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Reporting Summary

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Please do not complete any field with "not applicable" or n/a. Refer to the help text for what text to use if an item is not relevant to your study. For final submission: please carefully check your responses for accuracy; you will not be able to make changes later.

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

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n/a	Cor	nfirmed
	X	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	X	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	X	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\times		A description of all covariates tested
	X	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	X	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	X	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
X		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
X		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\times		Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
'		Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

Raman measurements were acquired by WiRE 3.4 software. Ultraviolet-visible spectra were recorded by UVProbe 2.330 software. Hydrodynamic diameters and Zeta potentials were measured by Zetasizer 7.02 software. TEM images were taken with GMS 3 software. SEM images were measured by xT microscope Server 3.0.11 software. Fluorescence spectra were performed by FluorEssence V3.5 software. Electron paramagnetic resonance (EPR) measurements were recorded by ESR 2.0.0.0 software. Bruker the minispec 27010.0 software was used to measure T₂ relaxivity. T₂-weighted MR imaging was taken by Proc_Csharp 1.0.0.0 software. Data from Inductively-coupled plasma mass spectrometry were acquired using IcpMHLauncher 3.01.0005 software.

Data analysis

Origin 9.0 for Raman, UV, Hydrodynamic diameters and Zeta potentials, EPR measurements, T_2 relaxivity and ICP-MS measurements analysis. Image J 2006.02.01 software for T_2 -weighted MR imaging analysis. Statistical analysis were done using GraphPad Prism 6 (GraphPad Software, Inc.).

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

All data that support the findings of this study are available within the article and the Supplementary Information or from the corresponding author upon request. Source data are provided with this paper.

Field-spe	ecific reporting		
Please select the o	ne below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.		
X Life sciences	Behavioural & social sciences Ecological, evolutionary & environmental sciences		
For a reference copy of	the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf		
Life scier	nces study design		
	sclose on these points even when the disclosure is negative.		
Sample size	No specific sample-size calculation was performed during experiments. Adequate sample sizes are generally determined based on extensive laboratory experience and capable of statistical analysis and described in the figure legend.		
Data exclusions	No data were excluded from the analyses.		
Replication	Each experiment was performed 3-6 independent replications and the exact n-numbers were described in the figure legend.		
Randomization	All mice in the study were randomly assigned to the experimental groups. Enough replications and the control groups were carried out in consistent experimental condition.		
Blinding	The investigators were not blind to mice allocation during data collection and analysis. All the data of T ₂ -weighted MF imaging, Raman imaging, Gram staining and Relative 16S rRNA gene level were collected by the instruments directly leaving no room for subjectivity.		
Reportin	g for specific materials, systems and methods		
We require informat	ion from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, ted is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & ex	perimental systems Methods		
n/a Involved in th	ne study n/a Involved in the study		
Antibodies			
Eukaryotic			
	logy and archaeology MRI-based neuroimaging		
	nd other organisms search participants		
Clinical da			
	esearch of concern		
Eukaryotic c	rell lines		
Policy information	about <u>cell lines</u>		

Policy information about <u>cell lines</u>	
Cell line source(s)	The GES-1 cell line used in this study was obtained from iCell Bioscience Inc, Shanghai.
Authentication	Cell lines were authenticated using Short Tandem Repeat (STR) analysis.
Mycoplasma contamination	The cell line used in the study was tested for mycoplasma contamination and the result was negative.
Commonly misidentified lines (See ICLAC register)	No commonly misidentified lines were used.

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

BALB/c female mice 3-4 weeks old (housing conditions, dark/light cycle: 12/12 hours, temperature: 20°C, humidity: about 40%) were used in the study.

Wild animals The study did not involve wild animals.

Field-collected samples The study did not involve samples collected from the field.

Ethics oversight The study protocols were approved by the Institutional Animal Care and Use Committee of Hunan University.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

