

Supplementary Material

Table S1. Amino acid sequences used for phylogenetic tree construction.

Source	Abbreviation	GH13 subfamily number	Sequence similarity ^a (%)	GenBank or PDB accession number
Fungi (GH13_1)				
<i>Aspergillus oryzae</i>	AmyA	1	64.4	2GUY_A
<i>Malbranchea cinnamomea</i>	McAmyA	1	59.0	3VM7_A
<i>Aspergillus niger</i>	α -amylase	1	55.3	2AAA_A
Bacteria (GH13_2)				
<i>Bacillus stearothermophilus</i>	Novamyl	2	26.9	1QHO_A
<i>Thermoanaerobacterium thermosulfurigenes</i>	CGTase	2	23.1	3BMV_A
Fungi (GH13_5)				
<i>Paecilomyces variotii</i>	alpha-amylase		77.6	XP_028489551.1
<i>Aspergillus niger</i>	AmyD	5	64.4	XP_001389762.2
<i>Aspergillus niger</i>	AmyF	5	54.5	XP_001393627.2
<i>Paracoccidioides brasiliensis</i>	Amy1	5	39.1	ABS11196.1
Bacteria (GH13_5)				
<i>Bacillus</i> sp. 707	G6-amylase	5	47.5	1WP6_A
<i>Bacillus licheniformis</i>	alpha-amylase	5	47.0	1OB0_A
<i>Bacillus</i> sp. Ksm-1378	AmyK	5	46.8	2DIE
<i>Bacillus halmapalus</i>	alpha-amylase	5	46.7	1W9X
<i>Bacillus amyloliquefaciens</i>	BAA	5	46.4	3BH4_A
<i>Geobacillus stearothermophilus</i>	AmyS	5	45.3	4UZU_A
<i>Bacillus</i> sp. strain KSM-K38	AmyK38	5	44.5	1UD2_A
<i>Alicyclobacillus</i> sp. 18711	AliC	5	44.2	6GXV_A
<i>Halothermothrix orenii</i>	AmyB	5	31.7	3BC9_A
Plants (GH13_6)				
<i>Hordeum vulgare</i>	Amy1	6	26.6	2QPS_A
<i>Hordeum vulgare</i>	Amy2	6	22.6	1AMY_A
Archaea (GH13_7)				
<i>Pyrococcus woesei</i>	PWA	7	45.0	1MWO_A
<i>Pyrococcus woesei</i>	alpha-amylase	7	28.2	3QGV_A
Bacteria (GH13_10)				
<i>Deinococcus radiodurans</i> R1	MTHase	10	33.8	2BHU_A
Bacteria (GH13_20)				
<i>Thermus</i> sp. IM6501	ThMA	20	24.7	1SMA_A
<i>Geobacillus stearothermophilus</i>	Neopullulanase	20	20.9	1J0H_A

Archaea (GH13_20)				
<i>Thermococcus kodakarensis</i> KOD1	TK-PUL	20	37.7	5OT1_A
Bacteria (GH13_31)				
<i>Erwinia rhamontici</i>	NX-5	31	24.5	4HOW_A
<i>Listeria monocytogenes</i> EGD-e	Lmo0184	31	23.9	5DO8_A
Yeast (GH13_40)				
<i>Saccharomyces cerevisiae</i>	Ima1	40	28.7	3AXH_A
Bacteria (GH13_?)				
<i>Anoxybacillus ayderensis</i>	ASKA	unknown	33.2	5A2A_A
<i>Geobacillus thermoleovorans</i>	GTA	unknown	32.2	4E2O_A

^a The similarities of α -amylases were compared to NFAmy13B, except for those in group GH13_1, which were compared to NFAmy13A.

Table S2. The hydrolysis of NFAmy13B on various substrates.

Substrate	Main linkage/monomer	Glucose equivalents (mM) (mean \pm standard deviation)
Soluble starch	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	1.23 \pm 0.06
Amylopectin	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	11.6 \pm 0.3
Amylose	α -(1 \rightarrow 4) glucose	5.73 \pm 1.24
Wheat starch	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	6.06 \pm 1.06
Potato starch	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	5.42 \pm 0.71
Corn starch	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	7.09 \pm 0.66
Xylan	β -(1 \rightarrow 4) xylose	0
Laminarin	β -(1 \rightarrow 3) and β -(1 \rightarrow 6)-glucose	0
Dextrin	α -(1 \rightarrow 4) and α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	6.79 \pm 0.85
Glycogen	α -(1 \rightarrow 4)- α -(1 \rightarrow 6) glucose	0.225 \pm 0.030
Pullulan	α -(1 \rightarrow 6) glucose	0.0692 \pm 0.0948
Arabinoxylan	β -(1 \rightarrow 4) arabinose and xylose	0
Cellulose	β -(1 \rightarrow 4) glucose	0

Each experiment was performed in triplicate.

Table S3. Degradation capacity of NFAmy13A and NFAmy13B on wheat starch.

Enzyme	Concentration (nM)	Reducing ends (mM)
NFAmy13B	20	6.71±0.46
	900	4.67±0.30
NFAmy13A	1000	6.63±1.16
	1100	7.70±1.75
	1200	7.18±1.62

Each experiment was performed in triplicate.

Table S4. The hydrolysis of NFAmy13A and NFAmy13B on maltooligosaccharides (M3~M5).

Substrate	Enzyme	Product component	The yield of reducing ends (mM) (mean ± standard deviation)		
			5 min	20 min	60 min
M3	NFAmy13A	M1	1.52±0.11	2.47±0.05	3.29±0.09
		M2	1.96±0.11	2.97±0.05	3.84±0.02
	NFAmy13A	M1	0.727±0.024	1.34±0.05	2.17±0.08
		M2	7.32±0.07	7.79±0.21	8.53±0.17
		M3	1.43±0.06	0.657±0.176	0
	NFAmy13B	M1	0.0984±0.0394	0.208±0.024	0.335±0.061
		M2	0.269±0.058	1.01±0.06	2.66±0.05
		M3	0.0867±0.1279	0.483±0.063	0.817±0.195
M4	NFAmy13A	M1	1.02±0.17	2.99±0.04	5.33±0.02
		M2	8.93±0.39	11.6±0.1	13.5±0.1
		M3	5.59±0.07	3.65±0.15	1.07±0.06
		M4	0	0	0
	NFAmy13B	M1	0	0.0951±0.770	0
		M2	5.81±0.03	6.94±0.07	7.18±0.04
		M3	7.05±0.22	8.18±0.09	7.99±0.21
		M4	0	0	0
M5	NFAmy13A	M1	0	0.0951±0.770	0
		M2	8.93±0.39	11.6±0.1	13.5±0.1
		M3	5.59±0.07	3.65±0.15	1.07±0.06
		M4	0	0	0
		M1	0	0.0951±0.770	0

Each experiment was performed in triplicate.

Table S5. Comparison of alkali-resisting α -amylases.

Amylase	Source	Optimum pH	pH stability ^b	References
NFAmy13B	Unknown	5.5~6.0	pH 5.5~12.5, 1 h, >90%	This study
G6-amylase	<i>Bacillus</i> sp.	8.8	pH 4.7~10.8, 30 min, >50%	[1]
TdAmyA	<i>Thermomyces dupontii</i>	6.5	pH 4.5~10.0, 30 min, >80%	[2]
Amy1	<i>Bacillus amyloliquefaciens</i>	6.0	pH 7.0~9.0, 1 h, 90%	[3]
α -amylase ^a	<i>Bacillus</i> sp.	5.0	pH 4.5~11.0, 1 h, 80%	[4]
α -amylase ^a	<i>Bacillus</i> sp.	10.5	pH 10.5, 15 h, 55%	[5]
α -amylase ^a	<i>Halorubrum xinjiangense</i>	8.5	pH 7~11, 1 h, >45%	[6]
α -amylase ^a	<i>Bacillus</i> sp.	8.0	pH 6~13, 30 min, >67%	[7]
TfAmy48	<i>Tepidimonas fonticaldi</i>	8.0	pH 11, 6 h, 50%	[8]
McAmyA ^a	<i>Malbranchea cinnamomea</i>	6.5	pH 5.0~10.0, 30 min, >90%	[9]

^a Amylases were produced by the originated organism.

^b This column indicates the residual activities of α -amylases pre-treated in the range of specified pH with designated time.

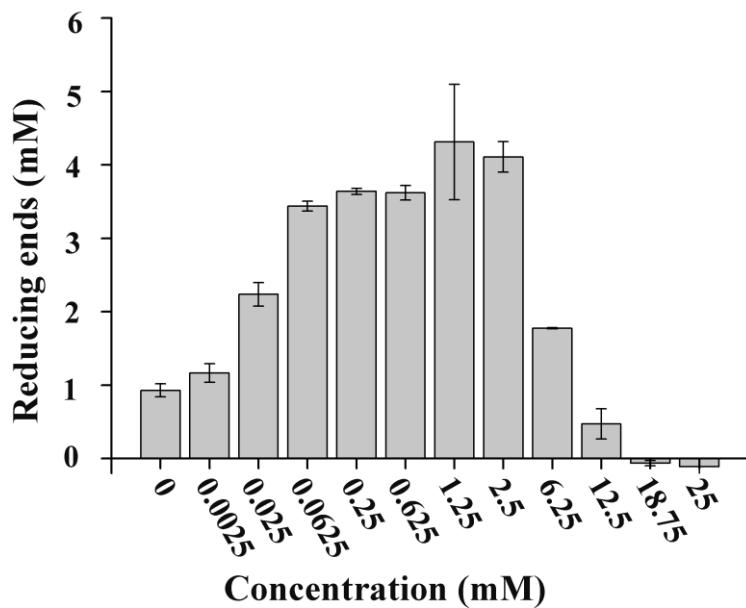


Fig. S1 Effect of the concentrations of calcium divalent ion on NFAmy13B thermostability. Enzyme (75 nM) was incubated with different concentrations of CaCl_2 at pH 6.0 and 55 °C for 30 min. The residual activities were determined by incubating 30 nM enzyme with 5 mg/mL potato starch at 54 °C and pH 6.0 for 30 min. Each experiment was performed in triplicate.

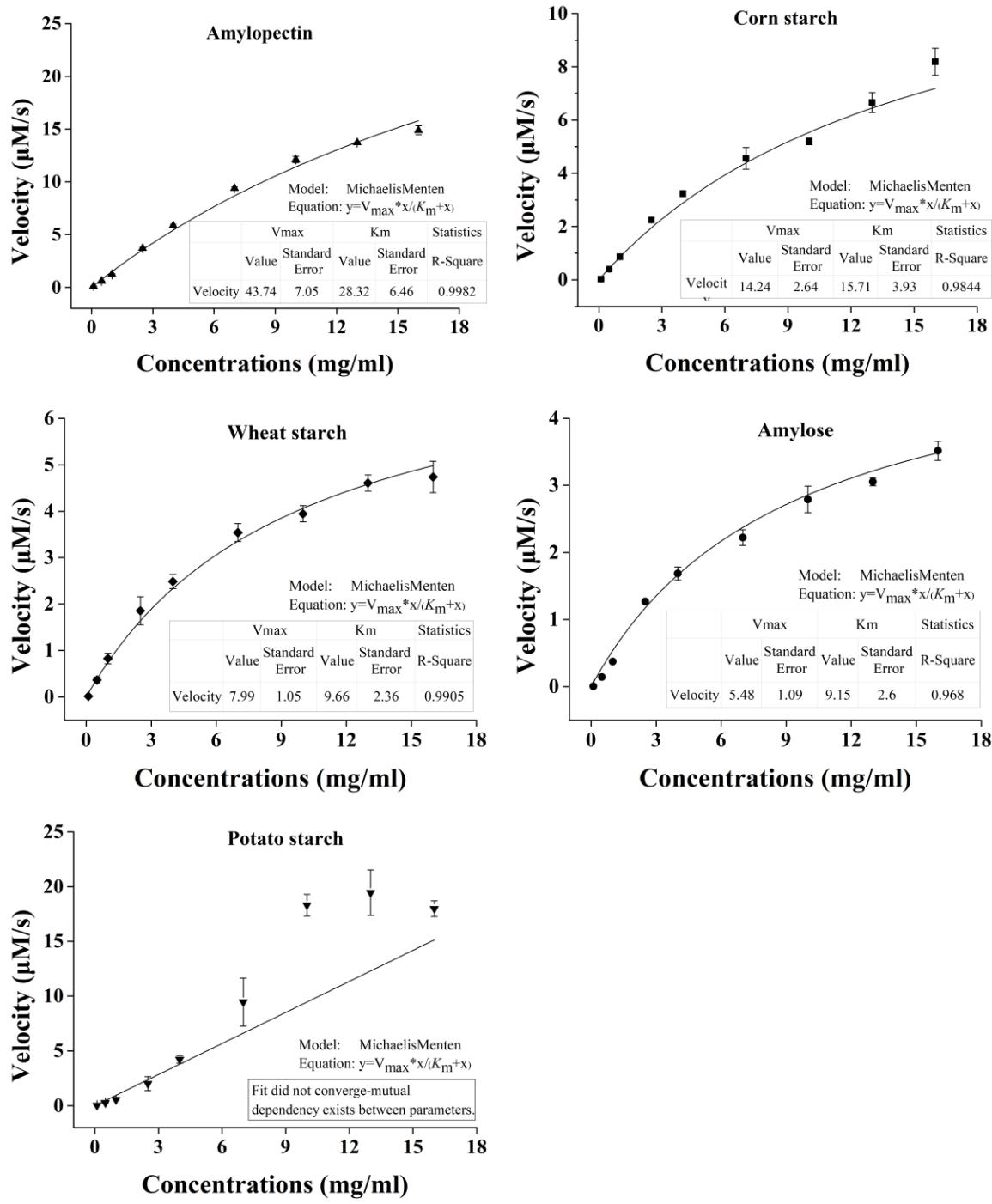


Fig. S2 Kinetic studies of the NFAmy13B on the different substrates. 20 nM enzyme with 1.25 mM CaCl₂ was incubated with various concentrations substrate between 0.1 and 16 mg/mL at 54 °C for 20 min. Each experiment was performed in quadruplicate.

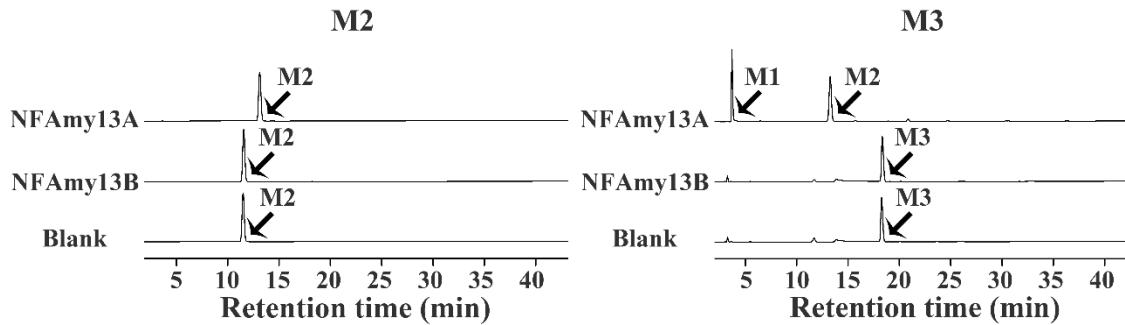


Fig. S3 Hydrolysis of maltooligosaccharides by NFAmy13A and NFAmy13B. 5 mM M2 and M3 were hydrolyzed by 1 μ M NFAmy13A or NFAmy13B in the presence of 1.25 mM CaCl_2 at pH 6.0 and 54 °C for 2 h. The reaction products were analyzed by HPAEC-PAD. M1, glucose; M2, maltose; M3, maltotriose.

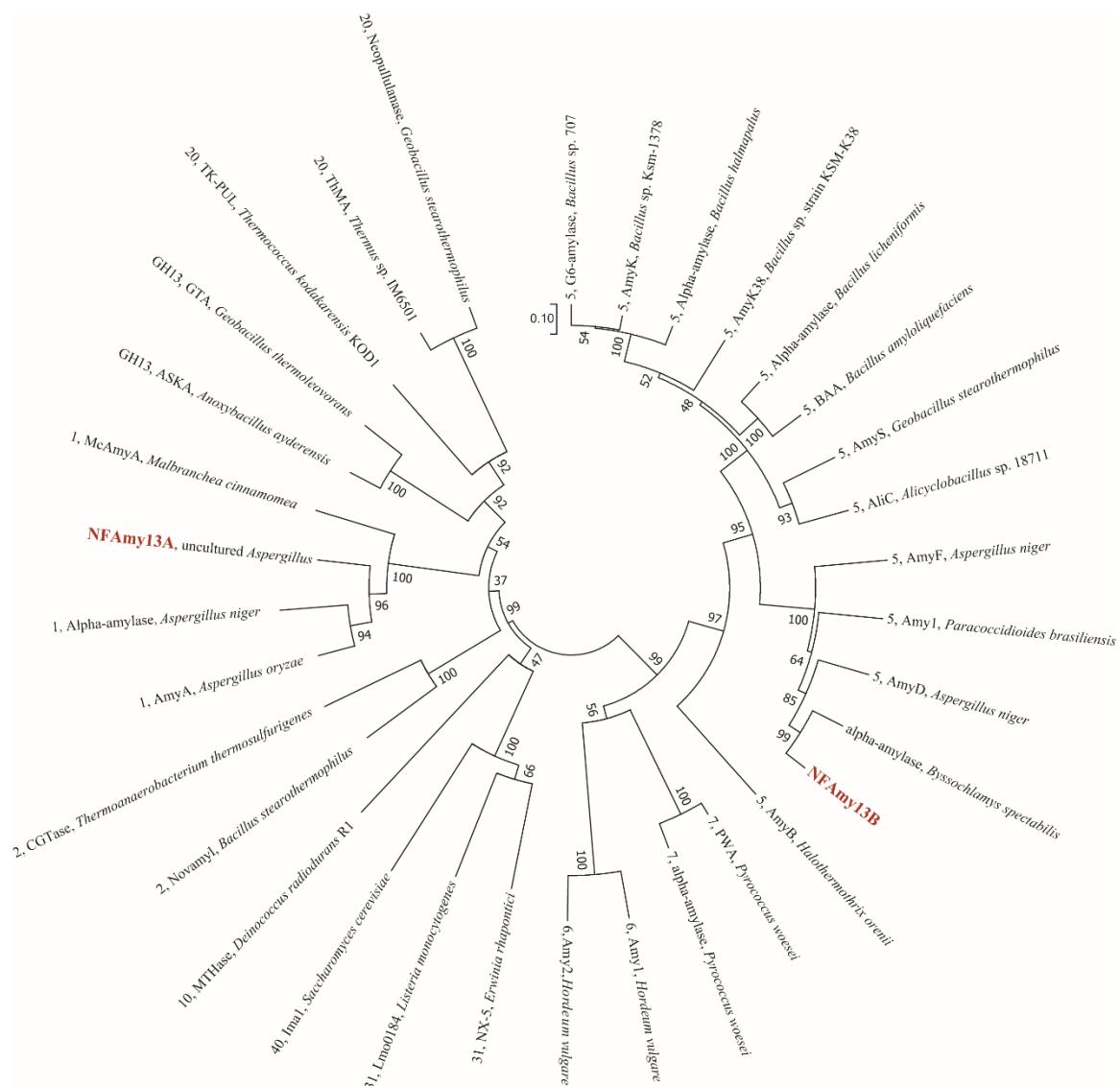
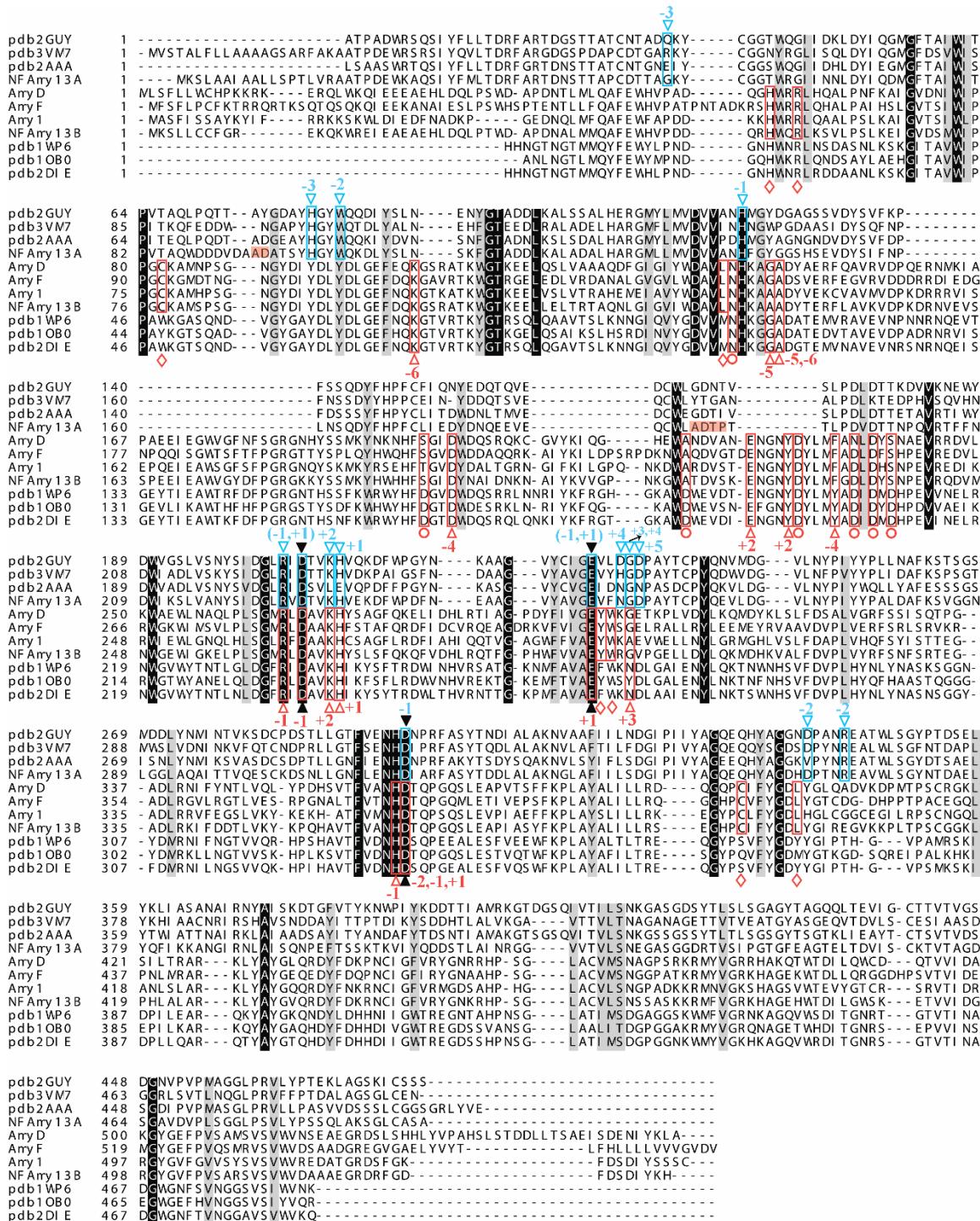


Fig. S4 Phylogenetic analysis of NFAmy13A and NFAmy13B. The amino acid sequences of enzymes from GH13 used for the construction of the phylogenetic tree were selected according to the similarities with NFAmy13A or NFAmy13B. Each enzyme was indicated a number corresponding to

the GH13 subfamily, its abbreviation, and source. Details are provided in Table S1.



\blacktriangledown Catalytic sites \triangle NFAmy13B binding sites \triangledown NFAmy13A binding sites

\circ NFAmy13B metal ions binding sites \blacksquare NFAmy13A blocks residues

\diamond Conserved amino acids of fungal α -amylase in group GH13_5

Fig. S5 Amino acid sequences alignment in NFAmy13A/B and high-similarity α -amylases.

References

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