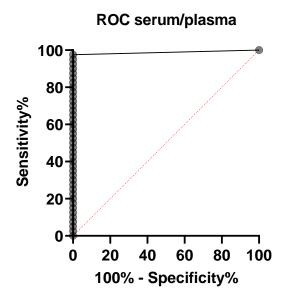
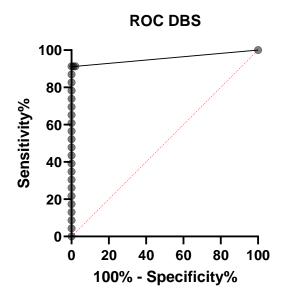
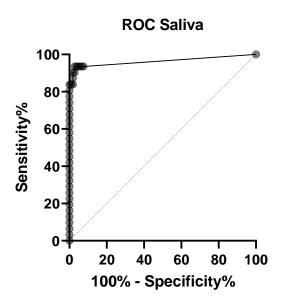
Supplementary Information.

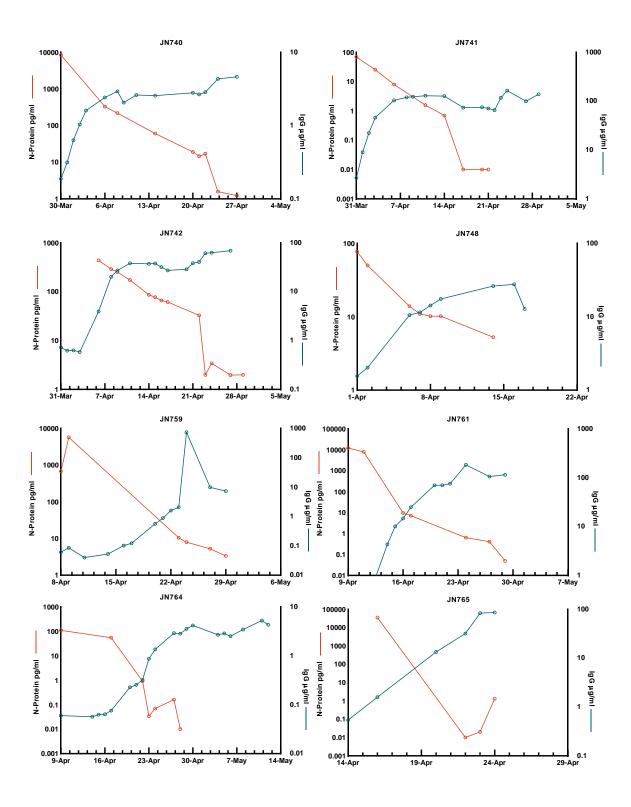
N-protein presents early in blood, dried blood and saliva during asymptomatic and symptomatic SARS-CoV-2 infection.

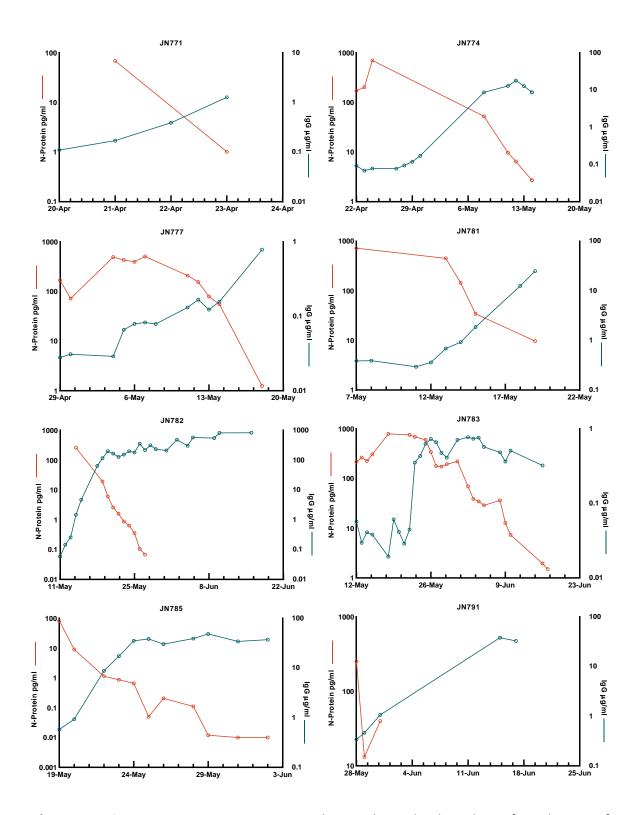






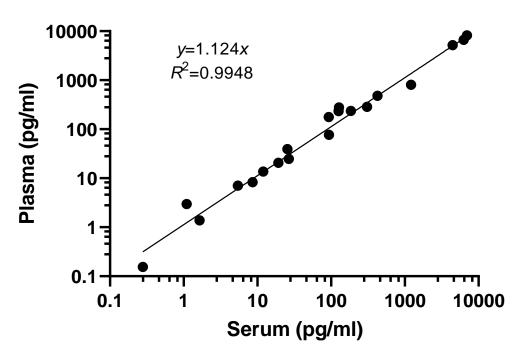
Supplementary Figure 1 – ROC curves for serum/plasma, DBS (within 14 days of positive PCR) and saliva. See Positive / Negative Cutoff in Methods for more information.





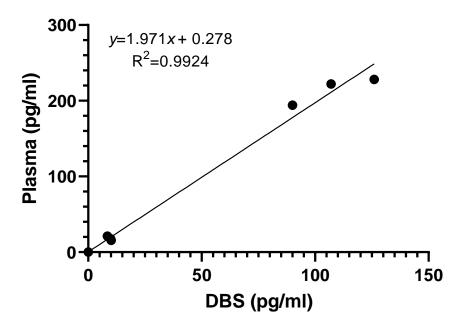
Supplementary Figure 2 – SARS-CoV-2 N-protein and anti-spike IgG levels in plasma from the Univ of Bonn cohort. Sixteen individual donors were sampled over multiple timepoints for a total of 141 data points. N-Protein is shown in red, and IgG in blue.

N-Protein Serum-Plasma Correlation



Supplementary Figure 3. Matched serum and plasma samples from the same donors were found to have correlation in N antigen levels between matrices. Twenty matched samples from BocaBio confirmed to be PCR+ were tested in both serum and K2 EDTA plasma.

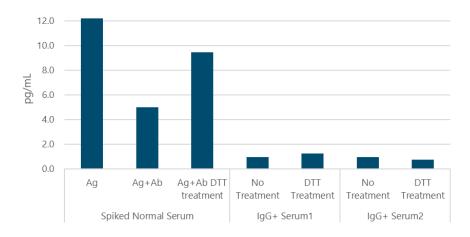
N-Protein DBS-Plasma Correlation



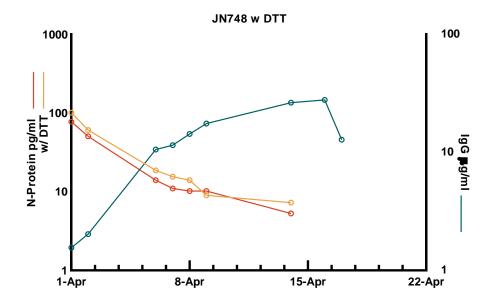
Supplementary Figure 4. Whole blood drawn into a K2 EDTA plasma tube (3 donors) was spiked with known levels of recombinant N-protein. It was then processed into neat plasma and in parallel into Dried Blood Microsamples (DBS) using Neoteryx Mitra tips. After extraction, both sample types were measured, showing a correlation of 0.99. The concentration in DBS was approximately ½ of that in plasma, as expected due to the excluded volume of hematocrit which is separated from plasma.

A.

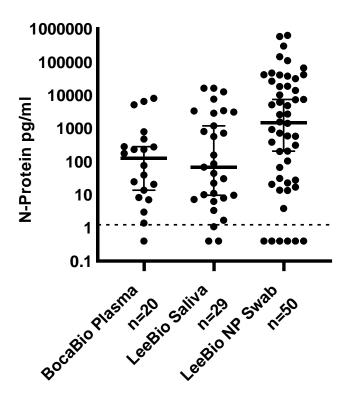
Nucleocapsid Level in Serum Samples



В.



Supplementary Figure 5. Panel A. A DTT reduction protocol was established to unmask N-protein bound by antibody in serum by doing a control experiment with recombinant antigen and capture antibody spiked into sample matrix. N-protein concentration measured in serum was reduced after co-spiking with antibody, indicative of epitope masking. Adding DTT to the sample rescued 63% of the signal loss, indicating that this treatment could unmask antigen in seroconverted samples. **Panel B.** Longitudinal plasma samples from Donor 4 Univ of Bonn, treated with and without DTT before testing. Samples were not treated with DTT before testing for IgG. Negligible decrease in N-protein levels were observed with DTT treatment, suggesting that antibody-antigen complexes were not masking N-protein signal, or causing the observed decrease over time. N-Protein is shown in red, N-Protein from samples treated with DTT is shown in orange, and IgG is shown in blue.



Supplementary Figure 6. N-protein measured in plasma, saliva and nasopharyngeal swabs from commercial sources. The solid lines represent median values (+/- 95% confidence interval).

Table S1. Inactivated, cultured virus was purchased from Zeptometrix, and tested for cross-reactivity at the TCID₅₀ levels listed. No cross-reactivity was observed.

Virus Description	Vendor	Cat#	Titer Tested TCID ₅₀ /mL	Conc Meas antigen Assa Serum	sured by N y Plasma
Adenovirus Type 07 (Species B) Culture Fluid	Zeptometrix	0810021CF HI	3.52E+04	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Enterovirus Type 68 (2007 Isolate) Culture Fluid	Zeptometrix	0810237CF HI	3.78E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Influenza A H1N1 (New Cal/20/99) Culture Fluid	Zeptometrix	0810036CF HI	2.88E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Influenza B (Florida/02/06) Culture Fluid	Zeptometrix	0810037CF HI	3.52E+04	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Parainfluenza Virus Type 1 Culture Fluid	Zeptometrix	0810014CF HI	2.28E+06	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Parainfluenza Virus Type 2 Culture Fluid	Zeptometrix	0810015CF HI	2.88E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Parainfluenza Virus Type 3 Culture Fluid	Zeptometrix	0810016CF HI	1.65E+06	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Parainfluenza Virus Type 4A Culture Fluid	Zeptometrix	0810060CF HI	7.05E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Respiratory Syncytial Virus Type A (Isolate: 2006 Isolate) Culture Fluid	Zeptometrix	0810040AC FHI	9.50E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Rhinovirus Type 1A Culture Fluid	Zeptometrix	0810012CF NHI	8.88E+04	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Coronavirus (Strain: 229E) Culture Fluid	Zeptometrix	0810229CF HI	1.04E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Coronavirus (Strain: OC43) Culture Fluid	Zeptometrix	0810024CF HI	2.63E+05	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>
Coronavirus (Strain: NL63) Culture Fluid	Zeptometrix	0810228CF HI	4.25E+04	<lod< td=""><td><lod< td=""></lod<></td></lod<>	<lod< td=""></lod<>