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Supporting Information

Strategic Synthesis of 'Picket Fence' Porphyrins Based on Nonplanar Macrocycles**

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Experimental Procedures

General Materials and Methods

All chemicals were supplied by Sigma Aldrich, Acros Organics, Fluka, Frontier Scientific, and Fischer and handled without further purification unless otherwise stated. Anhydrous DCM used in large scale reactions, obtained *via* drying with phosphorus pentoxide followed by distillation, while smaller amounts of EtOH and toluene were used as commercially available HPLC grade solvents. Chloroform used in titration studies was neutralized with potassium carbonate and filtered over celite before carrying out any measurements. Condensation reactions were carried out under an argon atmosphere. Reactions involving moisture and/or air-sensitive reagents were carried out in pre-dried glassware and with standard Schlenk line techniques. The 3,4-diethyl-1H-pyrrole **11**^[1] and [2,3,7,8,12,13,17,18-octaethyl-5,10,15,20-tetrakis(2-aminophenyl)porphyrin] **1**^[2] was prepared *via* known procedures. Dichloromethane (CH₂Cl₂) was dried over P₂O₅. Yields refer to chromatographically and spectroscopically (¹H NMR) homogeneous material, unless otherwise noted. Reactions were monitored by thin layer chromatography (TLC) and absorption spectroscopy.

Analytical thin layer chromatography was performed using silica gel 60 (fluorescence indicator F254, precoated sheets, 0.2 mm thick, 20 cm × 20 cm; Merck) or aluminum oxide 60 (neutral, F254; Merck) plates and visualized by UV irradiation (λ = 254 nm).

Column chromatography was carried out using Fluka Silica Gel 60 (230–400 mesh; Merck) or aluminum oxide (neutral, activated with 6% H₂O, Brockman Grade III). Mobile phases are given as (v/v).

Mass spectrometry was performed with a Q-Tof Premier Waters MALDI quadrupole time-of-flight (Q-TOF) mass spectrometer equipped with Z-spray electrospray ionization (ESI) and matrix-assisted laser desorption ionization (MALDI) sources in positive mode with DCTB *trans*-2-[3-(4-*tert*-butylphenyl)-2-methyl-2-propenylidene]malononitrile (DCTB) as the matrix. ESI mass spectra were acquired in positive modes as required, using a Micromass time-of-flight mass spectrometer (TOF) interfaced to a Waters 2960 HPLC or a Bruker microTOF-Q III spectrometer interfaced to a Dionex UltiMate 3000 LC. Atmospheric pressure chemical ionization (APCI) experiments were performed on a Bruker microTOF-Q III spectrometer interfaced to a Dionex UltiMate 3000 LC.

Melting points are uncorrected and were measured with a Stuart SP-10 melting point apparatus.

NMR spectra were recorded on a Bruker Advance III 400 MHz, a Bruker Advance HD 400 and an Agilent 400 spectrometer for ¹H (400.13 MHz) and ¹³C (100.61 MHz) NMR spectra. A Bruker Ultrashield 600 spectrometer was employed for ¹H (600.13 MHz) and ¹³C (150.90 MHz) NMR spectra. All NMR experiments were performed at 25 °C. Resonances δ are given in ppm units and referenced to the deuterium peak in the NMR solvents, d_{4} -methanol ($\delta_{H} = 4.87$, 3.31 ppm, $\delta_{C} = 49.1$ ppm). CDCl₃ ($\delta_{H} = 7.26$ ppm, $\delta_{C} = 77.2$ ppm). Signal multiplicities are abbreviated as follows: singlet = s, doublet = d, triplet = t, multiplet = m.

Single crystal X-ray crystallography: Diffraction data for all compounds were collected on a a Bruker D8 Quest ECO or Bruker APEX 2 DUO CCD diffractometer using graphite-monochromated Mo- K_{α} ($\lambda = 0.71073$ Å) or Incoatec IµS Cu- K_{α} ($\lambda = 1.54178$ Å) radiation. Crystals were mounted on a MiTeGen MicroMount and collected at 100(2) K using an Oxford Cryosystems Cobra low-temperature device. Data were collected using omega and phi scans and were corrected for Lorentz and polarization effects using the APEX software suite.^[3] Data were corrected for absorption effects using the multi-scan method (SADABS).^[4]

UV-Vis absorption measurements were recorded in solutions using a Specord 250 spectrophotometer from Quartz Glass 10mm 6030-UV (1 cm path length quartz cell).

IR measurements were done on a PerkinElmer Spectrum 100 FT-IR.

SheInutt's NSD (normal structural decomposition) method was used to delineate, quantify and illustrate the various distortions modes present in the tetrapyrrole macrocycles.^[5] NSD calculations were performed with the NSD online interface, available at https://www.sengegroup.eu/nsd.^[6]

Synthesis and Characterization of Compounds

The general procedure of porphyrin condensation to synthesize 3, 5, 7, 9 and 10.

To a pre-dried 2L round-bottomed flask, anhydrous dichloromethane (500 mL - 1 L) was added and purged with argon. Benzaldehyde **3A**, **5A**, **7A**, **9A or 10A** (0.9 - 1.1 eq.) and 3,4-diethyl-1H-pyrrole **11** (1.00 eq.) were added and the reaction mixture was stirred at room temperature under a slow steady flow of argon. After 15 min, BF₃xEt₂O (0.1 eq.) was added and the reaction flask was shielded from ambient light. After stirring for 16 h DDQ was added and 3 h later triethylamine (0.1 eq.) was added to neutralize BF₃xEt₂O. The solution was concentrated *in vacuo* to yield the crude product mixture, washed with NaOH (1 M), water, and dried over anhydrous magnesium sulfate. The organic extract was concentrated *in vacuo* for further purification by silica gel flash column chromatography.



Scheme S1. Synthetic scheme for the preparation of the target compounds.

2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(2,6-dimethoxyphenyl)porphyrin [3]

Synthesized *via* the General Procedure from 3,4-diethyl-1H-pyrrole **11** (1 mL, 8.12 mmol, 1 eq.), 2,6-dimethoxybenzaldehyde **3A** (1.21 g, 7.31 mmol, 0.9 eq.), BF₃×OEt₂ (0.1 mL, 0.812 mmol, 0.1 eq.) and DDQ (1.84 g, 8.12 mmol, 1 eq.). The reaction mixture was filtered through a silica gel using CH₂Cl₂ to remove the unreacted aldehyde. The compound was then removed from the silica using CH₂Cl₂/MeOH (9:1) and gave compound **3** as a green powder [1.352 g, 1.28 mmol, 70%] upon evaporation of the solvent under reduced pressure. M.p. <149–175 °C; $R_f = 0.60$ (SiO₂, CH₂Cl₂:MeOH = 9:1, v/v); ¹H NMR (600 MHz, CDCl₃, 25 °C): $\delta = 0.28$ (broad s, 24H, CH₂CH₃), 2.23 (broad s, 8H, *CH*₂CH₃), 2.46 (broad s, 8H, *CH*₂CH₃), 3.89 (s, 24H, o-OCH₃), 6.89-6.91 (d, *J* = 8.3 Hz 8H, H_{aryl}), 7.66-7.68 ppm (t, *J* = 8.2 Hz, 4H, H_{aryl}); ¹³C NMR (100 MHz, CDCl₃, 25 °C): $\delta = 160.9$, 143.5, 136.9, 132.6, 116.3, 107.2, 104.3, 56.0, 18.4, 14.3 ppm; UV-vis (CHCl₃ + 1% NEt₃): λ_{max} (log ε) = 463 (4.81), 591 (3.64), 635 (3.72), 692 nm (3.57); IR (ATR): $\tilde{v} = 3206$, 2935, 2833, 2219,



1579, 1472, 1451, 1252, 1107, 1018, 888, 775, 743, 715 cm⁻¹; HRMS (MALDI) m/z calcd. for C₆₈H₇₈N₄O₈ [M+H]⁺: 1079.5892, found 1079.5898.



Figure S1. UV-vis spectrum of 3 in CHCl₃.



Figure S2. ¹H NMR spectrum of 3 (600 MHz, *d*-chloroform, 25 °C).



Figure S3. ¹³C NMR spectrum of 3 (100 MHz, *d*-chloroform, 25 °C).

Minimum: Maximum:		5.0	100.0	-1.5 200.0						
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Form	ula		
1079.5887	1079.5898	-1.1	-1.0	31.5	39.2	0.0	C68	H79	N4	08

Figure S4. HRMS (MALDI) of 3.



Figure S5. FTIR spectrum of 3.

Synthesis and characterization of 5

2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(2,4,6-trimethoxyphenyl)porphyrin [5]

Synthesized via the General Procedure from 3,4-diethyl-1H-pyrrole 11 (1 mL, 8.12 mmol, 1eq.), 2,4,6-trimethoxybenzaldehyde 5A (1.43 g, 7.31 mmol, 0.9 eq.), BF₃×OEt₂ (0.1 mL, 0.812 mmol, 0.1 eq.) and DDQ (1.84 g, 8.12 mmol, 1 eq.). The reaction mixture was filtered through a silica gel using DCM to remove the unreacted aldehyde. The compound was then removed from the silica using DCM/MeOH (9:1) and lead, after evaporation of the solvent under reduce pressure to a green powder, compound 5 [0.217 mg, 0. 181 mmol, 10%]. M.p. = 186 - 199 °C; R_f = 0.58 (SiO₂, CH₂Cl₂:MeOH = 9:1, v/v); ¹H NMR (600 MHz, CDCl₃, 25 °C): δ = 0.44 (broad s, 24H, CH₂CH₃), 2.32 (broad s, 8H, CH₂CH₃), 2.48 (broad s, 8H, CH2CH3), 3.79 (s, 24H, o-OCH3), 4.06 (s, 12H, o-OCH3), 6.47 ppm (d, 8H, H_{arvl}), ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ = 163.69, 161.64, 144.77, 11.09, 106.31, 90.96, 56.08, 55.79, 18.49, 15.30, 14.34 ppm; UV-Vis (CHCl₃): λ_{max} (log ϵ) = 472 (4.84), 594 (3.62), 643 (3.73), 703 nm (3.64); IR (ATR): $\tilde{v} = 2933, 2830, 2202, 1599, 1578, 1453, 1412, 1331, 1224, 1204, 1154,$ 1123, 1123, 1056, 1033, 951, 812 cm-1; HRMS (MALDI) m/z calcd. for $C_{40}H_{38}N_4O_6$ [M+H]⁺: 1199.6368, found 1199.6321.





Figure S6. UV-vis spectrum of 5 in chloroform.







Figure S9. ¹³C NMR spectrum of 5 (100 MHz, *d*-chloroform, 25 °C).

Minimum: Maximum:		5.0	20.0	-1.5 200.0						
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm) Formu	ıla		
1199.6368	1199.6321	4.7	3.9	31.5	55.4	0.0	C72	H87	N4	012

Figure S10. HRMS (MALDI) of 5.



Figure S11. FTIR spectrum of 5.

2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(2,6-dimethoxy-4-hydroxyphenyl)porphyrin [6]

In a predried 50 mL round bottomed flask, 2,3,7,8,12,13,17,18-octaethyl-5,10,15,20tetrakis(2,4,6-trimethoxyphenyl)porphyrin (5) (17.2 mg, 14.34 µmol, 1.0 eq.) was dissolved in dry CHCl₃ (4 mL). Under an argon atmosphere the mixture was treated dropwise with BBr₃ (1M in CH₂Cl₂, 0.1 mL, 100 µmol, 7 eq). After 2 h of stirring the reaction at 25 °C, the solvent was removed under vacuum. The reaction mixture was dissolved in DCM/MeOH (9:1) and filtered through a silica gel using DCM/MeOH (9:1) to remove the impurities. The compound was then collected using MeOH, after evaporation of solvent under reduced pressure provided compound 6 as dark green powder [14.1 mg, 12.33 µmol,, 86%]; M.p.: >300 °C; R_f = 0.2 (SiO₂, MeOH); ¹H NMR (400 MHz, *d*-methanol, 25 °C) δ= 6.36 (s, 8H, *m*phenyl-H), 3.76 (s, J = 13.5 Hz, 24H, -OCH₃), 2.84 - 2.52 (m, 16H, -CH₂), 0.65 (t, J = 7.4 Hz, 24H, -CH₂CH₃).¹³C NMR (100 MHz, *d*-methanol, 25 °C): δ = 161.79, 160.48, 146.06, 138.17, 108.63, 107.36, 94.87, 55.19, 18.28, 14.71 ppm; UV/Vis (methanol): λ_{max} [nm] (log ε [L·mol⁻¹ cm⁻¹]) = 488 (4.97), 695 (4.11); IR (ATR) \tilde{v} = 3369.4 (br), 3194.8 (br), 2970,







Figure S12. UV-vis spectrum of 6 in methanol.







Figure S15. ¹³C NMR spectrum of 6 (100 MHz, *d*-methanol, 25 °C).



Figure S16. FTIR spectrum of 6.



Figure S17. Mass spectrum of 6, HRMS (MALDI) m/z calcd. for C₆₈H₇₉N₄O₁₂ [M+H]⁺: 1143.5689, found 1143.5625, recorded by Bruker Daltonics autoFlex.

Synthesis and characterization of 7

2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(3,5-dimethoxyphenyl)porphyrin [7]

Synthesized via the General Procedure from 3,5-dimethoxybenzaldehyde 7A (1.95 g, 11.7 mmol, 1.11 eq.), 3,4-diethyl-1H-pyrrole 11 (1.3 g, 10.55 mmol, 1.00 eq.), BF₃×Et₂O (0.01 mL, 0.8 mmol) and DDQ (2.76 g, 12.18 mmol). After aqueous work up, the organic extract was transferred directly to the column (Al₂O₃). Less polar side products and starting material were eluted first (CH₂Cl₂), followed by the collection of the major green band (CH₂Cl₂:MeOH = 50:1 to 150:1 v/v). Removal of solvent *in vacuo* yielded the product **3** as a dark purple coloured solid [0.545 g, 0.505 mmol, 19%]; M.p.: 260 °C (decomposition); $R_f = 0.25$ (SiO₂, Ethyl acetate); ¹H NMR (400 MHz, CDCl₃, 25 °C) δ= 7.71 (s, 8H, o-phenyl-H), 6.95 (s, 4H, p-phenyl-H), 4.07 (s, 24H, - OCH_3), 2.62 – 2.08 (m, 16H, -CH₂), 0.30 ppm (t, J = 7.3 Hz, 24H, -CH₃); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3, 25 \text{ °C})$: $\delta = 160.26, 143.90, 138.39, 118.74, 115.32, 102.56, 56.25, 56.25$ 18.55, 15.83 ppm; UV/Vis (chloroform): λ_{max} [nm] (log ε [L·mol⁻¹ ·cm⁻¹]) = 454.9 (5.28), 552.3 (4.11), 604.4 (3.78), 638.6 (3.62), 697.7 (3.48); HRMS (MALDI) m/z calcd. for $C_{68}H_{79}N_4O_8$ [M+H]⁺: 1079.5909, found 1079.5898. IR (ATR) $\tilde{v} = 2971.7, 2933.58,$ 2875.7, 2838.29, 1665.29, 1586.79 (s), 1452.86, 1421.62, 1355.18, 1314.41, 1290.24, 1199.57, 1153.91, 1132.14, 1059.16, 1021.31, 948.20, 835.80, 800.92, 717.10, 687.61 cm⁻¹





Figure S18. UV-vis spectrum of 7 in chloroform.



Figure S19. ¹H NMR spectrum of 7 (400 MHz, *d*-chloroform, 25 °C).



100- ₃		1079.	9.5909									
850.486	2 971.5754	1051.5646	1081.6038	1195.5005	1313.6516	1441.6283	1526.6720	1570.6	774 16	08.6536	1699.7222	m/7
0	900	1000	1100	1200	1300	1400	1500		1600		1700	- IIVZ
Minimum: Maximum:		5.0	50.0	-1.5 200.0								
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT	(Norm)	Formu	ıla			
1079.5909	1079.5898	1.1	1.0	31.5	50.6	0.0		C68	H79	N4 (8	

Figure S21. HRMS (MALDI) 7.



2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(3,5-dihydroxyphenyl)porphyrin [8]

In a predried 100 mL round bottomed flask, 2,3,7,8,12,13,17,18-octaethyl-5,10,15,20-tetrakis(3,5-dimethoxyphenyl)porphyrin **7** (550 mg, 0.506 mmol, 1.0 eq.) was dissolved in dry CH₂Cl₂ (60 mL). Under an argon atmosphere at 0 °C the mixture was treated dropwise with BBr₃ (1M in CH₂Cl₂, 10.5 mL, 10.5 mmol, 16 eq). The reaction flask was warmed to room temperature and after 18 h of stirring, the mixture was added dropwise to ice water (50 mL). NaOH (1 M, 100 mL) was slowly added and washed with dichloromethane (2 x 50 mL). The aqueous phase was acidified with conc. HCl until the product started to coagulate. The green solid was filtered and washed with water and dichloromethane. The green solid was dissolved with methanol and after evaporation of solvent under reduced pressure provided a dark green crystalline solid **8**. [519 mg, 0.499 mol, 76%]; M.p.: >300 °C; *R*_f = 0.63 (SiO₂, MeOH); ¹H NMR (400 MHz, d-methanol, 25 °C) δ = 7.40 (s, 8H, *o*-phenyl-*H*), 6.83 (s, 4H, *p*-phenyl-*H*), 2.67–2.42 (m, 16H, -*CH*₂), 0.52 ppm (t, *J* = 7.4 Hz, 24H, -CH₂*CH*₃); ¹³C NMR (100 MHz, d-methanol, 25 °C): δ = 158.41, 143.60, 139.10, 138.95, 118.60, 115.86, 104.08, 18.19, 14.96 ppm; UV/Vis (Methanol): λ_{max} [nm] (log ε [L·mol⁻¹ cm⁻¹])



= 475.9 (5.36), 689.2 (4.45); HRMS (MALDI) *m/z* calcd. for C₆₀H₆₃N₄O₈ [M+H]⁺: 967.4617, found 967.4646. IR (ATR) \tilde{v} = 3148.18 (br), 2972.31, 2933.66, 2873, 1587.88 (s), 1502.13, 1439.13, 1354.45, 1292.03, 1199.71, 1152.15 (s), 1054.38, 1003.66, 994.74, 980.09, 951.42, 886.88, 847.24, 809.12, 777.76, 685.77 cm⁻¹







Figure S24. ¹H NMR spectrum of 8 (400 MHz, CD₃OD, 25 °C).



100 684.3	96 4 ⁶⁸ 751 4489 887 470	969.4730	1111.3020	155	1570.6774 2.6740 1572.68	⁷⁶ 1715.7222.1	1861.6941		. m/z
600	800	1000	1200	1400	1600	1800	2000	2200	2400
Minimum: Maximum:		5.0	10.0	-1.5 200.0					
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (N	Norm) Formula		
967.4617	967.4646	-2.9	-3.0	31.5	38.2	0.0	C60 H63	3 N4 O	28

Figure S26. HRMS (MALDI) of 8.



Figure S27. FTIR spectum of 8.

Synthesis and characterization of 9A

3,5-Dipivaloyloxybenzaldehyde [9A]

In a 250 mL RBF at room temperature, 3,5-dihydroxybenzaldehyde (0.5 g, 3.62 mmol, 1.0 eq.) was dissolved in acetonitrile (100 mL). *N*,*N*-diisopropylethylamine (5 mL, 28.96 mmol, 8.0 eq) was added. The reaction was treated dropwise with pivaloyl chloride (1.75 mL, mol, eq) and stirred for 1 h. The crude mixture was extracted with dichloromethane, washed with NaHCO₃ (1 × 150 mL), brine (1 × 150 mL) and water (1 × 150 mL). The organic phase was dried with anhydrous magnesium sulfate, filtered and evaporated *in vacuo* to yield a white crystalline solid **9A**. [1.069 g, 3.49 mmol, 96%]; M.p.: 146.5 °C; $R_f = 0.89$ (SiO₂, EtOAc/*n*-hexane 1:1 v/v); ¹H NMR (400 MHz, CDCl₃, 25 °C) $\delta = 9.99$ (s, 1H, CO*H*), 7.51 (d, $J_m = 2.17$ Hz, 2H, *o*-phenyl-*H*), 7.17 (t, $J_m = 2.1$ Hz, 1H, *p*-phenyl-*H*), 1.39 ppm (s, 18H, *-CH*₃); ¹³C NMR (100 MHz, CDCl₃, 25 °C): $\delta = 190.29$, 176.47, 152.09, 138.00, 121.50, 119.79, 39.22, 27.06, 26.53 ppm; HRMS



(APCI) m/z calcd. for C₁₇H₂₃O₅ [M]⁺: 307.1535, found 307.1540; IR (ATR) $\tilde{v} = 2974.91$, 2937.33, 2874.84, 1752.35 (CO stretch), 1697.34, 1592.1, 1480.72, 1396.93, 1368.58, 1300.07, 1269.26, 1091.23 (s, br), 1030.07 (sh), 980.16, 941.15, 905.88, 803.73, 803.41, 757.89, 732.27, 672.11, 597.75, 552.37 cm⁻¹.



Figure S28. ¹H NMR spectrum of 9A (400 MHz, *d*-chloroform, 25 °C).





Figure S30. HRMS (APCI) of 9A.



Figure S31. FTIR spectrum of 9A.

Synthesis and characterization of 9

2,3,7,8,12,13,17,18-Octaethyl-5,10,15,20-tetrakis(3,5-dipivaloyloxyphenyl)porphyrin [9]

A: In a pre-dried 50 mL Schleck flask precursor porphyrin **8** (46 mg, 0.048 mmol, 1 eq.) was stirred in THF (8 mL) with pivaloyl chloride (0.53 mL, 3 mmol, 64 eq.) in the presence of *N*,*N*-diisopropylamine (0.374 mL, 3 mmol, 64 eq.) for 5 h. Aqueous work up with saturated NaHCO₃ (2 x 50 mL) and water (50 mL) followed by evaporation of solvent *in vacuo* yielded product **9** as green crystalline solid [52 mg, 0.032 mmol, 67%]. Introduction of eight nonpolar *tert*-butyl groups was easily monitored by TLC due to the large difference in R_f values between the highly polar starting material and product.

B: Synthesized *via* the General Procedure using 3,5-dipivaloyloxybenzaldehyde **9A** (1.069 g, 3.49 mmol, 1.0 eq.), 3,4-diethylpyrrole **11** (481 mg, 3.90 mmol, 1.1 eq.), BF₃×Et₂O (0.05 mL, 4.46 mmol) and DDQ (1.2 g, 5.29 mmol) in CH₂Cl₂ (500 mL). After aqueous work up, the residue was transferred directly to a silica gel column and non-polar impurities eluted with CH₂Cl₂. The crude mixture was acidified with 1% TFA and product was collected *via* silica gel column (CH₂Cl₂/EtOAc 4:1 v/v). Aqueous work up with NaOH (1 M, 2 x 150 mL) and water (150 mL) followed by evaporation of solvent *in vacuo* yielded the neutralized



product as green crystalline solid. This was recrystallized by slow diffusion in DMSO/acetonitrile to yield dark green crystals of **9** [0.809 g, 0.493 mmol, 57%]; M.p.: 276 °C (decomposition); $R_f = 0.72$ (SiO₂, *n*-hexane/EtOAc 1:1 v/v); ¹H NMR (600 MHz, CDCl₃, 25 °C) $\delta = 8.17$ (s, 8H, *o*-phenyl-*H*), 7.38 (s, 4H, *p*-phenyl-*H*), 2.56–2.30 (m, 16H, -*CH*₂), 1.51 (s, 72H, *t*-Bu), 0.42 ppm (t, *J*=7.4 Hz, 24H, -CH₂C*H*₃); ¹³C NMR (100 MHz, CDCl₃, 25 °C): $\delta = 176.77$, 151.21, 144.00, 139.30, 138.71, 127.20, 116.79, 116.52, 39.33, 27.24, 18.71, 15.73 ppm; UV/Vis (chloroform): λ_{max} [nm] (log *ɛ*[L·mol⁻¹ · cm⁻¹]) = 456.3 (5.17), 553.4 (4.10), 600.4 (3.48), 631.0 (3.62), 702.1 (3.62); HRMS (MALDI) *m/z* calcd. for C₁₀₀H₁₂₇N₄O₁₆ [M+H]⁺: 1639.9247, found 1639.9216; IR (ATR) $\tilde{v} = 2970.96$, 2934.29, 2873.32, 1754.12 (s, CO stretch) 1605.2, 1587.04, 1479.79, 1458.96, 1428.24, 1397.00, 1367.91 1268.48 1163.57, 1090.14 (s, br), 1028.82 (sh), 978.72, 945.95, 904.07, 862.88, 822.38, 798.92, 688.04, 617.29, 559.51 cm⁻¹.



Figure S32. UV-vis spectrum of 9 in chloroform.



Figure S33. ¹H NMR spectrum of 9 (600 MHz, *d*-chloroform, 25 °C).



Figure S34. ¹³C NMR spectrum of 9 (101 MHz, *d*-chloroform, 25 °C).



Figure S35. HRMS (MALDI) of 9.



Figure S36. FTIR spectrum of 9.

Synthesis and characterization of 10A

5-Pivaloyloxy-2-nitrobenzaldehyde [10A]

In a 250 mL RBF at room temperature, 5-hydroxy-2-nitrobenzaldehyde (0.500 g, 2.99 mmol, 1.0 eq.) was dissolved in acetonitrile (120 mL). *N*,*N*-Diisopropylethylamine (5 mL, 28.96 mmol, 8.0 eq) was added. The reaction was treated dropwise with pivaloyl chloride (1.46 mL, 11.98 mmol, 4 eq.) and stirred for 1h. The crude mixture was extracted with dichloromethane, washed with NaHCO₃ (1 x 150 mL), brine (1 x 150 mL) and water (1 x 150 mL). The organic phase was dried with anhydrous magnesium sulfate, filtered and concentrated *in vacuo* to yield a crystalline solid. Purification by silica gel column (CH₂Cl₂) collected a pale yellow fraction, which was dried to afford the product **10A** as a white crystalline solid [0.359 g, 1.43 mmol, 75 %]; M.p.: 92 - 96 °C; *R*t = 0.64 (SiO₂, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃,



25 °C) δ= 10.47 (s, 1H, CO*H*), 8.22 (d, J_0 = 8.9 Hz, 1H, *m*-phenyl-*H*), 7.65 (d, J_m = 2.4 Hz, 1H, *o*-phenyl-*H*), 7.49 (dd, J_0 = 9.0 Hz, J_m = 2.8 Hz, 1H, *p*-phenyl-*H*), 1.40 ppm (s, 9H, *t*-Bu); ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ = 187.30, 173.91, 155.38, 146.25, 133.23, 126.50, 122.63, 39.40, 26.98 ppm; HRMS (APCI) *m/z* calcd. for C₁₂H₁₂NO₅ [M]⁻: 250.0721, found 250.0718; IR (ATR) \tilde{v} = 2973.75, 2873.12, 1757.43 (s, C=O stretch.), 1610.98, 1578.45, 1525.08 (s, NO asym. stretch.), 1465.06, 1397.46, 1340.72 (s, NO sym. stretch), 1260.34, 1182.77, 1093.08 (s, C-O stretch.), 1023.02 (sh, C-O stretch.), 951.08, 900.54, 850.25, 751.93, 696.63, 652.16, 622.27, 574.91, 539.58 cm⁻¹.



Figure S37. ¹H NMR spectrum of **10A** with expansion of 8.4 -7.4 ppm (400 MHz, *d*-chloroform, 25 °C).





Figure S39. HRMS (APCI) of 10A.



Figure S40. FTIR spectrum of 10A.

2,3,7,8,12,13,17,18-Octaethylporphyrin-5,10,15,20-tetrakis(5-pivaloyloxy-2-nitrophenyl) [10]

Synthesized via the General Procedure from 5-pivaloyloxy-2-nitrobenzaldehyde 10A (0.934 g; 3.72 mmol; 1.0 eq.), 3,4-diethylpyrrole 9 (0.504 g; 4.09 mmol; 1.1 eq.), BF₃×Et₂O (0.075 mL, 0.66 mmol) and DDQ (1.2 g, 5.29 mmol). The crude reaction mixture was guenched with 10% triethylamine. After aqueous work up, the concentrated organic extract was purified by silica gel column (CH₂Cl₂/EtOAc 10:1 v/v) and eluted (CH₂Cl₂/TEA 10:1 v/v) as a dark brown/green fraction. The residue was transferred to a 250 mL RBF and dissolved in chloroform (100 mL). N,N-Diisopropylethylamine (1.39 mL, 7.97 mmol, 8.0 eq.) was added and the reaction was treated dropwise with pivaloyl chloride (0.5 mL, 3.98 mmol, 4 eq.) and stirred for 1 h. The crude mixture was quenched with a few drops of NaOH, followed by



extraction with dichloromethane, and washed with NaHCO3 (1 × 150 mL) and water (1 × 150 mL). The organic phase was dried with anhydrous magnesium sulfate, filtered and dried under reduced pressure. Purification by a silica gel column removed black impurities (CH₂Cl₂/acetone 10:1 v/v) and the desired intermediate was collected (CH₂Cl₂/TEA 10:1 v/v) as a brown coloured product 10 [0.558 g, 3.93 mmol, 20%] HRMS (MALDI) *m*/z calcd. for C₈₀H₉₁N₈O₁₆ [M+H]⁺: 1419.6553, found 1419.6549.

100 % 1412.6 0 1412. 1412.	318 1415.6384 5 1415.0	1417.6414 1418.64 1417.5	419.6549 184 1420.0	1420.6588 1421.6638 1422.66 	⁹⁶ 1424.6744 777777777777777777777777777777777	1427.6542	<u>1429.6549</u> 1430.0	1430.63	00 14	34.66	²¹ 1435.6177 1435.0	
Minimum: Maximum:		5.0	50.0	-1.5 400.0								
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT	(Norm)	Formu	la			
1419.6549	1419.6553	-0.4	-0.3	39.5	59.2	0.0		C80 1	Н91	N8	016	
Figuro S/1		I DI) of 10										

Figure S41. HRMS (MALDI) of 10.

5,10,15,20-Tetrakis(5-pivaloyloxy-2-nitrophenyl)-2,3,7,8,12,13,17,18-octaethylporphyrinato]nickel(II) [12]

In a 250 mL RBF, the atropisomeric mixture of 10 (0.335 g, 0.236 mmol,1.0 eq.) was dissolved in toluene (100 mL) at room temperature. Nickel(II) acetylacetonate (0.303 g, 1.180 mmol, 5.0 eq.) was added and the reaction was heated to reflux at 111 °C. After 2 h, the mixture was cooled and solvent was evaporated *in vacuo*. The mixture was dissolved in a small volume of dichloromethane and transferred to a silica gel column (CH₂Cl₂). Three green fractions (CH₂Cl₂) were collected and dried under reduced pressure to yield three products as dichromatic (green/purple) crystalline solids:

 $\alpha,\beta,\alpha,\beta$ -12 [87.2 mg, 0.059 mmol, 25%]; M.p.: >300°C; $R_{\rm f}$ = 0.82 (SiO₂, CH₂Cl₂);

¹H NMR (600 MHz, CDCl₃, 25 °C): δ = 8.35 (d, J_0 = 8.7 Hz, 4H, *m*-phenyl-*H*), 7.70 (d, J_m = 2.6 Hz, 4H, *o*-phenyl-*H*), 7.57 (dd, J_0 = 8.8 Hz, J_m = 2.2 Hz, 4H, *p*-phenyl-*H*), 2.40 (br, 16H, -CH₂), 1.45 (s, 36H, *t*-Bu), 0.62 (br, 24H, -CH₂CH₃) ppm; ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ = 176.45, 153.18, 148.90, 144.03, 136.41, 130.28, 125.60, 123.10, 111.40, 39.40, 27.09, 19.55 ppm; UV/Vis (chloroform): λ_{max} [nm] (log ε [L·mol⁻¹ ·cm⁻¹]) = 442.6 (5.00), 572.2 (4.07), 611.3 (4.04); HRMS (MALDI) *m*/z calcd. for C₈₀H₈₈N₈NiO₁₆ [M]⁺: 1474.5672, found 1474.5699; IR (ATR) \tilde{v} = 2972.50, 2934.84, 2873.17, 1757.33 (s, C=O stretch.), 1610.59, 1578.45, 1523.98 (s, -NO asym. stretch.), 1466.13 (m, CH₂ bend), 1397.58, 1342.75 (s, -NO sym. stretch.), 1261.73, 1184.14, 1081.52 (s, C-O stretch.), 1023.05 (sh, C-O stretch.), 950.37, 900.54, 853.95, 828.03, 801.43, 752.00, 699.37, 651.67, 617.94 cm⁻¹;

 $\alpha_2\beta_2$ -**12** [50.7 mg, 0.034 mmol, 15%]; M.p.: 279-280°C (decomposition); $R_f = 0.56$ (SiO₂, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃, 25 °C): $\delta = 8.35$ (d, $J_0 = 8.7$ Hz, 4H, *m*-phenyl-*H*), 7.90 (br, 4H, *o*-phenyl-*H*), 7.57 (dd, $J_0 = 8.9$ Hz, $J_m = 2.0$ Hz, 4H, *p*-phenyl-*H*), 2.40 (br, 16H, -CH₂), 1.46 (s, 36H, *t*-Bu), 0.63 (br, 24H, -CH₂CH₃) ppm; ¹³C NMR (100 MHz, CDCl₃, 25 °C): $\delta = 176.37$, 152.89, 149.31, 144.80, 144.18, 136.32, 129.92, 125.79, 122.95, 39.40, 27.11, 19.52; UV/Vis (chloroform): λ_{max} [nm] (log ε [L·mol⁻¹·cm⁻¹]) = 442.1 (4.97), 573.1 (4.05), 609.4 (4.01); HRMS (MALDI) *m/z* calcd. for C₈₀H₈₈N₈NiO₁₆ [M]⁺: 1474.5672, found 1474.5707; IR (ATR) $\tilde{v} = 2973.48$, 2934.36, 2873.37, 1758.47 (s, C=O stretch.), 1611.04, 1578.83, 1525.46 (s, -NO asym. stretch), 1465.10 (m, CH₂- bend.), 1397.75, 1341.15 (s, -NO sym. stretch.), 1260.08, 1182.72, 1092.89 (s, C-O stretch.), 1022.92 (sh, C-O stretch.), 951.10, 900.74, 851.23, 826.04, 800.37, 752.30, 697.90, 652.48, 620.27, 578.09 cm⁻¹;

 $\alpha_3\beta$ -12: [76.6 mg, 0.052 mmol, 22%]; M.p.: 272-276°C (decomposition); $R_f = 0.44$ (SiO₂, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃, 25 °C): δ = 8.44 – 8.32 (m, 4H, *m*-phenyl-*H*), 8.10 (s, 1H, *o*-phenyl-*H*), 8.04 – 7.68 (br, 2H, *o*-phenyl-*H*), 7.61 (s, 1H, *o*-phenyl-*H*), 7.60 – 7.53 (m, 4H, *p*-phenyl-*H*), 2.41 (br, 16H, -C*H*₂), 1.45 (s, 36H, *t*-Bu), 0.63 (br, 24H, -CH₂C*H*₃) ppm; ¹³C NMR (100 MHz, CDCl₃, 25 °C): δ = 176.36, 153.32, 152.86, 152.62, 149.59, 148.48, 143.92, 136.48, 136.30, 136.12, 130.70, 129.75, 129.57, 126.08, 125.90, 125.52, 123.12, 122.94, 122.80, 111.76, 111.27, 39.39, 27.10, 19.53; UV/Vis (chloroform): λ_{max} [nm] (log *ε*[L·mol⁻¹·cm⁻¹]) = 442.7 (5.01), 571.8 (4.10), 610.8 (4.06); HRMS (MALDI) *m/z* calcd. for C₈₀H₈₈N₈NiO₁₆ [M]⁺: 1474.5672, found 1474.5724; IR (ATR) $\tilde{\nu}$ = 2973.74, 2934.01, 2873.13, 1757.65 (s, C=O stretch.), 1610.96, 1578.41, 1525.07 (s, -NO asym. stretch), 1465.21 (m, C*H*₂ bend.), 1397.54, 1340.27 (s, -NO sym. stretch.), 1260.09, 1182.28, 1092.81 (s, C-O stretch.), 1022.79 (sh, C-O stretch.), 951.19, 900.55, 850.52, 826.47, 800.62, 752.16, 697.14, 651.97, 620.27, 575.07 cm⁻¹.



Figure S42. UV-vis spectrum of $\alpha_1\beta_1, \alpha_2\beta_2$ -12 and $\alpha_3\beta_1$ -12 in chloroform.





Figure S43. ¹H NMR spectrum of $\alpha,\beta,\alpha,\beta$ -12 with expansion of 8.5 – 7.4 ppm (600 MHz, *d*-chloroform, 25 °C).



Figure S44. ¹H NMR spectrum of $\alpha_2\beta_2$ -12 with expansion of 8.5 – 7.4 ppm (600 MHz, *d*-chloroform, 25 °C).



Figure S45. ¹H NMR spectrum of $\alpha_{3}\beta$ -12 with expansion of 8.5 – 7.5 ppm (600 MHz, *d*-chloroform, 25 °C).



Figure S46. ¹³C NMR spectrum of $\alpha,\beta,\alpha,\beta$ -**12** (101 MHz, *d*-chloroform, 25 °C).



Figure S47. ¹³C NMR spectrum of $\alpha_2\beta_2$ -12 (101 MHz, *d*-chloroform, 25 °C).



Figure S49. FTIR spectrum of α , β , α , β -12, α ₂ β ₂-12 and α ₃ β -12.



Figure S51. ¹³C–¹H HMBC spectrum with expansion in aromatic region of $\alpha,\beta,\alpha,\beta$ -**12** (*d*-chloroform, 25 °C).



Figure S53. ¹³C–¹H HMBC spectrum with expansion in aromatic region of $\alpha_2\beta_2$ -**12** (*d*-chloroform, 25 °C).



Figure S55. ¹³C–¹H HMBC spectrum with expansion in aromatic region of $\alpha_3\beta$ -12 (*d*-chloroform, 25 °C).





Figure S57. HRMS (MALDI) of $\alpha_2\beta_2$ -12.



Figure S58. HRMS (MALDI) of $\alpha_{3}\beta$ -12.

Results and Discussion

Structural Determination of Isolated Structures

Crystals were grown following the protocol developed by Hope, liquid-liquid diffusion in CHCl₃ and methanol or oversaturated solutions in DMSO.^[7] Using Olex2, the structure was solved with the XT structure solution program, using the intrinsic phasing solution method and refined against $|F^2|$ with XL using least squares minimization.^[8] The C and N bound H atoms were placed in their expected calculated positions and refined as riding model: N–H = 0.88 Å, C–H = 0.95–0.98 Å, with U_{iso} (H) = 1.5 U_{eq} (C) for methyl H atoms and 1.2 U_{eq} (C, N) for all other atoms other H atoms. Details of data refinements can be found in Table S1. All images were prepared by using Olex2.^[8a]

In the structure of **9A** one t-Bu acetate group modelled as disordered over two positions with almost equal occupancy (53/47%) using restraints (SADI and ISOR) and constraints (EADP/EXYZ for C17C17a and EADP C19/C19a).

In the structure of **9** a pivaloyl group at O114 was modelled over two positions using SIMU restraint in a 72:28% occupancy. Two of the trifluoroacetate molecules was modelled over two positions using rigid models in a 50:50% occupancies.

In the structure of **8** the phenyl moiety at C10_2 was modelled over two positions using restraints (SADI, SIMU, ISOR, AFIX 66) in a 54:46 % occupancy. The ethyl groups at C18_2, C7_2, C12_1 were modelled over two positions using restraints (SADI, SIMU) in 58:42 %, 54:46 %, 75:25 % occupancies respectively. DMSO molecules were modelled using rigid models, moreover, units at S26S, S7S and S1S were modelled over two positions in 25:75, 40:60, 40:60 % occupancies. In the structure there were solvent accessible voids that contained large amounts of solvent molecules, however, due to high disorder these could not be modelled reliably and were omitted using OLEX2 maps.

In the structure of $\alpha_{3}\beta$ -12 one ethyl group at C18 was disordered and modelled in two locations (36:54% occupied) with restraints (SADI, SIMU). Three pivaloate groups at C10, C15, C20 disordered and modelled in two locations (57:33%; 26:74%; 85:15% correspondingly) with restraints (SADI, SIMU). The solvent molecules in the lattice void were modelled with rigid groups and consist of DMSO

In the structure of $\alpha_2\beta_2$ -**12** one ethyl group at C2 was disordered and modelled in two locations (51:49% occupied) with restraints (SADI, SIMU). One pyrrole ring carbon and ethyl group at C13 was disordered and modelled in two locations (64:36% occupied) with restraints (SADI, SIMU). One pivaloate group at C15 disordered and modelled in two locations (52:48%) with restraints (SADI, SIMU). The solvents in the lattice void were modelled with rigid groups and consist of DMSO and H2O.

The structure $\alpha, \beta, \alpha, \beta$ -**12** was refined as a 2-component twin. In the first unit, two pivaloate groups at C15, C20 disordered and modelled in two locations (57:43%; 60:40% correspondingly) with restraints (SADI, SIMU). Two nitro groups at C5, C15 disordered and modelled in two locations (61:49%; 47:53% correspondingly) with restraints (SADI, SIMU). In the second unit, two pivaloate groups at C10B, C15B disordered and modelled in two locations (52:48%; 56:44% correspondingly) with restraints (SADI, SIMU). The solvent molecules in the lattice void were modelled with rigid groups and consist of chloroform and dichloromethane.



Figure S59. Molecular structures of isolated and analyzed compounds by X-ray crystallography. Hydrogen atoms and solvent molecules omitted for clarity; thermal ellipsoids give 50% probability.

Compound	α,β,α,β -12	α ₂ β2- 12	α ₃ β- 12	8	9	10A	9A
Internal code	KN016b	KN016	KN015	KN013	KN012	KN011	TCD1366
CCDC #	2058266	2058269	2058267	2058270	2058268	2058264	2058265
Empirical formula	$C_{162}H_{179}CI_5N_{16}Ni_2O_{32}$	$C_{86.56}H_{108.21}N_8NiO_{19.79}S_{3.29}$	$C_{85}H_{103}N_8NiO_{18.5}S_{2.5}$	$C_{134}H_{172}CI_4N_8O_{24}S_7$	$C_{56}H_{67}D_6F_3N_3O_{11}S$	$C_{12}H_{13}NO_5$	$C_{17}H_{22}O_5$
Formula weight	3156.87	1741.38	1671.61	2645.01	1059.27	251.23	306.34
Temperature/K	100(2)	100(2)	100(2)	100(2)	100(2)	100(2)	100(2)
Crystal system	Triclinic	Triclinic	Triclinic	Triclinic	Monoclinic	Triclinic	Monoclinic
Space group	ΡĪ	PĪ	ΡĪ	ΡĪ	C2/c	ΡĪ	C2/c
a/Å	14.6355(6)	15.936(3)	11.5285(3)	19.4431(8)	35.1721(10)	5.960(9)	18.2037(7)
b/Å	23.0234(10)	18.284(4)	12.6909(3)	19.5835(7)	15.7125(5)	8.693(9)	11.3258(4)
c/Å	23.9896(9)	19.337(4)	30.6892(9)	27.6688(11)	21.2195(6)	12.089(13)	17.6188(6)
α/°	89.203(3)	102.927(5)	84.5201(18)	97.5447(19)	90	80.713(18)	90
β/°	83.210(3)	105.949(7)	82.3776(18)	99.898(2)	98.0799(15)	80.22(4)	106.1224(15)
γ/°	79.715(3)	113.742(5)	77.4770(17)	117.2468(17)	90	75.41(4)	90
Volume/ų	7897.6(6)	4586.9(17)	4334.1(2)	8957.7(6)	11610.4(6)	592.7(13)	3489.6(2)
Ζ	2	2	2	2	8	2	8
D _{calc} g/cm ³	1.328	1.261	1.281	0.981	1.212	1.408	1.166
µ/mm⁻¹	1.719	0.357	1.472	1.799	1.062	0.111	0.702
F(000)	3320.0	1845.0	1770.0	2808.0	4488.0	264.0	1312
Crystal size/mm ³	0.52 × 0.05 × 0.05	0.242 × 0.232 × 0.062	0.15 × 0.06 × 0.02	0.35 × 0.11 × 0.09	0.25 × 0.17 × 0.13	0.28×0.15×0.03	0.2x0.2x0.09
Radiation	CuKα	ΜοΚα	CuKα	CuKα	CuKα	ΜοΚα	ΜοΚα
Wavelength/Å	1.54178	0.71073	1.54178	1.54178	1.54178	0.71073	0.71073
2 0 /°	3.71 to 139.574	5.262 to 53.138	5.824 to 139.928	3.338 to 134.998	5.076 to 141.472	5.614 to 55.132	4.651 to 68.294
Reflections collected	28881	75488	57091	103813	69660	8726	11132
Independent reflections	28881	18997	16155	31757	10915	2736	3192
Rint	merged	0.1702	0.0595	0.0510	0.0527	0.0445	0.0347
Rsigma	0.2389	0.1623	0.0640	0.0511	0.0337	0.0490	0.0327
Restraints	773	769	798	352	177	0	53
Parameters	2274	1372	1298	1865	763	166	248
GooF	1.022	1.018	1.162	1.084	1.128	1.033	1.053
R1 [l> 2σ (l)]	0.1316	0.1007	0.1158	0.0667	0.0684	0.0461	0.0380
wR ₂ [l> 2σ (l)]	0.3383	0.2502	0.3121	0.1958	0.1946	0.0979	0.1084
R1 [all data]	0.2334	0.2254	0.1561	0.0837	0.0816	0.0809	0.0412
wR ₂ [all data]	0.4267	0.3283	0.3495	0.2099	0.2077	0.1130	0.1118
Largest peak/e Å ⁻³	1.08	1.46	1.15	0.80	0.69	0.27	0.203
Deepest hole/e Å-3	-1.28	-0.75	-0.58	-0.75	-0.76	-0.27	-0.171

Table S1: Details of XRD data refinement of α , β , α , β -12, α ₂ β ₂-12, α ₃ β -12, 8, 9, 10A, 9A.

Demethylation and Substitution attempts

Elemental Composition Report N Single Mass Analysis Tolerance = 20.0 PPM / DBE: min = -1.5, max = 200.0 Element prediction: Off Number of isotope peaks used for i-FIT = 5 Monoisotopic Mass, Odd and Even Electron lons 19 formula(e) evaluated with 1 results within limits (up to 10 best isotopic matches for each mass) Elements Used: Chemical Formula: C75H90N8O3 C: 0-75 H: 0-91 N: 0-8 O: 0-3 Exact Mass: 1150.7136 Karolis Norvaisa (MSe), KN71B_crude Q-TOF20180409MF003 55 (1.185) AM (Cen,6, 80.00, Ht,10000.0,1570.68,0.70); Sm (SG, 2x3.00); Sb (15,10.00); Cm (8:97-(53:56+78:80)) TOF MS LD+ 1.43e+003 1151,7218 100-3550.6346 1153.7302 1570.6774 % 751,4539 1001.6010_1067.6660 1572.6871 1715.7446 1552.6721, 0 <u>----</u> m/z 800 1600 600 1000 1400 1800 1200 2000 2200 2400 Minimum: -1.5 200.0 5.0 20.0 Maximum: mDa PPM DBE i-FIT i-FIT (Norm) Formula Mass Calc. Mass 1151.7218 1151.7214 0.3 57.7 0.0 C75 H91 N8 0.4 34.5 03

Figure S60. HRMS (MALDI) of the reaction mixture in attempt to produce 2



% 75	51.4460 823.4778	1123.6885	1236.7832	15 7678	570.6774 1573.6843_1	716.7583			m/7
600	800	1000	1200	1400	1600	1800 2	000	2200	2400
Minimum: Maximum:		5.0	5.0	-1.5 200.0					
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula		
1235.7780	1235.7789	-0.9	-0.7	35.5	87.4	0.0	C80 H9	9 N8	04

Figure S61. HRMS (MALDI) of the reaction mixture in second attempt to produce 2

Entry	Temperature	Time	Solvent	Demethylating	Eq.	Number of methyl
				agent		groups cleaved*
1	20 °C	18 h	DCM	BBr ₃	24	4
2	50 °C	10	CHCl ₃	TMSI	40	0
2		min		TMCI	40	0
5	(Microwave)	min		TWO	40	0
4	50 °C (Microwave)	30 min	CHCl₃	TMSI	40	0
5	50 °C	24 h	CHCl ₃	TMSI	40	0
6	20 °C	18 h	DCM	BBr ₃	76	6, 5, 4
7	20 °C	48 h	DCM	BBr ₃	76	6, 5, 4 + side prod.
8	20 °C	96 h	DCM	BBr ₃	76	7, 6, 5 + side prod.
9	20 °C	8 d	DCM	BBr ₃	76	Majority of side prod.
10	20 °C	18 h	o-Dichlorobenzene	BBr ₃	16	4, 5
11	110 °C	2 h	o-Dichlorobenzene	BBr ₃	16	6
12	110 °C	6.5 h	o-Dichlorobenzene	BBr ₃	16	8 (trace), 7, 6
13	110 °C	10	o-Dichlorobenzene	BBr ₃	25	4
1/		30		BBr.	25	654
14	(Microwave)	min	0-Dicitioroberizerie	0013	20	0, 3, 4
15	110 °C	60	o-Dichlorobenzene	BBr ₃	25	6
	(Microwave)	min				
16	110 °C	3.5 h	o-Dichlorobenzene	BBr ₃	75	6
17	20 °C	18h	DCM	BBr₃	43	5, 4
	20 °C	48h	DCM	BBr ₃	43	7, 6

 Table S2. Demethylation attempts of nonplanar porphyrin 3.

* Indicated by mass spectrometry

Stability of Individual Atropisomers



Figure S62. Stability evaluation of $\alpha,\beta,\alpha,\beta$ -12, $\alpha_2\beta_2$ -12, and $\alpha_3\beta$ -12, by ¹H NMR recorded in CDCl₃. Top: over a period of 22h at 25 °C and 18h at 50°C (note, no spectroscopic changes was observed at room 25 °C). Bottom: enlargement of the aromatic region signals of the samples a) $\alpha,\beta,\alpha,\beta$ -12, b) $\alpha_2\beta_2$ -12, and c) $\alpha_3\beta$ -12 recorded after 18h at 50°C. In blue, the percentage value of the integrated characteristic signal corresponding to the domain atropisomer. In red, the predominant observable signal corresponding to the equilibrated atropisomer. Note, due to the very broad profile of $\alpha_2\beta_2$ -12 characteristic signal, the tracing the exact atropisomeric composition can be unattainable. Highlighted with star symbol is unidentified signal that could potentially correspond to the formation of α_4 -12.

References

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