Supplementary Table S1. Primers used in this study. For dCAPS genotyping, restriction enzymes are indicated as well as corresponding fragment sizes after digestion. Highlighted nucleotides are non-complementary bases introduced to generate corresponding restriction sites.

Como	.		Restriction enzyme	Size restriction fragments (bp)	
Gene	Primer name	Sequence 5' to 3'		WT	Mutant
dCAPS genotyping	3				
	PGF2	TGTAGATGGCCTTGTTCTCATACCTC	Yhai	224	106128
PCH2-9	P9R	GGCATGGACAACAGGCTTAACTT <u>T</u> C	Abai	224	196+28
DCH2 12	PGF2	TGTAGATGGCCTTGTTCTCATACCTC	Eco PV	260	222+28
PCH2-12	P12R	CAGTGCCTTATTCAACTTTTGCATA <u>G</u> ATA	ECORV	260	232+28
ACV1 12	A13F	GAACTTTTAACTCCACTGCTGACATTA <u>AT</u>	Pad	122	94+29
A371-13	A13R	GCACAATCACGAAAATTACAAAGCATG	Restriction enzyme Xbal EcoRV Pacl EcoRI	125	94+29
ACV1 14	A14F	CGGTTCCTGCTCTAGGTAACAAGC	Xbal EcoRV Pacl EcoRI	195	148+37
A371-14	A14R	GCTAAAATCATTAGAATACAAAAAACCTTTCT GA AT		185	
Reverse transcrip	tion-PCR				
1 at in	Actin_F	GAGCAACTGGGATGACATGG			
	Actin_R	GAAACCCTCGTAGATTGGCAC	Pacl EcoRl		
DCU2 12	ASY1-13 A13F GAACTTITAACTCCACTGCTGACATTAAT ASY1-13 A13R GCACAATCACGAAAATTACAAAGCATG ASY1-14 A14F CGGTTCCTGCTCTAGGTAACAAGC A14R GCTAAAATCATTAGAATACAAAAAACCTTTCTGAA Actin_F GAGCAACTGGGATGACATGG Actin_R GAAACCCTCGTAGGATGGCAC PCH2_RT_F1 CTCGTCTCTGTGGAGGTCTG PCH2_RT_R2 GACCCAGTTCCTCACACG Asy1-4 ASY1_F ASY1_F CAGCTGAGATTTTGCAAGCTGAAC Shoc1-1 SHOC1_F	CTCGTCTCTGTGGAGGTCTG			
	PCH2_RT_R2	GACCCAGTTCCTCACACG			
T-DNA genotyping	g				
PCH2-9 PCH2-12 ASY1-13 ASY1-14 Reverse transcrip Actin PCH2-12 T-DNA genotypin asy1-4 shoc1-1 mus81-2 pch2-1	ASY1_F	TCAGCTGGAGTTTTTAGCTCG			
usy1-4	ASY1_R	GAACCATTTTGCAAGCTGAAC	Restriction enzyme Xbal EcoRV Pacl EcoRI		
shact 1	SHOC1_F	AGCTTAAGCGAATCTTACCGG			
51001-1	SHOC1_R	me Sequence 5' to 3' Itematy TGTAGATGGCCTTGTTCTCATACCTC Xba GGCATGGACAACAGGCTTAACTTTC Xba GGCACTGGCCTTGTTCTCATACCTC EcoR CAGTGCCTTATTCAACTTTTGCATAGATA EcoR GAACTTTTAACTCCACTGCTGACATTAAT Pac GCACAATCACGAAAATTACAAAGCATG Pac GCACAATCACGAAAATTACAAAAGCC EcoF GCTAAAATCATTAGAATACAAAAAACCTTTCTGAAAT EcoF GAGCAACTGGGGATGACATGG GAAACCCTCGTAGGATGACATGG GAAACCCTCGTAGGATGACATGG GAAACCCTCGTAGGATGGCAC F1 CTCGTCTCTGTGGGAGGTCTG R2 GACCCAGTTCCTCACACG TGAGCTGGAATCTTACCGG GAACCATTTTGCAAGCTGAAC AGCTTAAGCGAATCTTACCGG TGGTGAAATCTAGCAACCCAG AGCTTAAGCGAATCTTACCGG TGGTGAAATCAAGCAACCCAG AGCTTAAGCGAATCTTACCGG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCCAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCAAATAGCCGTCGCTGAG CAGTGCCAATTCCGGAAC CAGTGCCAATTCCACAGC GAACCATTCTCCACAAGCCACCAG CAGTGCCAATAGCCGTCGCTGAG			
muc 81 2	MUS81_F	TGGTGAAATCTAGCAACCCAG			
mus81-2	MUS81_R	AATTTTCCACAAACCCTTTGG			
nah2 1	PCH2_F	CAGTGCAAATAGCCGTCGCTGAG			
	PCH2_R	CTCACATGGTCCTTCTTCAATGAGC			
T-DNA	LB_salk_LB1.3	ATTTTGCCGATTTCGGAAC			

Supplementary Table S2. Plant fertility in *B. rapa* **WT**, *asy1* and *pch2* mutants. Silique length and seed setting in *B. rapa* WT and in TILLING plants WT, heterozygous or mutant for *ASY1* and *PCH2* including allelic mutants. N = total number of siliques (10 siliques per plant).

	Average silique length (cm)	Average seed set	Ν
WT	7.60 ± 0.61	37.73 ± 6.73	120
ASY1-13 +/+	7.10 ± 0.60	37.84 ± 4.94	50
ASY1-13 +/-	7.48 ± 0.51	36.02 ± 5.63	50
asy1-13	2.80 ± 0.34	0 ± 0	50
ASY1-14 +/+	7.49 ± 0.55	37.32 ± 5.52	50
ASY1-14 +/-	7.86 ± 0.65	38.7 ± 6.58	50
asy1-14	3.05 ± 0.67	0 ± 0	50
asy1-13/14	3.16 ± 0.20	0 ± 0	50
PCH2-9 +/+	7.40 ± 0.50	36.38 ± 4.22	50
PCH2-9 +/-	7.17 ± 0.41	34.16 ± 5.39	50
pch2-9	4.41 ± 1.01	2.74 ± 3.60	50
PCH2-12 +/+	6.80 ± 0.56	35.32 ± 8.09	50
PCH2-12 +/-	6.56 ± 0.54	33.20 ± 6.34	50
pch2-12	3.19 ± 0.79	1.02 ± 2.32	50
pch2-9/12	4.28 ± 1.08	3.22 ± 1.01	50

Supplementary Table S3. Relative occurrence (%) of 5S/45S-labelled chromosomes as either univalents or bivalents. Chromosome #3 formed a bivalent with significantly higher frequency than all other discernible chromosomes. N = 73 *asy1* diakinesis cells.

	#1 (5S)	#3 (5S+45S)	#4 (5S+45S)	#5 (5S+45S)	#6 (45S)	#10 (5S)
Univalent	84.93	36.76	79.45	84.93	86.30	82.19
Bivalent	15.07	63.24	20.55	15.07	13.70	17.81

Supplementary Table S4. ASY1 immunofluorescence relative signal intensity in WT and *pch2* cells. A. Asynapsed vs synapsed regions. In *pch2*, ASY1 is not depleted from synapsed regions and remains brightly stained in pachytene cells. B. Whole leptotene nuclei. Initial ASY1 loading is impaired in *pch2*. Independent experiments performed with two independent microscopes. Values represent nuclei fluorescence intensity after deduction of background signal intensity (see methods). N = number of cells. Asterisks indicate significant differences (Student's *t* test, P < 0.05).

А

W	/Т		pcl	n-2	
Asynapsed	Synapsed	Difference	Asynapsed	Synapsed	Difference
1922.41 N=11	846.38 N=10	-55% (*)	1758.09 N=16	2137.14 N=17	+21% (*)

В

Whole nuclei	WT	pch2	Difference
Microscope 1	447.09 N=21	348.79 N=31	-22% (*)
		N=8	-2270
Microscope 1	1392.20 N=23	331.07 N=23	-76% (*)
Microscope 2	3943.67 N=43	3388.21 N=21	-14%



Supplementary Fig. S1. Alternative splicing of *PCH2* in *pch2-12*. Reverse transcription-PCR for *PCH2* in two independent mutant plants, one sibling TILLING plant WT for *PCH2* and one WT plant.
A. Primer pair spanning the mutation site. Note alternative splicing variants in mutant plants only. B. Expression of actin as technical control. B: bud; L: leaf; W: water control.



Supplementary Fig. S2. Male meiotic chromosome behaviour in *B. rapa*. A. Meiotic stages in *B. rapa* WT. a. Leptotene. b. Zygotene. c. Pachytene. d. Diakinesis. e. Metaphase I. f. Telophase I. g. Metaphase II. h. Anaphase II. i. Telophase II. j. Tetrad. B. WT meiosis in TILLING plants WT or heterozygous for *ASY1* and *PCH2*. C. Meiosis in the allelic mutants *asy1-13/14* and *pch2-9/12*. DNA counterstained with DAPI and shown in grey. Bars = 10µm.



Supplementary Fig. S3. Localization defect of ASY1 in Arabidopsis *pch2*. **A**. Staining of ASY1 at leptotene in WT and *pch2*. ASY1 relative signal intensity indicated in the bottom right corner. Bars = $10\mu m$. **B**. Plot showing relative ASY1 signal intensity in each background. Asterisks represent statistically significant differences (MWW test).