

Supplementary Figure Legends

Supplementary Figure 1. High frequencies of circulating B cells in patients with CLL. Frequencies of T and B cells in patients with CLL and HDs (A) (patients with CLL n=12, HDs n=9). Representative flow cytometric analysis of CD5+ B cells in patients with CLL, identified in the lymphocyte and blasts cells gates (B).

Supplementary Figure 2. *In vitro* expanded CD19 CAR T cells from patient with CLL kill specifically CD19+ cells. (A) Frequencies of CD3+ T cells and CD19+ B cells three days after CD19 CAR retroviral transduction of a patient with CLL cells (top panel), and in untransduced control cells (bottom panel). (B) CD19 CAR T cells produced from a patient with CLL were incubated for 24hrs with either CD19+ K562 cells (top panel), or NGFR+ K562 cells (bottom panel) before analysis.

Supplementary Figure 3. CD4/CD8 ratio of *in vitro* expanded CD19 CAR+ and CAR- T cells. CD4/CD8 ratio in CD19 CAR+ and CD19 CAR- T cells in patients with CLL (n=9) and healthy donors (n=9). Wilcoxon test.

Supplementary Figure 4. *In vitro* expanded CD19 CAR- T cells phenotype. CD45RA and CCR7 expression in (A) patients with CLL (n=9), and (B) in HDs (n=9). Squares represent CD8+ CD19 CAR- T cells, and triangles CD4+ CD19 CAR- T cells. Wilcoxon test, * P < 0.05. ** P < 0.004.

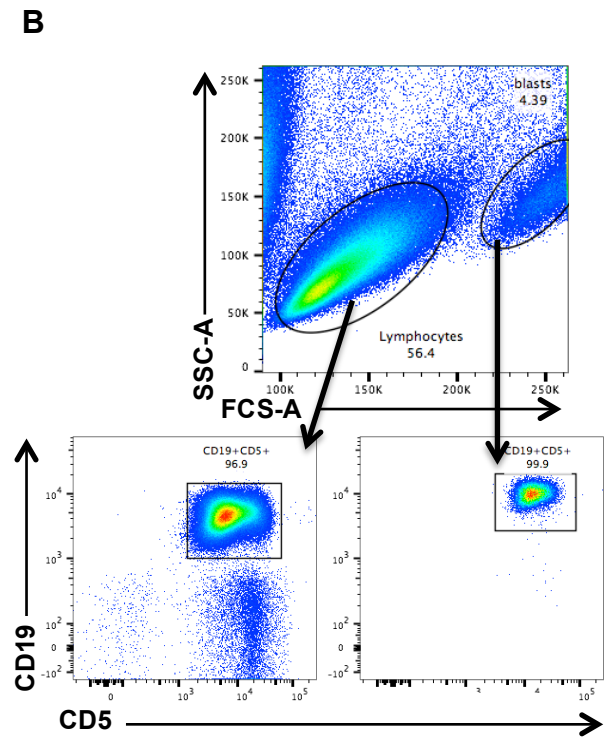
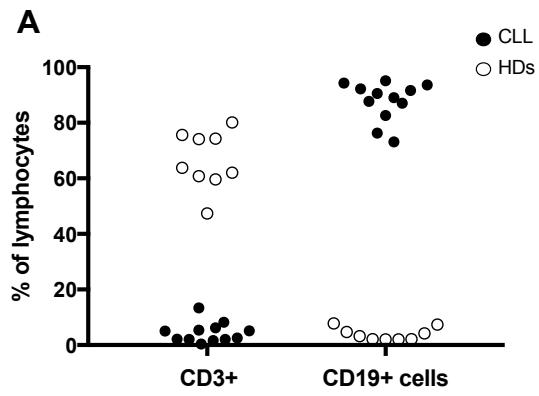
Supplementary Figure 5. Cytokine production in response to NGFR+ K562 cells, and after PMA/Ionomycin stimulation of *in vitro* expanded CD19 CAR transduced T cells. Frequencies within (A) CD8+ and (B) CD4+ T cells after stimulation with NGFR+ K562 cells in patients with CLL (n=9) and HDs (n=7), and after PMA/Ionomycin stimulation within (C) CD8+ T cells and (D) CD4+ T cells in

patients with CLL (n=7) and HDs (n=5). Black circles represent patients with CLL, and open circles HDs. Mann-Whitney test, * P < 0.04. ** P < 0.006.

Supplementary Figure 6. CD19+ K562 cells express high levels of cell surface CD19. Overlay of CD19 cell surface expression from CD19+ K562 cells, CLL cells and HD (EBV-transformed) B cells using a FITC-conjugated CD19 antibody (A), from CLL cells (n=4) and HD (EBV-transformed) B cells (n=3) using an APC-conjugated CD19 antibody (B).

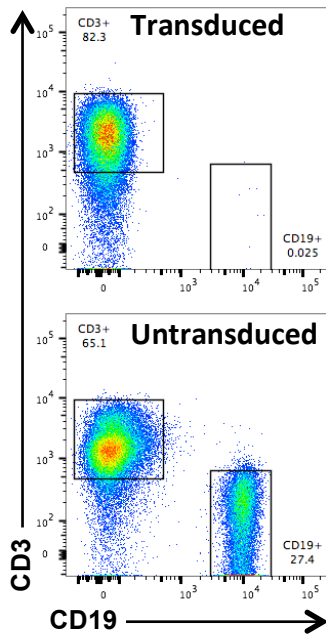
Supplementary Figure 7. CD19+ K562 and autologous B cells induce a different effector profile on *in vitro* expanded and stimulated CD19 CAR T cells. The different combinations of CD107a expression and IFN- γ /IL-2/TNF production median values within activated (displaying at least one of the function measured) CD8+ and CD4+ CD19 CAR T cells in response to either CD19+ K562 cells or autologous B cells (A and B) in patients with CLL (n=9), and (C and D) in HDs (n=7-8). Mann-Whitney test, * P < 0.05. ** P < 0.008.

Supplementary Figure 1.

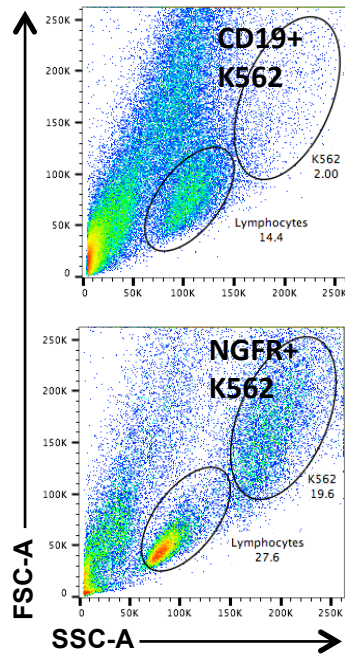


Supplementary Figure 2.

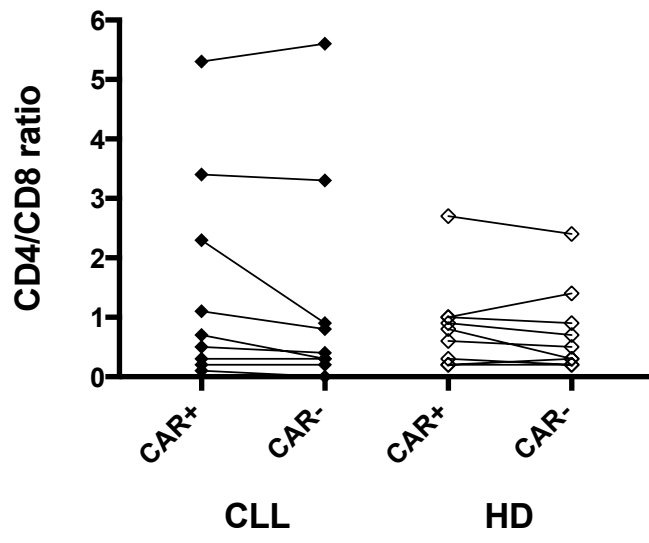
A



B

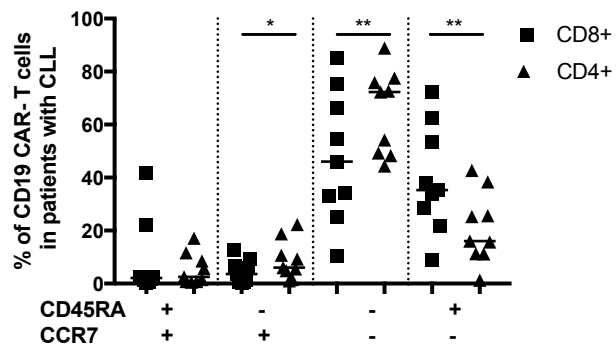


Supplementary Figure 3.

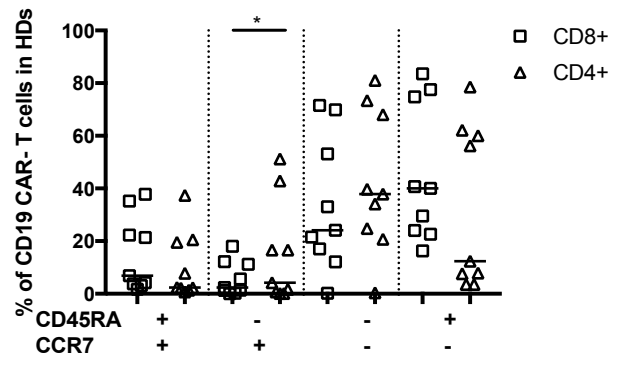


Supplementary Figure 4.

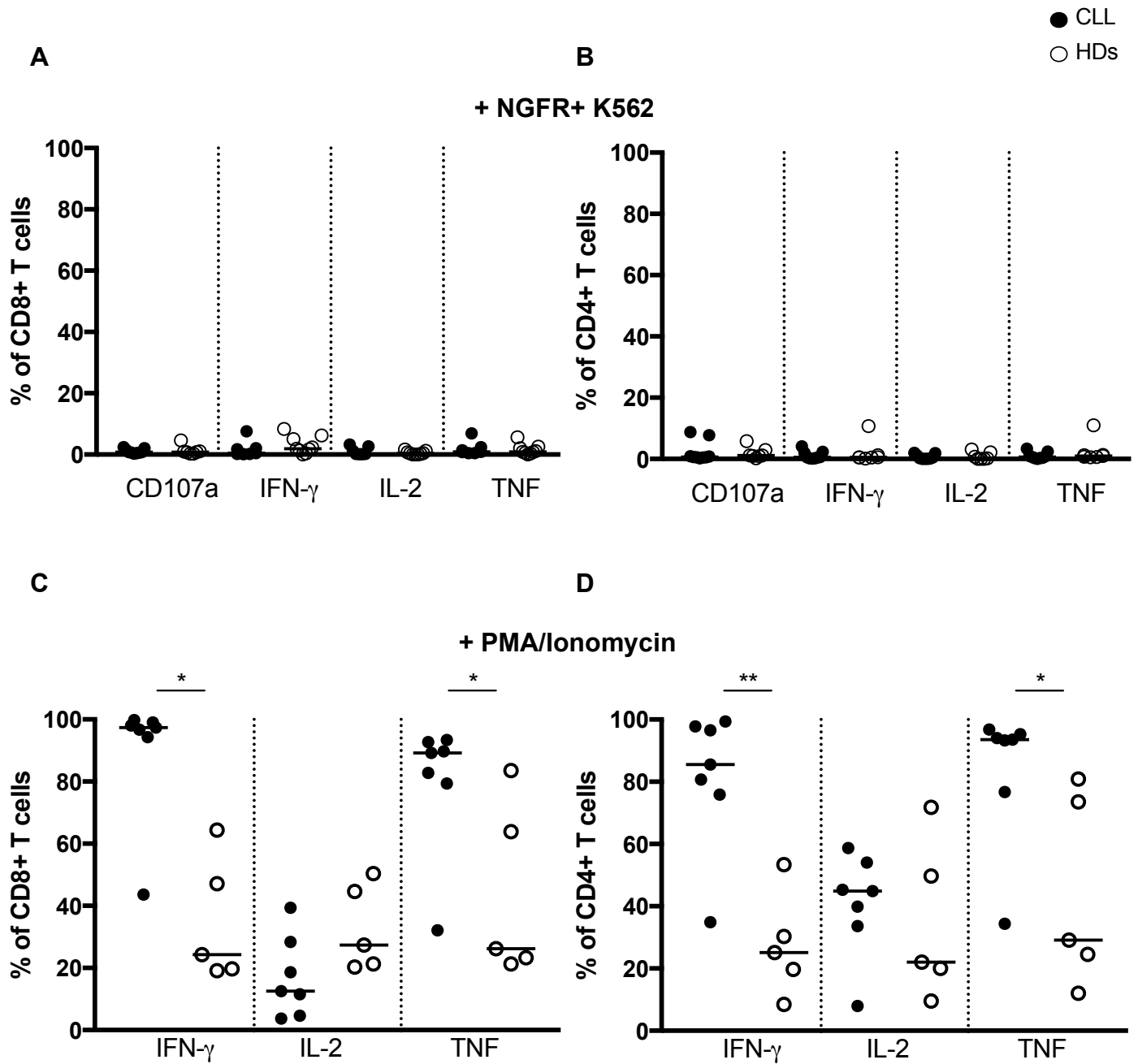
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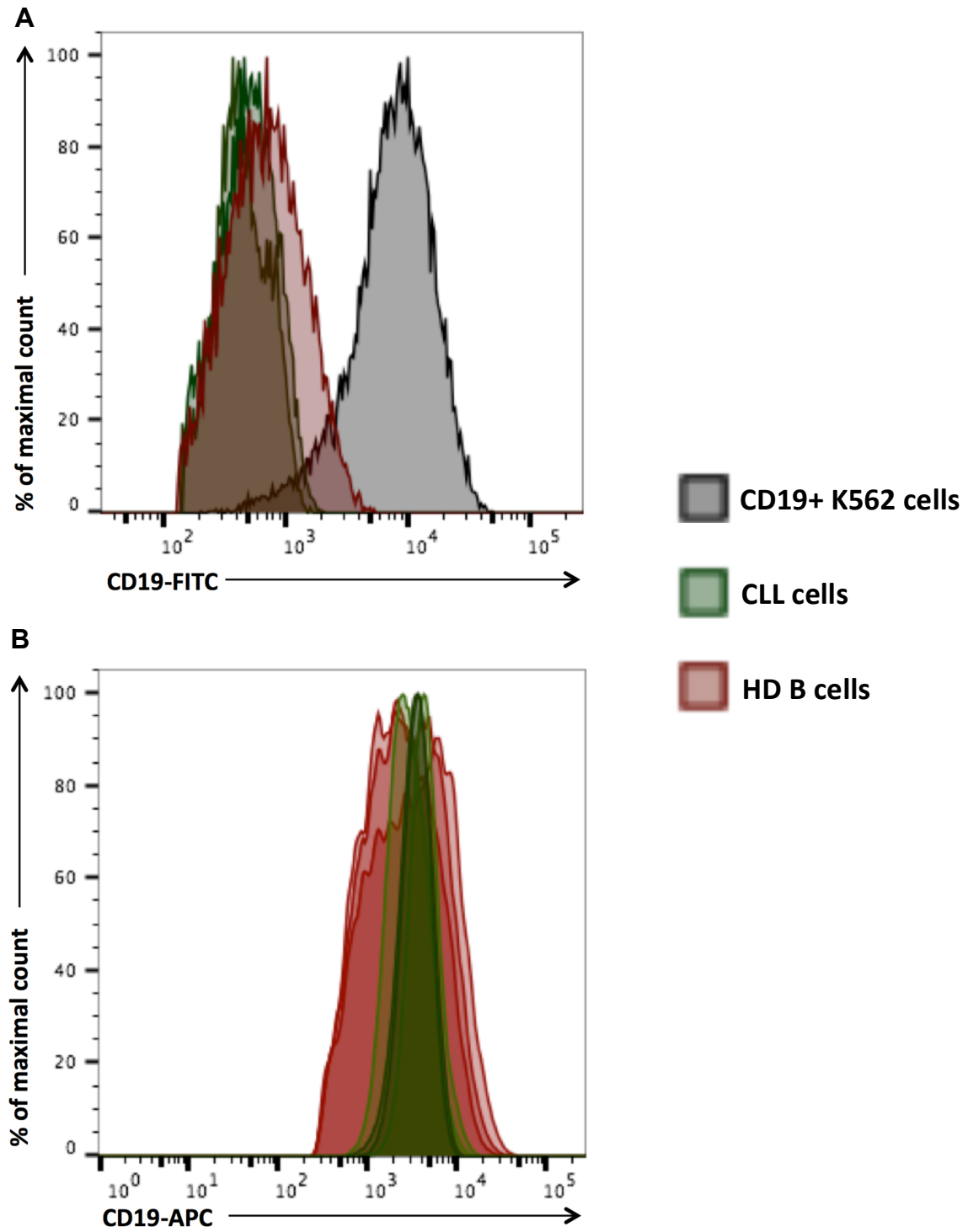
B



Supplementary Figure 5.



Supplementary Figure 6.



Supplementary Table 1. Characteristics of patients with CLL

Patient	WBC (10⁹/L)	Lymphocytes (10⁹/L)	Rai stage *	FISH	Clinical phase**	Previous therapy	Ongoing therapy	Response to last therapy
#01	27.2	23	-	13q-	Stable PR after therapy	Yes, FC (2010)	No	PR
#02	170	167	3	ND	Progressive disease	No	No	-
#03	56	51.2	4	11q-	Progressive disease	Yes, FC (2008)	No	PR
#04	23	19.3	1	ND	Indolent		No	-
#05	287.6	279.2	3	Normal	Progressive disease	No	No	-
#06	106.9	105.9	3	ND	Indolent	No	No	-
#07	281.3	280.5	4	13q-	Progressive disease	No	No	-
#08	102.9	97.8	3	13q-	Progressive disease	No	No	-
#09	62.7	60.5	0	ND	Indolent	No	No	-
#10	161.2	160.7	4	13q-	Progressive disease	No	No	-
#11	121.8	118	1	17q-	Progressive disease	No	No	-
#12	157.5	154.2	2	13q-	Indolent	No	No	-

WBC= whole blood count, PR= partial response, FC= fludarabine cyclophosphamide, ND= not determined, (-)= not applicable. * For untreated patients only. ** At time of blood collection.