

## Supporting information

# Gold Nanorod-Melanin Hybrids for Enhanced and Prolonged Photoacoustic Imaging in the Near-Infrared-II Window

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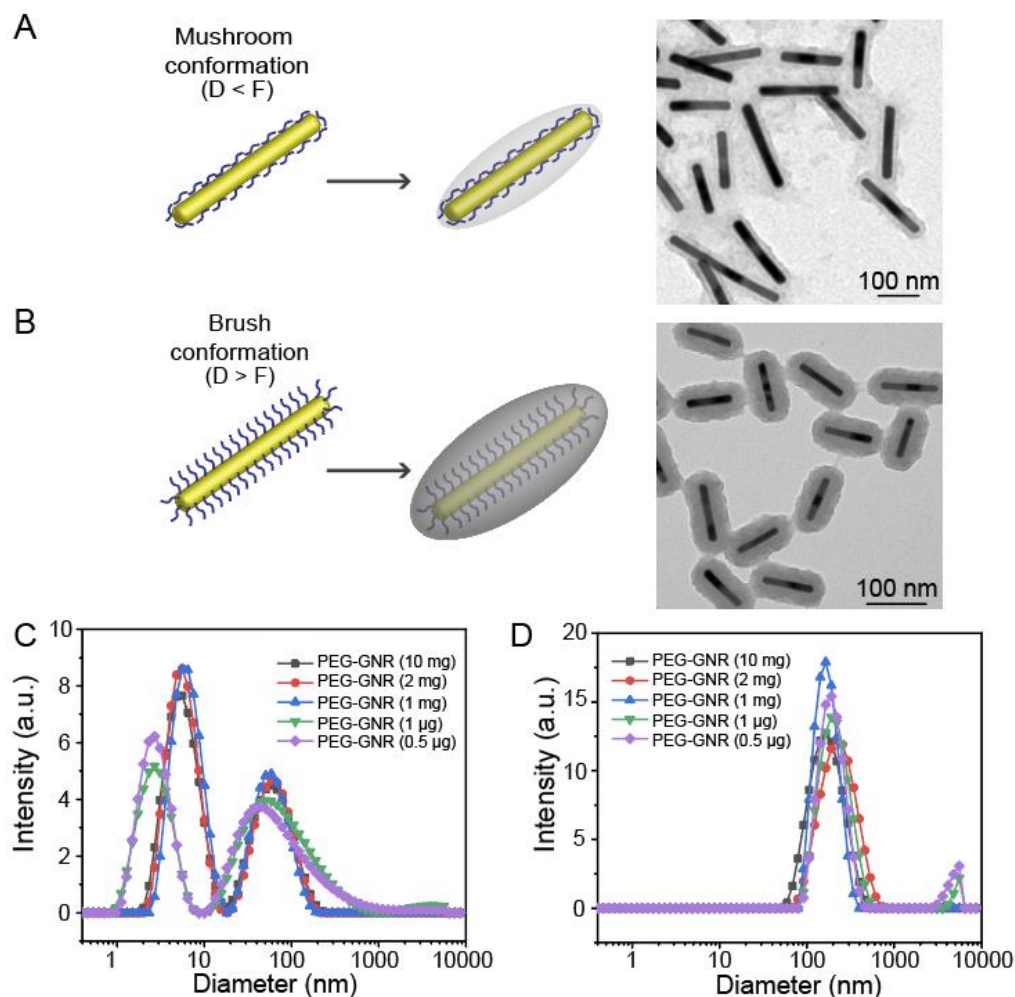
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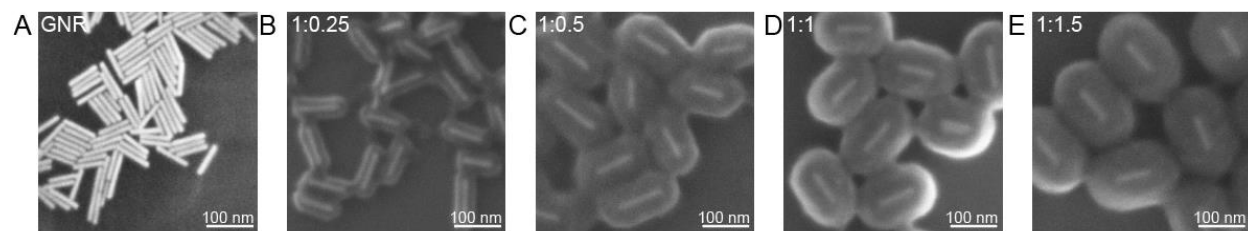
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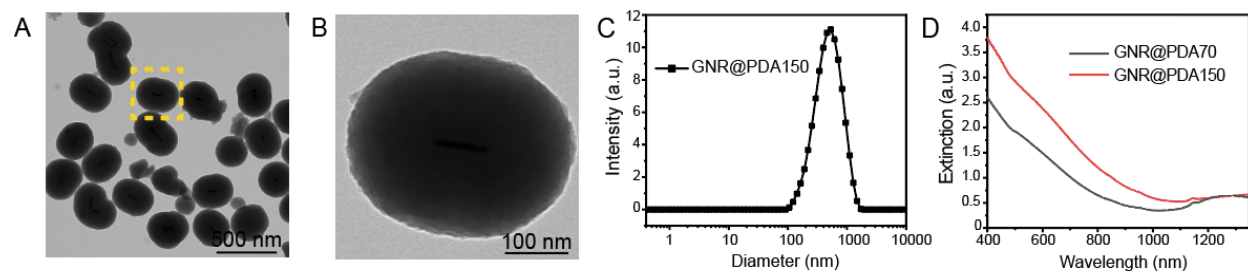
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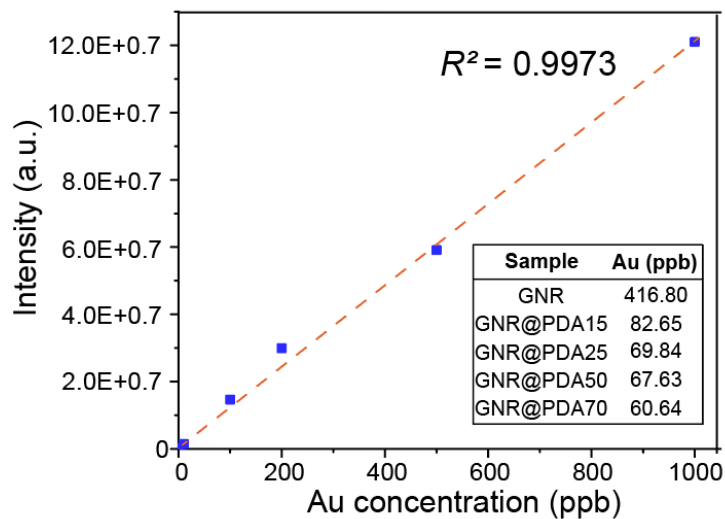
**Figure S1.** Mushroom and brush conformation. PEG chains can form either mushroom or brush conformations. (A) PEG chains formed the mushroom conformation and PDA coating was instable when we used a small amount of PEG-SH. (B) PEG chains made the brush conformation when we used large amount of PEG-SH ( $>1$  mg). PDA coating became stable and uniform, showing colloidal stability. (C) DLS data of PEGylated GNRs at different concentration of PEG-SH: 10 mg, 2 mg, 1 mg, 1  $\mu\text{g}$ , and 0.5  $\mu\text{g}$ . (D) DLS data of PDA coated GNRs replaced by different amounts of PEG chain. GNRs were aggregated, and PDA coating was unstable when we used 1  $\mu\text{g}$ , and 0.5  $\mu\text{g}$  (mushroom conformation). The mass concentration of GNR was 200  $\mu\text{g}/\text{mL}$ , and we used 1:1 ratio of GNR to dopamine concentration (4 mg/mL).



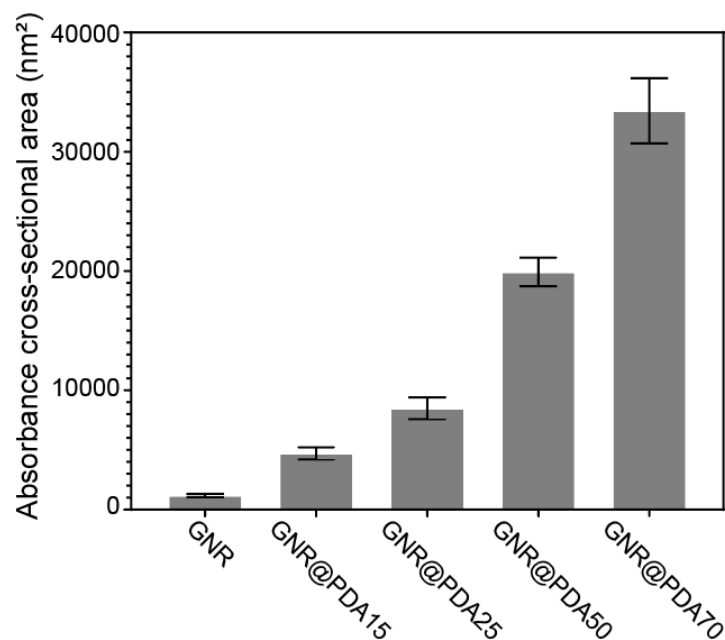
**Figure S2.** SEM images of GNR@PDAs. (A) GNR, (B) GNR@PDA15, (C) GNR@PDA25, (D) GNR@PDA50, and (E) GNR@PDA70.



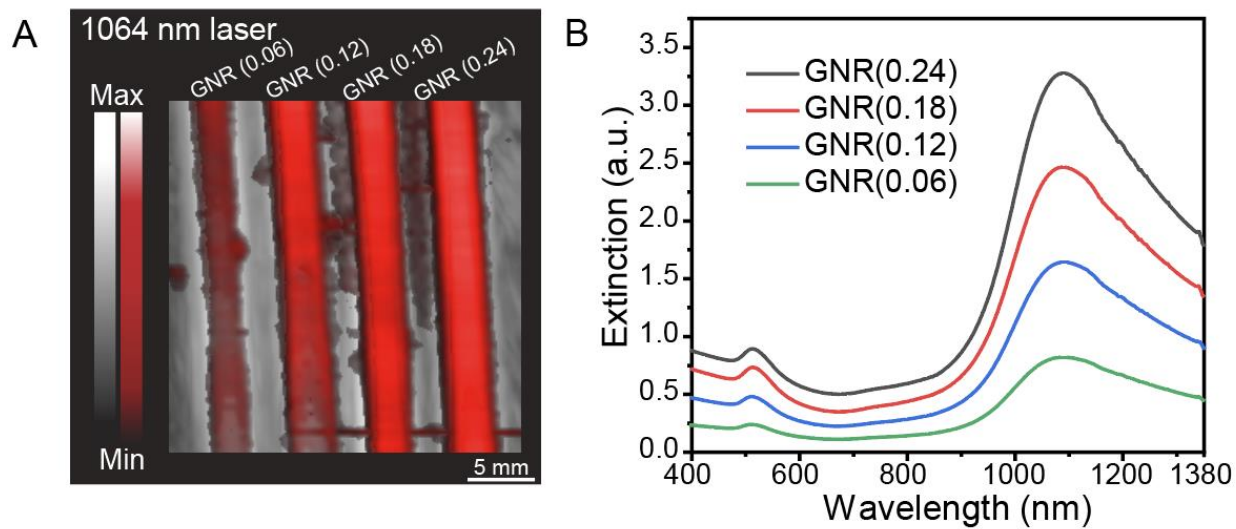
**Figure S3.** (A) GNRs aggregated when we used an excess of dopamine. (B) The yellow dotted square shows TEM image of a single GNR@PDA particle with PDA thickness of 150 nm (referred as GNR@PDA150). (C) DLS data of GNR@PDA150 showed a large size distribution (PDI  $\sim$  0.2). (D) Optical extinction of GNR@PDA70 and GNR@PDA150. Longitudinal peak of GNR@PDA150 was not shown while GNR@PDA70 exhibited longitudinal peak around 1300 nm. This result indicated that longitudinal peak of GNR@PDA was no longer distinguishable when the PDA coating thickness became 150 nm.



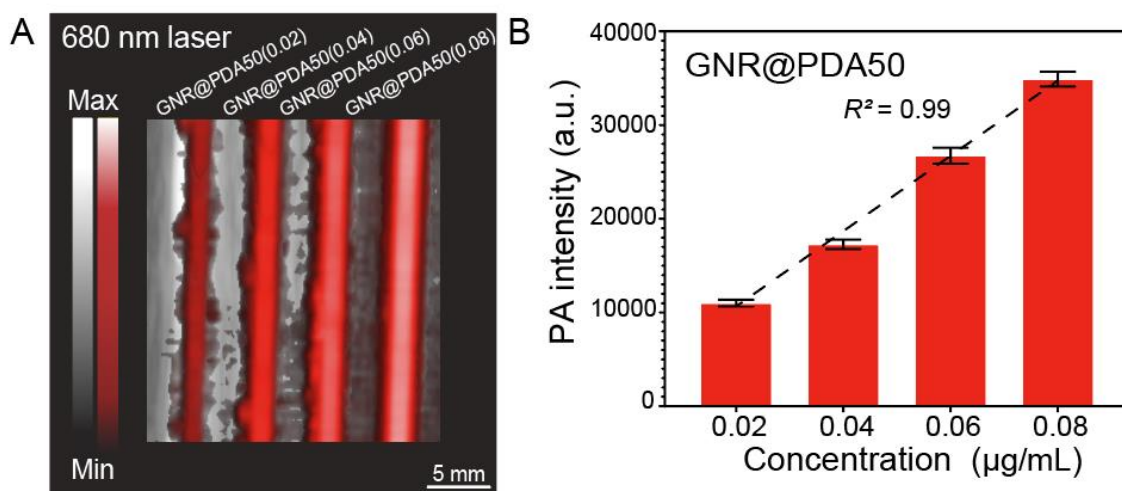
**Figure S4.** ICP-MS data. The number of GNRs in GNR, GNR@PDA15, GNR@PDA25, GNR@PDA50, and GNR@PDA70 was calculated by using ICP-MS method. Inset table indicates the concentration of Au ions in each sample solution.



**Figure S5.** Absorption cross-sectional area of GNR@PDAs. Absorption cross-sectional area was exponentially increasing as PDA coating became thicker. Absorption cross-sectional area (nm<sup>2</sup>) of GNR@PDA nanoparticles was calculated by the formula of diameter (D) × length (L) based on the TEM images. Error bar represents the standard deviation of fifteen GNR@PDA nanoparticles.

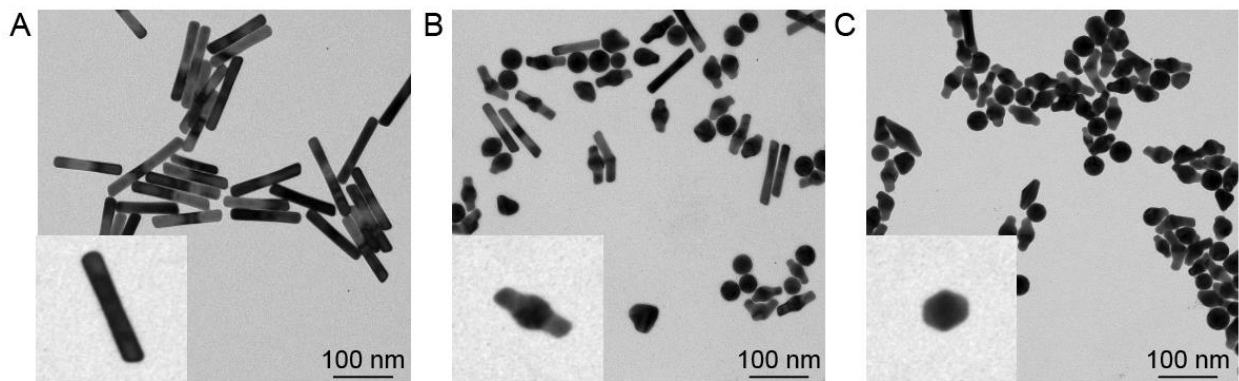


**Figure S6.** (A) PA image when illuminated with a 1064 nm laser, and (B) optical extinction of GNR at various concentrations (240, 180, 120, and 60  $\mu\text{g/mL}$ ). The optical absorption is proportional to the concentration leading to higher PA signal.

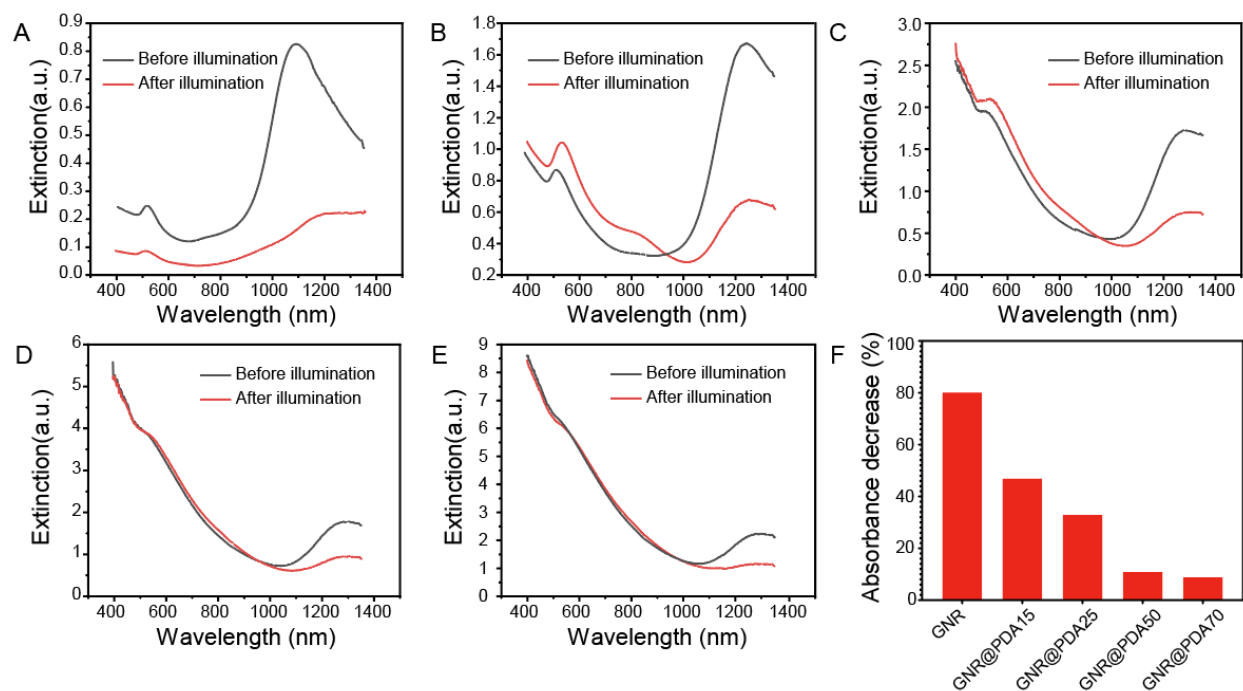


**Figure S7.** (A) PA image and (B) its corresponding quantitative PA signal amplitude of GNR@PDA50 when illuminated with a 680 nm laser. The increased concentration led to higher PA signal due to the increased absorbance. The error bar represents the standard deviation of five regions of interest.

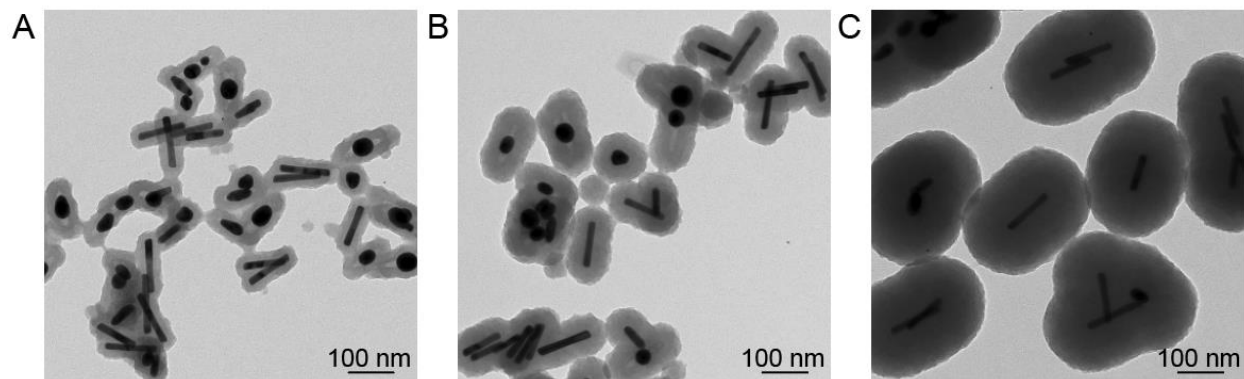




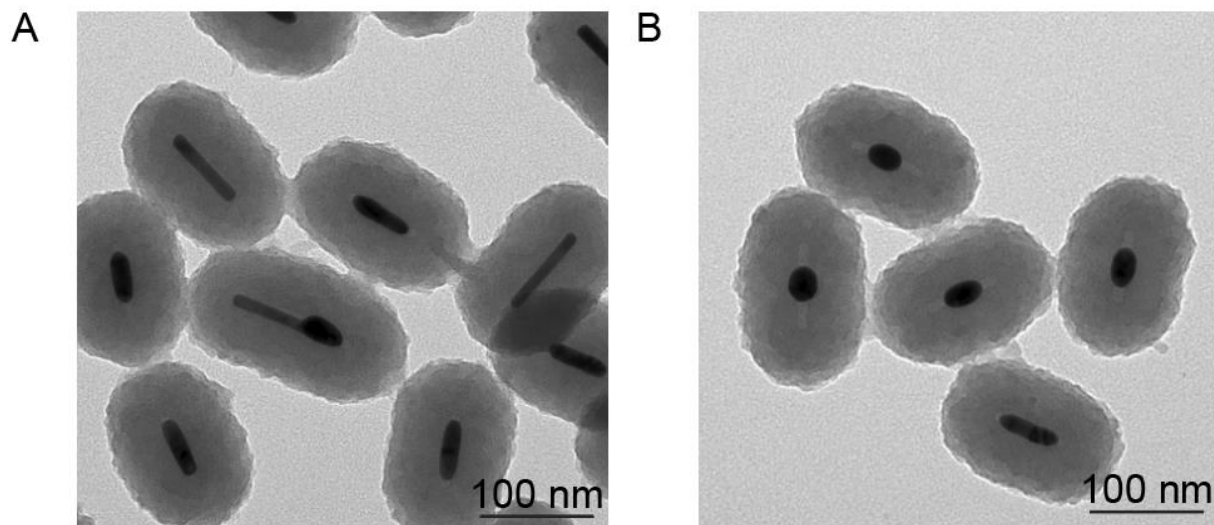
**Figure S8.** Shape transition of GNR under the laser illumination: (A) pristine GNRs, (B) shape transition of GNRs after 1 min of laser illumination, and (C) shape transition of GNRs after 5 min of laser illumination. Atomic rearrangement occurs at  $\{110\}$  facets of GNRs to minimize their surface energy, converting into spherical shape.



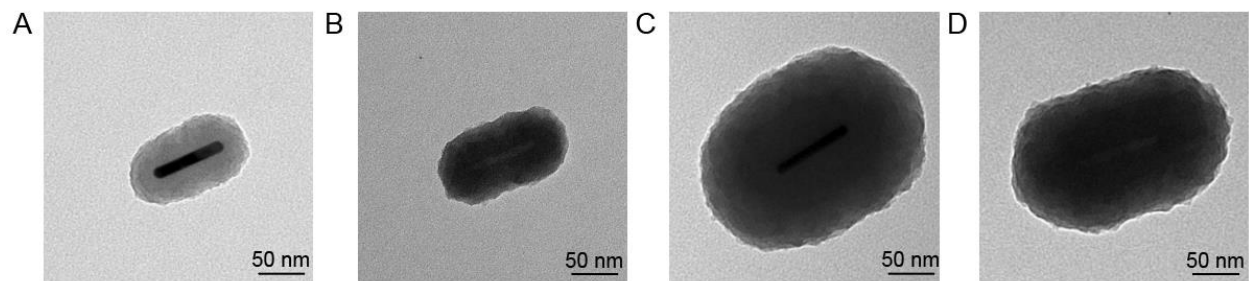
**Figure S9.** Changes of UV-vis-NIR spectra of GNR and GNR@PDAs before and after 5 min of the laser illumination. (A) GNR, (B) GNR@PDA15, (C) GNR@PDA25, (D) GNR@PDA50, (E) GNR@PDA70, and (F) longitudinal absorption peaks of GNR, GNR@PDA15, GNR@PDA25, GNR@PDA50, and GNR@PDA70 decreased by 80%, 47%, 33%, 11%, and 9%, respectively.



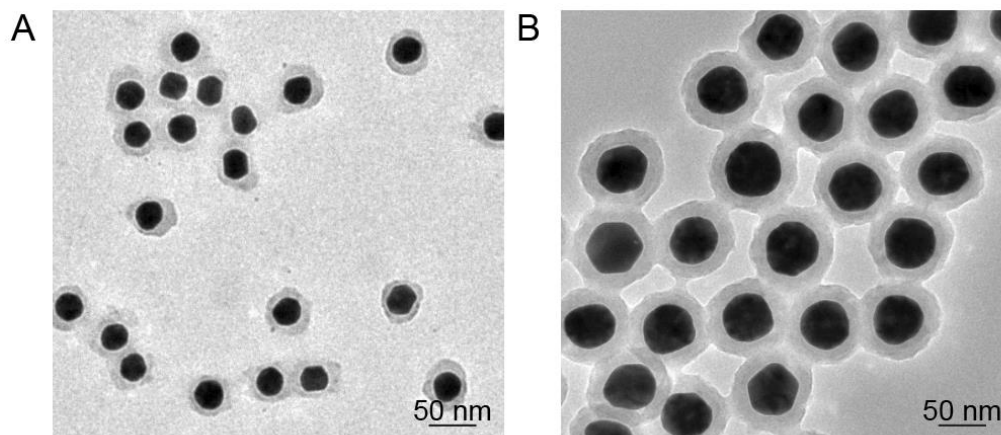
**Figure S10.** Shape transition of GNR@PDAs. (A) GNR@PDA15, (B) GNR@PDA25, and (C) GNR@PDA70 after 5 min of the laser illumination.



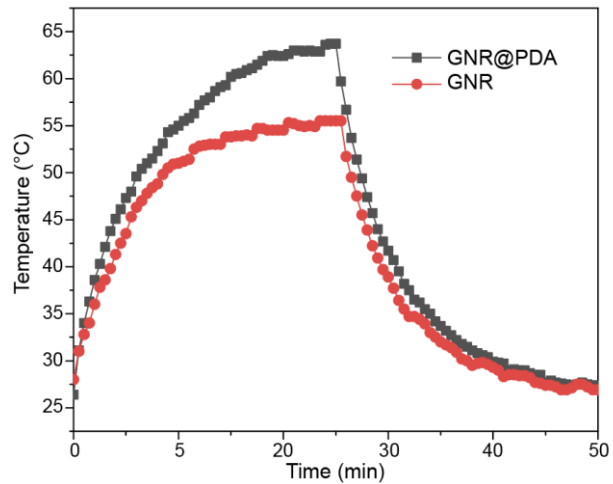
**Figure S11.** Shape transition of PDA@GNR50 after 5 min (A) and 15 min (B) of the laser exposure. PDA coating maintained its original structure while most of the coated GNRs melt, showing that PDA is highly robust.



**Figure S12.** TEM images of single PDA capsule particle. (A) GNR@PDA15, (B) PDA<sub>capsule</sub>15, (C) GNR@PDA70, and (D) PDA<sub>capsule</sub>70.



**Figure S13.** PDA coated gold nanospheres (GNSs): (A) 30 nm and (B) 60 nm GNSs covered by PDA shell. These images indicate that PDA coating strategy can be applied to various types of nanoparticles for improving PA performance.



**Figure S14.** Photothermal conversion efficiency of GNR and GNR@PDA. Photothermal conversion efficiency of GNR@PDA (40 %) was 2 % higher than that of GNR (38 %), indicating that PDA coating can be used for photothermal therapy. 1064 nm laser with 1 W/cm<sup>2</sup> was used for the measurement.

### Calculation of mushroom and brush conformation of PEG chain.

Based on the Flory – Huggins theory, brush conformation will be formed when the graft density is high ( $D > F$ ). Thus, we need to calculate Flory radius (F) by using below equation (1).<sup>1</sup>

$$F = n^{3/5}\alpha \quad (1)$$

where n is the number of monomers per chain, and a is a monomer size ( $\alpha = 3.5 \text{ \AA}$  for PEG).<sup>2</sup> After calculating F, we can estimate the minimum D to have brush conformation ( $D > F$ ). For example,  $F = 120^{3/5} * 3.5^2$  (n is about 120 when a molecular weight of PEG is 5000). Thus,  $D > F \approx 216.6 \text{ \AA}$  is required to make brush conformation.

Next, the surface area of single gold nanorod (GNR) nanoparticle can be calibrated by following equation (with an assumption that GNR has a perfect cylindrical shape.)

$$A = \pi r^2 + 4rL \quad (2)$$

where A is surface area of GNR, r is half diameter of GNR, and L is length of GNR. Then, we can divide the surface area of a single GNR nanoparticle with  $D^2$  to know the minimum number of PEG-chains to cover a single GNR nanoparticle. For example, r and L of GNR were  $6.23 \pm 0.68$ , and  $95.07 \pm 6.92 \text{ nm}$  respectively (see **Table 1**). Surface area of a single GNR nanoparticle was about  $2489.33 \text{ nm}^2$ , and  $D^2$  is  $470.89 \text{ nm}^2$ . Thus, minimum number of PEG-SH molecule was about 20 to cover a single GNR nanoparticle under the premise that all the PEG-SH molecule is substituted with CTAB. With this information, we can estimate the minimum amount of PEG-SH solution to make brush conformation when we know the concentration of GNR by ICP-MS method (**Figure S5**).



## Detailed information about Equation 5

Shape transition of GNRs can be also explained by absorbance decrease according to equation 1.<sup>3</sup>

$$[(A_{\lambda,0} - A_{\lambda,1})/\sigma b] = \rho\sigma^{(n)}I^{(n)} \quad (1)$$

where  $A_{\lambda,0}$  is the sample absorbance at a given wavelength ( $\lambda$ ) before the laser illumination,  $A_{\lambda,1}$  is the sample absorbance at a given wavelength ( $\lambda$ ) after the laser illumination,  $\sigma$  is the absorption cross-section of the nanoparticles,  $b$  is the path length,  $\rho$  is the density of the nanoparticles, and  $n$  is the number of photons absorbed.  $I$  is laser fluence.

$$[(A_{\lambda,0} - A_{\lambda,1}/b)] = \rho\sigma^{(n)+1}I^{(n)} \quad \text{---- multiply } \sigma$$

$$[(A_{\lambda,0} - A_{\lambda,1}/c\epsilon b)] = (\rho\sigma^{(n)+1}I^{(n)})/c\epsilon \quad \text{---- (2)}$$

$c$ : concentration,  $\epsilon$ : molar extinction coefficient

Applying the Beer-Lambert Law ( $A_{\lambda,0} = c\epsilon b$ ) to equation (2)

$$(A_{\lambda,0} - A_{\lambda,1})/A_{\lambda,0} = (\rho\sigma^{(n)+1}I^{(n)})/c\epsilon \quad \text{---- (3)}$$

$$\ln [(A_{\lambda,0} - A_{\lambda,1})/A_{\lambda,0}] = \ln (\rho\sigma^{(n)+1}I^{(n)})/c\epsilon \quad \text{---- applying ln to both sides of equation (3)}$$

$$\ln [(A_{\lambda,0} - A_{\lambda,1})/A_{\lambda,0}] = \ln I^{(n)}/c\epsilon + \ln (\rho\sigma^{(n)+1})/c\epsilon \quad \text{---- } c\epsilon \ll I^{(n)}$$

Equation (1) can be rearranged to

$$\ln [(A_{\lambda,0} - A_{\lambda,1})/A_{\lambda,0}] = n \ln(I) + \text{constant}$$

## References

- (1) de Gennes, P. Conformations of Polymers Attached to an Interface. *Macromolecules* **1980**, *13* (5), 1069-1075.
- (2) Jokerst, J. V.; Lobovkina, T.; Zare, R. N.; Gambhir, S. S. Nanoparticle PEGylation for Imaging and Therapy. *Nanomedicine* **2011**, *6* (4), 715-728.
- (3) Gandhi, S. R.; Bernstein, R. B. Influence of the Focal Length of the Laser Beam Focusing lens on MPI yield. *Chem. Phys.* **1986**, *105* (3), 423-434.