# nature research

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Last updated by author(s):	Feb 10, 2021

# **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	ali statisticai an	alyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.			
n/a	Confirmed				
	The exact	sample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement			
	🔀 A stateme	ent on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
	The statis Only comm	tical test(s) used AND whether they are one- or two-sided non tests should be described solely by name; describe more complex techniques in the Methods section.			
$\boxtimes$	A descript	cion of all covariates tested			
	A descript	ion of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons			
	A full desc	full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) ND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)			
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.				
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings				
$\boxtimes$	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes				
	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated				
Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.					
Software and code					
Poli	cy information	about <u>availability of computer code</u>			
Dá	ata collection	Andor iQ software was used to collect data			
Da	ata analysis	ImageJ-FIJI was used to prepare microscope images for analysis MATLAB (Mathworks) version R16 was used for anlaysing images AFM data were analysed using JPK Analysis software (JPK DP, JPK Instruments) Western blots were analysed using Image Studio Lite (LI-COR Biotechnology, USA)			

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data sets generated and analysed during the current study are available from the corresponding author on reasonable request.

Field-specific reporting				
<u>.</u>	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.			
Life sciences	Behavioural & social sciences			
	the document with all sections, see <a href="mailto:nature.com/documents/nr-reporting-summary-flat.pdf">nature.com/documents/nr-reporting-summary-flat.pdf</a>			
Life scier	nces study design			
All studies must dis	sclose on these points even when the disclosure is negative.			
Sample size	Sample sizes were chosen such that the mean and standard deviation can be reliably estimated. Thus most of the experiments included 5 or more independent biological replicates.			
Data exclusions	No data were excluded			
Replication	Replication of the experiments were successful. Experiments were performed independantly atleast 3 times to test replicability			
Randomization	This is not relevant for this study as samples were not grouped			
Blinding	Blinding is not relevant for this study as the samples were not grouped			
Reporting for specific materials, systems and methods  We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.  Materials & experimental systems  Methods    Involved in the study				
Antibodies used  Lamin C: obtained from Developmental Studies Hybridoma Bank (DSHB), Cat No. LC28.26				
Antibodies dised	Lamin DM0: Obtained from DSHB, Cat No. ADL67.10			
	Lamin A: Obtained from Thermo Fischer, Cat No. MA1-06101, Clone Name: 133A2 ZO1: Obtained from Thermo Fischer, Cat No. 40-2200			
	phospho Ser22 LamA: Obtained from Cell Signaling Technologies, cat No. 2026S  Lam B1: Obtained from Abcam, Cat No. ab8982			
	Actin: Obtained from Abcan, Cat No. 8227			
	Polyclonal phospho-Histone H3: obtained from Sigma Aldrich, Cat No. 06-570 Goat anti-mouse Alexa 647: Obtained from Thermo Fischer, Cat No. A28181			
	Goat anti-mouse Alexa 488: Obtained from Thermo Fischer, Cat No. A-21121 Donkey anti-Rabbit IgG IRdye 680 : Obtained from LiCor, Cat No. 926-68073			
	Donkey anti-Mouse IRdye 800: Obtained from LiCor, Cat No. 926-32212			
Validation	Lamin A: The antibody was verified by relative expression to ensure that the antibody binds to the antigen stated. ZO-1: This Antibody was verified by Knockdown to ensure that the antibody binds to the antigen stated.			
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## Eukaryotic cell lines

Policy information about <u>cell lines</u>

Cell line source(s)

Madin Darby Canine Kidney cells (MDCK II), Public Health England, cat#00062107

Authentication

The cell line was not authenticated

Mycoplasma contamination

Commonly misidentified lines (See <u>ICLAC</u> register)

No

### Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

The cell line is tested negative for mycoplasma

Laboratory animals

Drosophila melanogaster. Following strains were used:

w-: DECADGFP: Obtained from Hong Lab

w-;UAS-CD8::Cherry;Dpp-GAL4,Gal80ts: Obtained from Dahmann Lab, TU Dresden

yw;vkg::GFP and w-;Ptc::Gal4,Gal80ts : Obtained from MPI\_CBG Fly Facilty

w-;UAS-mmp2: Obtained from Bloomington Stock Center (#58705)

w-;UAS-Lin: Obtained from Bloomington Stock Center(#7074)

w-;UAS-CDC42F89: Obtained from Bloomington Stock Center(#6286)

w-;UAS-CDC42L89: Obtained from Bloomington Stock Center(#6289)

w-;UAS-ykiGFP : Obtained from Bloomington Stock Center(#28815) w-;UAS-ykiS168A: Obtained from Bloomington Stock Center(#28818)

w-;trol::GFP: Obtained from Bloomington Stock Center(#60214)

w-;UAS-Stinger::GFP: : Obtained from Bloomington Stock Center (#84277)

w-;Df(2R)trix/CyO: Obtained from Bloomington Stock Center(#1896)

w-;Laminin-A::GFP: Obtained from Vienna Drosophila Stock Center (#318155)

UAS-LamC: Obtained from the Wallrath Lab

w-;LamCEX296/CyO: Obtained from the Wallrath Lab.

w-;UAS-LamCRNAi: Obtained from National Institute Genetics (#10119R-1)

w-;UAS-MMP2,Vkg::GFP : Generated in this work w-;CD8::Cherry; Ubx-GAL4: Generated in this work

Wild animals

The study did not involve wild animals

Field-collected samples

The study did not involve field collected samples

Ethics oversight

No ethical approval or guidance was required as the study used only Drosophila melanogaster

Note that full information on the approval of the study protocol must also be provided in the manuscript.