, Davies FE, Pawlyn C, et al. Lenalidomide maintenance versus observation for patients with newly diagnosed multiple myeloma (Myeloma XI): a multicentre, open-label, randomised, phase 3 trial. *The lancet oncology* 2019;20(1):57-73. doi: 10.1016/s1470-2045(18)30687-9 [published Online First: 2018/12/19]
21. Kuiper R, Broyl A, de Knecht Y, et al. A gene expression signature for high-risk multiple myeloma. *Leukemia* 2012;26(11):2406-13. doi: 10.1038/leu.2012.127
22. Decaux O, Lode L, Magrangeas F, et al. Prediction of survival in multiple myeloma based on gene expression profiles reveals cell cycle and chromosomal instability signatures in high-risk patients and hyperdiploid signatures in low-risk patients: a study of the Intergroupe Francophone du Myelome. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 2008;26(29):4798-805. doi: 10.1200/jco.2007.13.8545 [published Online First: 2008/07/02]
23. Dickens NJ, Walker BA, Leone PE, et al. Homozygous deletion mapping in myeloma samples identifies genes and an expression signature relevant to pathogenesis and outcome. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2010;16(6):1856-64. doi: 10.1158/1078-0432.ccr-09-2831 [published Online First: 2010/03/11]

- 1
2
3 24. Mulligan G, Mitsiades C, Bryant B, et al. Gene expression profiling and
4 correlation with outcome in clinical trials of the proteasome inhibitor
5 bortezomib. *Blood* 2007;109(8):3177-88. doi: 10.1182/blood-2006-09-044974
6 [published Online First: 2006/12/23]
7
- 8 25. Walker BA, Boyle EM, Wardell CP, et al. Mutational Spectrum, Copy Number
9 Changes, and Outcome: Results of a Sequencing Study of Patients With
10 Newly Diagnosed Myeloma. *Journal of clinical oncology : official journal of the*
11 *American Society of Clinical Oncology* 2015;33(33):3911-20. doi:
12 10.1200/jco.2014.59.1503 [published Online First: 2015/08/19]
13
- 14 26. Samur MK, Samur AA, Fulciniti M, et al. Genome-Wide Somatic Alterations in
15 Multiple Myeloma Reveal a Superior Outcome Group. 2020;38(27):3107-18.
16 doi: 10.1200/jco.20.00461
17
- 18 27. Perrot A, Lauwers-Cances V, Tournay E, et al. Development and Validation of a
19 Cytogenetic Prognostic Index Predicting Survival in Multiple Myeloma.
20 2019;37(19):1657-65. doi: 10.1200/jco.18.00776
21
- 22 28. Kumar SK, Uno H, Jacobus SJ, et al. Impact of gene expression profiling-based
23 risk stratification in patients with myeloma receiving initial therapy with
24 lenalidomide and dexamethasone. *Blood* 2011;118(16):4359-62. doi:
25 10.1182/blood-2011-03-342089 [published Online First: 2011/08/24]
26
- 27 29. Meissner T, Seckinger A, Reme T, et al. Gene expression profiling in multiple
28 myeloma--reporting of entities, risk, and targets in clinical routine. *Clinical*
29 *cancer research : an official journal of the American Association for Cancer*
30 *Research* 2011;17(23):7240-7. doi: 10.1158/1078-0432.CCR-11-1628
31
- 32 30. Stewart AK, Rajkumar SV, Dimopoulos MA, et al. Carfilzomib, lenalidomide, and
33 dexamethasone for relapsed multiple myeloma. *New England Journal of*
34 *Medicine* 2015;372(2):142-52.
35
- 36 31. Palumbo A, Gay F, Cavallo F, et al. Continuous Therapy Versus Fixed Duration
37 of Therapy in Patients With Newly Diagnosed Multiple Myeloma. *Journal of*
38 *Clinical Oncology*;33(30):3459-66.
39
- 40 32. Brioli A, Kaiser MF, Pawlyn C, et al. Biologically defined risk groups can be used
41 to define the impact of thalidomide maintenance therapy in newly diagnosed
42 multiple myeloma. *Leuk Lymphoma* 2013;54(9):1975-81. doi:
43 10.3109/10428194.2012.760736 [published Online First: 2012/12/29]
44
- 45 33. Morgan GJ, Gregory WM, Davies FE, et al. The role of maintenance thalidomide
46 therapy in multiple myeloma: MRC Myeloma IX results and meta-analysis.
47 *Blood* 2012;119(1):7-15. doi: 10.1182/blood-2011-06-357038 [published
48 Online First: 2011/10/25]
49
- 50 34. Attal M, Lauwers-Cances V, Marit G, et al. Lenalidomide maintenance after
51 stem-cell transplantation for multiple myeloma. *The New England journal of*
52 *medicine* 2012;366(19):1782-91. doi: 10.1056/NEJMoa1114138 [published
53 Online First: 2012/05/11]
54
- 55 35. Barlogie B, Anaissie E, van Rhee F, et al. Incorporating bortezomib into upfront
56 treatment for multiple myeloma: early results of total therapy 3. *Br J Haematol*
57 2007;138(2):176-85. doi: 10.1111/j.1365-2141.2007.06639.x [published
58 Online First: 2007/06/27]
59
60

- 1
2
3 36. Bergsagel PL, Mateos MV, Gutierrez NC, et al. Improving overall survival and
4 overcoming adverse prognosis in the treatment of cytogenetically high-risk
5 multiple myeloma. *Blood* 2013;121(6):884-92. doi: 10.1182/blood-2012-05-
6 432203 [published Online First: 2012/11/21]
- 7
8 37. Harousseau JL, Attal M, Avet-Loiseau H, et al. Bortezomib plus dexamethasone
9 is superior to vincristine plus doxorubicin plus dexamethasone as induction
10 treatment prior to autologous stem-cell transplantation in newly diagnosed
11 multiple myeloma: results of the IFM 2005-01 phase III trial. *Journal of clinical
12 oncology : official journal of the American Society of Clinical Oncology*
13 2010;28(30):4621-9. doi: 10.1200/jco.2009.27.9158 [published Online First:
14 2010/09/09]
- 15
16 38. Neben K, Lokhorst HM, Jauch A, et al. Administration of bortezomib before and
17 after autologous stem cell transplantation improves outcome in multiple
18 myeloma patients with deletion 17p. *Blood* 2012;119(4):940-8. doi:
19 10.1182/blood-2011-09-379164
- 20
21 39. Shaughnessy JD, Zhou Y, Haessler J, et al. TP53 deletion is not an adverse
22 feature in multiple myeloma treated with total therapy 3. *Br J Haematol*
23 2009;147(3):347-51. doi: 10.1111/j.1365-2141.2009.07864.x [published
24 Online First: 2009/08/26]
- 25
26 40. Roussel M, Lauwers-Cances V, Robillard N, et al. Front-line transplantation
27 program with lenalidomide, bortezomib, and dexamethasone combination as
28 induction and consolidation followed by lenalidomide maintenance in patients
29 with multiple myeloma: a phase II study by the Intergroupe Francophone du
30 Myelome. *Journal of clinical oncology : official journal of the American Society
31 of Clinical Oncology* 2014;32(25):2712-7. doi: 10.1200/jco.2013.54.8164
32 [published Online First: 2014/07/16]
- 33
34 41. Kumar S, Flinn I, Richardson PG, et al. Randomized, multicenter, phase 2 study
35 (EVOLUTION) of combinations of bortezomib, dexamethasone,
36 cyclophosphamide, and lenalidomide in previously untreated multiple
37 myeloma. *Blood* 2012;119(19):4375-82. doi: 10.1182/blood-2011-11-395749
- 38
39 42. Plesner T, Arkenau HT, Gimsing P, et al. Phase 1/2 study of daratumumab,
40 lenalidomide, and dexamethasone for relapsed multiple myeloma. *Blood* 2016
41 doi: 10.1182/blood-2016-07-726729 [published Online First: 2016/08/18]
- 42
43 43. Katja C. Weisel JSM, Gordon Cook, Merav Leiba, Kenshi Suzuki, Shaji Kumar,
44 Michele Cavo, Herve Avet-Loiseau, Hang Quach, Vania Hungria, Suzanne
45 Lentzsch, Roman Hajek, Pieter Sonneveld, Kaida Wu, Xiang Qin, Christopher
46 Chiu, David Soong, Ming Qi, Jordan Mark Schechter, Meletios A. Dimopoulos.
47 Efficacy of daratumumab in combination with lenalidomide plus
48 dexamethasone (DRd) or bortezomib plus dexamethasone (DVd) in relapsed
49 or refractory multiple myeloma (RRMM) based on cytogenetic risk status.
50 2017 ASCO Annual Meeting 2017; J Clin Oncol 35, 2017 (suppl; abstr 8006)
- 51
52 44. Farooqui AA, Tariq M, Nabeel S, et al. Daratumumab-based three drug regimens
53 for high-risk multiple myeloma: A meta-analysis. 2020;38(15_suppl):e20549-
54 e49. doi: 10.1200/JCO.2020.38.15_suppl.e20549
- 55
56 45. Cavo M, Tosi P, Zamagni E, et al. Prospective, randomized study of single
57 compared with double autologous stem-cell transplantation for multiple
58
59
60

- 1
2
3 myeloma: Bologna 96 clinical study. *Journal of clinical oncology : official*
4 *journal of the American Society of Clinical Oncology* 2007;25(17):2434-41.
5 doi: 10.1200/jco.2006.10.2509 [published Online First: 2007/05/09]
6
- 7 46. Cavo M, Gay F, Beksac M, et al. Autologous haematopoietic stem-cell
8 transplantation versus bortezomib–melphalan–prednisone, with or without
9 bortezomib–lenalidomide–dexamethasone consolidation therapy, and
10 lenalidomide maintenance for newly diagnosed multiple myeloma
11 (EMN02/HO95): a multicentre, randomised, open-label, phase 3 study. *The*
12 *Lancet Haematology* 2020;7(6):e456-e68. doi: [https://doi.org/10.1016/S2352-](https://doi.org/10.1016/S2352-3026(20)30099-5)
13 [3026\(20\)30099-5](https://doi.org/10.1016/S2352-3026(20)30099-5)
14
- 15 47. Barlogie B, Tricot GJ, van Rhee F, et al. Long-term outcome results of the first
16 tandem autotransplant trial for multiple myeloma. *Br J Haematol*
17 2006;135(2):158-64. doi: 10.1111/j.1365-2141.2006.06271.x [published
18 Online First: 2006/08/31]
19
- 20 48. Roussel M, Moreau P, Huynh A, et al. Bortezomib and high-dose melphalan as
21 conditioning regimen before autologous stem cell transplantation in patients
22 with de novo multiple myeloma: a phase 2 study of the Intergroupe
23 Francophone du Myelome (IFM). *Blood* 2010;115(1):32-7. doi: 10.1182/blood-
24 2009-06-229658 [published Online First: 2009/11/04]
25
- 26 49. Rawstron AC, Child JA, de Tute RM, et al. Minimal residual disease assessed by
27 multiparameter flow cytometry in multiple myeloma: impact on outcome in the
28 Medical Research Council Myeloma IX Study. *Journal of clinical oncology :*
29 *official journal of the American Society of Clinical Oncology* 2013;31(20):2540-
30 7. doi: 10.1200/JCO.2012.46.2119
31
- 32 50. Goicoechea I, Puig N, Cedena M-T, et al. Deep MRD profiling defines outcome
33 and unveils different modes of treatment resistance in standard and high risk
34 myeloma. *Blood* 2020 doi: 10.1182/blood.2020006731
35
- 36 51. Croft J, Riddell A, Koh D-M, et al. Inter-observer agreement of baseline whole
37 body MRI in multiple myeloma. *Cancer Imaging* 2020;20(1):48. doi:
38 10.1186/s40644-020-00328-9
39
- 40 52. Messiou C, Hillengass J, Delorme S, et al. Guidelines for Acquisition,
41 Interpretation, and Reporting of Whole-Body MRI in Myeloma: Myeloma
42 Response Assessment and Diagnosis System (MY-RADS). 2019;291(1):5-13.
43 doi: 10.1148/radiol.2019181949
44
- 45 53. Clinical Trials.gov website.
46
- 47 54. Harousseau JL, Avet-Loiseau H, Attal M, et al. Achievement of at least very good
48 partial response is a simple and robust prognostic factor in patients with
49 multiple myeloma treated with high-dose therapy: long-term analysis of the
50 IFM 99-02 and 99-04 Trials. *Journal of clinical oncology : official journal of the*
51 *American Society of Clinical Oncology* 2009;27(34):5720-6. doi:
52 10.1200/jco.2008.21.1060 [published Online First: 2009/10/15]
53
- 54 55. Moreau P, Hullin C, Garban F, et al. Tandem autologous stem cell
55 transplantation in high-risk de novo multiple myeloma: final results of the
56 prospective and randomized IFM 99-04 protocol. *Blood* 2006;107(1):397-403.
57 doi: 10.1182/blood-2005-06-2573 [published Online First: 2005/09/08]
58
59
60

- 1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
56. McCarthy PL, Owzar K, Hofmeister CC, et al. Lenalidomide after stem-cell transplantation for multiple myeloma. *The New England journal of medicine* 2012;366(19):1770-81. doi: 10.1056/NEJMoa1114083
 57. Thall PF, Simon RM, Estey EH. Bayesian sequential monitoring designs for single-arm clinical trials with multiple outcomes. *Stat Med* 1995;14(4):357-79. [published Online First: 1995/02/28]
 58. Thall PF, Simon RM, Estey EH. New statistical strategy for monitoring safety and efficacy in single-arm clinical trials. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 1996;14(1):296-303. [published Online First: 1996/01/01]
 59. Thall PF, Sung HG. Some extensions and applications of a Bayesian strategy for monitoring multiple outcomes in clinical trials. *Stat Med* 1998;17(14):1563-80. [published Online First: 1998/08/12]
 60. Wale A, Pawlyn C, Kaiser M, et al. Frequency, distribution and clinical management of incidental findings and extramedullary plasmacytomas in whole body diffusion weighted magnetic resonance imaging in patients with multiple myeloma. *Haematologica* 2016;101(4):e142-4. doi: 10.3324/haematol.2015.139816 [published Online First: 2016/01/29]
 61. Messiou C, Kaiser M. Whole body diffusion weighted MRI--a new view of myeloma. *Br J Haematol* 2015;171(1):29-37. doi: 10.1111/bjh.13509 [published Online First: 2015/05/28]
 62. Brown SR, Sherratt D, Booth G, et al. Experiences of establishing an academic early phase clinical trials unit. *Clinical trials (London, England)* 2017;1740774517710250. doi: 10.1177/1740774517710250 [published Online First: 2017/05/24]
 63. Rajkumar SV, Harousseau JL, Durie B, et al. Consensus recommendations for the uniform reporting of clinical trials: report of the International Myeloma Workshop Consensus Panel 1. *Blood* 2011;117(18):4691-95. doi: blood-2010-10-299487 [pii];10.1182/blood-2010-10-299487 [doi]
 64. Office for National Statistics. Office for National Statistics: Cancer Statistics Registrations, England (Series MB1) , No. 42, 2011 2013 [
 65. Kaiser MF, Walker BA, Hockley SL, et al. A TC classification-based predictor for multiple myeloma using multiplexed real-time quantitative PCR. *Leukemia* 2013;27(8):1754-7. doi: 10.1038/leu.2013.12 [published Online First: 2013/01/16]

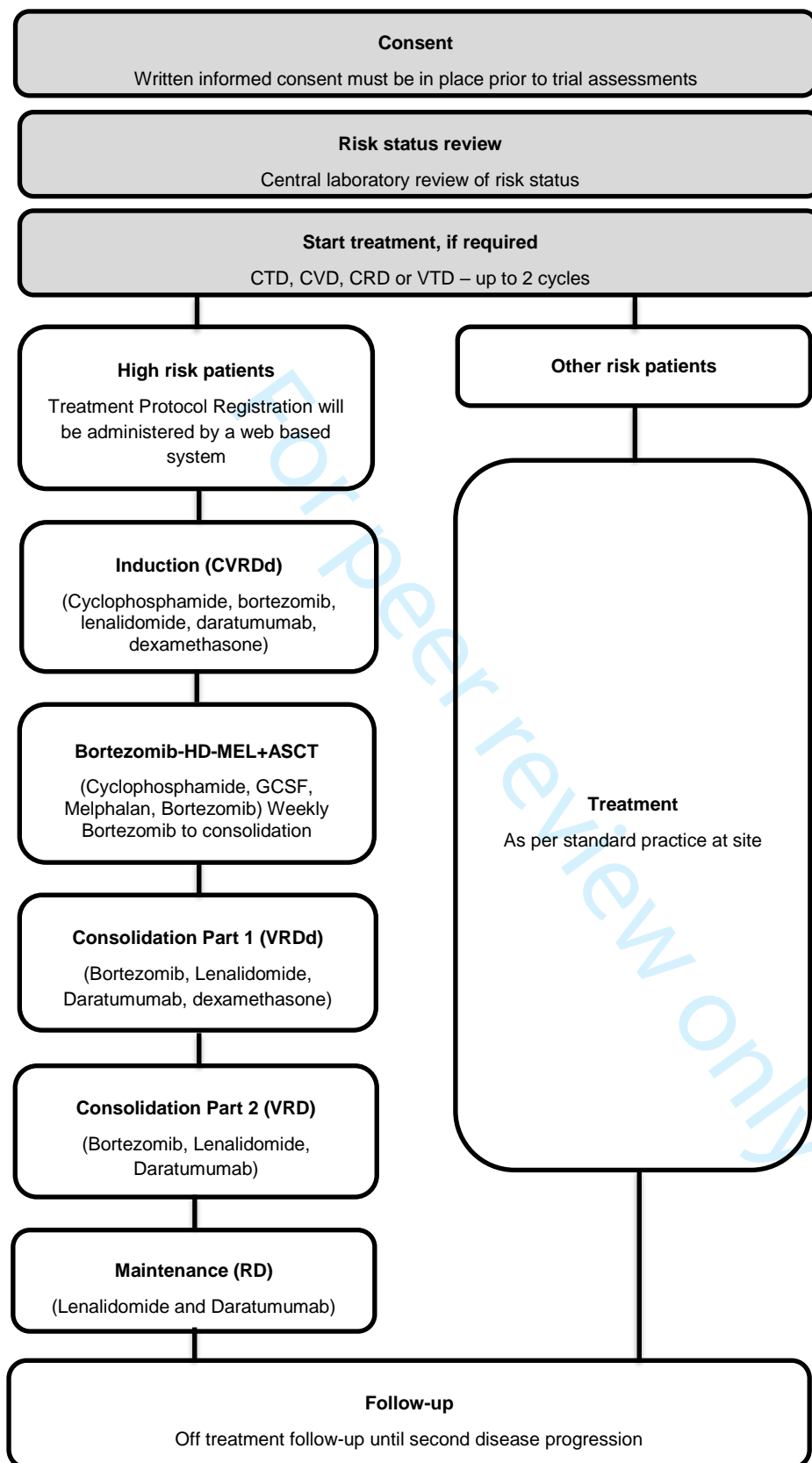


Figure 1. MUKnine OPTIMUM trial design

Statistical analysis

MUK *nine a*

Analyses using response and progression data will be performed using the data recorded on the CRF. The data recorded on the CRF may be centrally reviewed to assess the quality of these data.

Screening population

The screening population will include all participants who are registered into MUK *nine a*, regardless of their risk assessment. Participants for whom we do not receive a risk assessment result will be included in this population.

Non-MUK *nine b* population

The Non-MUK *nine b* population will include all participants who are registered into MUK *nine a*, regardless of their risk assessment, who are not registered into the MUK *nine b* trial. Participants for whom we do not receive a risk assessment result will be included in this population.

Non-MUK *nine b* high risk population

The high risk population will include all participants who are assessed as being high risk, but who are not registered into the MUK *nine b* trial.

Non high risk population

The non-high risk population will include all participants who are assessed as being not high risk. Participants for whom we do not receive a risk assessment result will not be included in this population.

Analyses will be performed for the four populations separately, unless specified.

Imaging study population

The imaging study population will contain all participants who have entered the imaging sub-study and had a diffusion-weighted whole body MRI scan at baseline.

Primary endpoint analysis

The number and proportion of molecular risk-defining investigations performed within 8 weeks will be reported. Summary statistics of the length of time taken to turn around molecular risk-defining investigations will also be reported, including median, mean, standard deviation, inter-quartile range (IQR). This will be summarised for the screening population.

Secondary endpoint analysis

Recruitment rates

The number of participants registered overall and the average rate per month will be reported. The number and proportion of participants identified as high risk (out of both the number of participants registered to MUK *nine a* and the number of participants registered and with a corresponding sample taken for risk definition) overall will also be reported, as well as the number and proportion of participants accepting registration to MUK *nine b* (out of the number identified as high risk). Summaries will be provided overall and by site.

Progression-free survival

Progression-free survival curves will be calculated using the Kaplan-Meier method and the median progression-free survival estimates and progression-free survival estimates at yearly time-points with corresponding 95% confidence intervals will be presented for the Non-MUK *nine b* population, the Non-MUK *nine b* High risk population and the Non high risk population..

The Cox proportional hazards model (if appropriate), adjusting for treatment received first-line, will also be used to summarise progression-free survival for each population. Covariate estimates, standard errors, hazard ratios, 95% confidence intervals, as well as p-values will be presented for all variables incorporated in the model.

The Cox proportional hazards model, adjusting for treatment received first-line and other pre-specified baseline characteristics, may be used to further summarise progression-free survival, after discussion with the MUK *nine* Trial Management Group (TMG). The number of characteristics (and their factors) included in the model will depend on the number of participants in the relevant population.

Second progression-free survival (PFS2)

PFS2 curves will be calculated using the Kaplan-Meier method and the median second progression-free survival estimates and progression-free survival estimates at appropriate time-points with corresponding 95% confidence intervals will be presented for the Non-MUK *nine b* population, the Non-MUK *nine b* High risk population and the Non high risk population..

The Cox proportional hazards model (if appropriate), adjusting for treatment received first and second-line, will also be used to summarise PFS2 within each population. Treatment received second-line will be included using a time-dependant covariate to incorporate timing of treatment. Covariate estimates, standard errors, hazard ratios and 95% confidence intervals, as well as p-values will be presented for all variables incorporated in the model.

1
2
3 The Cox proportional hazards model, adjusting for treatment received first-line and other pre-
4 specified baseline characteristics, may be used to further summarise second progression-free
5 survival, after discussion with the MUK *nine* TMG. The number of characteristics (and their
6 factors) included in the model will depend on the number of participants in the relevant
7 population.
8
9
10

11 **Treatment received first and second-line**

12
13 Treatment received first and second-line will be summarised in a tabular form, along with
14 reasons for stopping treatment. This will be summarised for the Non-MUK *nine b* population,
15 the Non-MUK *nine b* High risk population and the Non high risk population.
16
17
18

19 **Response to first and second-line treatment**

20
21 The number and proportion of participants in each response category post first and second-
22 line treatment will be presented with corresponding 95% confidence intervals for each
23 population and type of treatment for the Non-MUK *nine b* population, the Non-MUK *nine b*
24 High risk population and the Non high risk population.
25
26
27

28 **Overall survival**

29
30 Overall survival curves will be calculated using the Kaplan-Meier method and median overall
31 survival estimates and overall survival estimates at yearly time-points with corresponding 95%
32 confidence intervals will be presented for the Non-MUK *nine b* population, the Non-MUK *nine*
33 *b* High risk population and the Non high risk population.
34
35
36

37 The Cox proportional hazards model (if appropriate), adjusting for treatment received first and
38 second-line will also be used to summarise overall survival within each population. Treatment
39 received second-line will be included using a time-dependant covariate to incorporate timing
40 of treatment. Covariate estimates, standard errors, hazard ratios and 95% confidence
41 intervals, as well as p-values will be presented for all variables incorporated in the model.
42
43
44

45 The Cox proportional hazards model, adjusting for treatment received first-line and other pre-
46 specified baseline characteristics, may be used to further summarise overall survival, after
47 discussion with the MUK *nine* TMG. The number of characteristics (and their factors) included
48 in the model will depend on the number of participants in the relevant population.
49
50
51

52 **MUK*nine b***

53
54 Analyses using response and progression data will be performed using both the data
55 recorded on the eCRF, and data from analysis of central samples. Primacy will be given to the
56 central sample assessment of response, and local data recorded on the eCRF will be used
57
58
59
60

1
2
3 where a central assessment is not available. Differences between the local eCRF data and
4 the central assessment data will be summarised.
5

6 7 **Analysis population**

8
9 The analysis population, as well as the safety population, will include all participants who
10 receive at least one dose of any trial treatment.
11

12 13 **Imaging study population**

14
15 The imaging study population will contain all participants who have entered the imaging sub-
16 study and had a diffusion-weighted whole body MRI scan at baseline.
17

18 19 **Interim analyses**

20
21 Interim assessments will be performed after cohorts of 10 participants have been registered
22 to treatment and followed up to 120 days post-ASCT, until all participants have been recruited
23 and received induction treatment.
24

25
26 The trial may be terminated early for futility based on MRD status at 100 days post-ASCT and
27 PFS at 100 days post-ASCT. If a participant does not receive an ASCT, the 100 days post-
28 ASCT time-point becomes 12 months post registration as this is approximately equivalent.
29

30
31 At each interim analysis, the posterior probabilities of the two events in terms of pre-defined
32 cut-points are calculated in order to determine whether the trial should be stopped for futility.
33

34 35 **Primary endpoint analysis**

36
37 At the end of the study, the experimental treatment arm will be compared to the historical
38 control prior in terms of PFS at 18 months post-registration/randomisation for the MUK *nine b*
39 analysis population. If the proportion of participants who are alive and progression-free at 18
40 months post-registration is higher in the treatment arm than in the control prior with 85%
41 probability, the treatment arm will be deemed efficacious.
42
43

44
45
46 Further analyses of progression-free survival at 18 months post-registration will also be
47 performed outside of the Bayesian framework. PFS curves will be calculated using the Kaplan-
48 Meier method and median progression-free survival estimates and progression-free survival
49 estimates at 6, 12 (corresponding approximately to the 100 days post-ASCT time-point) and
50 18 months with corresponding 95% confidence intervals will be presented.
51
52

53
54
55 At the end of the study, the experimental arm will be independently compared to historical
56 control data using molecularly matched individual participant data (IPD) from Myeloma XI/XI+
57 and assessed for superiority in terms of progression-free survival (PFS) at 18 months post-
58 registration/randomisation in an exploratory analysis.
59
60

Secondary endpoint analysis

Although not an endpoint, baseline characteristics will be summarised for participants in the analysis population. All MUK nine b secondary endpoint analyses will be performed for the MUK nine b analysis/safety population.

Safety

The number of SAEs will be summarised and presented by MedDRA system organ class. In addition, information will be given on the number of SAEs per participant, together with details on the causality, expectedness and outcome of each SAE experienced. Summaries of SARs and SUSARs will also be presented.

Toxicity

Summaries will be produced to show the proportion of participants experiencing each grade of toxicity overall, presented overall and by other groupings, such as stage of treatment, as necessary.

Progression-free survival at 100 days post-ASCT

Progression-free survival at 100 days post-ASCT is used to determine whether the trial should be stopped for futility, as detailed above. The 100 days post-ASCT time-point is approximately equivalent to 12 months post-registration, and progression-free survival estimates at this time-point are presented as part of the primary endpoint, detailed above.

Minimal residual disease (MRD) at the end of induction therapy, 100 days post-ASCT and post-consolidation part 2

MRD status will be assessed at the end of induction therapy, 100 days post-ASCT and 1 year post-ASCT for all participants, regardless of their categorical paraprotein response.

MRD status will be summarised at each time-point. These summaries will be obtained using multi-level repeated measures models accounting for data at all time-points, regardless of timing of sample for the time-point not of interest, assuming missing data at random [MAR], allowing for time and adjusting for pre-specified clinically important baseline characteristics [all fixed effects] and for participant and participant-time interaction [random effects] where appropriate.

Data will also be summarised descriptively using plots of proportion of participants with MRD negative disease over time and summary tables. Missing data patterns will be examined carefully and sensitivity analyses using different missing data assumptions will be performed if appropriate.

1
2
3
4
5 Sensitivity analyses may be carried out using methods such as multiple imputation or pattern-
6 mixture multi-level models categorising participants into strata based on clinical information
7 which is believed to represent the reasons for missing data (assuming MAR data conditional
8 upon participants' clinical data).
9

11 **Overall survival**

12
13
14 Overall survival curves will be calculated using the Kaplan-Meier method and median overall
15 survival estimates and overall survival estimates at 12, 24 and 36 months with corresponding
16 95% confidence intervals will be presented by treatment group.
17

18
19 The Cox proportional hazards model (if appropriate), adjusting for pre-specified baseline
20 characteristics, may be used to further summarise overall survival, after discussion with the
21 MUK *nine* TMG. The number of characteristics (and their factors) included in the model will
22 depend on the number of participants in the analysis population.
23
24

25 **Maximum response at the end of induction therapy, 100 days post-ASCT and post- 26 consolidation part 2**

27
28 The number and proportion of participants in each maximum response category will be
29 presented with corresponding 95% confidence intervals at each time-point. Participants who
30 do not achieve a maximum response will be summarised as 'no maximum response'.
31
32

33 **Overall response at the end of induction therapy, 100 days post-ASCT and post- 34 consolidation part 2**

35
36 The proportion of participants achieving at least PR will be summarised with corresponding
37 95% confidence intervals, at each time-point.
38

39 **Time to progression**

40
41 Time to progression will be summarised using cumulative incidence function curves, and
42 median time to progression estimates with corresponding 95% confidence intervals will be
43 presented.
44

45
46 An assessment based on the number of participants who die with no previous evidence of
47 disease progression (i.e. the number of competing risk events) will be made as to whether
48 time to progression should be summarised using the Kaplan-Meier method (i.e. not
49 incorporating competing risks).
50
51

52 **Time to maximum response**

1
2
3 Time to maximum response curves will be calculated using the Kaplan-Meier method, and the
4 median time to maximum response estimates with corresponding 95% confidence intervals
5 will be presented.
6
7

8 As maximum response is defined as SD or better, the number of participants who progress or
9 die without achieving a maximum response is expected to be small. This will be monitored on
10 an ongoing basis via DMEC reports and an assessment will be made as to whether analyses
11 that take into account competing risks (for example cumulative incidence function curves) are
12 required.
13
14
15

16 **Second progression-free survival (PFS2)**

17
18 PFS2 curves will be calculated using the Kaplan-Meier method and the median second
19 progression-free survival estimates and progression-free survival estimates at appropriate
20 time-points with corresponding 95% confidence intervals will be presented.
21
22
23

24 The Cox proportional hazards model, adjusting for pre-specified baseline characteristics, may
25 be used to further summarise second progression-free survival, after discussion with the MUK
26 *nine* TMG. The number of characteristics (and their factors) included in the model will depend
27 on the number of participants in the analysis population.
28
29
30

31 **Overall treatment benefit and clinician assessment of treatment benefit at the end of 32 induction therapy and 100 days post-ASCT**

33
34 The proportion of participants achieving each score for overall treatment benefit, and each
35 response given to the clinician assessment of treatment benefit question, will be summarised
36 with corresponding 95% confidence intervals, at each time-point.
37
38
39

40 A cross-tabulation of overall treatment benefit and clinician assessment of treatment benefit
41 will be created to compare the two measures.
42
43

44 **Quality of life**

45
46 Quality of life will be summarised at each post-registration time-point, adjusting for baseline
47 mean scores and 95% CIs. These summaries will be obtained using multi-level repeated
48 measures models accounting for data at all post-baseline time points, regardless of time of
49 completion for the time-point not of interest, assuming missing data at random [MAR], allowing
50 for time and adjusting for baseline QoL and pre-specified clinically important baseline
51 characteristics [all fixed effects] and for participant and participant-time interaction [random
52 effects] where appropriate.
53
54
55
56
57
58
59
60

1
2
3 Data will also be summarised descriptively using bar charts, box plots, plots of mean QoL over
4 time and summary tables. Missing data patterns will be examined carefully and sensitivity
5 analyses using different missing data assumptions will be performed if appropriate.
6
7

8 **Treatment compliance**

9

10 Information on dose delays, reductions and omissions will be summarised by stage of
11 treatment. Information on the proportion of participants with at least one dose delay, reduction
12 or incidence of missed doses will also be presented overall and by stage of treatment.
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

For peer review only

Delete this line, then print on Trust/Hospital headed paper

Participant ID:	Initials:
Date of Birth:	Principal Investigator:

MUK *nine a*

A phase II study identifying and evaluating high risk (HR) myeloma patients suitable for novel treatment approaches

SAMPLE PARTICIPANT CONSENT FORM

Participant initial after each statement

1. I confirm that I have read and understand the information sheet for the above trial and have had the opportunity to ask questions.
2. I understand that taking part in this trial is voluntary and that I am free to withdraw at any time without my medical care or legal rights being affected. I understand that even if I withdraw from the above trial, the data and samples and MRI, CT scan and x-ray images collected from me will be used in analysing the results of the trial and in some cases further information about any unwanted effects of my treatment may need to be collected by the trial team.
3. I understand that my healthcare records may be looked at by authorised individuals from the trial team, the NHS Trust, regulatory bodies or Sponsor in order to check that the trial is being carried out correctly.
4. I agree to allow any information or results arising from this trial to be used for healthcare and/or further medical research upon the understanding that my identity will remain anonymous wherever possible.
5. I agree to a copy of this Consent Form, detailing my full name, being sent to the Clinical Trials Research Unit (CTRU) at the University of Leeds.
6. I agree that my GP, or any other doctor treating me, will be notified of my participation in this trial.
7. I agree that samples of blood, urine and bone marrow taken from me during the course of the trial may be used for additional research investigations that form part of this trial and that the samples will be sent to laboratories outside my hospital.
8. I understand that some of the research studies using my samples may include genetic research aimed at understanding the genetic influences on predicting response to treatments and predisposition to multiple myeloma,

1
2
3 but the results of these investigations are unlikely to have any implications
4 for me personally.
5

6 9. I agree to take part in the trial.
7
8

9 **The following points are OPTIONAL. Even if you agree to take part in this trial,**
10 **you do not have to agree to this part**

Participant initial
Yes or No

Yes No

11
12
13 1. If I am diagnosed with any disease other than symptomatic multiple myeloma
14 or plasma cell leukaemia, I give permission to be contacted through my hospital
15 about other research that may be available to me in the future
16

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

17
18
19 **Consent for storage and use in possible future research projects**

20 1. I agree that the samples I have given and anonymised information stored
21 about me can be stored for possible use in future projects. I understand
22 that some projects may be carried out by researchers at different
23 institutions other than the trial team who ran the first trial. I understand that
24 any future research using these samples would require further ethical
25 approval.
26

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

27 2. I agree that information obtained through this research about the
28 molecular features of my multiple myeloma or plasma cell leukaemia, e.g.
29 findings about tumour mutations, may be fed back to my treating doctor if
30 they are of potential relevance for my future treatment.
31

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

32 3. If I am diagnosed with any disease other than symptomatic multiple myeloma
33 or plasma cell leukaemia, I give permission for the samples sent to the central
34 laboratories to be stored and used in future research that receives ethical
35 approval. I understand that the samples and data collected from them may be
36 shared with researchers, possibly outside the European Economic Area (EEA).
37

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

38
39
40
41 **Consent for the imaging study**

42 My hospital is not taking part in the imaging study

43
44 1. I agree to take part in the whole body MRI study.

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

45 2. I agree that the whole body MRI images I have given and anonymised
46 information stored about me can be stored for possible use in future
47 projects. I understand that some projects may be carried out by
48 researchers at different institutions other than the trial team who ran the
49 first trial. I understand that any future research using these scans would
50 require further ethical approval.
51
52
53
54
55
56
57
58
59
60

<input type="checkbox"/>	<input type="checkbox"/>
--------------------------	--------------------------

Participant:

Signature.....

Name (block capitals).....

Date.....

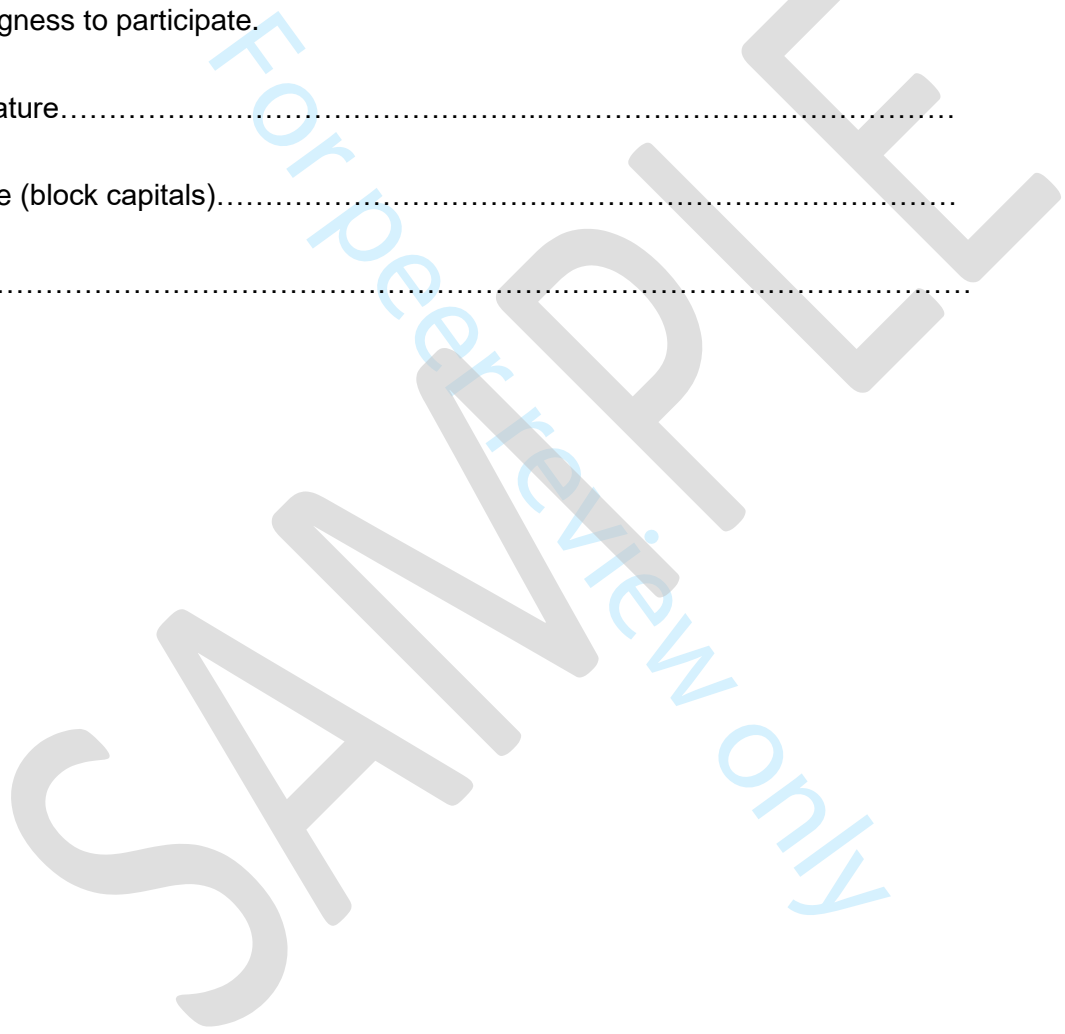
Investigator:

I have explained the trial to the above named participant and he/she has indicated his/her willingness to participate.

Signature.....

Name (block capitals).....

Date.....



1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



STANDARD PROTOCOL ITEMS: RECOMMENDATIONS FOR INTERVENTIONAL TRIALS

SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	1
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	2
	2b	All items from the World Health Organization Trial Registration Data Set	1, 2, 4-9, 11-15, Tables 1 and 2
Protocol version	3	Date and version identifier	6
Funding	4	Sources and types of financial, material, and other support	14, 15
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	1,14
	5b	Name and contact information for the trial sponsor	15
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	14, 15
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	12, 13, 14

1 **Introduction**

2

3 Background and rationale 6a Description of research question and justification for undertaking the trial, including summary of relevant studies (published and unpublished) examining benefits and harms for each intervention 4-7

4

5

6 6b Explanation for choice of comparators 3, 8, 12

7

8 Objectives 7 Specific objectives or hypotheses 7, 8

9

10 Trial design 8 Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group), allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory) 2, 8

11

12

13

14 **Methods: Participants, interventions, and outcomes**

15

16 Study setting 9 Description of study settings (eg, community clinic, academic hospital) and list of countries where data will be collected. Reference to where list of study sites can be obtained 8, 9

17

18

19 Eligibility criteria 10 Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and individuals who will perform the interventions (eg, surgeons, psychotherapists) Table 2

20

21

22 Interventions 11a Interventions for each group with sufficient detail to allow replication, including how and when they will be administered Table 1

23

24

25 11b Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose change in response to harms, participant request, or improving/worsening disease) 9, 10 and Table 3

26

27

28 11c Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence (eg, drug tablet return, laboratory tests) 9

29

30

31 11d Relevant concomitant care and interventions that are permitted or prohibited during the trial Table 2, Table 3

32

33

34 Outcomes 12 Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended 11, 12, supplementary file 1

35

36

37

38

39

40 Participant timeline 13 Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure) 9-11, Figure 1, Table 4

41

42

43

44

45

46

1 Sample size 14 Estimated number of participants needed to achieve study objectives and how it was determined, including 8, 9
 2 clinical and statistical assumptions supporting any sample size calculations

4 Recruitment 15 Strategies for achieving adequate participant enrolment to reach target sample size 8

7 **Methods: Assignment of interventions (for controlled trials)**

8 Allocation:

10 Sequence 16a Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any NA, not
 11 generation 16a factors for stratification. To reduce predictability of a random sequence, details of any planned restriction randomised
 12 (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants
 13 or assign interventions

16 Allocation 16b Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, NA, not
 17 concealment 16b opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned randomised
 18 mechanism

20 Implementation 16c Who will generate the allocation sequence, who will enrol participants, and who will assign participants to NA, not
 21 interventions randomised

24 Blinding (masking) 17a Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome NA, not
 25 assessors, data analysts), and how randomised or
 26 blinded

28 17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's NA, not
 29 allocated intervention during the trial randomised or
 30 blinded

34 **Methods: Data collection, management, and analysis**

35 Data collection 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related 9-13, Table 4
 36 methods 18a processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of
 37 study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known.
 38 Reference to where data collection forms can be found, if not in the protocol

1		18b	Plans to promote participant retention and complete follow-up, including list of any outcome data to be collected for participants who discontinue or deviate from intervention protocols	10
2				
3				
4	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality (eg, double data entry; range checks for data values). Reference to where details of data management procedures can be found, if not in the protocol	13, 14
5				
6				
7				
8	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the statistical analysis plan can be found, if not in the protocol	11, 12, Supplementary file 1
9				
10				
11				
12				
13		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	Supplementary file 1
14				
15				
16		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation)	Supplementary file 1
17				
18				
19				
20				
21	Methods: Monitoring			
22				
23	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed	12, 13
24				
25				
26				
27				
28				
29		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	9, 12, supplementary file 1
30				
31				
32				
33	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse events and other unintended effects of trial interventions or trial conduct	Table 4, 10, 11, 12
34				
35				
36	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	13
37				
38				
39				

40 Ethics and dissemination

1	Research ethics approval	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	2, 13
2				
3				
4	Protocol amendments	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals, regulators)	12
5				
6				
7				
8	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and how (see Item 32)	8-10, 13.
9				
10				
11		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary studies, if applicable	9, 10.
12				
13				
14	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained in order to protect confidentiality before, during, and after the trial	12
15				
16				
17				
18	Declaration of interests	28	Financial and other competing interests for principal investigators for the overall trial and each study site	14
19				
20				
21	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that limit such access for investigators	12, 13, 14.
22				
23				
24	Ancillary and post-trial care	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial participation	12
25				
26				
27	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals, the public, and other relevant groups (eg, via publication, reporting in results databases, or other data sharing arrangements), including any publication restrictions	2, 13
28				
29		31b	Authorship eligibility guidelines and any intended use of professional writers	13
30				
31		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	13
32				
33				
34				
35				
36	Appendices			
37				
38	Informed consent materials	32	Model consent form and other related documentation given to participants and authorised surrogates	Supplementary file 2
39				
40				
41				
42				
43				
44				
45				
46				

1 Biological 33 Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular 9, 10, Table 4.
2 specimens analysis in the current trial and for future use in ancillary studies, if applicable
3

4 *It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items.
5 Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons
6 [“Attribution-NonCommercial-NoDerivs 3.0 Unported”](#) license.
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46

For peer review only