

# **Analytical and Bioanalytical Chemistry**

## **Supplementary Information**

### **An integrated strategy of MS-network- based offline 2DLC-QTOF-MS/MS coupled with UHPLC-QTRAP<sup>®</sup>-MS/MS for the characterization and quantification of the non-polysaccharides in Sijunzi Decoction**

Bangjian Dong<sup>1</sup>, Chongsheng Peng<sup>1</sup>, Ping Ma, Xiaobo Li\*

School of Pharmacy, Shanghai Jiao Tong University, No. 800 Dongchuan Road, Minhang  
District, Shanghai, 200240, China

## **Index of Contents**

**Fig. S1** The first dimensional separation: HPLC-DAD chromatograms of NPSs in SJZD (A) at 210 nm; (B) at 254 nm.

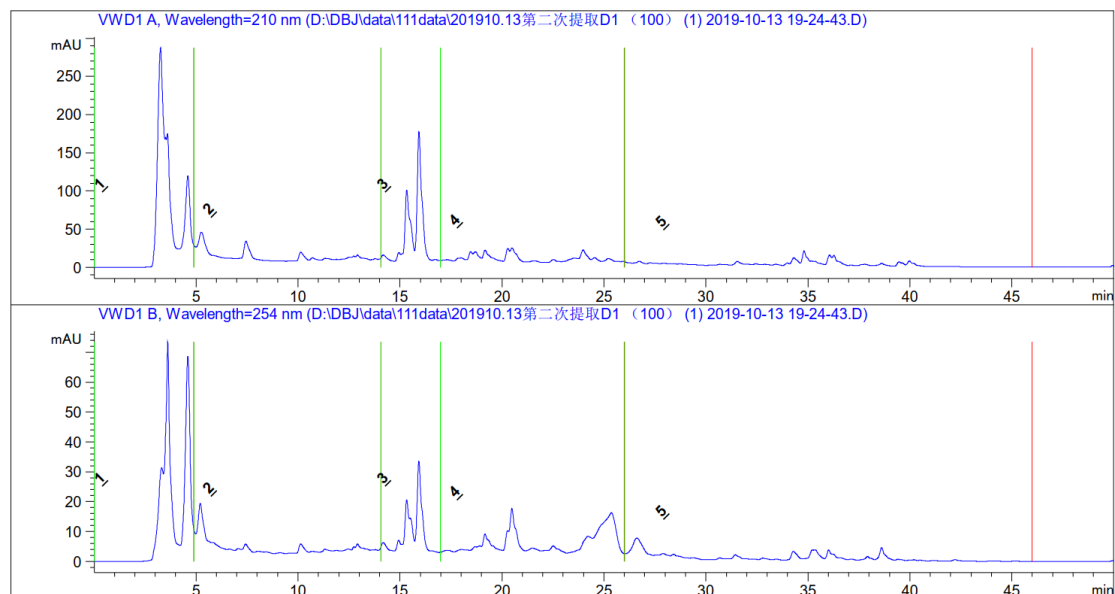
**Fig. S2** Comparison of the separation selectivity of four RP columns for SJZD components using CH<sub>3</sub>CN-0.1% water-containing formic acid for as the mobile phase (data recorded on an Agilent UHPLC-QTOF-MS instrument. Sample A was used for optimization. a is liquiritin; b is liquiritin apioside ).

**Fig. S3** Base peak intensity chromatograms (BPCs) in negative and positive ion mode of Fractions 1–5.

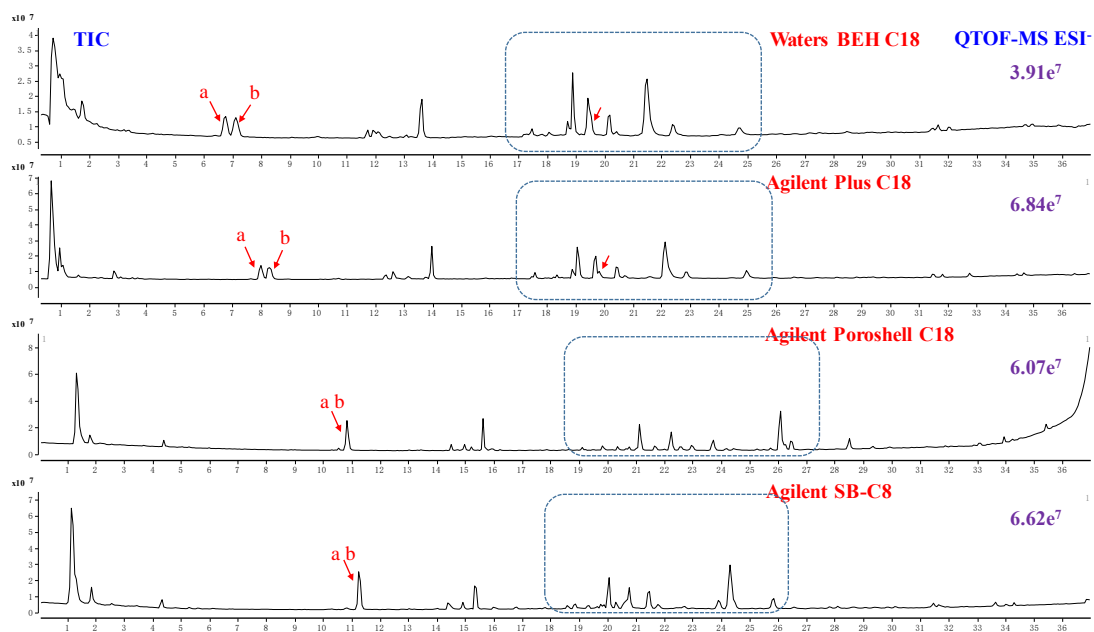
**Fig. S4** Tandem mass spectra of six potential new compounds **A25** (A), **A52**(B), **C73** (C), **C94** (D), **C154** (E), and **C210** (F).

**Fig. S5** Representative MRM chromatograms of 19 analytes from Sample B1 (A) and Sample B2 (B).

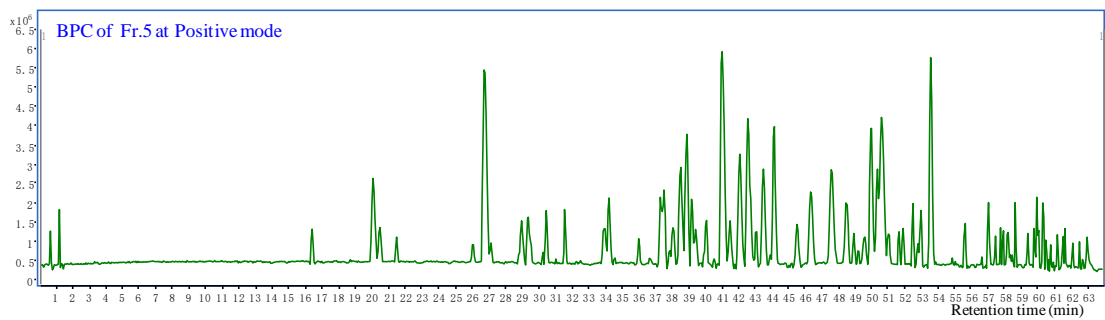
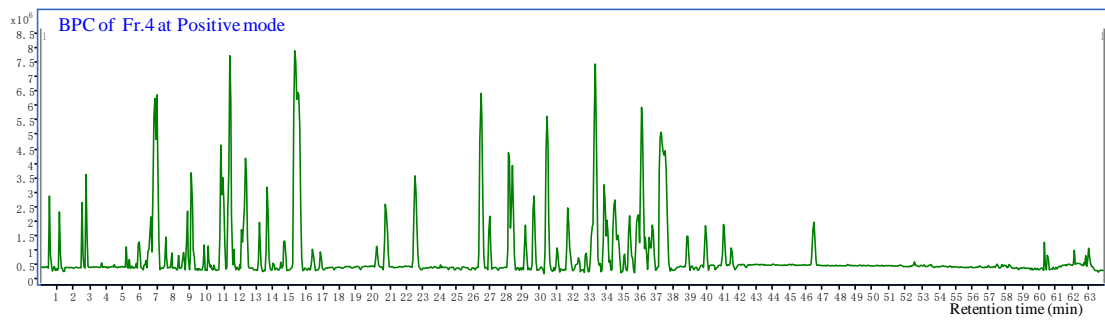
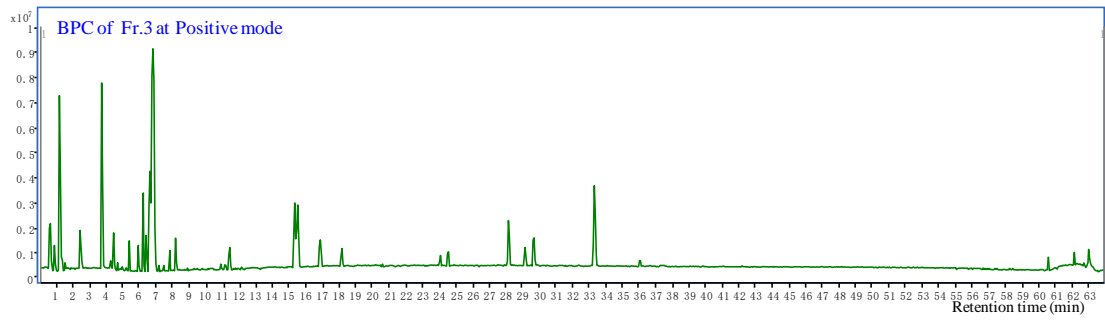
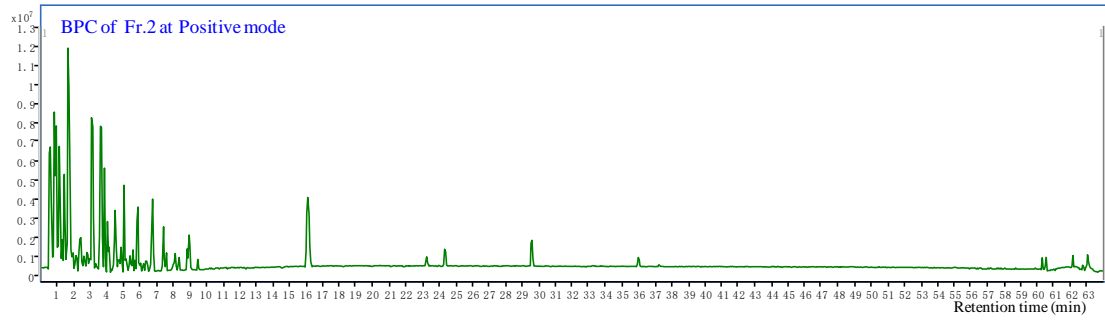
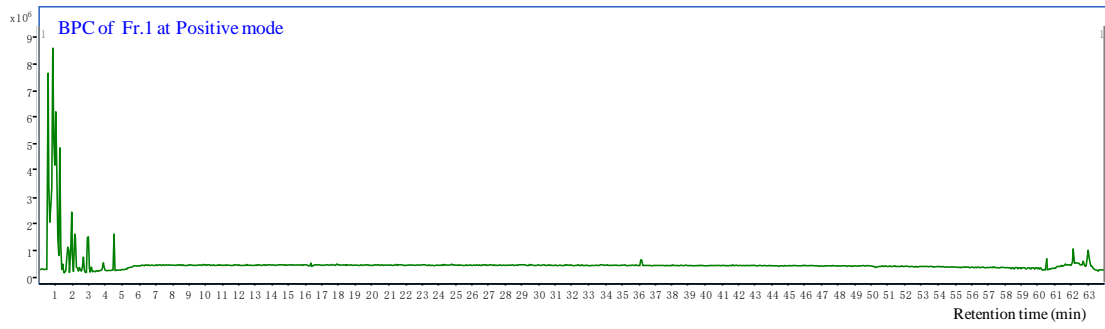
**Table S1** UHPLC-QTRAP<sup>®</sup>-MS/MS parameters for the quantification.

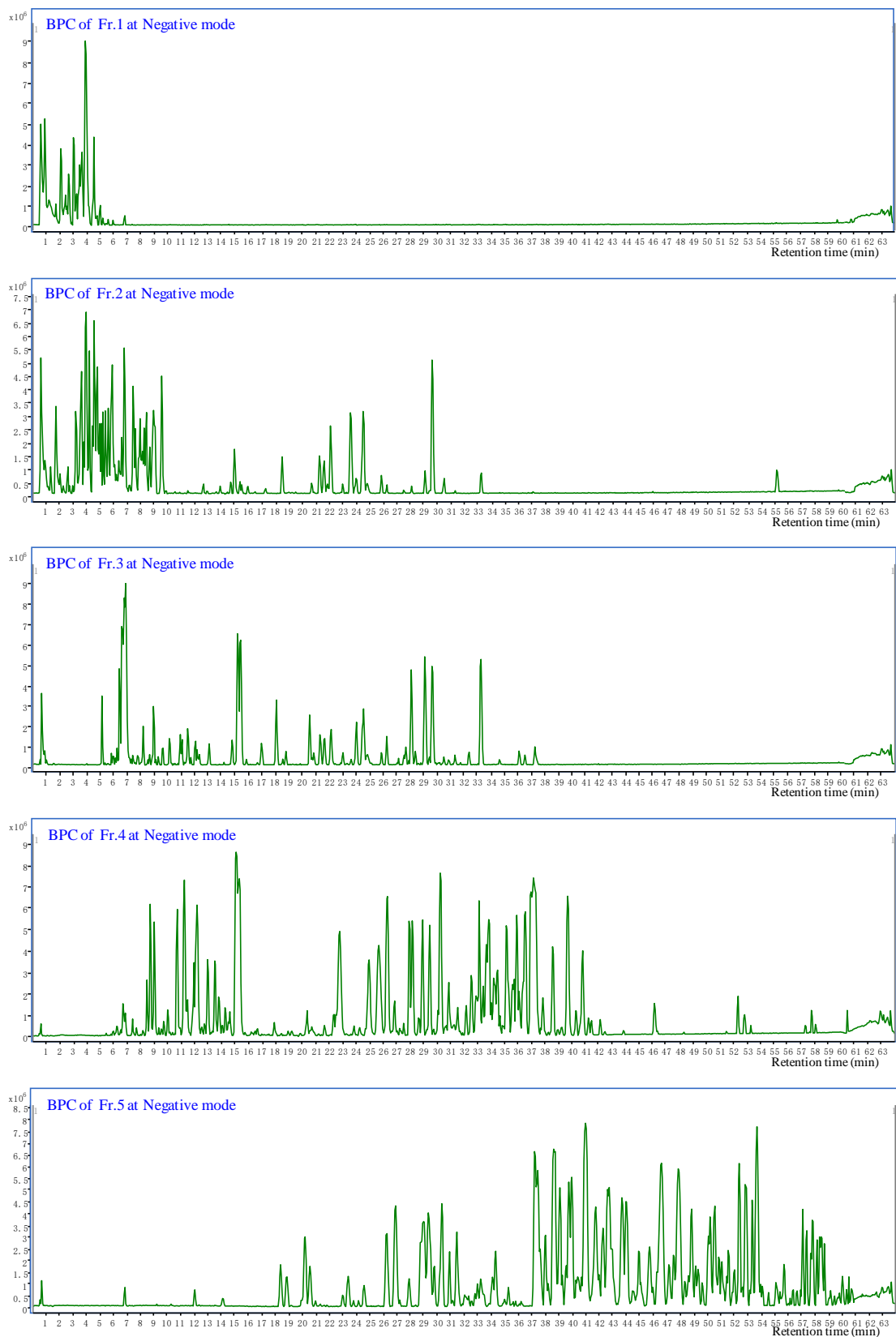


**Fig. S1** The first dimensional separation: HPLC-DAD chromatograms of NPS in SJZD (A) at 210 nm; (B) at 254 nm.

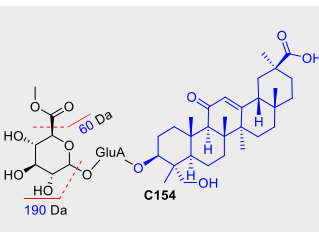
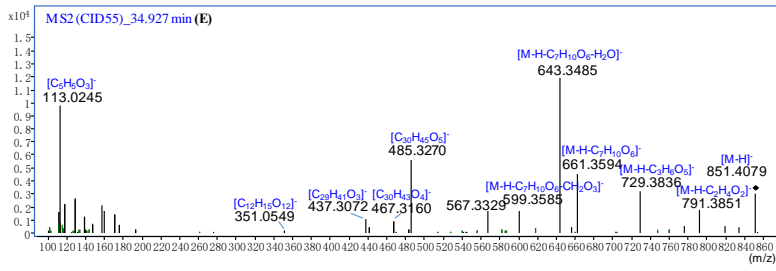
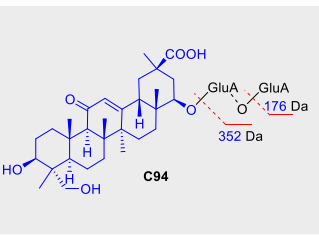
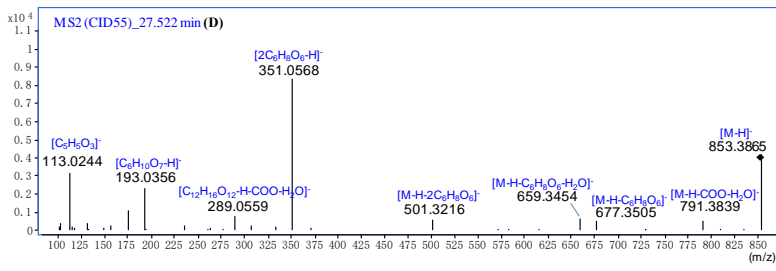
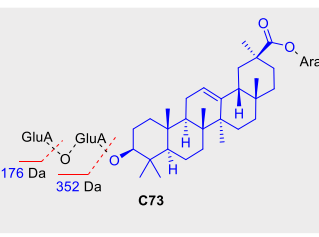
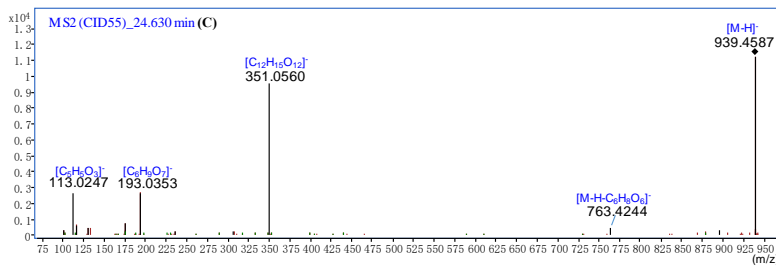
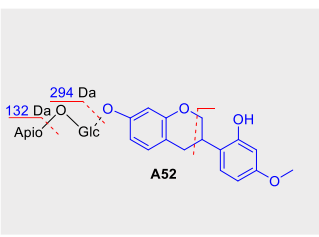
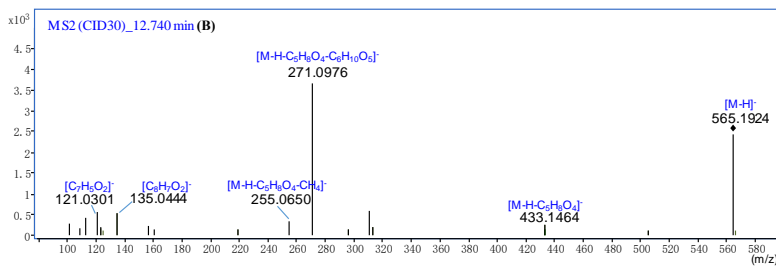
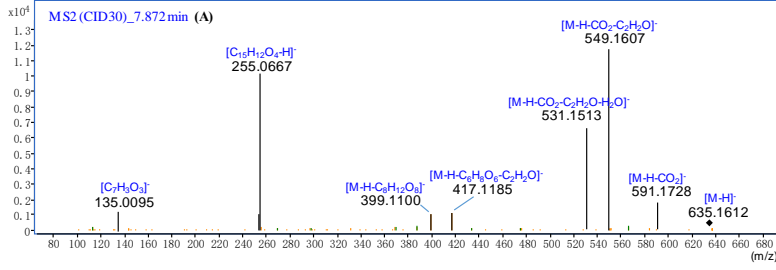
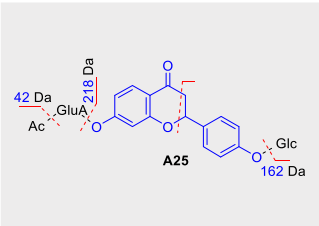
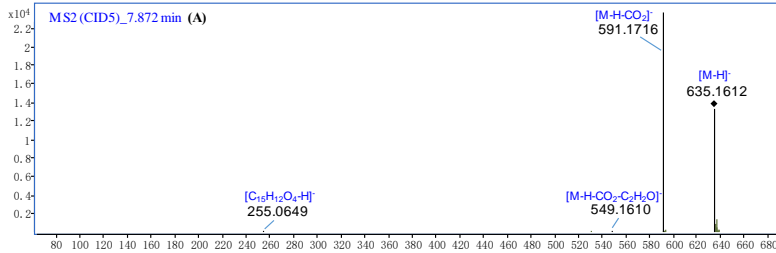


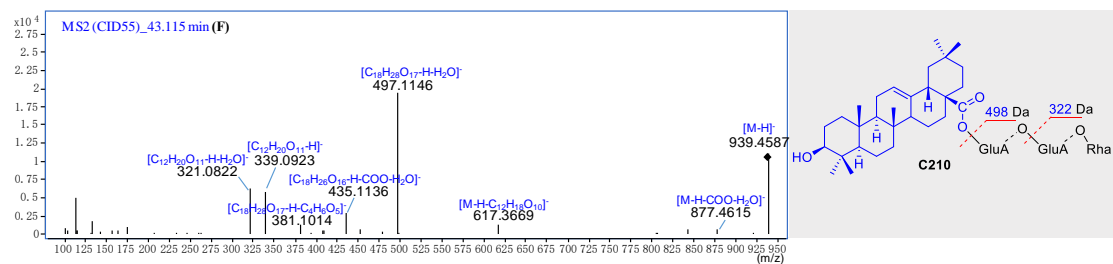
**Fig. S2** Comparison of the separation selectivity of four RP columns for SJZD components using  $\text{CH}_3\text{CN}$ -0.1% water-containing formic acid for as the mobile phase (data was recorded on an Agilent UHPLC-QTOF-MS instrument. Sample A was used for optimization. a is liquiritin; b is liquiritin apioside).



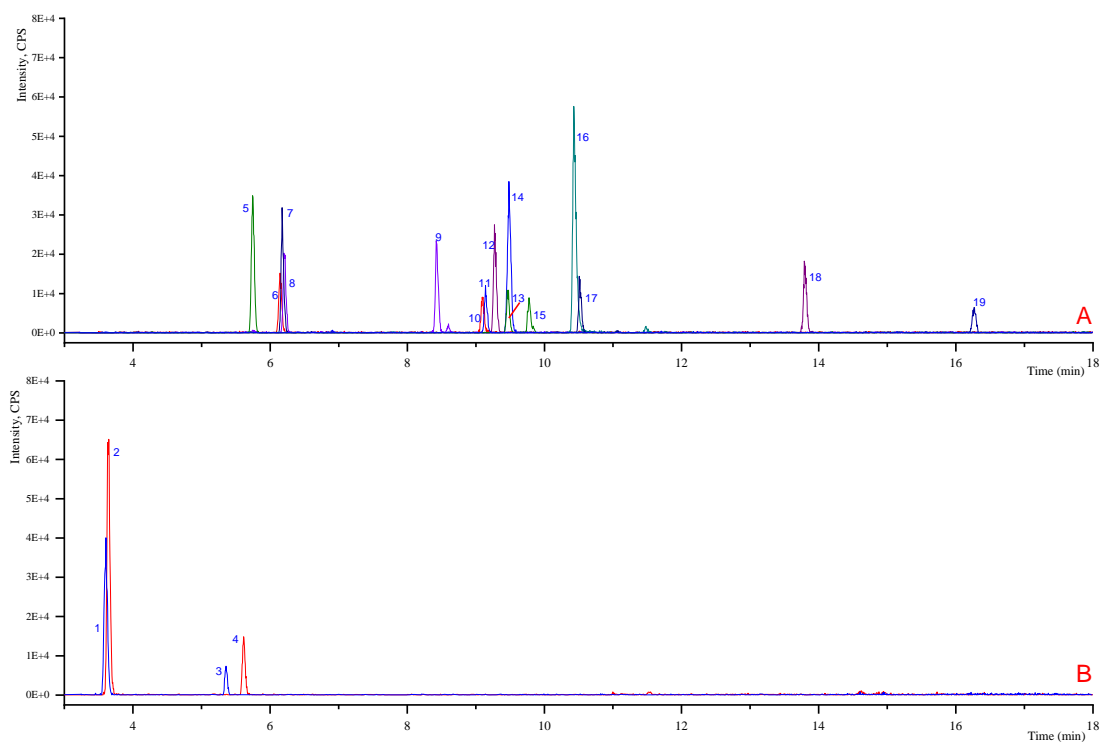


**Fig. S3** Base peak intensity chromatograms (BPCs) in positive and negative ion mode of Fractions 1–5.





**Fig. S4** Tandem mass spectra of six potential new compounds **A25** (A), **A52**(B), **C73** (C), **C94** (D), **C154** (E), and **C210** (F).



**Fig. S5** Representative MRM chromatograms of 19 analytes from Sample B1 (A) and Sample B2 (B).

**Table S1** UHPLC/QTRAP<sup>®</sup>-MS/MS parameters for the quantification.

NO.	Compound	Formula	RT (min)	Quantifier ( <i>m/z</i> )	Qualifier ( <i>m/z</i> )	CE <sup>a</sup> (eV)	DP (eV)	CXP (eV)
1	Liquiritin apioside	C <sub>26</sub> H <sub>30</sub> O <sub>13</sub>	3.59	549.2→255.1	549.2→135.0	-50	-100	-15
2	Liquiritin	C <sub>21</sub> H <sub>22</sub> O <sub>9</sub>	3.63	417.1→255.1	417.1→135.0	-28	-100	-15
3	Isoliquiritin apioside	C <sub>26</sub> H <sub>30</sub> O <sub>13</sub>	5.35	549.2→255.1	549.2→135.0	-50	-100	-15
4	Isoliquiritin	C <sub>21</sub> H <sub>22</sub> O <sub>9</sub>	5.60	417.1→255.1	417.1→135.0	-28	-100	-15
5	Ononin	C <sub>22</sub> H <sub>22</sub> O <sub>9</sub>	5.75	475.1→267.1	475.1→252.0	-30	-100	-15
6	Liquiritigenin	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	6.15	255.1→135.0	255.1→119.2	-30	-100	-15
7	Ginsenoside Re	C <sub>48</sub> H <sub>82</sub> O <sub>18</sub>	6.17	991.6→945.5	991.6→799.5	-30	-100	-15
8	Ginsenoside Rg1	C <sub>42</sub> H <sub>72</sub> O <sub>14</sub>	6.20	845.5→799.5	845.5→637.6	-30	-100	-15
9	Ginsenoside Rf	C <sub>42</sub> H <sub>72</sub> O <sub>14</sub>	8.42	845.5→799.5	845.5→637.6	-30	-100	-15
10	Isoliquiritigenin	C <sub>15</sub> H <sub>12</sub> O <sub>4</sub>	9.10	255.1→135.0	255.1→119.2	-30	-100	-15
11	Ginsenoside Rb1	C <sub>54</sub> H <sub>92</sub> O <sub>23</sub>	9.12	1153.6→1107.6	1153.6→945.5	-30	-100	-15
12	Ginsenoside Rg2	C <sub>42</sub> H <sub>72</sub> O <sub>13</sub>	9.22	829.5→783.5	829.5→637.6	-30	-100	-15
13	Ginsenoside Rc	C <sub>53</sub> H <sub>90</sub> O <sub>22</sub>	9.46	1123.6→1077.6	1123.6→945.5	-30	-100	-15
14	Formononetin	C <sub>16</sub> H <sub>12</sub> O <sub>4</sub>	9.48	267.1→252.0	267.1→223.0	-30	-100	-15
15	Ginsenoside Rb2	C <sub>53</sub> H <sub>90</sub> O <sub>22</sub>	9.76	1123.6→1077.6	1123.6→945.5	-30	-100	-15
16	Glycyrrhizic acid	C <sub>42</sub> H <sub>62</sub> O <sub>16</sub>	10.43	821.4→351.1	821.4→193.1	-58	-100	-15
17	Ginsenoside Rd	C <sub>48</sub> H <sub>82</sub> O <sub>18</sub>	10.51	991.6→945.5	991.6→783.5	-30	-100	-15
18	Ginsenoside Rg3	C <sub>42</sub> H <sub>72</sub> O <sub>13</sub>	13.75	829.5→783.5	829.5→621.5	-30	-100	-15
19	Ginsenoside Rk1	C <sub>42</sub> H <sub>70</sub> O <sub>12</sub>	16.26	811.5→765.5	811.5→161.3	-30	-100	-15

<sup>a</sup> the CE values was applied for quantifier ions