

Supplementary material

Drivers of ESBL-producing *Escherichia coli* dynamics in calf fattening farms: a modelling study

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SM1. Longitudinal data of ESBL-EC carriage in calves

Table S1. ESBL-EC carriage across time in calves. The last sampling day was day 161 in farms A and B, and day 147 in farm C. ESBL-EC positive (resp. negative) samples are represented as 1 (resp. 0).

Farm	Pen	Calf	Sampling days											
			7	21	35	49	63	77	91	106	119	133	147	161
A	A-1	1	1	1	1	1	1	0	0	0	0	0	0	0
A	A-1	2	1	1	1	1	0	0	0	0	0	0	0	0
A	A-1	3	1	1	1	1	1	1	0	0	0	0	0	1
A	A-1	4	1	1	1	1	1	0	0	0	0	0	0	0
A	A-1	5	1	1	1	1	0	0	0	0	0	0	0	0
A	A-2	6	1	1	1	1	1	0	0	0	0	0	0	0
A	A-2	7	1	1	1	1	1	0	0	0	0	0	0	0
A	A-2	8	1	1	1	1	1	1	0	0	0	0	0	0
A	A-2	9	1	1	1	1	1	0	0	0	0	0	0	0
A	A-2	10	1	1	1	1	1	1	0	0	0	0	0	0
A	A-3	11	1	1	1	1	1	1	0	0	1	0	1	0
A	A-3	12	1	1	1	1	1	0	0	0	0	0	0	0
A	A-3	13	1	1	1	1	1	1	1	0	0	0	1	0
A	A-3	14	1	1	1	1	1	0	0	0	0	0	0	0
A	A-3	15	1	1	1	1	1	1	0	0	0	0	0	0
B	B-1	16	0	0	0	0	0	0	0	0	0	0	0	0
B	B-1	17	0	0	0	0	0	0	0	0	0	0	0	0
B	B-1	18	0	0	0	0	0	0	0	0	0	0	0	0
B	B-1	19	0	0	0	0	0	0	0	0	0	0	0	0
B	B-1	20	0	0	0	0	0	0	0	0	0	0	0	0

B	B-2	21	1	0	0	1	0	0	0	0	0	0	0	0
B	B-2	22	1	0	0	0	0	0	0	0	0	0	0	0
B	B-2	23	1	1	1	0	0	0	0	0	0	0	0	0
B	B-2	24	1	0	0	0	0	0	0	0	0	0	0	0
B	B-2	25	1	1	0	0	0	0	0	0	0	0	0	0
B	B-3	26	1	0	0	0	0	0	0	0	0	0	0	0
B	B-3	27	1	0	1	0	0	0	0	0	0	0	0	0
B	B-3	28	1	0	0	0	0	0	0	0	0	0	0	0
B	B-3	29	1	0	0	0	0	0	0	0	0	0	0	0
B	B-3	30	1	0	0	0	0	0	0	0	0	0	0	0
C	C-1	31	0	1	1	0	0	0	0	0	1	1	1	/
C	C-1	32*	0	1	1	0	/	/	/	/	/	/	/	/
C	C-1	33	0	0	1	0	0	0	0	0	1	1	0	/
C	C-1	34	0	0	1	0	1	0	0	0	1	0	0	/
C	C-2	35	1	1	1	0	1	0	0	0	1	1	1	/
C	C-2	36	1	1	1	0	0	0	1	0	1	0	0	/
C	C-2	37	1	1	1	0	0	0	0	0	1	1	0	/
C	C-2	38	1	1	1	0	0	0	0	1	1	1	1	/
C	C-2	39	1	0	1	0	1	0	0	1	1	1	0	/
C	C-3	40	1	1	1	1	1	0	0	0	1	1	0	/
C	C-3	41	1	0	1	0	0	0	0	0	1	1	0	/
C	C-3	42	1	0	1	0	1	0	0	0	1	1	0	/
C	C-3	43	1	0	1	0	1	0	0	1	0	1	0	/
C	C-3	44	1	0	1	1	1	1	0	1	1	1	0	/
C	C-3	45	1	0	1	0	0	0	0	0	1	1	1	/

9 * Calf #32 died between the 4th and the 5th sampling days.

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15 SM2. Details on the models

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17 A schematic representation of the model is presented in Figure 1 of the main text. We assumed
18 the dynamics of ESBL-EC were independent of other *E. coli*. At time t , each ESBL-EC negative
19 calf could acquire an ESBL-EC with a probability

$$20 \quad P = 1 - e^{-A(t)}$$

21 where $A(t)$ was the acquisition rate, which formula depended on the model variant as detailed
22 in Table S2. $A(t)$ was generally defined as:

$$23 \quad A(t) = [A_{trans}(t) + A_{spor}(t)] \times \lambda_a(t)$$

24 with, at time t , $A_{trans}(t)$ the acquisition term related to between-calves transmission, $A_{spor}(t)$ the
25 term related to sporadic contaminations and $\lambda_a(t)$ the multiplicative effect related to AMU.

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28 *Transmission model.* Transmission was assumed to occur either homogeneously between calves
29 of the same farm F , with rate β_r^F , or between calves depending on their attributed pen,
30 assuming two transmission rates, within (β_w^F) and between (β_b^F) pens of a farm F . The
31 transmission rate between individual pens was assumed to be the same as between collective
32 pens. Transmission was supposed to be proportional to the proportion of calves carrying
33 ESBL-EC. As a null hypothesis, we also investigated models, which did not include any
34 transmission between calves, but instead a constant, farm-specific ESBL-EC acquisition rate
35 β_0^F . Therefore, at time t :

$$36 \quad A_{trans}(t) = \beta_r^F \left(\frac{I}{S+I}\right)_{F, t-1} \text{ for homogeneous mixing (models 1a, 3a, 5a, 1b, 3b and 5b)}$$

$$37 \quad = \beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1} \text{ for pen-specific mixing (models 2a, 4a, 6a, 2b, 4b and}$$

38 6b)

$$39 \quad = \beta_0^F \text{ for the baseline acquisition rate (models, 0a, 7a, 8a, 0b, 7b and 8b)}$$

40 where I was the number of ESBL-EC positive calves in the farm (within the pen, outside the
41 pen or globally in the farm) and S the number of ESBL-EC negative calves.

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43 *Sporadic contamination.* We also assumed that sporadic contamination could occur, depicting
44 the possible acquisition of ESBL-EC by the calves on farm F on some specific days from another
45 (unknown) source other than the colonised calves. To model this process, we assumed that
46 sporadic contamination events in a given farm increased the acquisition rate $A(t)$ of all calves
47 on the farm on the day of contamination by a factor μ . We estimated N^F , the number of days
48 with contamination events that occurred across the follow up, and D^F , the specific dates of
49 contamination. This mechanism was activated in models 0a to 8a.

$$50 \quad \text{We defined: } A_{spor}(t) = \mu \text{ if } t \in D^F \quad (= 0 \text{ otherwise})$$

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52 *Clearance.* ESBL-EC positive calves could clear carriage with a probability $1 - e^{-C(t)}$, where $C(t)$
53 was the clearance rate at time t . In the baseline model, we assumed a natural clearance rate, ν_0
54 (inverse of the baseline carriage duration).

55 *Impact of antibiotics.* We modelled the possible effect of AMU through two mechanisms. On the
 56 one hand, we assumed that AMU could affect the acquisition rate $A(t)$. For a given antibiotic
 57 class i , this was modelled by a multiplicative factor $\alpha_{a,i}$. After the end of exposure, this effect
 58 was supposed to persist [1], but decrease exponentially [2] with a rate τ , common to all
 59 antibiotic classes, until it reaches a saturation at the value of 1 (see SM7 of this Supplementary
 60 material for illustration). This mechanism was active for models 3a, 4a, 7a, 3b, 4b and 7b (Table
 61 1, main text).

62 Therefore, in these models, at time t ,

$$63 \quad \lambda_a(t) = \prod_i \alpha_{a,i}^{\max(0; 1-\tau E_i(t))} \quad (= 1 \text{ otherwise})$$

64 where $E_i(t)$ was the number of days since the end of the last exposure of the calf to antibiotic
 65 class i . Therefore, if a calf was exposed to antibiotic class i on day t , $E_i(t)=0$. During model
 66 fitting, the antimicrobial exposure considered was only that observed in the longitudinal
 67 study.

68 On the other hand, we assumed that antibiotics could impact clearance by modulating the
 69 baseline clearance rate v_0 as follows (activated in models 5a, 6a, 8a, 5b, 6b and 8b):

$$70 \quad C(t) = v_0 \cdot \lambda_c(t) = v_0 \cdot \prod_i \alpha_{c,i}^{\max(0; 1-\tau E_i(t))} \quad (= v_0 \text{ otherwise})$$

71 where $E_i(t)$ and τ were as defined above, and $\alpha_{c,i}$ was a multiplicative factor analogous to $\alpha_{a,i}$,
 72 representing the effect of exposure to antibiotic class i on the clearance rate $C(t)$.

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90 **Table S2. Expressions of the acquisition rate $A(t)$ and clearance rate $C(t)$, for the considered**
 91 **models.**

92 $\lambda_a(t) = \prod_i \alpha_{a,i}^{\max(0; 1-\tau E_i(t))}$ and $\lambda_c(t) = \prod_i \alpha_{c,i}^{\max(0; 1-\tau E_i(t))}$. Parameter $\alpha_{a,i}$ (resp. $\alpha_{c,i}$) corresponds
 93 to the effect of antimicrobial class i on acquisition (resp. clearance). $E_i(t)$ is the number of days since the
 94 end of the last exposure to antibiotic class i , while τ is the rate of the exponential decay of the
 95 antimicrobial's effect over time, after the end of exposure. I (resp. S) is the number of ESBL-EC positive
 96 (resp. negative) calves, while t represents the day and F is either farm A, B or C.

Model	Acquisition rate $A(t)$	Clearance rate $C(t)$
0a	$A_{spor}(t) + \beta_0^F$	ν_0
1a	$A_{spor}(t) + \beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}$	ν_0
2a	$A_{spor}(t) + \beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}$	ν_0
3a	$[A_{spor}(t) + \beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}] \times \lambda_a(t)$	ν_0
4a	$[A_{spor}(t) + \beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}] \times \lambda_a(t)$	ν_0
5a	$A_{spor}(t) + \beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}$	$\nu_0 \times \lambda_c(t)$
6a	$A_{spor}(t) + \beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}$	$\nu_0 \times \lambda_c(t)$
7a	$[A_{spor}(t) + \beta_0^F] \times \lambda_a(t)$	ν_0
8a	$A_{spor}(t) + \beta_0^F$	$\nu_0 \times \lambda_c(t)$
0b	β_0^F	ν_0
1b	$\beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}$	ν_0
2b	$\beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}$	ν_0
3b	$[\beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}] \times \lambda_a(t)$	ν_0
4b	$[\beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}] \times \lambda_a(t)$	ν_0
5b	$\beta_f^F \left(\frac{I}{S+I}\right)_{F, t-1}$	$\nu_0 \times \lambda_c(t)$
6b	$\beta_w^F \left(\frac{I}{S+I}\right)_{in\ pen, t-1} + \beta_b^F \left(\frac{I}{S+I}\right)_{out\ pen, t-1}$	$\nu_0 \times \lambda_c(t)$
7b	$\beta_0^F \times \lambda_a(t)$	ν_0
8b	β_0^F	$\nu_0 \times \lambda_c(t)$

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100 **SM3. Code sharing and details on modelling assumptions and on the estimation using**
101 **MCMC**

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103 We explored the space of possibilities for acquisition and clearance dates for each individual
104 using data augmentation [3, 4]. Based on previous estimates of the time to clearance of ESBL-
105 producing *Enterobacteriales* [5-7], we assumed that, for a given calf, if two consecutive samples
106 were positive (resp. negative) for ESBL-EC carriage, the calf was ESBL-EC positive (resp.
107 negative) everyday between the two sampling dates.

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109 Model parameters were estimated in a Bayesian framework, using a Markov Chain Monte
110 Carlo (MCMC) algorithm, implemented with the R package *rjags* [8]. For each model variant,
111 3 chains were run for 6,000 iterations each. Depending on the model variant, from 2,000 to
112 10,000 values were discarded as burn-in, such that convergence of the Markov chains was
113 reached. We sampled every 40th value because this was enough to avoid autocorrelation within
114 the Markov chains. The effective sample size was at least 328 for posteriors of each model (953
115 for model 5a, the model selected). A Gelman-Rubin statistic below 1.1 for all parameters was
116 considered to indicate that between-chain variance was low enough compared to the within-
117 chain variance [9]. The exception was model 4 where, if convergence of each chain was
118 reached, the between-chain variance was still high, no matter the length of the burn-in. Non-
119 informative uniform priors were used for all parameters (Table 2, main text). The 18 models
120 were compared using the Deviance Information Criterion (DIC) [10]. The model for which the
121 DIC was the lowest was selected as the best model (Table S3).

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123 All analyses of this paper, including models estimation and simulation, were performed using
124 R version 3.6.1. The code is available on the following link:

125 https://github.com/JonathanBas/model_ESBL_calves.

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138 **SM4. Antimicrobial use observed in farms**

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140 In our study, no individual treatment was observed: antibiotics were always administered to
141 all calves on each farm simultaneously.

142 On farm A, five treatments were observed: colistin and sulfonamides between days 1 and 10,
143 tetracycline between days 11 and 20, doxycycline on day 53, and amoxicillin on day 136 (Figure
144 2, main text). On farm B, calves were treated with doxycycline and erythromycin on day 26,
145 tetracycline on day 90, and amoxicillin on day 101, resulting in four one-day treatments. On
146 farm C, calves were exposed to five treatments: sulfonamide-trimethoprim combination
147 between days 3 and 8, followed by tetracycline between days 10 and 16, doxycycline between
148 days 20 and 24, spiramycin between days 25 and 27, and tetracycline between days 80 and 82.

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172 **SM5. DIC of models**

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174 *Table S3. Deviance Information Criterion (DIC) of models. Model 5a presented the lowest DIC.*

Model	0a	1a	2a	3a	4a	5a	6a	7a	8a
DIC	223.6	217.7	225.5	224.0	246.2	196.3	212.0	230.3	201.9
Model	0b	1b	2b	3b	4b	5b	6b	7b	8b
DIC	256.1	238.5	248.0	236.6	250.9	219.1	232.6	251.6	239.2

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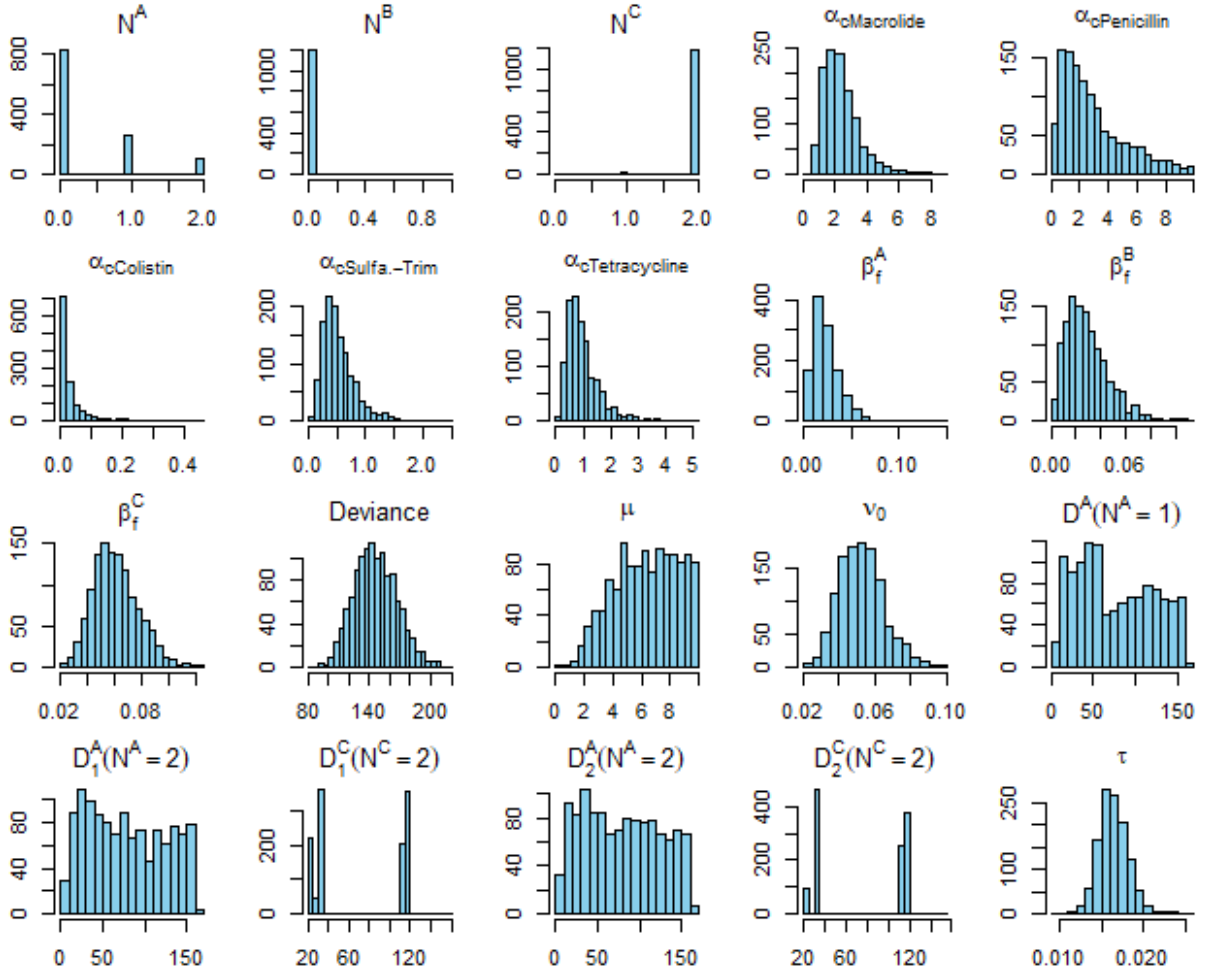
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196 **SM6. Posterior distributions in model 5a**

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200 **Figure S1. Posterior distributions for the best model, model 5a.** Parameters (and their prior
 201 distributions) are defined in Table 2 of the main text. $D^F(N^F=1)$ is the date of occurrence of the sporadic
 202 contamination event in farm F when N^F (number of sporadic contamination events in farm F) is
 203 estimated to be 1. $D_1^F(N^F=2)$ and $D_2^F(N^F=2)$ are the dates of the two sporadic contamination events in
 204 farm F when N^F is estimated to be 2. The posterior distributions of the dates of sporadic contamination
 205 events in farm A, i.e. $D^A(N^A=1)$, $D_1^A(N^A=2)$ and $D_2^A(N^A=2)$, were close to their prior distributions,
 206 because a majority (69.3%) of N^A posterior samples were 0 (see Table 3 of the main text). Therefore,
 207 these dates parameters were hardly affected by the data. Similarly, the posterior distributions of
 208 $D^B(N^B=1)$, $D_1^B(N^B=2)$, $D_2^B(N^B=2)$ corresponded to their prior distributions and were not displayed. On
 209 the contrary, these dates were clearly estimated around days 22, 35, 114 and 117 for farm C.

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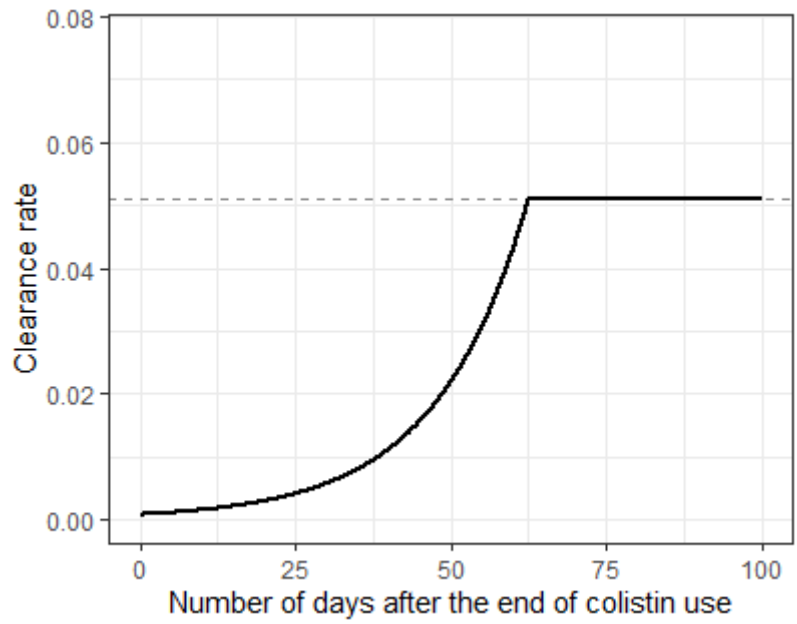
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214 SM7. Evolution of the clearance rate $C(t)$ after the end of colistin use

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217 **Figure S2.** Evolution of the clearance rate after the end of colistin use (solid line), using median
218 estimated values of parameters in Model 5a (Table 3, main text). After a delay, the clearance rate $C(t)$
219 equals v_0 (dashed line).

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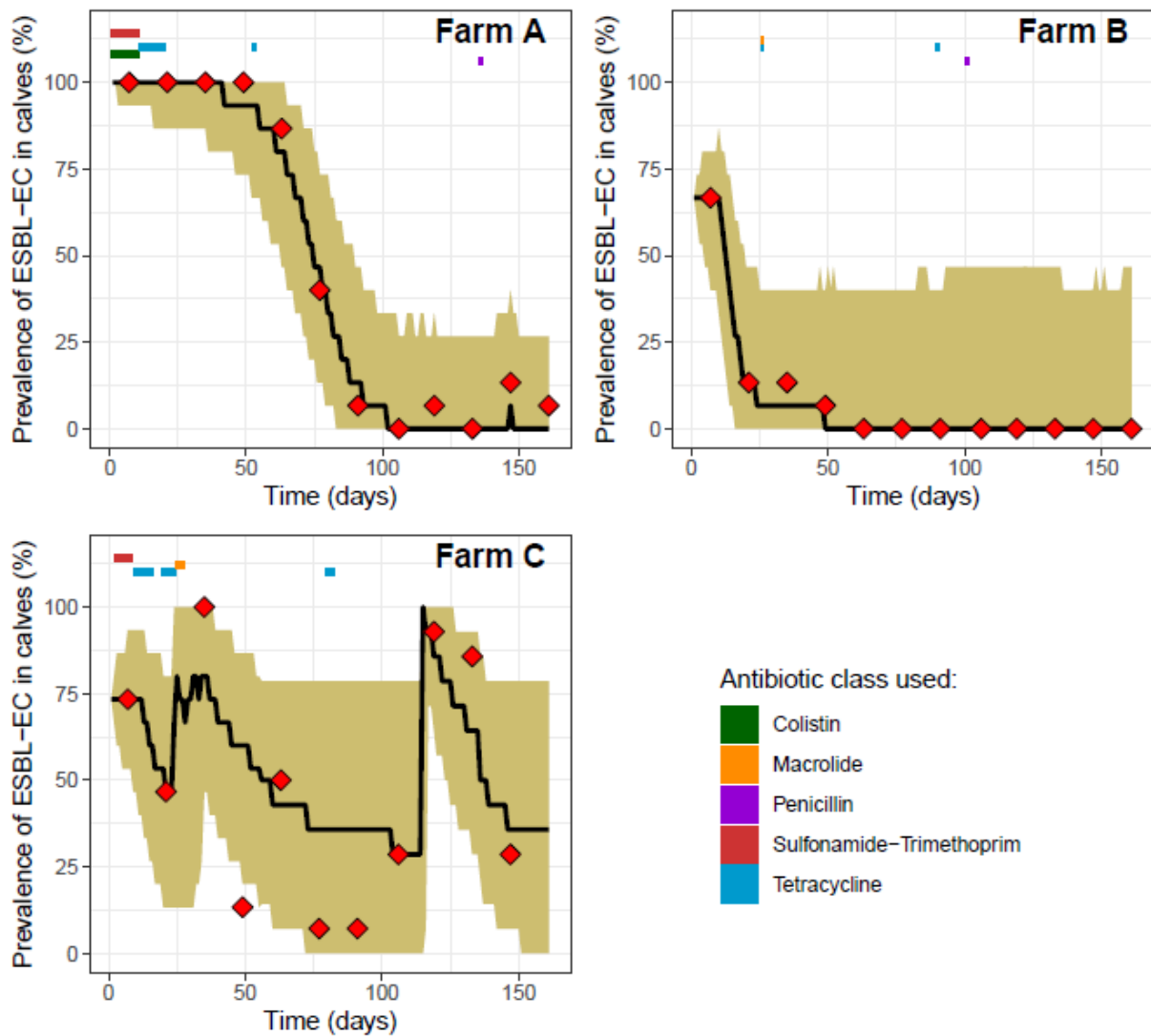
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236 SM8. Separate fits of each farm using model 5a

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240 *Figure S3. Observed (red dots), median predicted ESBL-EC prevalence (black line) and 95% prediction*
241 *interval using model 5a for each farm separately (5,000 repetitions of the model). Observed periods of*
242 *antimicrobial use are represented for each antibiotic class.*

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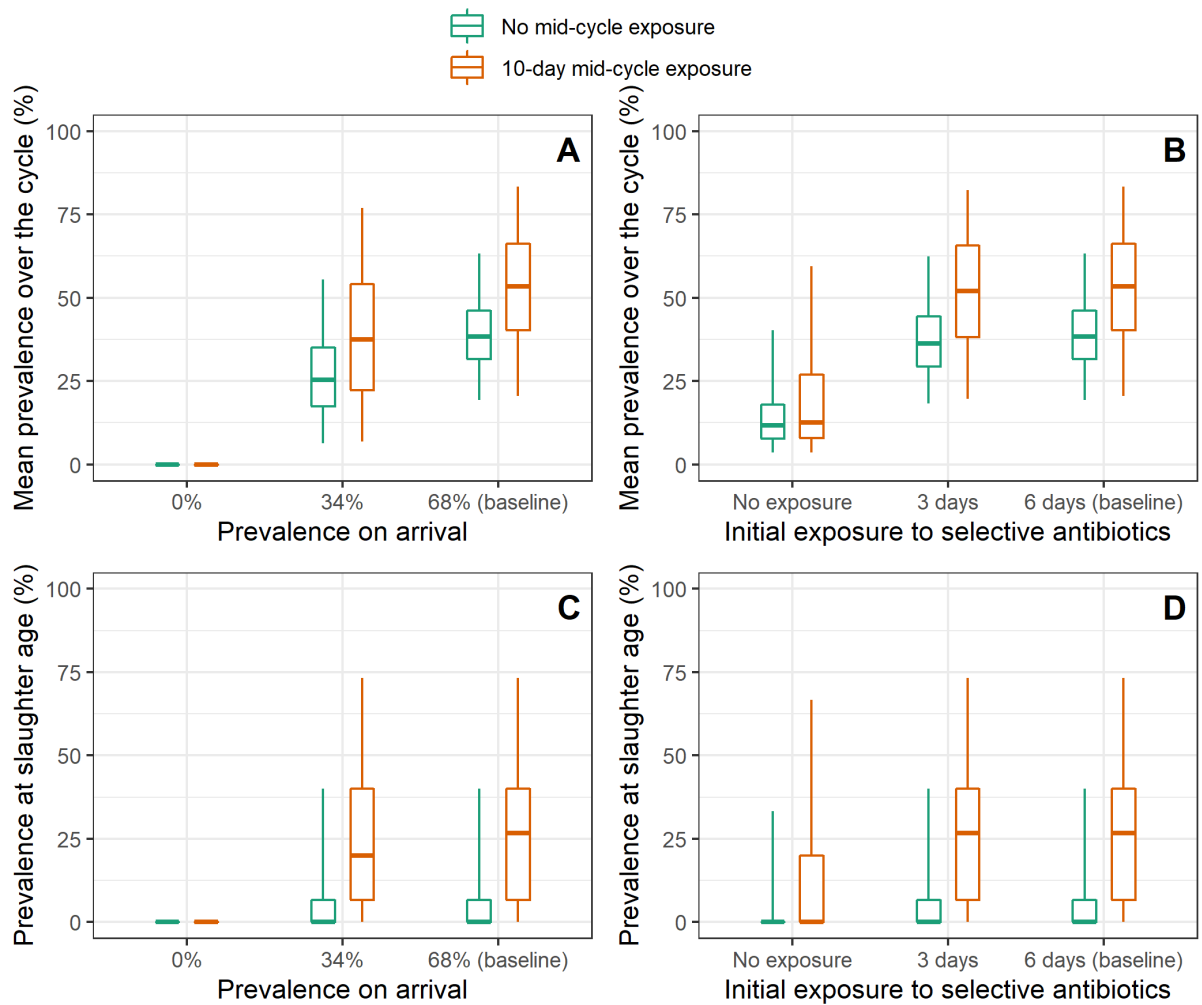
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250 **SM9. Sensitivity analysis on the farm selected for the simulations study**

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253 **Figure S4. Simulations of ESBL-EC mitigation strategies in farm B.** Mean ESBL-EC prevalence
 254 over the production cycle (panels A and B) and prevalence at slaughter age (panels C and D) predicted
 255 in farm B by model 5a (5,000 repetitions of the model), when ESBL-EC prevalence on arrival (panels A
 256 and C) and the duration of the initial antibiotic exposure (panels B and D) are changed from their
 257 baseline values. Scenarios without (turquoise) or with (orange) a 10-day antibiotic exposure in the
 258 middle of the fattening cycle (between days 81 and 90) are explored. Values represented by the boxes are
 259 the predicted median, and the 50% and 95% prediction intervals.

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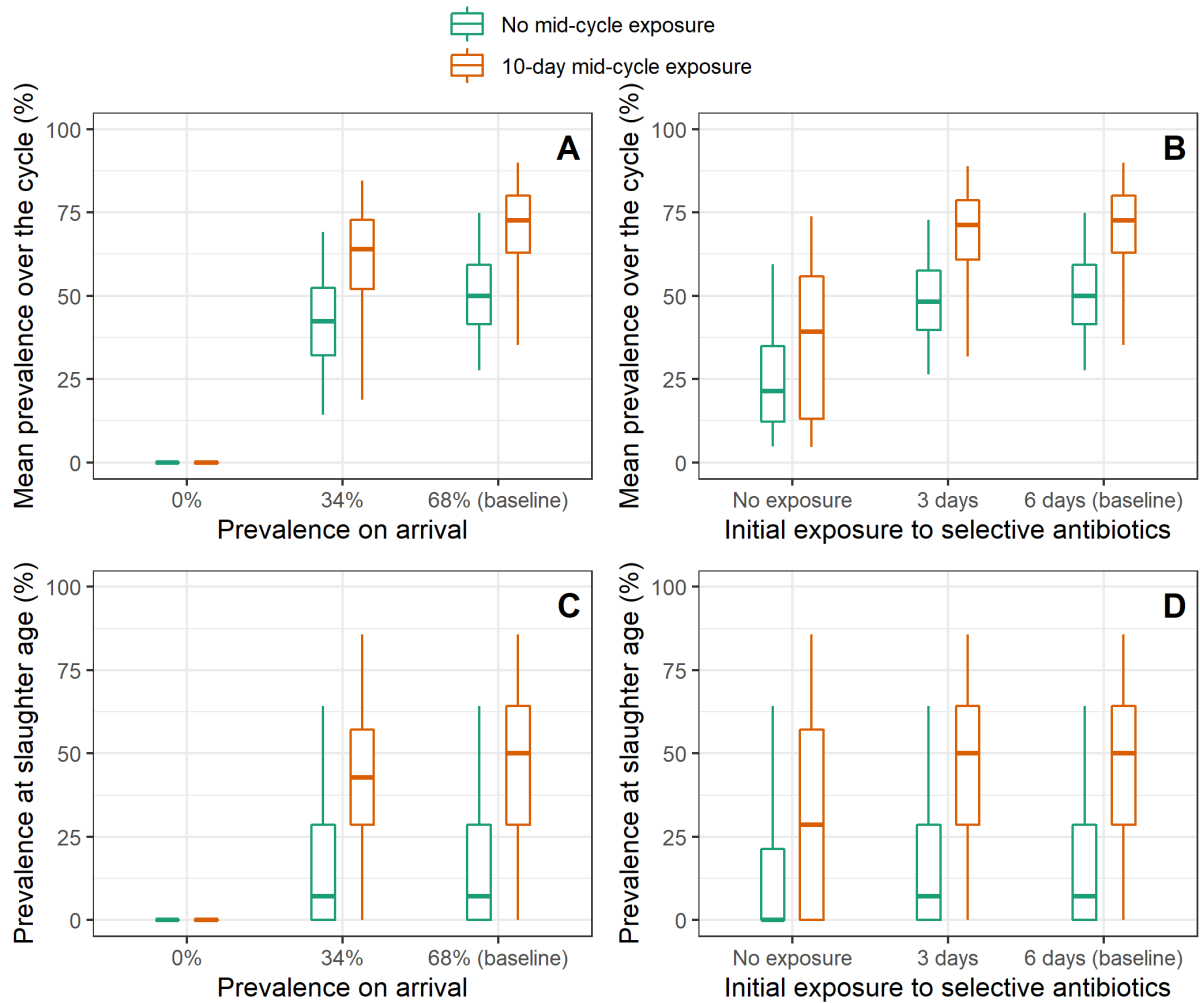
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267 **Figure S5. Simulations of ESBL-EC mitigation strategies in farm C.** Mean ESBL-EC prevalence
 268 over the production cycle (panels A and B) and prevalence at slaughter age (panels C and D) predicted
 269 in farm C by model 5a (5,000 repetitions of the model), when ESBL-EC prevalence on arrival (panels A
 270 and C) and the duration of the initial antibiotic exposure (panels B and D) are changed from their
 271 baseline values. Scenarios without (turquoise) or with (orange) a 10-day antibiotic exposure in the
 272 middle of the fattening cycle (between days 81 and 90) are explored. Values represented by the boxes are
 273 the predicted median, and the 50% and 95% prediction intervals.

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282 **References of the Supplementary material**

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- 284 [1] S. Caballero, S. Kim, R.A. Carter, I.M. Leiner, B. Sušac, L. Miller, et al. Cooperating
285 Commensals Restore Colonization Resistance to Vancomycin-Resistant *Enterococcus faecium*.
286 *Cell Host Microbe*. 2017;21(5):592-602.e4.
- 287 [2] E.N. Abatih, L. Alban, A.K. Ersbøll, D.M. Lo Fo Wong. Impact of antimicrobial usage on
288 the transmission dynamics of antimicrobial resistant bacteria among pigs. *Journal of*
289 *Theoretical Biology*. 2009;256(4):561-73.
- 290 [3] S. Cauchemez, L. Temime, A.-J. Valleron, E. Varon, G. Thomas, D. Guillemot, et al. S.
291 pneumoniae transmission according to inclusion in conjugate vaccines: Bayesian analysis of a
292 longitudinal follow-up in schools. *BMC Infect Dis*. 2006;6:14-.
- 293 [4] B.S. Cooper, G.F. Medley, S.J. Bradley, G.M. Scott. An Augmented Data Method for the
294 Analysis of Nosocomial Infection Data. *American Journal of Epidemiology*. 2008;168(5):548-
295 57.
- 296 [5] M.S. Arcilla, J.M. van Hattem, M.R. Haverkate, M.C.J. Bootsma, P.J.J. van Genderen, A.
297 Goorhuis, et al. Import and spread of extended-spectrum beta-lactamase-producing
298 Enterobacteriaceae by international travellers (COMBAT study): a prospective, multicentre
299 cohort study. *The Lancet Infectious Diseases*. 2017;17(1):78-85.
- 300 [6] G. Birgand, L. Armand-Lefevre, I. Lolom, E. Ruppe, A. Andreumont, J.-C. Lucet. Duration
301 of colonization by extended-spectrum beta-lactamase-producing Enterobacteriaceae after
302 hospital discharge. *Am J Infect Control*. 2013;41(5):443-7.
- 303 [7] S.B. Jørgensen, A. Søråas, A. Sundsfjord, K. Liestøl, T.M. Leegaard, P.A. Jenum. Fecal
304 carriage of extended spectrum β -lactamase producing *Escherichia coli* and *Klebsiella*
305 pneumoniae after urinary tract infection - A three year prospective cohort study. *PLOS ONE*.
306 2017;12(3):e0173510-e.
- 307 [8] M. Plummer, A. Stukalov, M. Denwood, M.M. Plummer. Package 'rjags'. 2018.
- 308 [9] S.P. Brooks, A. Gelman. General Methods for Monitoring Convergence of Iterative
309 Simulations. *Journal of Computational and Graphical Statistics*. 1998;7(4):434-55.
- 310 [10] D.J. Spiegelhalter, N.G. Best, B.P. Carlin, A.J.J.o.t.R.S.S.S.B. Van Der Linde. Bayesian
311 measures of model complexity and fit. 2002;64(4):583-639.

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