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Supplemental information

**The mitochondrial carrier SFXN1
is critical for complex III integrity
and cellular metabolism**

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A

Sequence alignment

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SFXN1 -----MSGELPPNINIKPRWDQSTFIGRANHFPTVTDPRNILLTNEQLESARKIVHDYRQGI VPPGLTENELWRKYIYDSAFHPD
SFXN3 -----MGELPLDINIQEPRWDQSTFELGARHFPTVTDPRNLLSQAQLEASRNIVQNYRAGVVTGITEDQLWRKYVYDSAFHPD
SFXN2 -----MEADLSGFNIDAPRWQDQRTFLGKVKHFLNITDPRTFVFSERELDWAKVMVEKSRMGVVPFGTQVEQLLYAKKLYDSAFHPD
SFXN5 --MADTATTASAAAASASDAPPFQLGKPRFQVTSFYGRFRHFLDIIDPRTLFVTERRLEAVQLLEDYKHGTLRFGVTNEQLWSQKIKQAILHPD
SFXN4 MSLEQEETQPGRLGRDAVPAFIEPNVRFWITTEQSFIRRLQWTELLDPTNVFISVESIENSRLQCTN-EDVSSASADQRIQEWKRSLATVHPD

SFXN1 TGEKMLIGMSAQVPMNMTITGCMMTFYRTPAVLFWQINQSFNAVVNYSNRSGDAPLTVNELGTAYVSAITGAVATALGLNALTKHVS-----P
SFXN3 TGEKVVIGMSAQVPMNMTITGCMMLTFYRKTPTVVFVWVWVNSFNALVNYNSRSGDPTITVRQLGTAYVSAITGAVATALGLKSLTKHLP-----P
SFXN2 TGEKMNVIIGMSFQLPGCMIIITGFMLOFYRTMPAVIFWQVWVNSFNALVNYNSRSGDPTITVRQLGTAYVSAITGAVATALGVMNMLTKKAP-----P
SFXN5 TNEKLFMPFPMGSGYIPFGTPIVVGILLPNQTLASTVFWQVWVNSFNALVNYNSRSGDPTITVRQLGTAYVSAITGAVATALGVMNMLTKKAP-----P
SFXN4 SSLNLPKLFPPAALFFMAPTVFLSMTPLKGIKSVLPLVFLCAYMAAFNSINGNRSYCKPLERSLLMAGVASSSTFLGVIPOFVQMKYGG-----LTGP

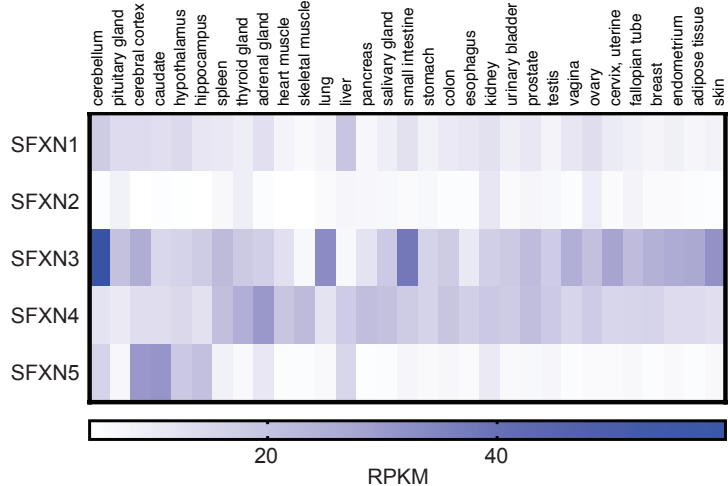
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SFXN3 LVGRFVPPFAAVAAANCINIPLMRQRELKVGIPVDAEAGORLQVSVTAAKQGIQVVISRIICMAIPAMAIPPLIMDTLEKDFLKRFPWLGAPLQVGLVGF
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SFXN5 LIQRFVPPFAVAAANCINIVMLFYGLEEELVLDSDGNLVGSSKIAARHALLETALTRVLPMPILVLPPIVMSMLEKATLLOARPRLLLPVQSLVCLAA
SFXN4 WIKRLPVIPLVQASGMVYMSLSLSIKGIAMVMEKGNVLGHSRIATKAVRETLASRIVLFGTSALTPVFTYFFKRTQYFRKNPGLWLLKLSCTVLA

SFXN1 LVFATPLCCALFPQKSSMSVTSLEAELOAKIQESHPELR-RVYFNKGL
SFXN3 LVFATPLCCALFPQKSSIHISNLEPELRAQIHEQNPSVE-VVYFNKGL
SFXN2 LIFMVVACGLFPQKCELPVSYLEPKLQDTIKAKYGELEPYVYFNKGL
SFXN5 FGLALPLAISLFPQMSSEIETSQLEPEIAQATSSR-----TVYFNKGL
SFXN4 MGLMVVFSFSIFPQIGITIQYCSLEEKIQSPTEET-----EIFYHGRV

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B

Tissue expression



C

SFXN1 across species

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Saccharomyces cerevisiae MASSVPGPIDLPESRYDLSTYWGRIHCAEISDPTMLLTTEKDLAHAREIISAYRHGELKE--TTFEWFRAKKQLDSTVHPDGTGKVLPL
Homo sapiens MSGELPPNINIKPRWDQSTFIGRANHFPTVTDPRNILLTNEQLESARKIVHDYRQGI VPPGLTENELWRKYIYDSAFHPDGTGKMLILI
Bos taurus MSGELPPNINIKPRWDQSTFIGRANHFPTVTDPRNILLTNEQLEAARKVHDYRQGIIPSGLTENELWRKYIYDSAFHPDGTGKMLILI
Ovis aries MSGELPPNINIKPRWDQSTFIGRANHFPTVTDPRNILLTNAQLEAARKVHDYRQGI VPPGLTENELWRKYIYDSAFHPDGTGKMLILI
Sus scrofa MSGELPPNINIKPRWDQSTFVGRANHFPTVTDPRNILLTNEQLENARKVHDYRQGI VPPGLTENELWRKYIYDSAFHPDGTGKMLILI
Mus musculus MSGEVPPNINIKPRWDQSTFIGRASHFPTVTDPRNILLTNEQLENARKVHDYRQGI VPAGLTENELWRKYAYDSAFHPDGTGKMTLI
Rattus norvegicus MSGEVPPNINIKPRWDQSTFIGRASHFPTVTDPRNILLTNEQLENARKVHDYRQGI VPAGLTENELWRKYAYDSAFHPDGTGKMTLI

Saccharomyces cerevisiae FRMSSNVLSNLVVTGMLTGPLGCTAGTVFQWQANQSLNAVNSANANKSHPMSTSQLLTNYAAAVTASCGVALGLNLLVPRLKNISPHSK
Homo sapiens GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKHVS
Bos taurus GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKHVS
Ovis aries GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKRVS
Sus scrofa GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKHVS
Mus musculus GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKRVS
Rattus norvegicus GRMSAQVPMNMTITGCMMTFYRTPAVLFWQWVQVNSFNNAVNYNTRSGDAPLTVNELGTAYVSATTGAVATALGLN-----ALTKHVS

Saccharomyces cerevisiae LILGRLVPPFAAVSAGIVNVFLMRGNEIRKIGSVFDSNGDEVGKSKKAAMFVGETALSRVINATPTMVIPLLILVLRQGRVGLKSKLGV
Homo sapiens PLIIGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESANAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM
Bos taurus PLIIGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESANAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM
Ovis aries PLVGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESASAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM
Sus scrofa PLIIGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESANAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM
Mus musculus PLIIGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESANAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM
Rattus norvegicus PLIIGRFVPPFAAVAAANCINIPLMRQRELKVGIPVTDENGRLGESANAQAQITQVVSRILMAAPGMAIPPFIMNTLEKAFKLRFPWM

Saccharomyces cerevisiae QTLANLGLISVTMFSALPFALGIFPQQAHLNKLLEPELHGKDKDGKPIEKVYFNKGI
Homo sapiens SAPIQVGLVGFCLVFATPLCCALFPQKSSMSVTSLEAELOAKIQESHPELRVYFNKGL
Bos taurus SAPVQVIGVGFCLVFATPLCCALFPQKSSMSVTSLEAELOAKIARETYPELRRVYFNKGL
Ovis aries SAPVQVIGVGFCLVFATPLCCALFPQKSSMSVTSLEAELOAKIARETYPELRRVYFNKGL
Sus scrofa SAPIQVGLVGFCLVFATPLCCALFPQKSSMSVTSLEAELOAKIARETYPELRRVYFNKGL
Mus musculus SAPIQVTLVGFCLVFATPLCCALFPQKSSMSVTSLEDELQASIQRTHPELRRVYFNKGL
Rattus norvegicus SAPIQVTLVGFCLVFATPLCCALFPQKSSMSVTSLEDDLQASIQKSHPELRRVYFNKGL

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Figure S1

Figure S1, related to Figure 1. SFXN isoforms.

(A) Alignment of SFXN isoforms. Identical amino acids are boxed whereas highly similar ones are highlighted blue. Predicted transmembrane domains are in pink.

(B) Tissue distribution of SFXN isoforms. Gene expression is reported as median RPKM, generated by the GTEx project and accessed via the Human Protein Atlas.

(C) SFXN1 is highly conserved in eukaryotes. *Saccharomyces cerevisiae* possesses a related protein (Fsf1p).

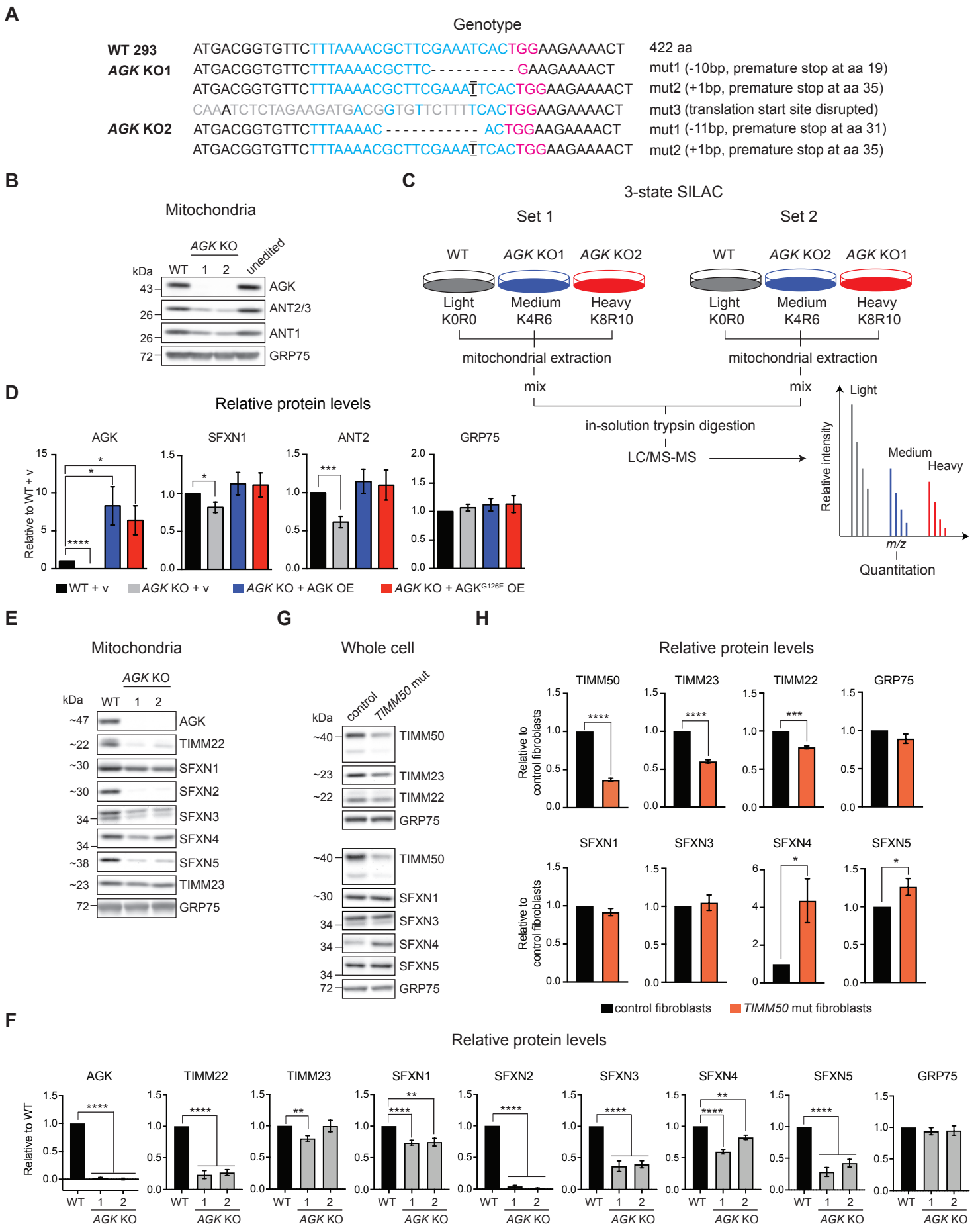


Figure S2

Figure S2, related to Figure 1. AGK disruption reduces steady-state protein levels of mitochondrial carriers including SFXNs.

(A) Genomic mutations documented in two HEK293-based *AGK* KOs generated by CRISPR/Cas9. gRNA target site is in blue, while the protospacer-adjacent motif (PAM) sequence is in red.

(B) Immunoblotting using mitochondrial isolates for AGK, ANT1, and ANT2/3 in WT, AGK-deficient clones, and an unedited clone that went through the transfection process with the CRISPR construct. GRP75 served as a loading control.

(C) SILAC scheme related to Figure 1J.

(D) Densitometric analysis of bands for proteins in Figure 1K. Protein levels in WT were set to 1.0. (mean±SEM, $n \geq 4$).

(E) Immunoblotting for different SFXN isoforms and select TIM subunits using mitochondrial isolates from WT and AGK-null clones. GRP75 served as a loading control.

(F) Densitometric analysis of bands for proteins in (E). Protein levels in WT were set to 1.0. (mean±SEM, $n \geq 4$).

(G) Immunoblotting for different SFXN isoforms and select TIM subunits using cell extracts from control and *TIMM50* mutation-containing patient fibroblasts. GRP75 served as a loading control. (H) Densitometric analysis of bands for proteins in (G). Protein levels in WT were set to 1.0. (mean±SEM, $n \geq 3$).

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, unpaired Student's t-test

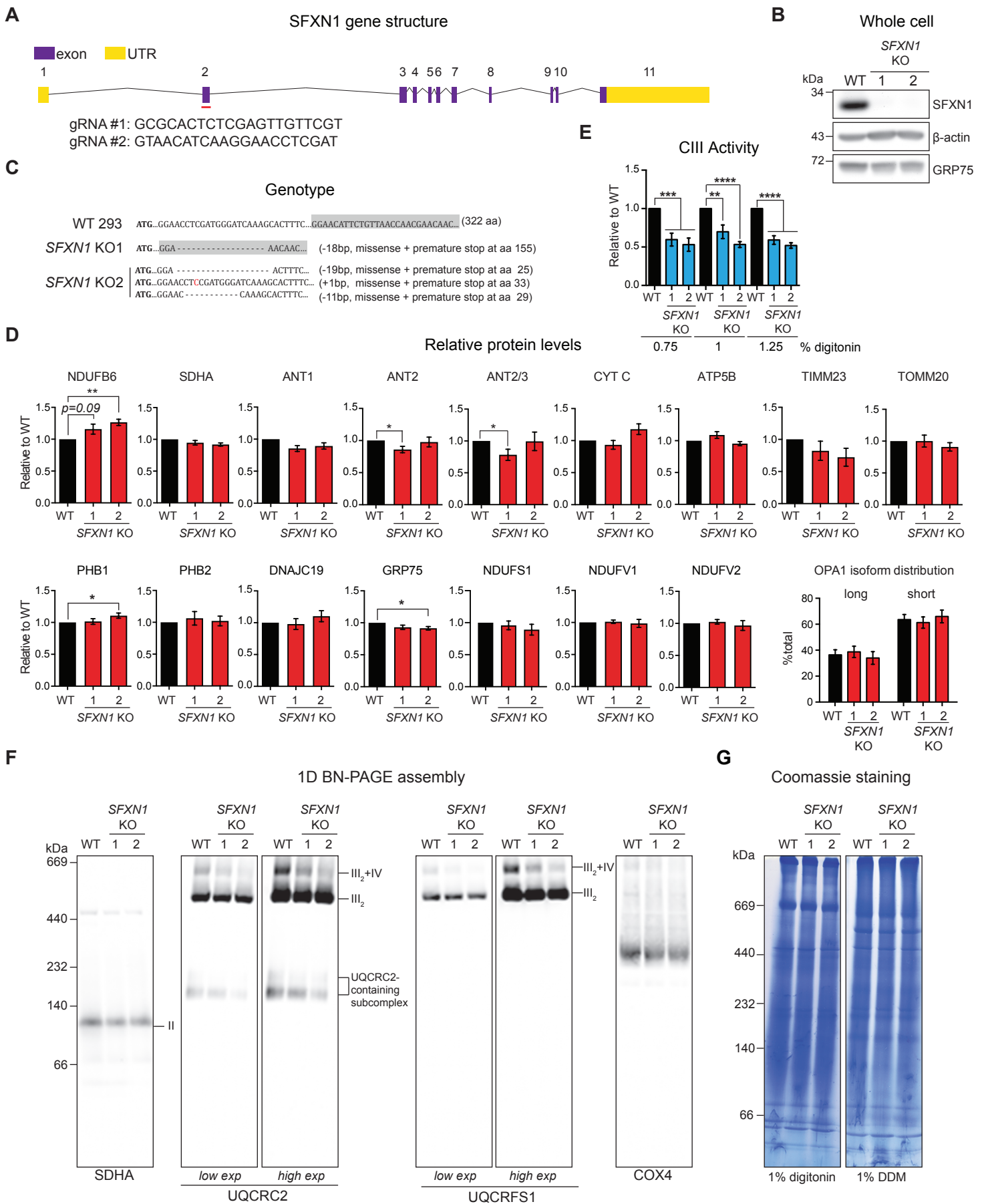


Figure S3

Figure S3, related to Figures 2 and 3. Effects of SFXN1 ablation on the levels, function and assembly of respiratory complexes.

(A) *SFXN1* gene structure showing the target sites of gRNAs used for CRISPR/Cas9.

(B) Whole cell extracts from WT and *SFXN1* KO clones were analyzed by SDS-PAGE and immunoblotting.

(C) Genotyping shows genetic lesions resulting in premature stop codons in both clones.

(D) Densitometric analysis of bands for other proteins immunoblotted as shown in Figure 2A. Protein levels in WT were set to 1.0. (mean±SEM, $n \geq 4$).

(E) Complex III activity was measured by solubilizing mitochondria with increasing concentrations of digitonin and monitoring cytochrome *c* reduction at 550 nm (mean±SEM, $n=6$).

(F) 1D BN assembly of respiratory complexes I-IV was assessed by solubilizing mitochondria in 1% (w/v) DDM, resolving in a 4-16% BN gel, and immunoblotting for the indicated complex subunits.

(G) Coomassie staining of mitochondria solubilized with 1% (w/v) digitonin or 1% (w/v) DDM and resolved on 6-16% BN gel.

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, unpaired Student's t-test

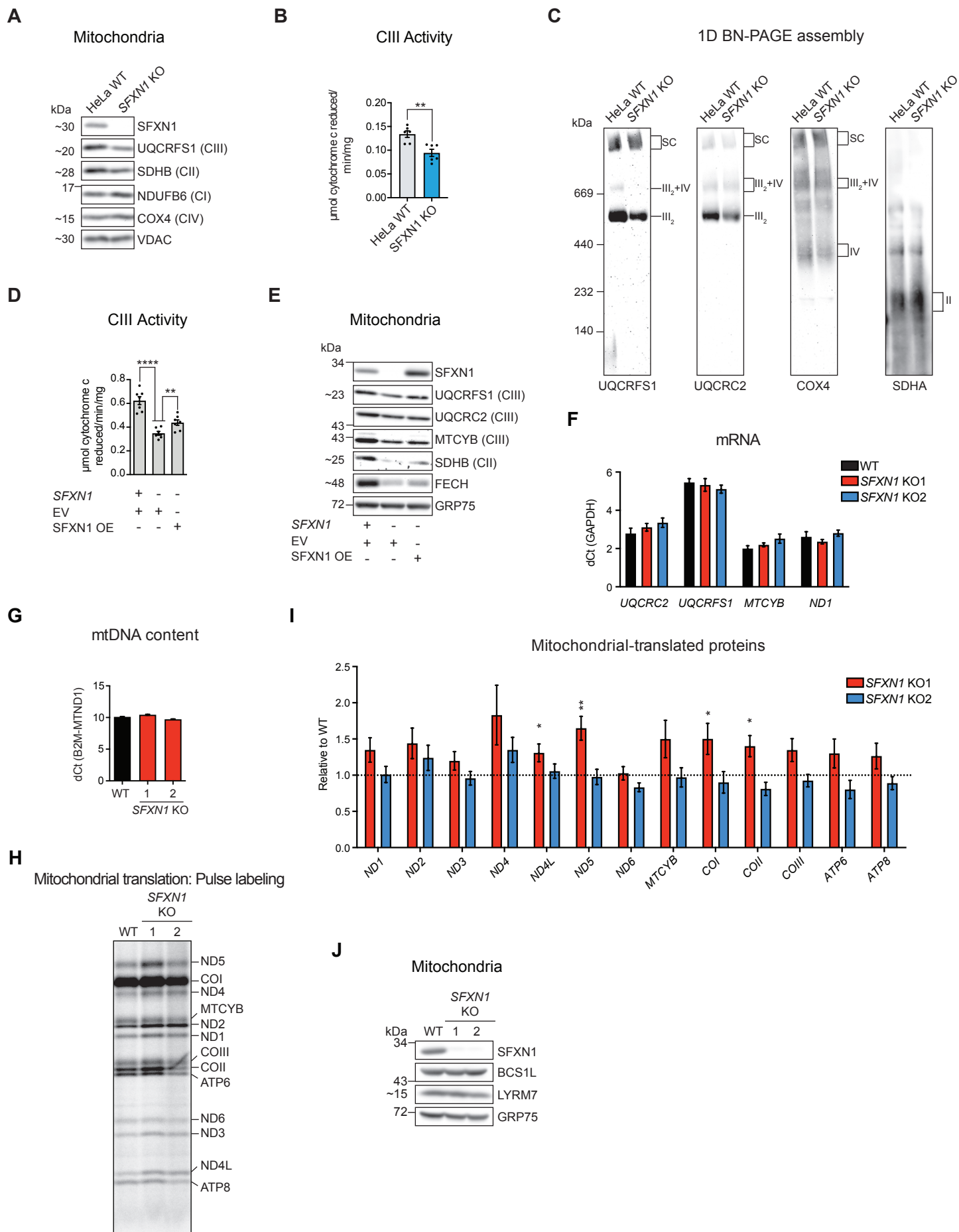


Figure S4

Figure S4, related to Figure 3. Complex III-associated function of SFXN1 does not derive from changes in mtDNA content, its transcription or translation, or altered CIII assembly factor levels.

HeLa was used for (A) to (C). HEK293 was used for the remainder of the experiments.

(A) Immunoblotting for select OXPHOS subunits using mitochondrial extracts.

(B) Complex III activity was measured after solubilizing mitochondria with 0.5% (w/v) DDM and monitoring cytochrome *c* reduction at 550 nm (mean±SEM, *n*=6).

(C) 1D BN assembly of Complexes II-IV was analyzed by solubilizing mitochondria in 1% (w/v) digitonin, resolving clarified extracts on a 4-16% BN gel, and immunoblotting for the indicated subunits.

(D) Complex III activity was measured after solubilizing mitochondria with 0.5% (w/v) DDM and monitoring cytochrome *c* reduction at 550 nm (mean±SEM, *n*=7).

(E) Immunoblotting for select mitochondrial proteins using mitochondrial isolates.

(F) Gene expression analysis of CIII subunits *UQCRC2*, *UQCRFS1* and *MTCYB* by quantitative PCR (qPCR). *ND1*, a Complex I subunit, served as a control. Data presented as mean fold-change (FC)±SEM (*n*=6) relative to HEK293 WT. FC was calculated using the $2^{-\Delta\Delta CT}$ method. GAPDH served as the housekeeping gene.

(G) mtDNA content determination by qPCR. Genomic DNA isolated from the indicated cell lines served as template. mtDNA levels are presented as mean dC_T ±SEM (*n*=6) between the mitochondria-encoded *ND1* and the nuclear-encoded reference gene β -2-microglobulin (*β 2M*).

(H) Mitochondrial translation experiments. Cytosolic translation was inhibited by anisomycin (100 μ g/mL), and mitochondrially-encoded polypeptides were pulse-labeled for 1 hr with 35 S-methionine/cysteine (200 μ Ci/mL). Bands were detected by phosphoimaging.

(I) Densitometric band analysis was performed and the distribution of signals from mitochondrial-translated products was calculated as a percentage of the total signal per experiment (mean % total signal±SEM, *n*=6).

(J) Immunoblotting for CIII assembly factors using mitochondrial extracts.

p*<0.05, *p*<0.01, ****p*<0.001, *****p*<0.0001, unpaired Student's t-test

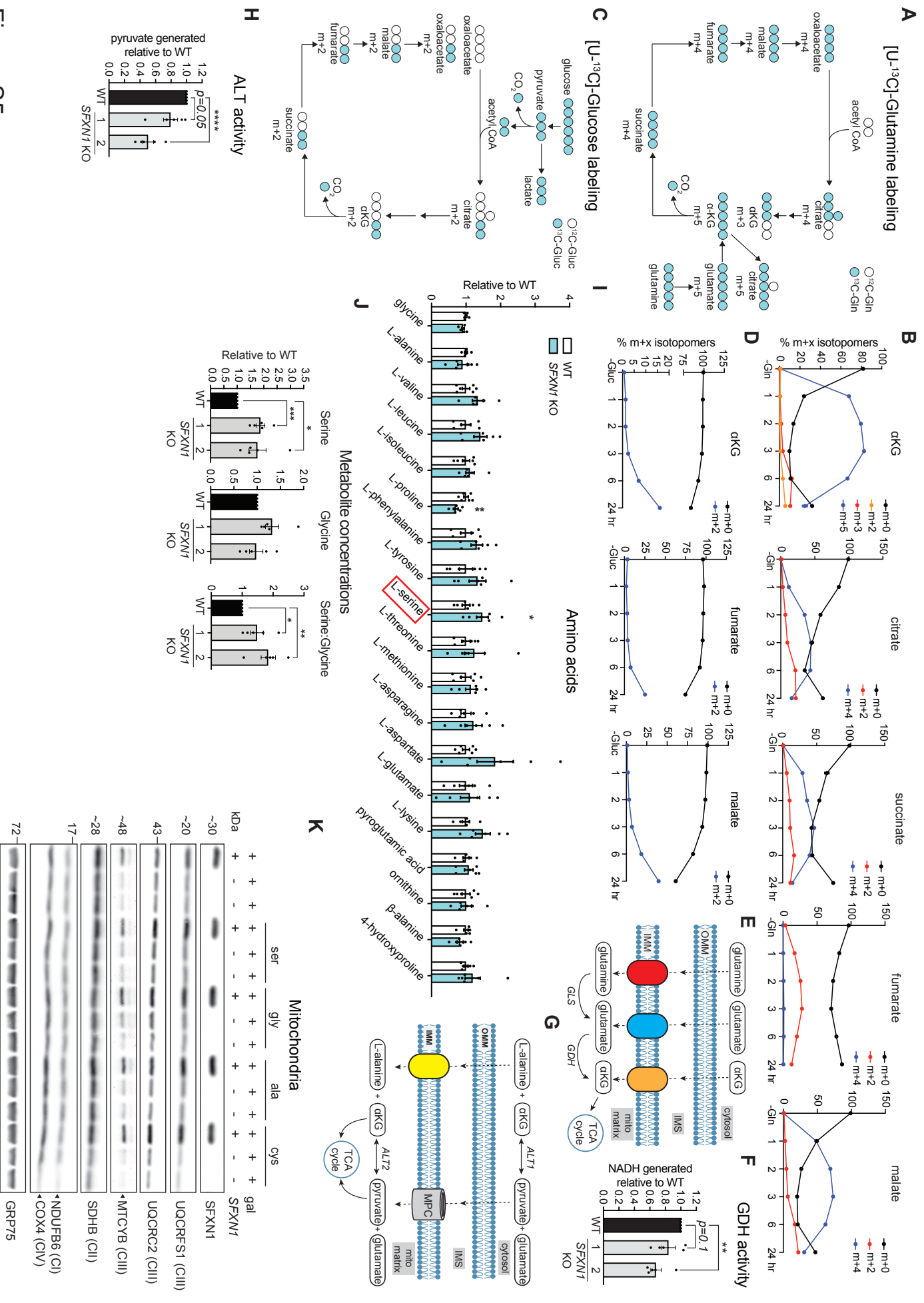


Figure S5

Figure S5, related to Figures 4, 6 and 7. Assessment of central carbon and amino acid metabolism in *SFXN1* KOs.

- (A) Schematic diagram of [U-¹³C]-glutamine labeling of TCA intermediates.
 - (B) Time-course experiment for [U-¹³C]-glutamine labeling.
 - (C) Schematic diagram of [U-¹³C]-glucose labeling of TCA intermediates.
 - (D) Time-course experiment for [U-¹³C]-glucose labeling.
 - (E) Reaction mediated by glutamine dehydrogenase (GDH).
 - (F) GDH activity in cellular extracts of HEK293 WT and *SFXN1* KOs.
 - (G) Reaction mediated by alanine aminotransferase (ALT).
 - (H) ALT activity in cellular extracts of HEK293 WT and *SFXN1* KOs.
 - (I) Relative quantification of amino acids by GC-MS (mean±SEM, *n*=6), related to Figure 4C.
 - (J) Serine and glycine abundance in cellular extracts relative to WT as determined by fluorimetric-based assays. Serine:glycine ratio is also presented (mean±SEM, *n*=5).
 - (K) Immunoblotting for select respiratory complex subunits using mitochondrial isolates from cells grown for 2 days in galactose-based media only or supplemented with 2 mM serine, glycine, alanine or cystine.
- **p*<0.05, ***p*<0.01, ****p*<0.001, *****p*<0.0001, unpaired Student's t-test

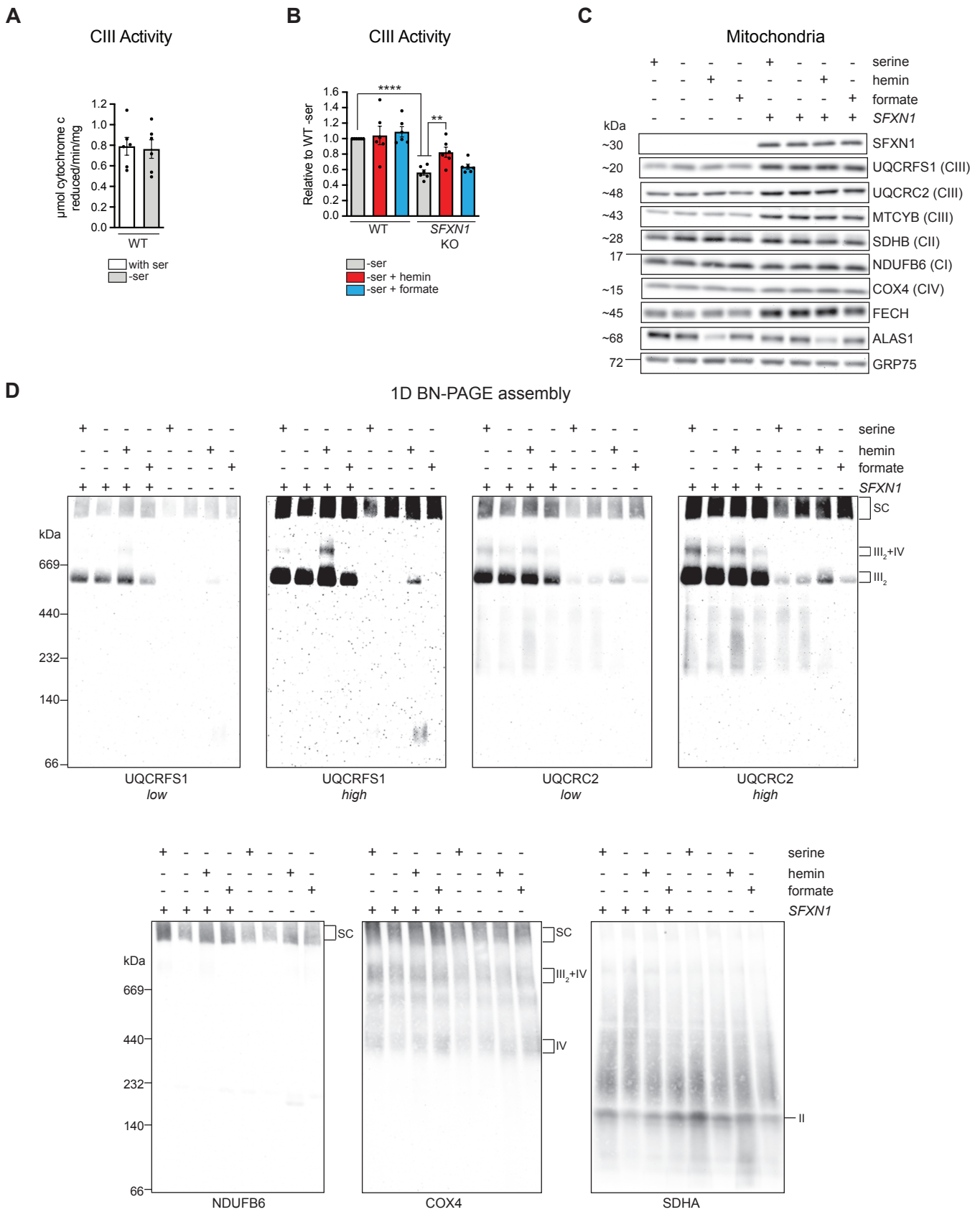


Figure S6

Figure S6, related to Figure 7. Complex III function in the absence of serine and upon hemin and formate supplementation.

(A) Complex III activity in DDM-solubilized mitochondria from cells grown in glucose-containing media with or without serine (mean±SEM, $n=6$).

Mitochondrial isolates from cells grown in glucose-containing media without serine, or with addition of hemin or formate were used to assess:

(B) Complex III activity (mean±SEM, $n=6$),

(C) Steady-state abundance of select proteins, and

(D) 1D BN assembly of respiratory complexes.

* $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$, unpaired Student's t-test

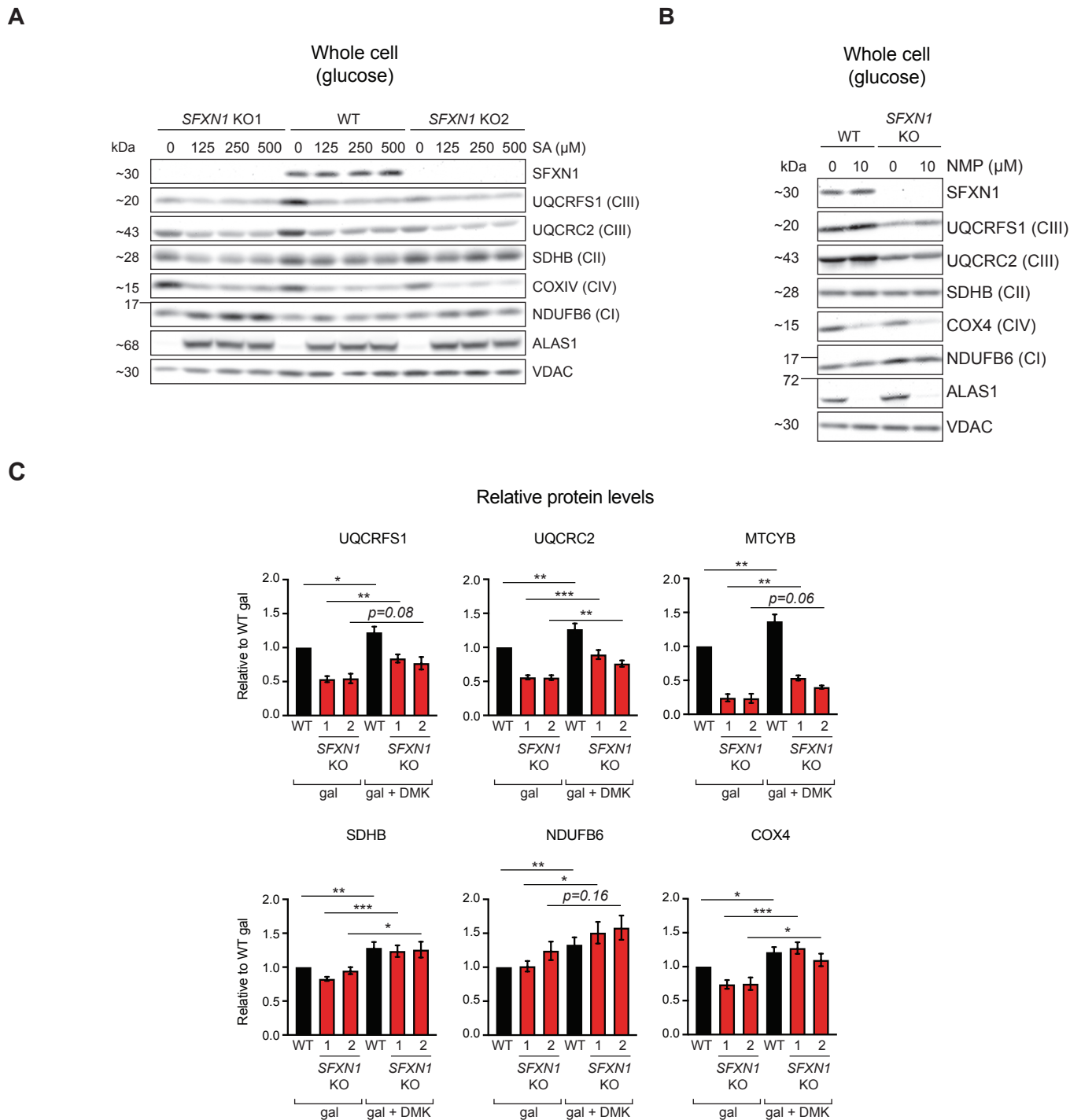


Figure S7, related to Figure 7. Effect of heme biosynthetic inhibitors or DMK supplementation on steady-state abundance of respiratory complex subunits.

(A) Immunoblotting for select respiratory complex subunits using cell lysates. Cells were grown in glucose-containing media with the indicated SA concentration for 2 days. ALAS1 protein is elevated in response to SA-mediated inhibition of heme biosynthesis.

(B) Immunoblotting for select respiratory complex subunits using cell lysates. Cells were grown in glucose-containing media with 10 mM NMP for 2 days. ALAS1 protein is decreased in response to NMP-mediated inhibition of heme biosynthesis.

(C) Densitometric analysis of bands for select proteins in (H). Protein levels in WT were set to 1.0 (mean±SEM, n≥5).

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$, unpaired Student's t-test