

Figure S1. Transfection of a FoxP1 plasmid induces fiber atrophy. *Tibialis anterior* muscles co-transfected with a GFP empty vector and either a FoxP1 expression plasmid or empty vector show reduced (A) muscle mass and (B) fiber cross-sectional area (CSA). Minimum feret diameters (MFD) were binned, fit with a Gaussian least squares regression and significance was determined by calculating the extra sum-of-squares F test. Data are shown as Mean \pm SEM.

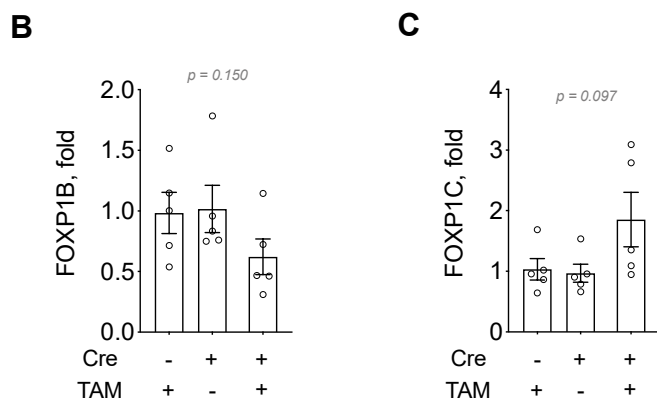
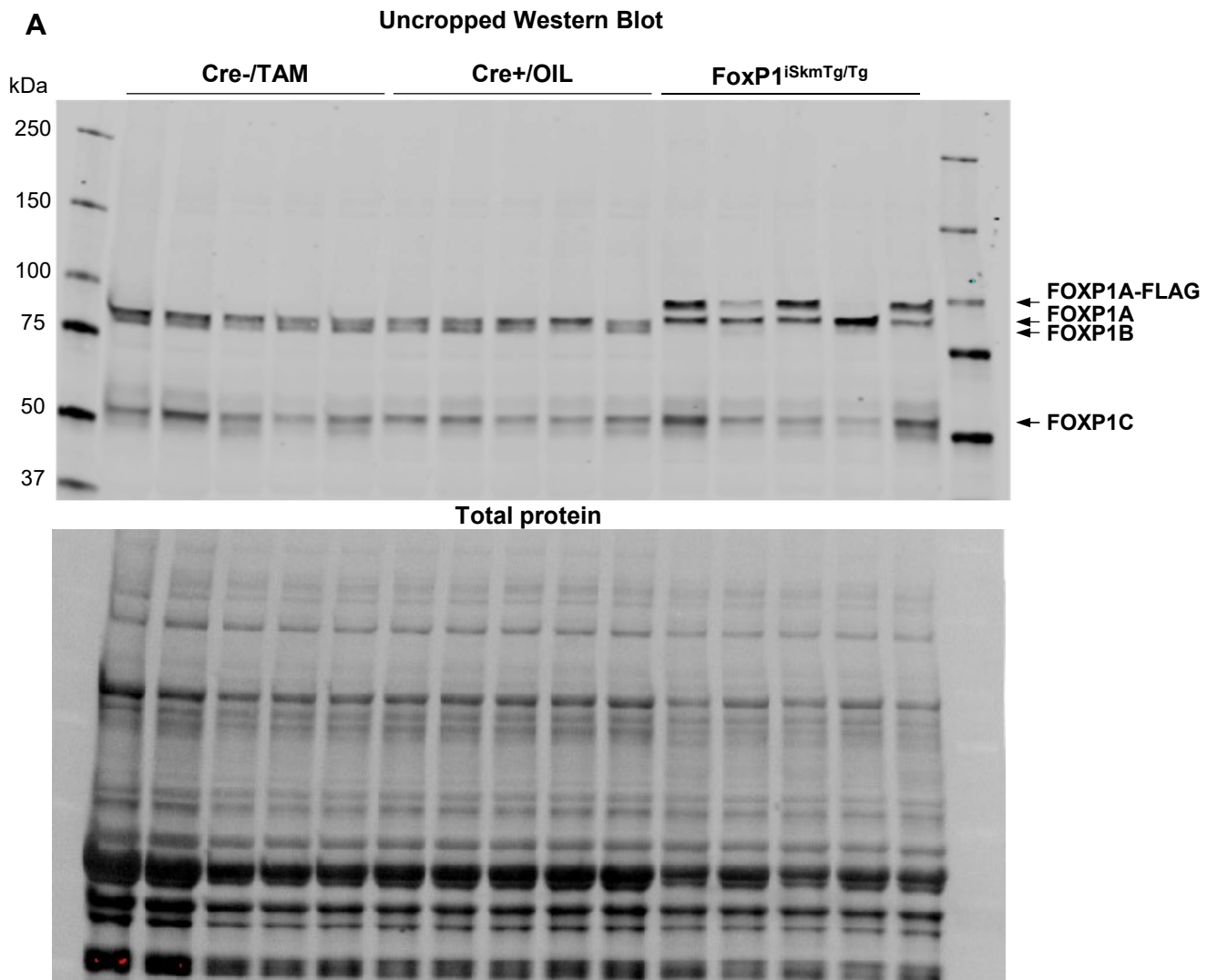


Figure S2. Tamoxifen treatment increases FoxP1A expression in FoxP1^{iSkmTg/Tg} *triceps surae* muscles. (A) Uncropped images of the western blot presented in Fig. 2B. (B-C) Quantification of FOXP1B (B) and FOXP1 C (C) expression in CRE+ FoxP1^{iSkmTg/Tg}, and CRE- littermate and genetic controls, normalized to total protein. Data are shown as Mean ± SEM.

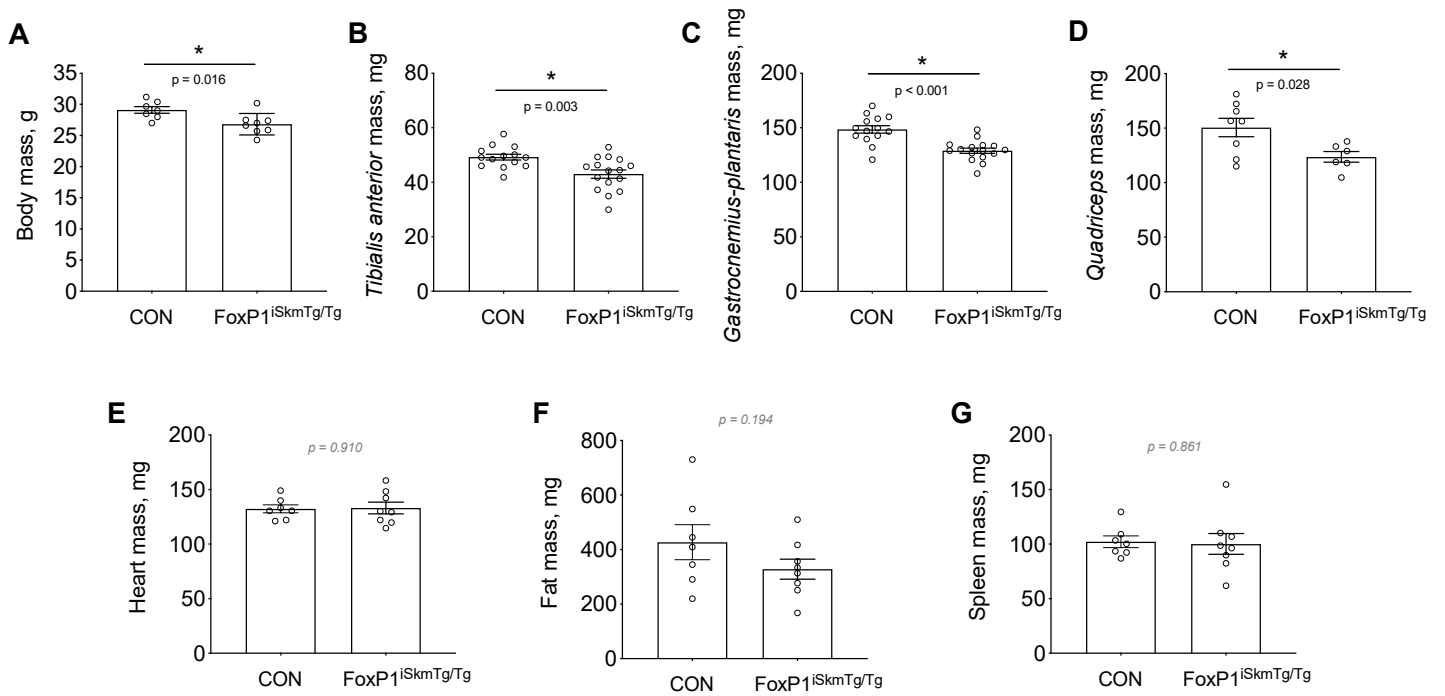


Figure S3. Skeletal muscle-specific over-expression of FoxP1 leads to muscle wasting in male mice. Tamoxifen treatment (started at ≥ 9 weeks of age) induces body wasting (**A**) and skeletal muscle wasting (**B-D**) but not heart (**E**) or fat (**F**) wasting. (**G**) Skeletal muscle-specific over-expression of FoxP1 does not affect spleen mass. CON = CRE-littermates. Data are shown as Mean \pm SEM.

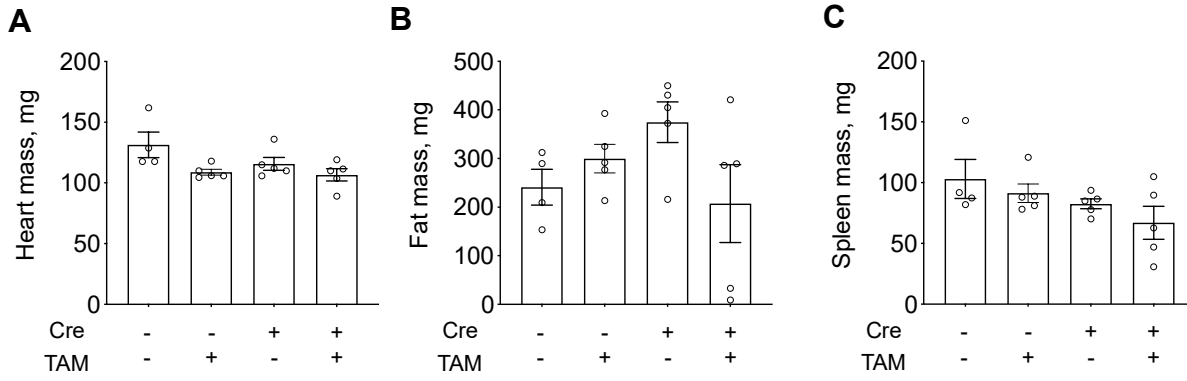


Figure S4. Skeletal muscle-specific over-expression of FoxP1 does not induce changes in (A) heart, (B) gonadal fat and (C) spleen masses in female mice. Note, however that several FoxP1^{iSkmTg/Tg} mice showed fat wasting. Data are reported as mean ± SEM.

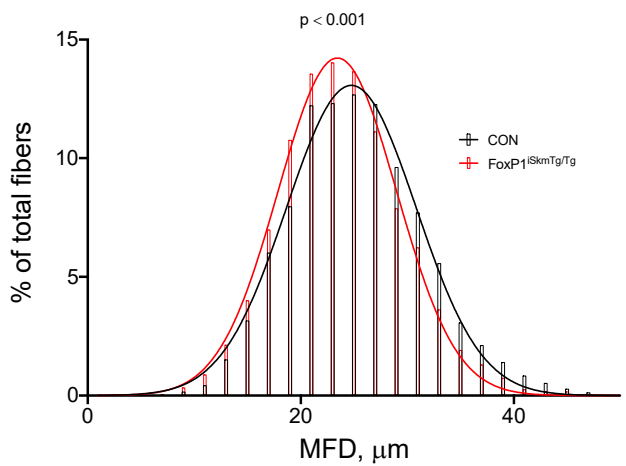


Figure S5. Skeletal muscle-specific over-expression of FoxP1 induces diaphragm fiber atrophy as indicated by the leftward shift observed in fiber size distribution in FoxP1^{iSkmTg/Tg} mice vs. genetic controls (CON). Minimum feret diameters (MFD) were binned, fit with a Gaussian least squares regression and significance was determined by calculating the extra sum-of-squares F test.

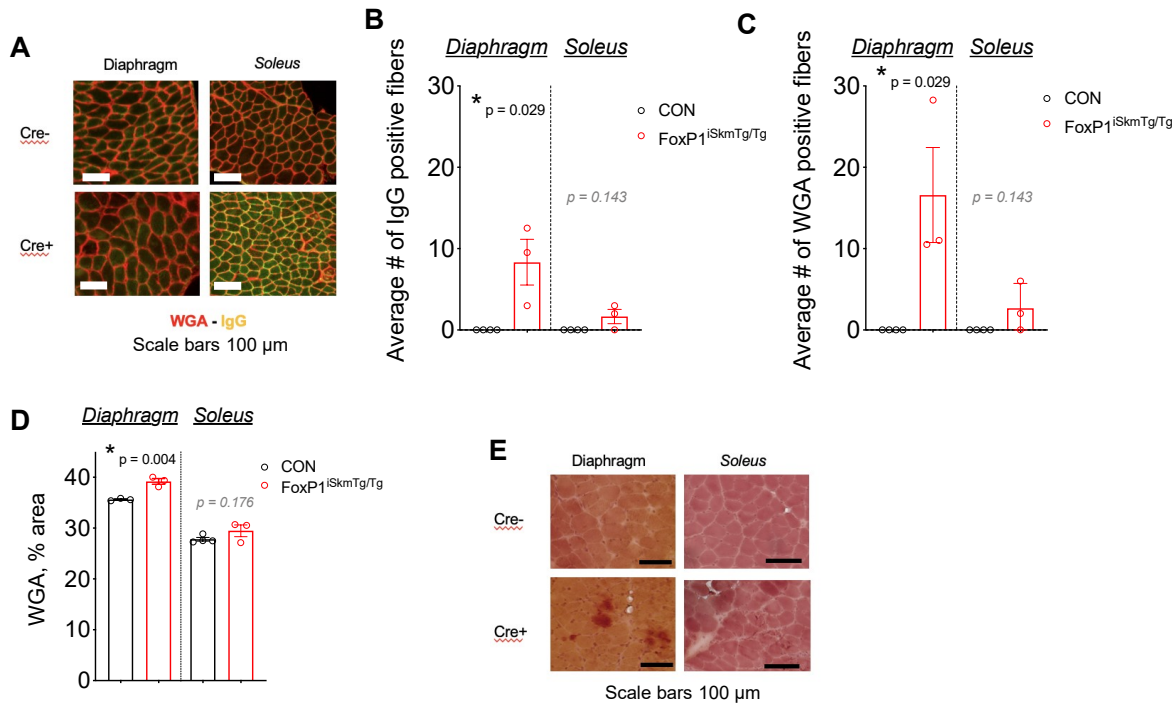


Figure S6. Skeletal muscle-specific FoxP1 over-expression also leads to fiber damage in male mice. (A) Representative images of diaphragm and *soleus* skeletal muscle cross sections from FoxP1^{iSkmTg/Tg} mice and CRE-littermate controls (CON) stained with wheat germ agglutinin (WGA) and endogenous immunoglobulin G (IgG). (B-D) Quantification of IgG (B) and WGA (C) positive fibers as well as of the percent area showing WGA staining per 10X field of view (D). (E) Representative images of diaphragm and *soleus* skeletal muscle cross sections from FoxP1^{iSkmTg/Tg} mice and CRE-littermate controls stained with Alizarin Red. Data are reported as mean \pm SEM.

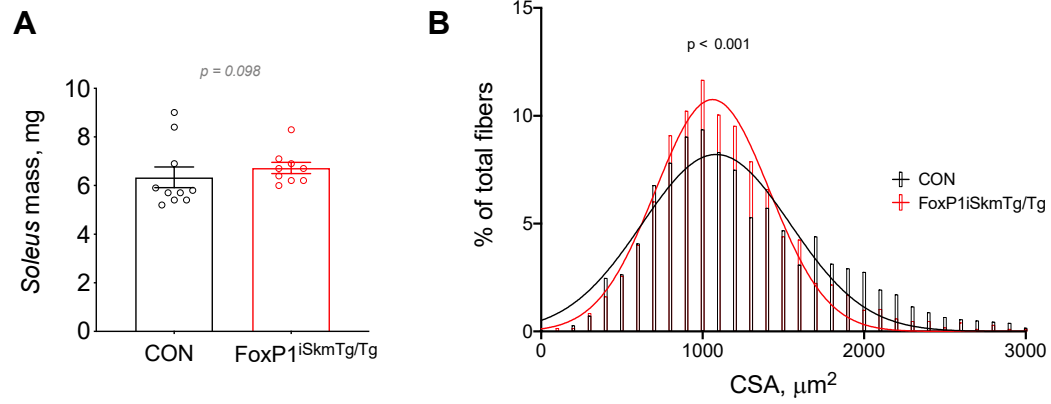


Figure S7. Female *soleus* (A) muscle mass and (B) fiber size distribution in response to skeletal muscle specific FoxP1 over-expression. Fiber cross-sectional areas (CSA) were binned, fit with a Gaussian least squares regression and significance was determined by calculating the extra sum-of-squares F test.

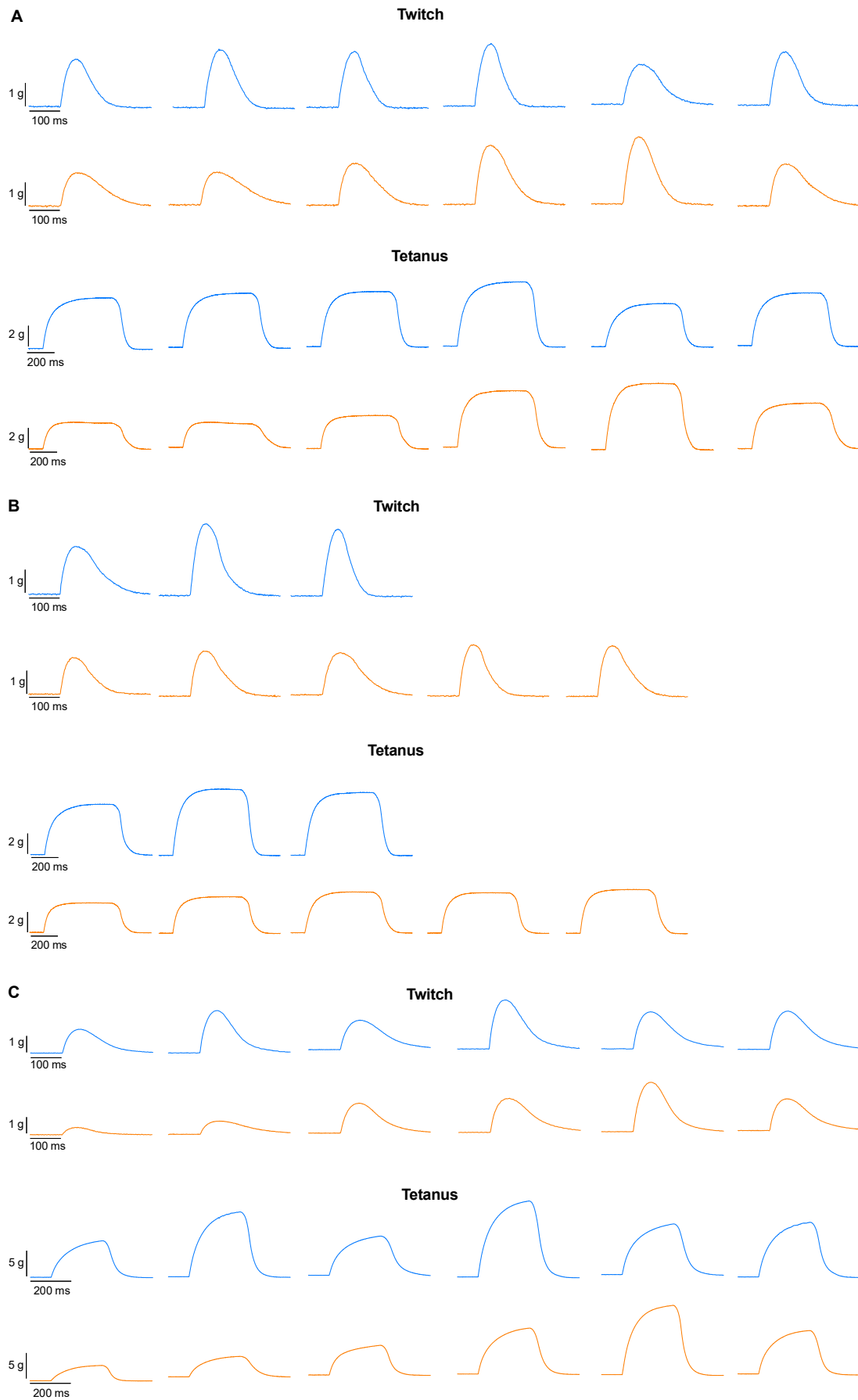


Figure S8. Original traces of twitch and tetanic forces recorded in (A) female and (B) male diaphragm strips as well as in (C) female *so/ei* of *FoxP1*^{ISkmTg/Tg} (orange) and CRE- littermate (CON, blue) mice.

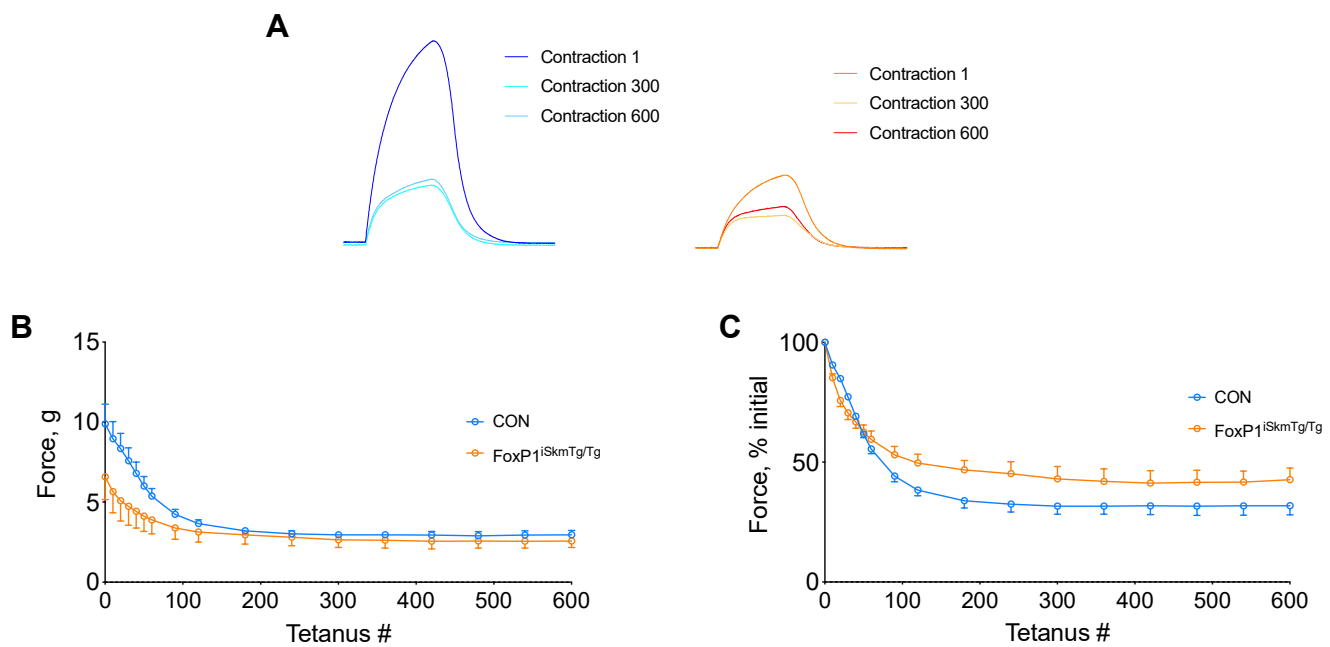


Figure S9. Skeletal muscle specific FoxP1 over-expression does not affect skeletal muscle fatigability. (A) Typical traces of force evoked by the 1st, 300th and 600th contraction and average of (B) absolute and (C) relative evoked forces in FoxP1^{iSkmTg/Tg} (orange) and CRE- littermates (CON, blue). Data are reported as mean \pm SEM.

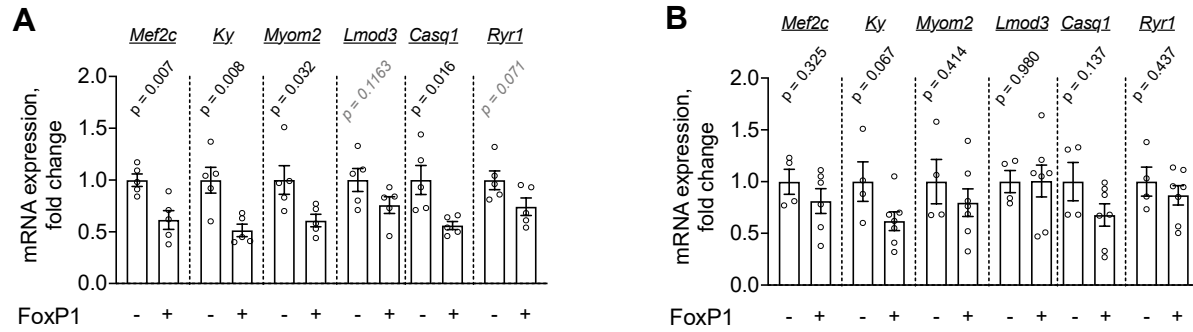


Figure S10. FoxP1 transcriptional repression is mediated, at least in part, by HDACs. (A-B) Mouse *tibialis anterior* muscles were transfected with a FoxP1 plasmid or empty vector. Subsequently, a subset of these mice received daily intraperitoneal (IP) injections of trichostatin A (TSA, a pan HDAC inhibitor) for four consecutive days (B). RT-qPCR shows that FoxP1-induced repression of several genes (A) is mediated by HDAC activity (B). Data are reported as mean \pm SEM.