

Supporting Information for *Adv. Sci.*, DOI: 10.1002/advs.202004044 A Cyclodextrin-hosted Ir(III) Complex for Ratiometric Mapping of Tumor Hypoxia *in vivo*

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Supporting Information

A Cyclodextrin-hosted Ir(III) Complex for Ratiometric Mapping of Tumor Hypoxia in vivo

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Synthesis of NaGdF₄:Yb,Tm,Ca@NaGdF₄ nanoparticles. Yb,Tm-doped NaGdF₄ nanoparticles were prepared according to literature method. Typically, GdCl₃·6H₂O (0.70 mmol), YbCl₃·6H₂O (0.18 mmol), TmCl₃·6H₂O (0.02 mmol), and CaCl₂ (0.01mmol) were mixed with 4 mL of OA and 16 mL of ODE in a 100 mL flask. After being heated to 150 °C to form a homogeneous solution under nitrogen protection, the solution was cooled down to 50°C and 10 mL of a methanol solution containing NaOH (1.25 mmol) and NH₄F (2 mmol) was added dropwise. The reaction system was then kept under stirring at 50°C for 30 min. Subsequently, methanol in the system was removed under vacuum at 100°C for 10 min, and the resulting reaction mixture was quickly heated to 300 °C under atmospheric pressure. The reaction system was not protection and then terminated by cooling the reaction mixture to room temperature. The resultant nanoparticles were precipitated by ethanol, collected by centrifugation, washed with ethanol for three times, and finally redispersed in cyclohexane for further experiments.

The following growth of the NaGdF₄ shell was carried out by similar procedures for the preparation of NaGdF₄: Yb,Tm core particles. Briefly, 9 mL of cyclohexane solution of the purified NaGdF₄: Yb,Tm,Ca core nanocrystals was mixed with GdCl₃·6H₂O (0.50 mmol), 4 mL of OA, and 16 mL of ODE in a 100 mL flask. The growth of the NaGdF₄ shell and the following purification procedures for the core@shell particles were the same as those for the core nanocrystals. The purified nanoparticles were also redispersed in cyclohexane for further experiments.

Ligand Approximately 10 of exchange. mg the purified particles (NaGdF₄:Yb,Tm@NaGdF₄) and 100 mg of COOH-PEG-dp were dissolved in 5 mL of THF, and the mixture was kept under stirring overnight at 40°C. After that, the PEGylated particles were precipitated by cyclohexane, washed with cyclohexane for three times, and finally dried under vacuum at room temperature. The obtained nanoparticles were further purified through ultrafiltration with 30 kDa MWCO centrifugal filter (Millipore YM-50) for 4 cycles at 6000 g to remove the free ligand. The nanoparticles finally obtained were characterized by TEM and a representative TEM image is shown in Figure S1.

Synthesis of Ir-BTPHSA-NH₂. Ir-BTPHSA (5.1 mg, 0.007 mmol) and EDC·HCl (2.0 mg, 0.010 mmol) was dissolved in 1 mL CH₂Cl₂. Then 20 μL of 2.2'-(ethylenedioxy)bis(ethylamine) was added into the above solution. The resulting mixture was maintained at room temperature under overnight stirring, and diethyl ether was introduced at 4°C to precipitate the product that was subsequently washed twice by CH₂Cl₂.

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Synthesis of Ir-BTPHSA/UCNP conjugate. The Ir-BTPHSA was covalently attached on the surface of the PEGylated UCNPs through the following procedures. Typically, 825 μ L of UCNPs aqueous solution (2.1×10⁻⁷ mol/L) containing 3 mg of EDC·HCl (0.015 mmol) and 4 mg of Sulfo-NHS (0.018 mmol) was prepared and stirred at room temperature for 10 min. Then, 200 μ L solution of Ir-BTPHSA in CH₃CN (1.4×10⁻³ mol/L) was added into the above reaction mixture that was then kept at 40 °C under overnight stirring. After that, low speed centrifugation was adopted to remove insoluble impurities, and the resulting solution was subjected to ultrafiltration for 2 cycles with 30 kDa MWCO centrifugal filter to obtain the Ir-BTPHSA/UCNP conjugate.



Figure S1. TEM image of the PEGylated NaGdF₄:Yb,Tm,Ca@NaGdF₄ nanoparticles.



Figure S2. The UV-Vis absorption spectra of Ir-BTPHSA in DMSO, Ir-BTPHSA-jeffamine conjugate in DMSO, Ir-BTPHSA attached on the surface of the PEGylated UCNPs (Ir-BTPHSA/UCNPs) in aqueous solution, and Ir-BTPHSA encapsulated by CDs (Ir-BTPHSA/CD) in aqueous solution.

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Figure S3. The most possible binding situation between Ir-BTPHSA and β -CD.



Figure S4. Photographs of Ir-BTPHSA in DMSO (left), Ir-BTPHSA in H_2O (middle), and Ir-BTPHSA/CD in H_2O (right) captured right after the preparation of the solution (left image) and 24 h after the solutions were kept under ambient conditions (right image), respectively.



Figure S5. PL spectra of Ir-BTPHSA encapsulated by CDs (ex=488 nm) in aqueous solutions containing different levels H_2O_2 (a), GSH (b), or hydrogen ions (c).

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Figure S6. The absorption spectra of Ir-BTPHSA recorded at different the oxygen levels.



Figure S7. The confocal images of LS180 cells imaged through channels for Ir-BTPHSA (a) and Cy7 (b), respectively, together with the merged image (c) and the intensity correlation plot (d).



Figure S8. Cytotoxicity test for Ir-BTPHSA/CD-Cy7



Figure S9. ¹H NMR of $[(BTP)_2Ir(\mu-Cl)]_2$.



Figure S11. COSY spectra of Ir-BTPHSA