







Supplemental Figure 1: FIPs and APCs gene profile from wild-type mice, Related to Figure 1

FIPs and APCs were sorted from epididymal WAT from 4-6 week-old male C57BL/6 mice using FACS by selecting CD45⁻, CD31⁻, CD140b⁺, Lyc6⁺ and CD9⁺ (FIPs) and CD45⁻, CD31⁻, CD140b⁺, Lyc6⁻ and CD9⁻ (APCs). **(A)** Flow cytometry graph representing the gating for FIPs and APCs for sorting **(B)** Gene expression of adipogenic, inflammatory and fibrosis markers compared between FIPs and APCs from WT mice. Data are presented in mRNA relative to FIPs **(C)** Mitochondrial DNA in FIPs and APCs from WT mice. Data are presented as the ratio between mtDNA and nuclear DNA. **(D-G)** Gene expression of FIPs and APCs from WT mice. Data are presented in mRNA relative to FIPs. **(D)** Gene expression of mitochondrial markers. **(E)** Gene expression of markers oxidative phosphorylation **(F)** Gene expression of fatty acid oxidative metabolism markers **(G)** Gene expression of glycolytic markers. Significance between FIPs and APCs from WT mice was calculated using a two-tailed student's t-test. Error bars represent mean \pm S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001), **** (p<0.00001).





<u>Supplemental Figure 2:</u> FIPs and APCs have different metabolic capacities and are regulated differentially after 3 days of HFD feeding or by BSA-palmitate treatment, Related to Figure 1

FIPs and APCs were sorted from epidydimal WAT (eWAT) from 4-6 week-old male C57BL/6 mice using FACS by selecting CD45, CD31, CD140b⁺, Lyc6⁺ and CD9⁺ (FIPs) and CD45, CD31, CD140b⁺, Lyc6⁻ and CD9⁻ (APCs). (A) Protein content of 80,000 FIPs or APCs plated in a seahorse plate for 24h (n=6). Data are presented in µg/µL. (B-C) Oxygen consumption rate of 80,000 FIPs and APCs measured. Data are presented in pmol/min (D) Glycolysis of 80,000 FIPs and APCs evaluated by extracellular acidification rate (ECAR). Data are presented in mpH/min. (E-F) Glycolysis stress analysis of 80,000 FIPs and APCs. Data are presented in mpH/min. (G-J) FIPs and APCs treated with 7 µM BSA or 0.2mM palmitate combined with 7µM BSA for 4 hours and then evaluated by Seahorse analysis. (G-H) Oxygen consumption rate of 80,000 FIPs and APCs pretreated with BSA or BSA-palmitate for 4 hours. Data is presented in pmol/min. (I-J) Glycolysis of 80,000 FIPs and APCs pretreated 4 hours with BSA or BSA-palmitate evaluated by ECAR. Data are presented in mpH/min. (K-N) FIPs and APCs sorting by FACS from the eWAT of wild type mice fed either control diet (CD) or high-fat diet (HFD) for 3 days. (K-L) Oxygen consumption rate of freshly isolated FIPs and APCs following 3 days of CD or HFD feeding. Data are presented in pmol/min per protein content (µg/µL). (M-N) Glycolysis of freshly isolated FIPs and APCs following 3 day chow or HFD feeding evaluated by ECAR. Data are presented in mpH/min. Significance in (B, E,G,I,K,M) was calculated usin a 2-way Anova with Tukey's posttest for multiple comparisons. Significance in (A,C,D,F,H,J, L,N) was calculated using a two-tailed student's t-test. Error bars represent mean ± S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001), **** (p<0.00001).



<u>Supplemental Figure 3:</u> Validation of PdgfR β -rtTA TRE-MitoNEET (MitoNEET ^{TG}) mouse model. Related to Figure 1 and Figure 3

(A) Breeding strategy of the mouse model PdgfR β -rtTA (Control) or PdgfR β -rtTA x TRE-MitoNEET (MitoNEET ^{TG}) mice. (B) In vivo validation: Control and MitoNEET^{TG} were fed 2 weeks with chow diet containing 600 mg/kg of doxycycline and tissues were harvest to evaluate mRNA expression of *Cisd1* (MitoNEET) gene between control and MitoNEET^{TG} mice (n=6). (C-I) Control or MitoNEET^{TG} cells were isolated by FACS from eWAT, plated at confluency and treated with or without 4.4 µM doxycycline for 24 hours. (C-D) In vitro evaluation of Cisd1 (MitoNEET) gene expression of FIPs (C) or APCs (D) from control or MitoNEET ^{TG} mice (n=6). Data are presented in mRNA expression relative to untreated control cells. (E) Protein expression of MitoNEET in vitro from PdgfRβ+ cells from control or MitoNEET^{TG} mice (n=3). Data are presented as a ratio between MitoNEET and β -actin protein expression. (F-G) In vitro death rate of FIPs (F) or APCs (G) from control or MitoNEET^{TG} mice (n=4). Data are presented as the ratio between intranuclear staining and DAPI. (H-I) In vitro protein content of FIPs (H) or APCs (I) from control or MitoNEET^{TG} mice plated in seahorse plate (n=4-5). (J-L) Control or MitoNEET^{TG} cells were isolated by FACS from eWAT of mice fed 1 week with chow diet containing 600 mg/kg of doxycycline. (J-K) Gene expression of Cisd1 (MitoNEET) from FIPs (n=4) (J) or APCs (n=5) (K). Data are presented in mRNA expression relative to control cells. (L) Protein expression of MitoNEET in vivo from PdgfR β + cells from control or MitoNEET^{TG} mice (n=4). Data are presented as a ratio between MitoNEET and β -actin protein expression. (M-R) Control or MitoNEET^{TG} cells were isolated by FACS from sWAT, plated at confluency and treated with or without 4.4 µM doxycycline for 24 hours. (M-N) In vitro mRNA expression of Cisd1(MitoNEET) of DPP4+ (n=6) (M) or DPP4- cells (n=6) (N) from control or MitoNEET^{TG} mice. Data are presented in mRNA expression relative to untreated control cells. (O-P) In vitro death rate of DPP4+ (n=3) (F) or DPP4- cells (n=3) (G) from control or MitoNEET^{TG} mice. Data are presented as the ratio between intranuclear staining and

DAPI. (Q-R) *In vitro* protein content of DPP4+ (n=4-5) (Q) or DPP4- cells (n=5) (R) from control or MitoNEET^{TG} mice plated in seahorse plate. (S-U) Control or MitoNEET^{TG} cells were isolated by FACS from sWAT of mice fed 1 week with chow diet containing 600 mg/kg of doxycycline. (S-T) Gene expression of Cisd1 (MitoNEET) from DPP4+ (n=5) (S) or DPP4- cells (n=4-5) (T). Data are presented in mRNA expression relative to control cells. (U) Protein expression of MitoNEET *in vivo* from PdgfRβ+ cells from control or MitoNEET^{TG} mice (n=4). Data are presented as a ratio between MitoNEET and β-actin protein expression. Significance in (C-I; M-R) was calculated using a Kruskal-Wallis (One-Way Anova) test with Dunn's post-test for multiple comparisons. Significance in (B; J-L; S-U) was calculated using a two-tailed student's t-test. Error bars represent mean \pm S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001), **** (p<0.0001).



















<u>Supplemental Figure 4:</u> DPP4⁺ and DPP4⁻ gene profile from wild-type mice, Related to Figure 3

DPP4⁺ and DPP4⁻ cells were sorted from subcutaneous WAT (sWAT) from 4-6 week-old male C57BL/6 mice (WT mice) using FACS by selecting for CD45⁻, CD31⁻, CD140b⁺, DPP4⁺ and CD45⁻, CD31⁻, CD140b⁺, DPP4⁻. (**A**) Gating strategy to sort DPP4+ and DPP4- cells based on the Cd26 (DPP4) marker (**B**) Gene expression of adipogenic and inflammatory markers in DPP4⁺ and DPP4⁻ cells isolated from the sWAT of WT mice. (**C**) Gene expression of mitochondrial markers in DPP4⁺ and DPP4⁺ cells from the sWAT of WT mice. (**C**) Mitochondrial DNA in DPP4⁺ and DPP4⁻ cells from the sWAT of WT mice. Data are expressed as the ratio of mtDNA to nuclear DNA. (**E**) Gene expression of oxidative phosphorylation markers in DPP4⁺ and DPP4⁻ cells from the sWAT of WT mice (**G**) Gene expression of glycolytic markers. Data are presented as relative expression of markers in DPP4⁺ cell to DPP4⁻ cells. Significance between FIPs and APCs from WT mice was calculated using a two-tailed student's t-test. Error bars represent mean ± S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001).



<u>Supplemental Figure 5</u>: DPP4⁺ and DPP4⁻ metabolic status in response to palmitate or three days of HFD, Related to Figure 3

DPP4⁺ and DPP4⁻ cells were sorted from sWAT from 4-6 week-old male C57BL/6 mice FACS by selecting CD45⁻, CD31⁻, CD140b⁺, CD26⁺ (DPP4⁺) cells and CD45⁻, CD31⁻, CD140b⁺, CD26⁻ (DPP4⁻) cells. (A) Protein content of DPP4+ and DPP4- cells plated in Seahorse plate. (B-C) Oxygen consumption rate of 80,000 DPP4⁺ and DPP4⁻ cells. Data are presented in pmol/min (D) Glycolysis of 80,000 DPP4⁺ and DPP4⁻ cells evaluated by extracellular acidification rate (ECAR). Data are presented in mpH/min. (E-F) Glycolysis stress analysis of 80,000 DPP4⁺ and DPP4⁻ cells. Data are presented in mpH/min. (G-J) DPP4⁺ and DPP4⁻ cells pretreated with 7µM BSA or 0.2mM palmitate combined with 7µM BSA for 4 hours: (G-H) Oxygen consumption rate of 80,000 DPP4⁺ and DPP4⁻ cells. Data are presented in pmol/min. (I-J) Glycolysis of 80,000 DPP4⁺ and DPP4⁻ evaluated by ECAR. Data are presented in mpH/min. (K-N) DPP4⁺ and DPP4⁻ cells sorted by FACS from the sWAT of WT mice fed either control diet (CD) or high-fat diet (HFD) for 3 days: (K-L) Oxygen consumption rate of freshly isolated DPP4⁺ and DPP4⁻ cells following 3 days of chow or HFD feeding. Data are presented in pmol/min per protein content (H) Glycolysis of freshly isolated DPP4⁺ and DPP4⁻ cells following 3 days of chow or HFD feeding evaluated by ECAR. Data are presented in mpH/min. Significance in (B, E,G,I,K,M) was calculated using 2-way Anova with Tukey's post-test for multiple comparisons. Significance in (A,C,D,F,H,J, L,N) was calculated using a two-tailed student's t-test. Error bars represent mean ± S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001), **** (p<0.00001).







<u>Supplemental Figure 6:</u> Mitochondrial activity greatly influences mesenchymal lineage determination in DPP4+ and DPP4-, Related to Figure 3

DPP4+ or DPP4- cells were isolated by FACS from sWAT of control or MitoNEET^{TG} mice. DPP4+ or DPP4- cells were plated at confluency and treated with or without 4.4 μ M doxycycline for 24 hours. Cells were then differentiated into osteoblast for 14 days as described in Methods. **(A)** Gene expression of osteogenic markers in DPP4⁺ cells from control or MitoNEET^{TG} mice. Data are presented in mRNA relative expression to untreated control cells. **(B)** Alizarin Red S staining for evaluation of calcium deposits in DPP4+ cells. **(C)** Gene expression of osteogenic markers in DPP4⁻ cells from control or MitoNEET^{TG} mice. Data are presented in mRNA relative expression to untreated control cells. **(D)** Alizarin Red S staining for evaluation of calcium deposit in DPP4cells. Significance in (A;C) was calculated using a Kruskal-Wallis (one-way Anova) test with Dunn's post-test for multiple comparisons. Error bars represent mean ± S.E.M. * (P<0.05), ** (p<0.01), *** (p<0.0001), **** (p<0.00001).