Structure of 3-mercaptopropionic acid dioxygenase with a substrate analog reveals bidentate substrate binding at the iron center

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Running title: *Mode of substrate binding in a thiol dioxygenase*

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SUPPLEMENTAL TABLES

Wilson *B* factor (\AA^2)

Data collection and processing¹ Crystal 3-HPA complex (crystal form A) Thiocyanate complex (crystal form B) X-ray source NSLS-II (FMX 17-ID-2) APS (NE-CAT 24-ID-E) Wavelength (Å) 0.979339 0.979180 **Space group** *P*3₁ *P*₃²² Unit cell lengths (\AA) $a = 178.22$, $c = 75.92$ $a = 102.18$, $c = 301.93$ Resolution $(\hat{A})^{\dagger}$ 50 – 2.25 (2.39 – 2.25) 50 – 2.95 (3.13 – 2.95) Unique reflections 127,973 (20,673) 20,505 (3,185) Multiplicity 12.9 (13.4) 12.9 (13.4)

Table S1. X-ray diffraction data collection and structure refinement statistics

Completeness (%) 99.9 (99.8) 99.8 (99.2)

<*I*/σ*I*> 6.35 (0.78) 12.9 (0.98)

*R*_{merge}*I* (%) 15.0 (164.2) 18.5 (288.3)

 $CC_{1/2}$ (%) 99.3 (48.4) 99.3 (48.4) 99.9 (46.8)

) $|55|$ 86

Refinement

1 Each data set was collected from a single crystal

† Values in parentheses are for the highest resolution shell of data

‡ Values in parentheses are the number of reflections used for cross-validation

[∀]Twin fraction was refined in REFMAC based on the observed and calculated amplitudes

* Evaluated using Molprobity

Table S2. Comparison of *Av*3MDO bond distances with other thiolate-bound non-heme iron complexes. Distances are shown for the iron to bound thiolate (Fe-S), either bidentate bound substrate carboxylate, amine, or equivalent (Fe-O or Fe-Namine), and average iron to histidine or equivalent nitrogen. All units are in Ångströms.

	Fe-S	Fe-O	$\rm Fe\text{-}N_{\rm amine}$	$\rm Fe\text{-}N_{his\;ave}$	ref
$AvMDO-3HPA complexa$	2.16^{b}	2.17		2.16	This work
Fe ^{II} structures					
$AvMDO 3HPA1$ -bound ^c	2.26^{b}	2.25		2.15	This work
$AvMDO 3HPA2$ -bound ^c	1.91^{b}	2.55^{d}		2.17	This work
$AvMDO$ 3MPA-bound ^c	2.33	2.51^{d}		2.19	This work
Rn CDO cysteine-bound ^a	2.35		2.35	2.17	(1)
$[Fe^{II}(2-MTS)(Ph2TIP)]BPh4a$	2.32	2.12		2.16	(2)
$[Fe^{II}(CysOEt)(Ph2TIP)]BPh4a$	2.31		2.26	2.17	(3)
$[Fe^{II}(Me3TACN)(abt)(OTf)]a$	2.44		2.23	2.25	(4)
$[Fe^{II}(Me3TACN)(abtCF3)(OTf)]a$	2.43		2.24	2.24	$\left(4\right)$
$[Fe^{II}(iPr3TACN)(abt)](OTf)a$	2.36		2.26	2.20	(4)
$[Fe^{II}(iPr3TACN)(abtCF3)](OTf)a$	2.38		2.27	2.20	(4)
Fe ^{III} structures					
$Av3MDO 3HPA1$ -bound ^c	2.23^{b}	1.97		2.13	This work
$Av3MDO3HPA2$ -bound ^c	1.87^{b}	2.14		2.18	This work
$Av3MDO3MPA-boundc$	2.32	2.08		2.20	This work
$[Fe^{III}(2-MTS)(Ph2TIP)]-CNc$	2.20	1.96		2.17	(2)
$[Fe^{II}(CysOEt)(Ph2TIP)]-CNc$	2.18		2.05	2.17	(2)
(L-Cys/CN)-CDO cross-linked ^c	2.28		2.09	2.07	(5)
$(L-Cys/CN)$ -CDO non-cross-linked ^c	2.26		2.11	2.05	(5)

^a Distances obtained from crystal structure. *^b*3-oxide of **3HPA** instead of sulfur. *^c* Distances obtained from computational methods. ^{*d*} To some extent, the longer Fe-O_(carboxylate) distance of 2.51 - 2.55 Å can be attributed to the is likely attributed to electrostatic interaction of the anionic carboxylate group with adjacent Arg168. However, a larger issue is the simultaneously accommodation of three negative charges [Cl- and **3HPA²** (or **3MPA²**)] directly coordinated to the Fe-site resulting in a monoanionic active site complex, $([Fe^{II}(His)_{3}Cl^{2}(X^{2})]^{1}$, $X = 3HPA^{2}$ or $3MPA^{2}$). This suggests that chloride is an unfavorable ligand for the catalytically relevant **3MPA**-bound Av3MDO Fe(II)-site.

Table S3. Geometric properties of the DFT optimized (**3MPA**/NO)-Fe(II) *Av*3MDO models. Three structures were optimized differing in the hydrogen bond donation of the Tyr159 hydroxyl group.

^a *X* refers to the hydrogen binding partner of the Tyr157 hydroxyl group, being either the nitrogen or oxygen of nitric oxide or the δ-nitrogen of His157.

Table S4. Calculated vs experimental Mössbauer parameters for wild-type **3MPA**-bound MDO with added nitric oxide (percent error shown in parentheses). Calculations are based on bidentate bound **3MPA** with nitrosyl group in the axial position *trans* to His90.

Table S5. The dipolar coupling in MHz and polar angles in degrees for each ¹H included in HYSCORE simulations. Shown below are the polar angles that relate a 1 H on **3MPA** to magnetic axis system. The angle φ represents rotation in x-y plane, whereas θ is the deviation from the z-axis defined by the Fe-NO bond.

SUPPLEMENTAL FIGURES

Figure S1. *Av*3MDO active site difference density modeling. In all panels the blue mesh represents sigma A-weighted $2F_0-F_c$ density for chain B of the structure contoured at 1 RMSD and the green and red mesh represent NCS-averaged, sigma A-weighted F_0 - F_c density contoured at $+8$ and -8 RMSD respectively. **A**. Map before modeling the open coordination sites. **B**. Updated maps following refinement with three aquo ligands modeled. The three aquo model poorly accounts for the difference map feature. **C**. Updated maps following refinement with two aquo ligands and one bicarbonate (**BCT**) modeled. **D**. Updated maps following refinement with one aquo ligand, one chloride (Cl⁻) and bicarbonate modeled. Chloride fully accounts for the difference density at the axial coordination site. Note the unacceptably close contact between bicarbonate and the modeled solvent and the residual strong positive density between these two moieties. E. Updated maps following refinement with one chloride (Cl⁻) and 3-hydroxypropionic acid (**3HPA**) modeled. This model fully accounts for the active site density. **F**. Updated maps following refinement with one chloride (Cl⁻) and 3-mercaptopropionic acid (3MPA) modeled. Note the difference map hole at the S atom confirming the presence of a lower *Z* atom (e.g. oxygen) at that site.

Figure S2. Mass spectrometry detection of **3HPA** in sodium polyacrylate 5100 solution used for *Av*3MDO crystallization. **A.** Negative ion-mode LC-MS of 3HPA standard with selective ion monitoring (SIM) of $m/z = 89$ species corresponding to deprotonated 3HPA. **B**. MS¹ spectrum of the SIM m/z 89 peak. C) MS² spectrum showing fragmentation of the m/z 89 precursor ion to form $m/z = 59$ (-H₂CO) and 43 (-HCOOH) species. **D**. LC-MS of the polyacrylate solution extract carried out in this same manner as (**A**) showing a single SIM m/z 89 peak with a retention time and fragmentation pattern (panels **E** and **F**) matching those of the 3HPA standard. No SIM m/z 89 peak was observed in control samples with acrylate extract or 3HPA standard omitted. By comparison of the SIM m/z 89 peak areas the concentration of 3HPA in the polyacrylate extract was estimated to be ~20 mM.

Figure S3. Inhibition of *Av*3MDO catalyzed oxidation of **3MPA** with increasing NaCl concentration. Assays were performed at fixed and saturated **3MPA** concentration (1 mM) in a buffered 20 mM HEPES pH 8 solution at fixed temperature (25 ± 1 °C). Salt concentrations were varied from 0 to 240 mM.

Figure S4. *Crystal structure of Av3MDO in complex with thiocyanate (green) and its comparison to the 3HPA-bound Av3MDO structure (grey).* **A**. Active site comparison between the two structure showing conformational differences in Gln63, Phe79, and Ile98. The green mesh represents unbiased, NCSaveraged, sigma A-weighted Fo-Fc electron density in the axial coordination position supporting the modeling of thiocyanate at this site. **B**. The tunnel providing access to the active site remains occluded in the thiocyanate-bound *Av*3MDO structure with the gatekeeper residues Tyr61, Pro88, and Phe180 found in nearly identical positions as compared to the **3HPA**-bound structure.

Figure S5. Selected viewpoints (**A**-**C**) of the *Av3MDO*-**3HPA** complex ([**1**], code 6XB9, *shown in white*) overlaid on DFT optimized structures for the **3HPA**-bound Fe-site. Structures for the deprotonated ([**2**], **3HPA2**- , *blue carbon atoms*) and protonated ([**3**], **3HPA1-** , *yellow carbon atoms*) inhibitor alcohol are presented for comparison. The top-down view shown in panel **A** aligns all structures along the [His90-Fe-Cl] axis. Panel **B** illustrates a 90° rotation of **A** along the perpendicular axis (*dashed line*). Clockwise rotation (-90°) of **B** around the[His90-Fe-Cl] axis results in Panel **C**. Selected distances, angles, and RMSD values are presented in **Table 2** for comparison.

Orientation of Y159 hydrogen bond - Multiple pH-dependent kinetic studies verify that both Tyr159 and His157 of *Av*3MDO are protonated in the catalytically active state (6-8). The crystal structures for the *Av*3MDO-**3HPA** complex presents three possible proton-acceptors for Tyr159. The Nδ-atom of His157 (2.86 Å), the chloride bound to the axial Fe- site (2.85 Å), and the proximal **3HPA**-carboxylate O-atom (3.01 Å). Alternatively, the directionality of the proton relay network could be reversed by N_{δ} -His157 donation of an H-bond to the Tyr159 phenol O-atom. This leaves Tyr159 free to donate a hydrogen bond to either the axial chloride or the **3HPA**-carboxylate.

Figure S6.

Figure S6. Comparison of **CYS**-bound rat CDO (**A**, PDB 4IEV) and **3HPA**-bound *Av*3MDO (**B**, PDB 6XB9) structures highlighting the extended proton relay network for each enzyme. **C**. Energy surface for Tyr159 H-bond donation/acceptor interactions as a function of dihedral bond angle. Given the ambiguous oxidation state of the Fe-site in the SXB9 structure, relaxed surface scan calculations were performed for both ferrous (*blue*) and ferric (*black*) states.

The energy surface for potential Tyr159 H-bond acceptors was evaluated using a relaxed surface scan (9- 11) of the *Av*3MDO-**3HPA** complex. Calculations included Fe-coordinated histidine residues (His90, His92, and His142), the 'SHY' motif (Tyr159, His157, and Ser155), Arg168, **3HPA-1** , and chlorine with coordinates taken from the *Av*3MDO crystal structure. All hydrogens were optimized while constraining the crystal structure coordinates. The Tyr159 phenol hydrogen was varied in orientation by varying the dihedral angle defined by the Tyr159 atoms Cε-Cξ-Oη-H. Where Cε denotes the Tyrosine ε-carbon in closest proximity to H157. The path of the relaxed surface scan placed the phenol hydrogen oriented towards the Nδ-atom of His157, axial iron-bound chlorine, and **3HPA**-carboxyl group while keeping the crystal structure rigid. This calculation was done for both Fe(II) and Fe(III) oxidation states to evaluate its influence (if any) on the preferred H-bond orientation. It should be noted that this model does not account for rotation of the tyrosine aromatic ring or other geometric perturbations and thus the values should be considered qualitative.

As illustrated above **(Figure S6C**, *black*), for the ferric state, an activation barrier of 2.8 kcal/mol is calculated for the Tyr159 \rightarrow His157 configuration from Tyr159 \rightarrow Cl. Given the small energy barrier and overall energy difference of >1 kcal/mol, it is reasonable that Tyr159 could alternate between donating a hydrogen bond to both His157 and the more preferred iron-bound chlorine. By contrast, the proximal Oatom of the **3HPA**-carboxylate is a poor H-bond acceptor for Tyr159 and is unfavorable in this model. The ferrous state (**Fig**. **S6C**, *blue*) calculations show a more dramatic bias towards Tyr159 → Cl donation. The energy difference between donation towards Cl or His157 is approximately 5 kcal/mol. Qualitatively the activation barrier is also approximately 5 kcal/mol as the Tyr159 \rightarrow His157 orientation is not in a local minimum. Based on these observations, it can be argued that Tyr159 donates an H-bond to the iron-bound Cl-atom in the *Av*3MDO-**3HPA** complex. Potentially, substitution of the axial chloride ligand for NO could alter the orientation of the Tyr159 H-bond to favor donation to H157. However, this would be inconsistent with previous EPR studies which demonstrate attenuated NO-binding for the Y159F variant (and to a lesser extent H157N). Moreover, as both chlorine and nitrosyl ligands have an equivalent formal charge (-1), an argument can be made that H-bond donation from Tyr159 would similarly favor axially bound NO over His157.

Figure S7

Figure S7. HYSCORE spectra and simulations of the 1 H on **3MPA** (*shown in magenta*) at 174, 176, 180, and 200 mT. Experimental spectra appear as color contours; simulations are overlaid as black contour lines. The anisotropic hyperfine couplings and the positions of each nucleus relative to the magnetic axis system are listed in **Table S5**.

Figure S8. HYSCORE spectra and simulations of the ¹H on His142 (*shown in magenta*) at 174, 176, 180, and 200 mT. Experimental spectra appear as color contours; simulations are overlaid as black contour lines. The anisotropic hyperfine couplings and the positions of each nucleus relative to the magnetic axis system are listed in **Table S5**.

Figure S9. HYSCORE spectra and simulations of the ¹H on His92 (*shown in magenta*) at 174, 176, 180, and 200 mT Experimental spectra appear as color contours; simulations are overlaid as black contour lines. The anisotropic hyperfine couplings and the positions of each nucleus relative to the magnetic axis system are listed in **Table S5**.

Figure S10

Figure S10. HYSCORE spectra and simulations of the ¹H on His90 (*shown in magenta*) at 174, 176, 180, and 200 mT. Experimental spectra appear as color contours; simulations are overlaid as black contour lines. The anisotropic hyperfine couplings and the positions of each nucleus relative to the magnetic axis system are listed in **Table S5**.

Figure S11

Figure S11. HYSCORE simulations of the closest ¹H on 3MPA at 176 mT. Color gradient represents the and θ in increments of 15 ° (center and right, respectively).

Figure S12

Figure S12. HYSCORE spectra (at 175 mT) for (**3MPA**/NO)-bound *Av*3MDO iron-nitrosyl prepared in ¹H₂O (*left*) buffer as compared to samples exchanged into ²H₂O buffer (*right*). For clarity, the HYSCORE data is focused entirely on the proton region of the spectra for comparison to ${}^{2}H_{2}O$ -data. The absence of ${}^{1}H$ peaks lost upon exchange into deuterated buffer suggests no solvent-derived ligands are coordinated to the Fe-site. The expanded HYSCORE spectra of the 1 H2O data at 175 mT (*left*) is presented in **Fig**. **8B**.

Figure S13

Figure S13. HYSCORE spectra and simulations of the 1 H on the hydroxyl group of Tyr159 (*shown in magenta*) at 174, 176, 180, and 200 mT. Experimental spectra appear as color contours; simulations are overlaid as black contour lines. The simulations were calculated with an anisotropic hyperfine coupling of 0.95 MHz and values of 40° and 20° for θ and φ , respectively.

CARTESIAN COORDINATES (Å) FOR SELECTED MODELS

3MPA-bound Fe(III) MDO: **Figure 6A-B (***white***)**

3MPA-bound Fe(II) MDO: **Figure 6C-D (***white***)**

3HPA-1 -bound Fe(III) MDO: **Figure S4 [3]**

(**3MPA**/NO)-bound *Av*3MDO: **Figure 7**

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