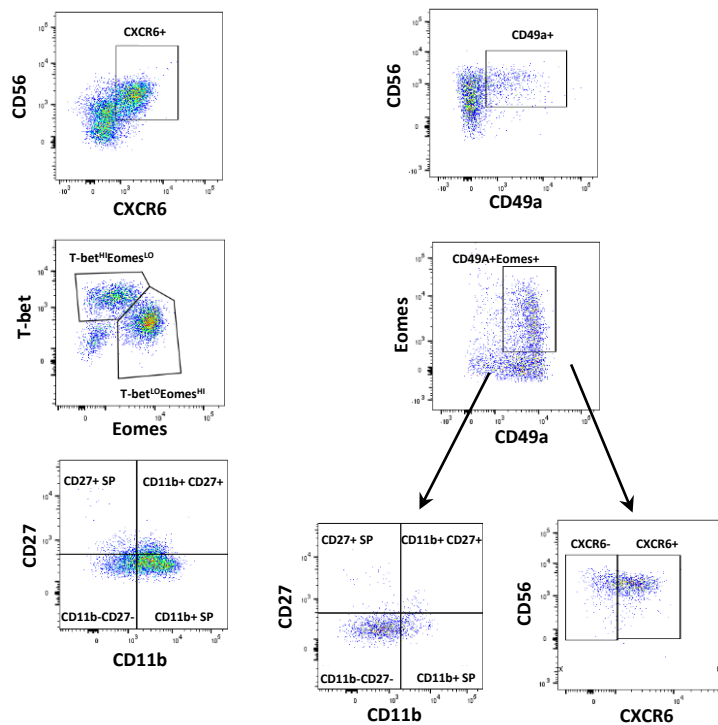
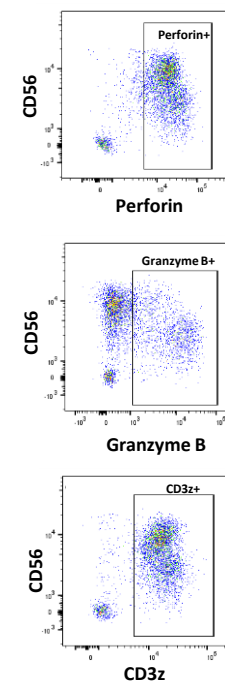


Liver residency, development and maturation markers

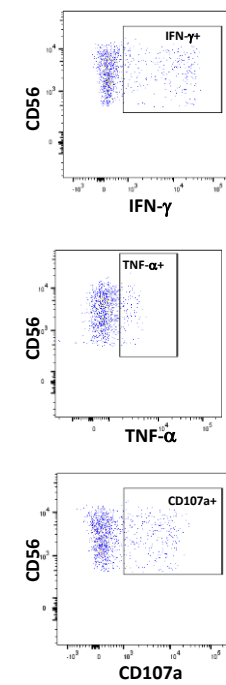


C

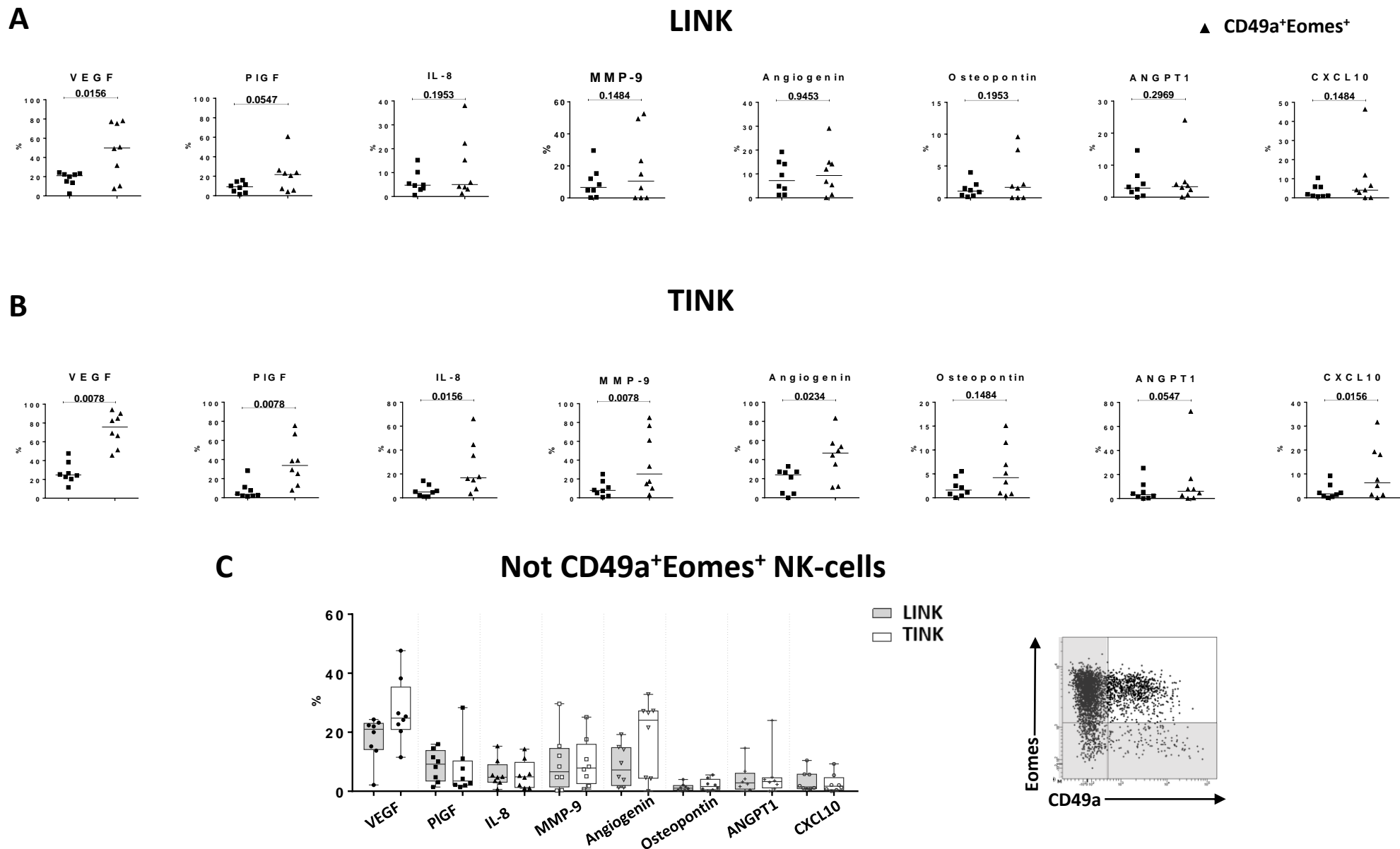
Cytotoxic granules and activating markers



Cytokine production and Degranulation potential



Supplementary Figure 1. Flow cytometry analysis representative dot plots. A: Gating strategy for the identification of infiltrating NK-cells (Lymphocytes, singlets, live cells, CD3-, total NK, CD56^{BRIGHT} and CD56^{DIM} and CD16+/-) using multicolor flow cytometry. **B:** Dot plots representing analysis example for each markers that have been used for phenotypical assessment (NKG2D, NKG2A, NKp30, NKp44, CXCR6, T-bet, Eomes, CD27, CD11b and CD49a) of NK-cells. **C:** Dot plots showing example of function analysis by Granzyme B, Perforin, CD3z, CD107a, TNF-α and IFN-γ staining.



Supplementary Figure 2. Pro-angiogenic factors production by CD49a⁺Eomes⁺ and NK-cells not co-expressing CD49a and Eomes. **A:** Frequency of cytokines positive cells in the two subsets (CD49a⁺Eomes⁺ and not CD49a⁺Eomes⁺) in the liver. **B:** Percentage of NK-cells producing angiogenic factors among CD49a⁺Eomes⁺ and NK-cells not co-expressing CD49a and Eomes in the tumor. **C:** Frequency of cytokines positive NK-cells not co-expressing CD49a and Eomes in the two compartments. On the right side, representative dot plot showing with grey background the not co-expressing CD49a and Eomes NK-cells. Pro-angiogenic factors staining was performed after overnight IL-12 and IL-18 stimulation. Horizontal lines represent median values, statistics by Wilcoxon matched pairs test (LINK vs TINK).