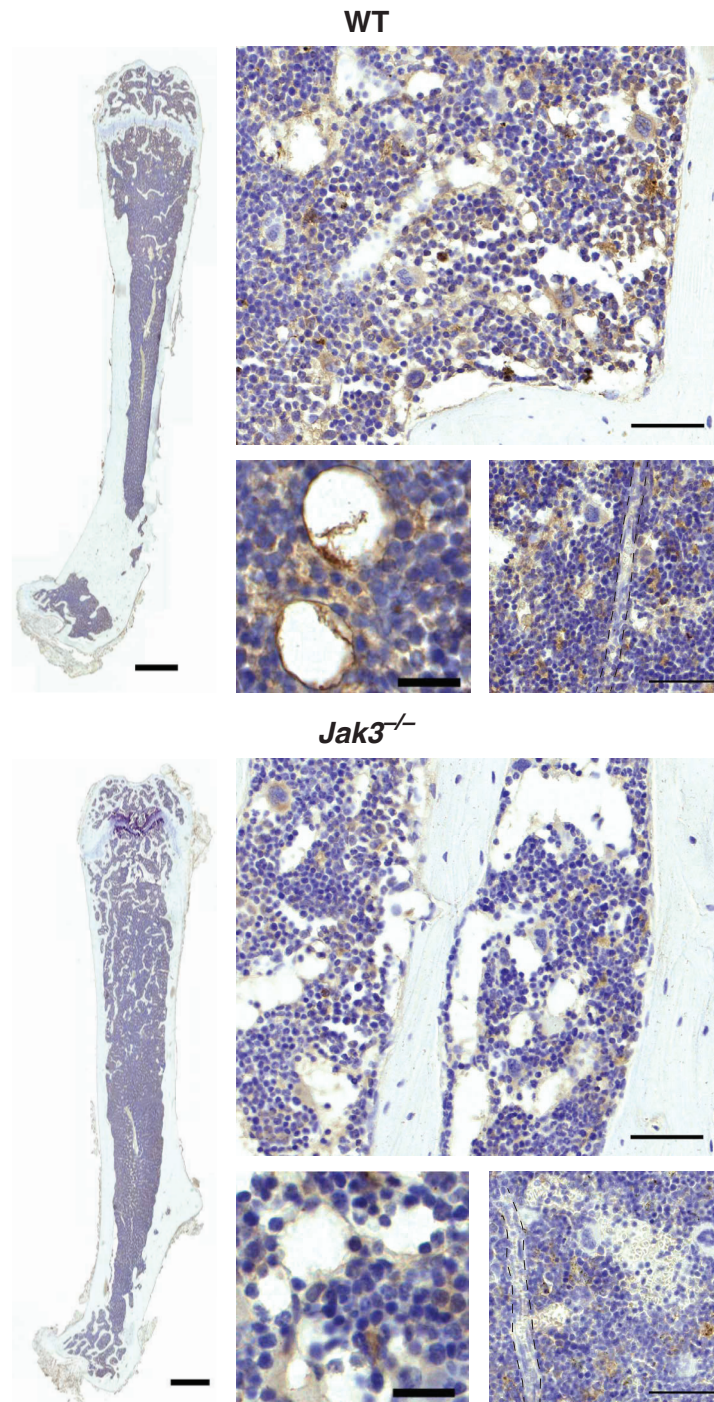


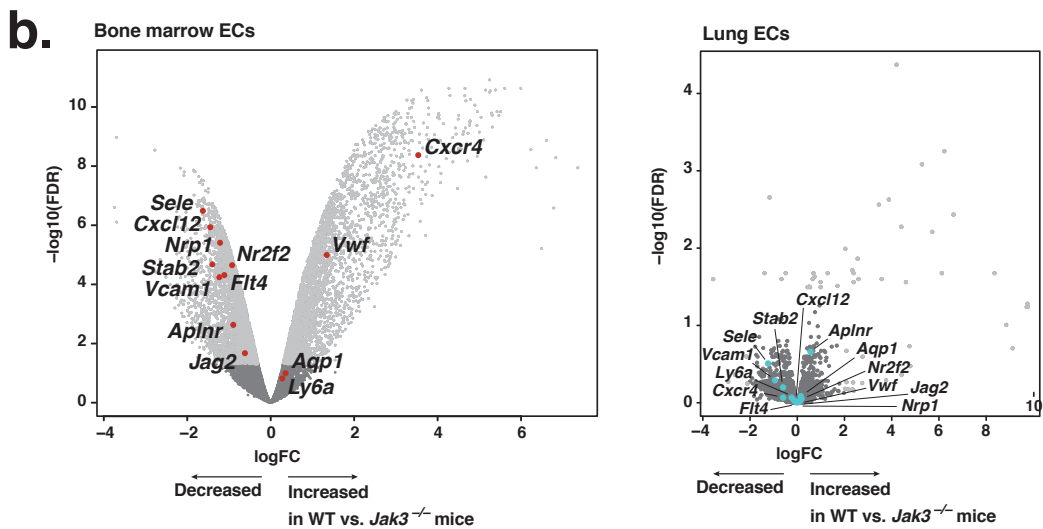
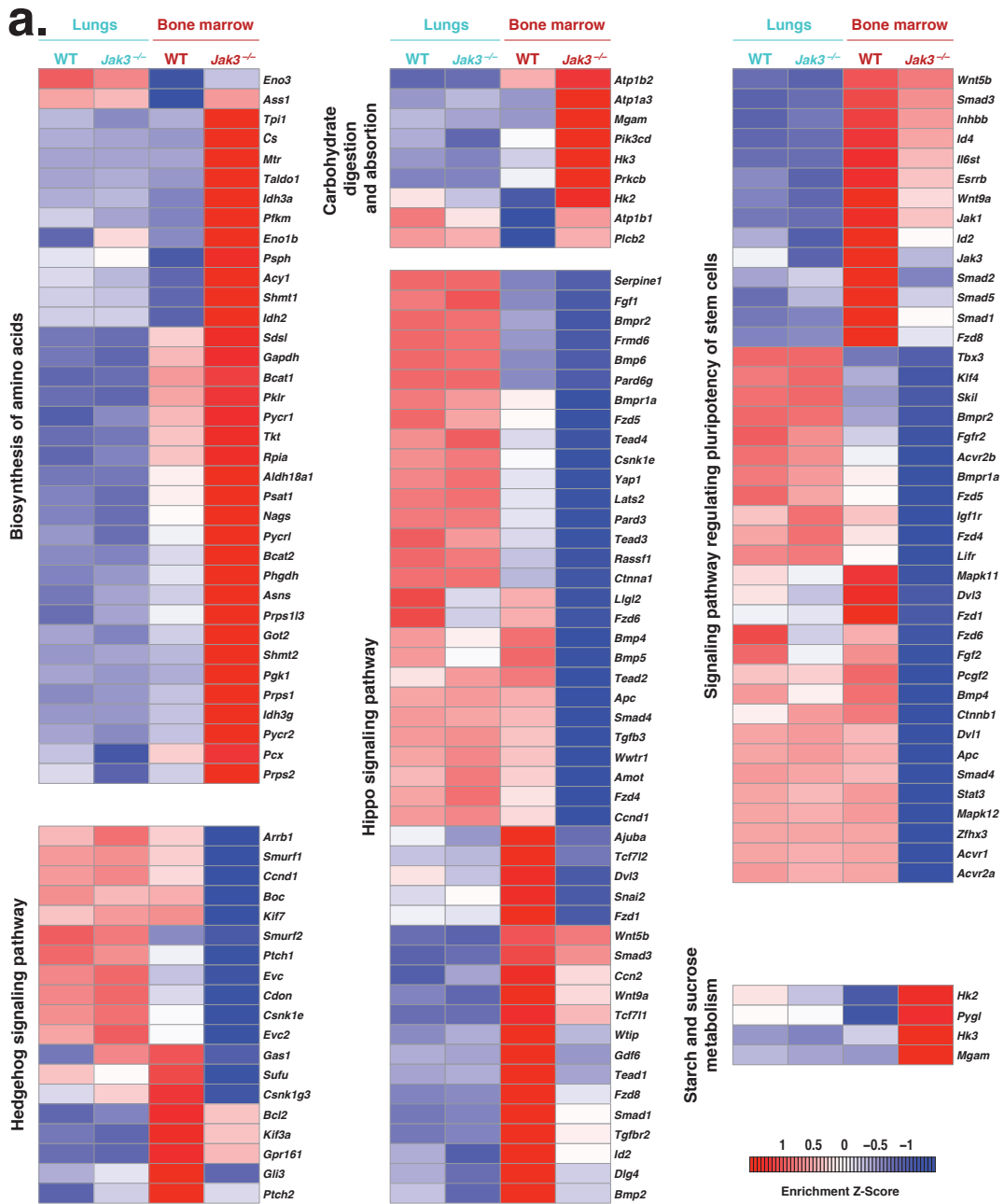
Supplementary Figure 1. Organotypic transcriptomic signatures of endothelial cells (ECs).

(a) Principal component analysis (PCA) performed on RNA sequencing (RNA-seq) counts of primary murine endothelial cell populations extracted from the indicated vascular beds where principal component 1 (PC1), PC2, and PC3 are illustrated plotted against each other. (b) Average FPKM from organotypic endothelial transcript expression across vascular beds obtained by RNA-seq (left, $n = 2$ or 3). Four asterisks (****), three asterisk (***), and two asterisks (**) denote, respectively, p values < 0.0001 , < 0.001 , and < 0.01 by one-way ANOVA. Each replicate is represented by an individual dot and error bars represent the standard deviation from the mean. Plots of 95% confidence intervals (right) generated by one-way ANOVA comparisons of indicated gene transcripts in every two vascular niches analyzed. (c) Network visualization of known and predicted protein-protein interactions between gene transcripts corresponding to the JAK-STAT signaling pathway following analysis of differentially expressed bone marrow EC gene transcripts against the Kyoto Encyclopedia of Genes and Genomes database⁵⁴⁻⁵⁶. (d) Volcano plot illustrating differential expression analysis of RNA-seq corresponding to bone marrow ECs vs. liver, lung, heart, and kidney ECs. Specific gene transcripts are highlighted with enlarged dots in red; respectively, light or dark gray dots indicate gene transcripts differentially expressed above or below cutoffs, $-\log_{10}(p \text{ value}) < 1$ and $|\log_2FC| < 2.5$.



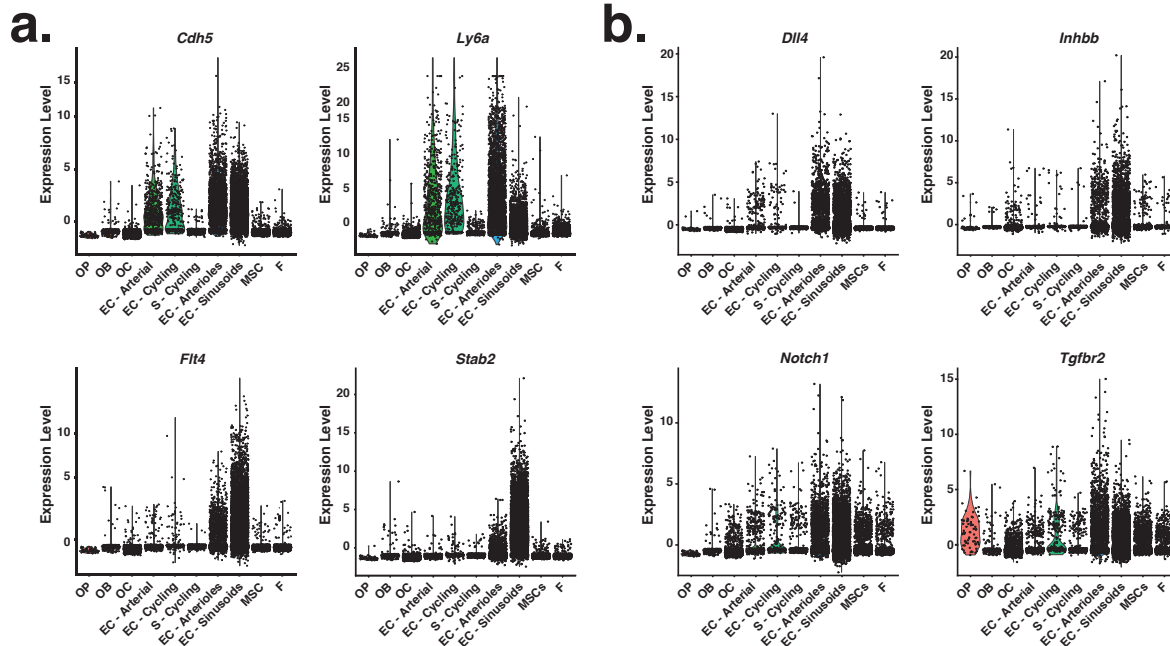
Supplementary Figure 2. *Jak3* protein expression in WT and *Jak3*^{-/-} mice.

Representative images of chromogen-based immunohistochemistry staining of WT (top) and *Jak3*^{-/-} (bottom) femurs ($n = 3$ per group). Dashed line indicates trajectory of arteriole. Clockwise from top left, scale bars are 1 mm, 50 μm , 50 μm , and 20 μm for each group.

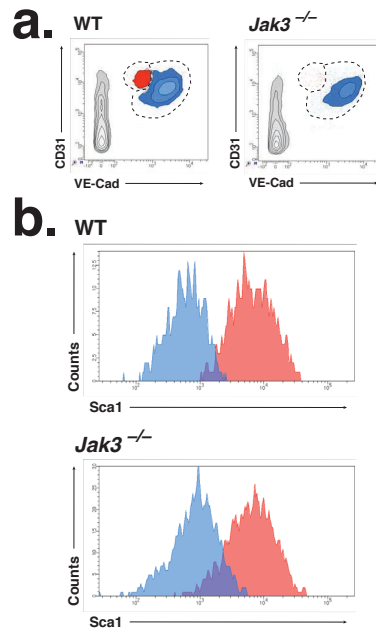


Supplementary Figure 3. Lung and bone marrow endothelial cell (EC) signatures in WT and *Jak3*^{-/-} mice.

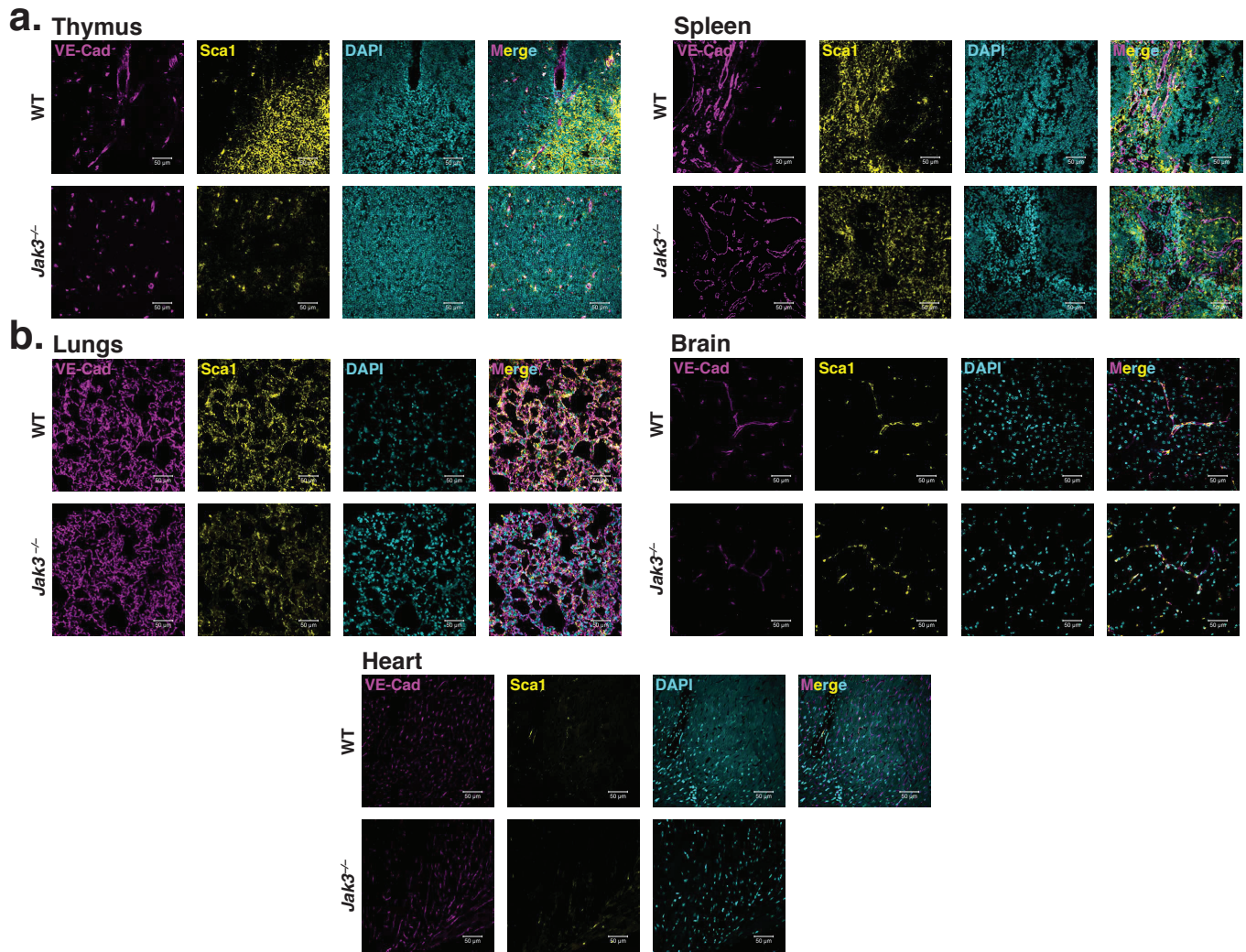
(a) Heatmaps depicting normalized transcript abundances of genes that contributed to the dysregulated Kyoto Encyclopedia of Genes and Genomes (KEGG)⁵⁴⁻⁵⁶ terms generated by Gene Set Enrichment Analysis (GSEA) as depicted in Fig. 2c that are missing from Fig. 2d ($n = 2$ or 3 per organ). (b) Volcano plot illustrating differential expression analysis of WT vs. *Jak3*^{-/-} bone marrow (top) and lung (bottom) ECs. Arterial and sinusoidal gene transcripts are highlighted with enlarged dots in red (bone marrow) and blue (lung); respectively, light or dark gray dots indicate gene transcripts differentially expressed above or below cutoffs, $-\log_{10}(\text{p value}) < 1$ and $|\log_2\text{FC}| < 2.5$.



Supplementary Figure 4. Specific transcript expression in the bone marrow microenvironment at the single-cell level. (a) Violin plots illustrating canonical pan-endothelial (*Cdh5*), arterial (*Ly6a*), and sinusoidal (*Fit4* and *Stab2*) transcript abundances. (b) Violin plots illustrating Notch signaling pathway ligand (*Dll4*) and receptor (*Notch1*) as well Tgf β signaling pathway ligand (*Inhbb*) and receptor (*Tgfr2*) transcript abundances. Data were obtained from combined single-cell RNA sequencing analysis of bone marrow cells from Tikhonova et al. (2019; GEO: GSE108892) and Baryawno et al. (2019; GEO: GSE128423). Each dot represents one cell.

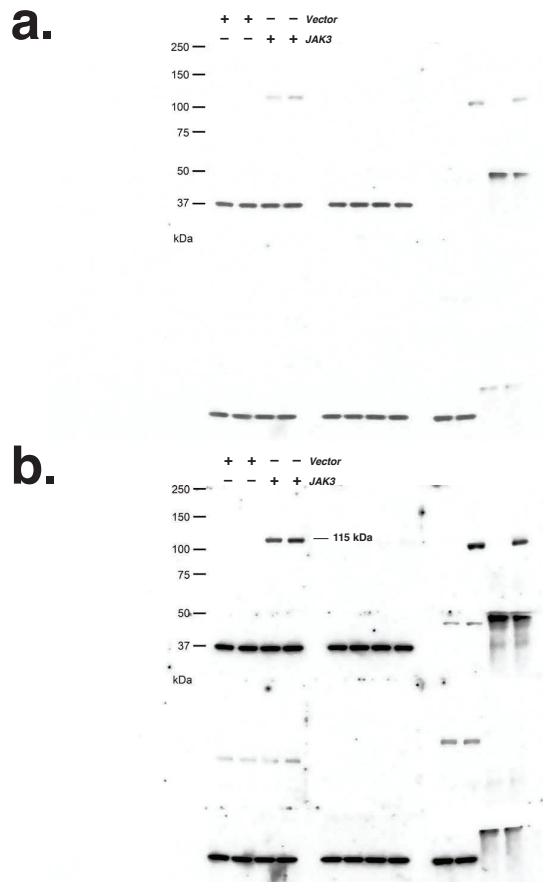


Supplementary Figure 5. Flow cytometry of primary WT and *Jak3*^{-/-} bone marrow endothelial cells (ECs). (a) Representative flow cytometry plots showing bone marrow EC surface expression of canonical pan-endothelial markers CD31 and VE-Cad in WT (left) and *Jak3*^{-/-} (right) mice ($n = 10$). (b) Representative flow cytometry histograms showing bone marrow EC surface expression of Sca1 in WT (top) and *Jak3*^{-/-} (bottom) mice ($n = 10$).

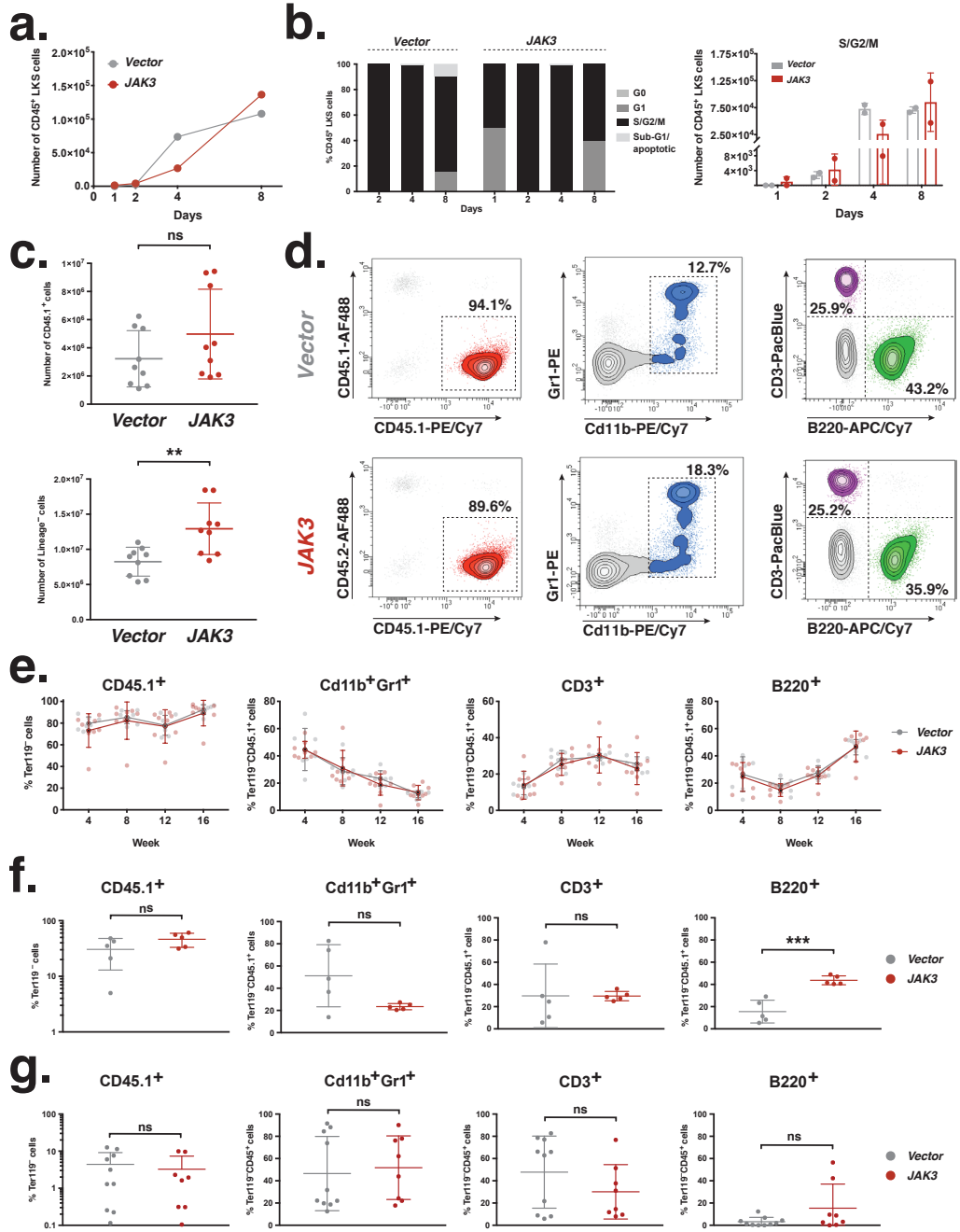


Supplementary Figure 6. Sca1 expression in various organs from WT and *Jak3^{-/-}* mice.

(a) Representative anti-VE-Cad (magenta), anti-Sca1 (yellow), and DAPI (cyan) immunofluorescent staining of WT (top) and *Jak3^{-/-}* (bottom) of indicated secondary hematopoietic organs ($n = 3$). (b) Representative anti-VE-Cad (magenta), anti-Sca1 (yellow), and DAPI (cyan) immunofluorescent staining of WT (top) and *Jak3^{-/-}* (bottom) of indicated non-hematopoietic organs ($n = 3$). Scale bars are 50 μm .

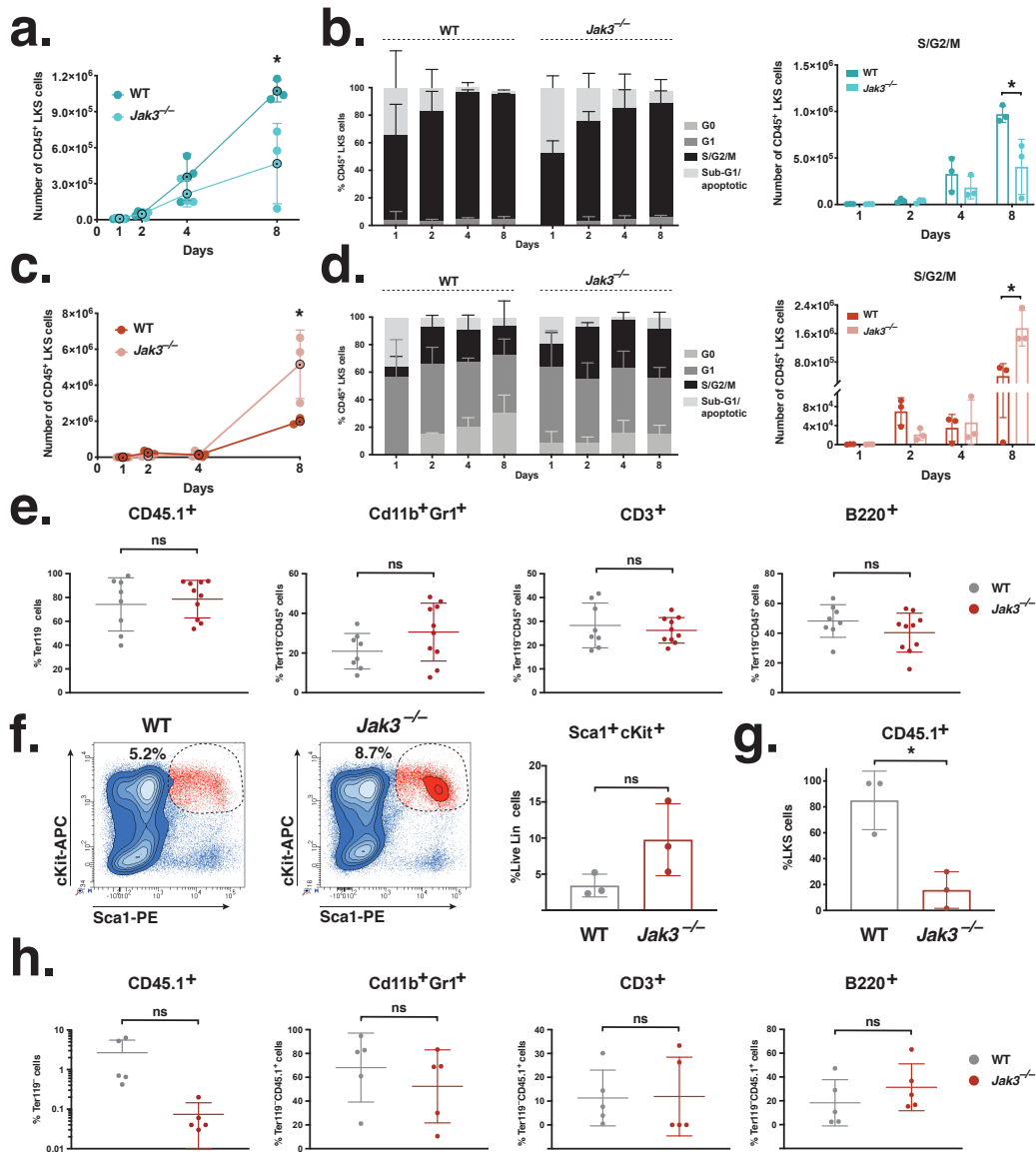


Supplementary Figure 7. Uncropped Western blots probing JAK3 protein in JAK3-transduced human ECs (ECs).
(a) Full, uncropped Western blot membrane developed with exposure to observe GAPDH protein expression at 37 kDa.
(b) Full, uncropped Western blot membrane developed with exposure to observe JAK3 protein expression at 115 kDa, as indicated on the right. The protein ladder used as reference has been recreated manually on the left. Both blots correspond to *Vector*- or *JAK3*-transduced ECs for in vitro co-culture experiments ($n = 2$). They have been cropped for display on Fig. 3a.



Supplementary Figure 8. Characterization of Lineage⁻cKit⁺Sca1⁺ (LKS) cell expansion on Vector- and JAK3-overexpressing human endothelial cells (ECs).

(a) Proliferation study quantifying the number of LKS cells in co-culture with *Vector-* (grey) or *JAK3-*(red) transduced ECs on days 1, 2, 4, and 8 of expansion ($n = 2$). (b) Cell cycle analysis of LKS cells in co-culture with *Vector-* (grey) or *JAK3-*(red) transduced endothelial cells (ECs) on days 1, 2, 4, and 8 of expansion (left) and total number of cycling (S/G2/M) LKS cells on each time point (right; $n = 2$). Each replicate is represented by an individual dot. (c) Measurement of CD45.1⁺ (top) and Lineage⁻ (bottom) cell expansion respectively gated on DAPI⁻ and DAPI⁻CD45.1⁺ single cells by flow cytometry. Center lines represent the average number of cells. (d) Representative flow cytometry plots depicting peripheral blood (PB) chimerism (left, red) as well as myeloid (center, blue), T lymphoid (right, purple), and B lymphoid (right, green) staining 16 weeks following transplantation of LKS cells into lethally irradiated CD45.2⁺ recipients after co-culture with *Vector-* (top) or *JAK3-* (bottom) transduced ECs ($n = 10$ per group). (e) Quantification of monthly flow cytometry analyses of chimerism and lineage contribution to the PB over the course of 4 months following competitive transplantation. From left to right: donor chimerism, myeloid cells, T lymphoid cells, and B lymphoid cells. (f) Chimerism and lineage distribution of PB from murine cohort that received highest cell dose 20 weeks after limiting-dilution transplantation of LKS cells following co-culture with *Vector-* (grey) or *JAK3-* (red) transduced ECs ($n = 10$ per group). From left to right: donor chimerism, myeloid cells, T lymphoid cells, and B lymphoid cells. Center lines represent the average number of cells. (g) Chimerism and lineage distribution of PB from secondary-transplanted mice that received whole bone marrow from highest-dose, primary-transplanted experimental (*Vector*, grey) and control (*JAK3*, red) groups 16 weeks after limiting-dilution transplantation. From left to right: donor chimerism, myeloid cells, T lymphoid cells, and B lymphoid cells. Center lines represent the average number of cells. Three asterisks (***) , two asterisks (**), and ns (not significant) denote, respectively, p values < 0.001 , < 0.01 , and > 0.05 by unpaired, two-tailed Student's t test. On all plots, each replicate is represented by an individual dot. On line plots, mean values are joined by a connecting line and every replicate is depicted in a lighter shade of its corresponding color. All error bars represent the standard deviation from the mean.



Supplementary Figure 9. Characterization of Lineage⁻cKit⁺Sca1⁺ (LKS) cell expansion on WT and *Jak3*^{-/-} lung and bone marrow endothelial cells (ECs).

(a) Proliferation study quantifying the number of LKS cells in co-culture with WT (dark blue) or *Jak3*^{-/-} (light blue) murine lung ECs on days 1, 2, 4, and 8 of expansion (*n* = 3). (b) Cell cycle analysis of LKS cells in co-culture with WT or *Jak3*^{-/-} murine lung ECs on days 1, 2, 4, and 8 of expansion (left) and total number of cycling (S/G2/M) LKS cells on each time point (right; *n* = 3). (c) Proliferation study quantifying the number of Lineage⁻cKit⁺Sca1⁺ (LKS) cells in co-culture with WT (dark red) or *Jak3*^{-/-} (light red) murine bone marrow ECs on days 1, 2, 4, and 8 of expansion (*n* = 3). (d) Cell cycle analysis of LKS cells in co-culture with WT or *Jak3*^{-/-} murine bone marrow ECs on days 1, 2, 4, and 8 of expansion (left) and total number of cycling (S/G2/M) LKS cells on each time point (right; *n* = 3). (e) Chimerism and lineage distribution of peripheral blood (PB) 20 weeks following primary transplantation into WT (grey) or *Jak3*^{-/-} (red) mice, prior to limiting-dilution secondary transplantation (*n* = 8 and 10, respectively). From left to right: donor chimerism, myeloid cells, T lymphoid cells, and B lymphoid cells. Center lines represent the average number of cells. (f) Representative flow cytometry plots depicting bone marrow-derived LKS cells from primary-transplanted mice 20 weeks following primary transplantation into WT (grey) or *Jak3*^{-/-} (red) mice, prior to limiting-dilution secondary transplantation (left and center; *n* = 3 per group) and averaged LKS cell percentages (right). (g) Averaged percent chimerism of LKS sub-population 20 weeks following primary transplantation into WT (grey) or *Jak3*^{-/-} (red) mice (*n* = 3 per group). (h) Chimerism and lineage distribution of PB from murine cohort that received highest cell dose 20 weeks after secondary transplantation of LKS in a limiting-dilution manner (*n* = 5 per group), including specimens that did not meet the threshold for engraftment (0.2% and higher percent chimerism). From left to right: donor chimerism, myeloid cells, T lymphoid cells, and B lymphoid cells. Center lines represent the average number of cells. One asterisk (*) and ns (not significant) denote, respectively, *p* values < 0.05 and > 0.05 by unpaired, two-tailed Student's *t* test. On all plots, each replicate is represented by an individual dot. On line plots, mean values are joined by a connecting line and every replicate is depicted in a lighter shade of its corresponding color. All error bars represent the standard deviation from the mean.