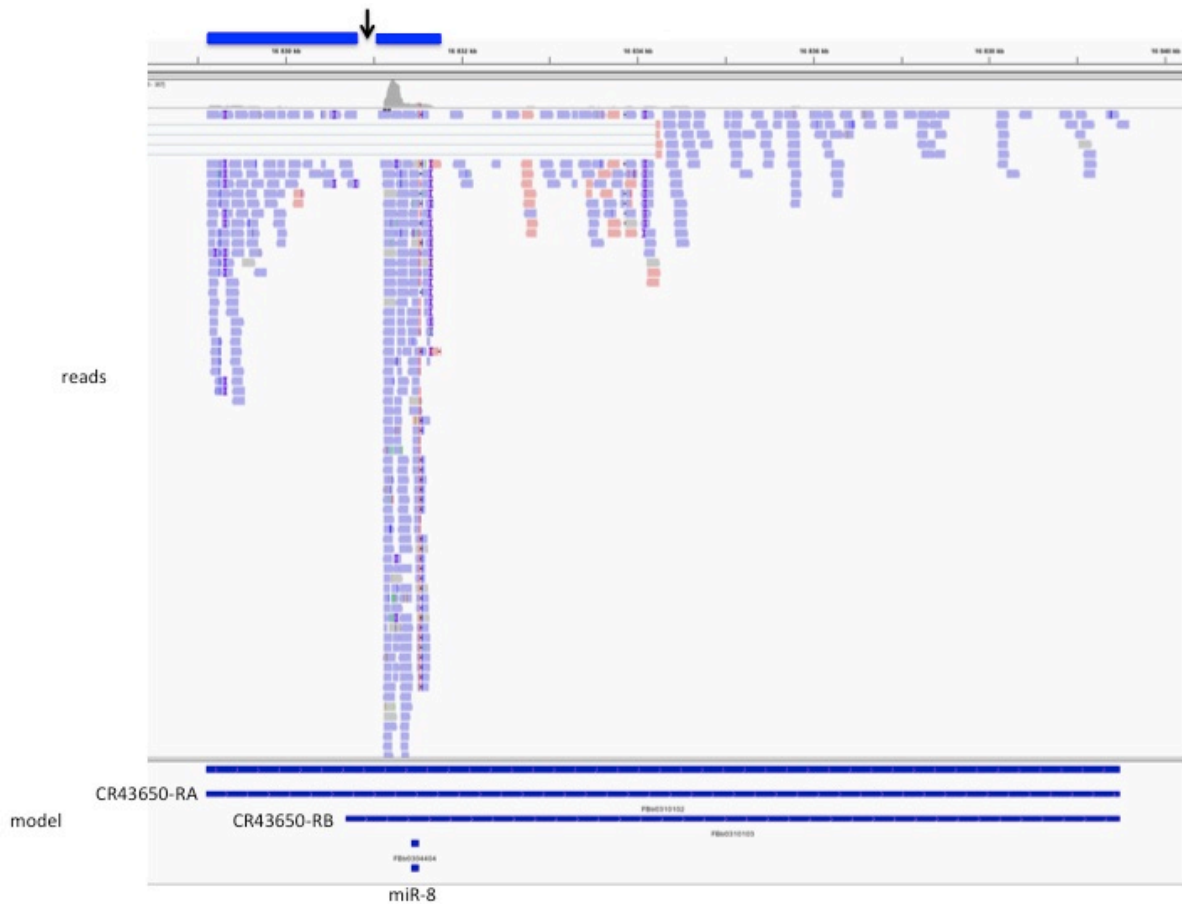


Figure S1 : Ribosome covering profiles on several *Drosophila* mi-R genes. Screen shot of examples of miR genes exhibiting ribosome profiling using the GWIPS-vis browser (<https://gwips.ucc.ie>) [65]. In green are shown the RNA-seq expression, in red ribosome profiling data. The coding gene CG12520 is shown for comparison.

A



B

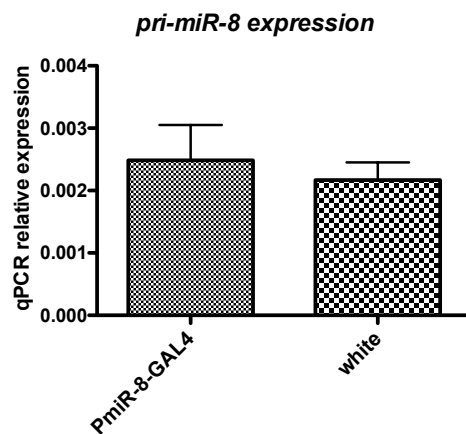
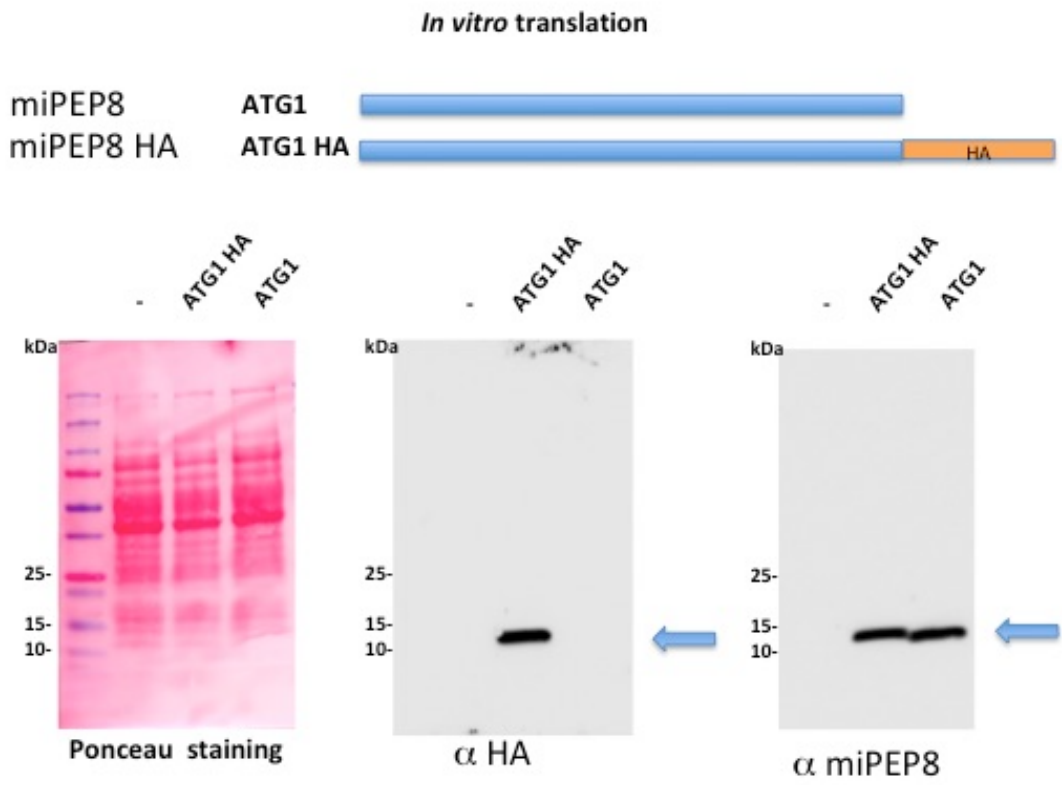
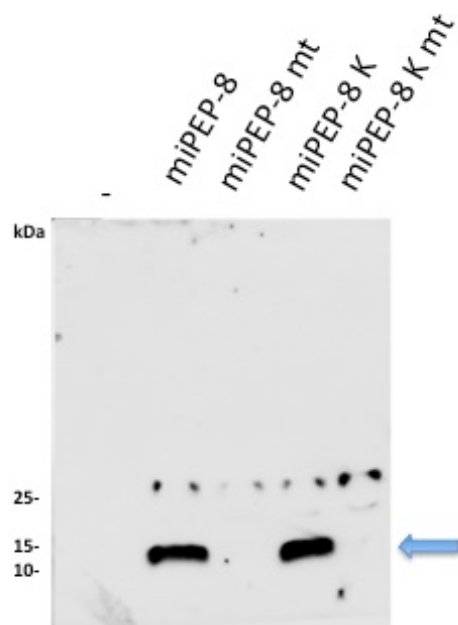


Figure S3 :

A : RNA-seq profil of the *miR-8* locus. Contrasting with the models of long RA and short RB CR43650 transcripts, no covering reads and overlapping reads were detected in the region located upstream the RB transcript suggesting that two different transcriptional units are present (schematized by blue rectangles on top). Arrowhead : Location of the *miR-8* GAL4 line insertion.

B : expression of *miR-8* gene in white recipient flies and *miR-8* GAL4 flies. Differences are non significant (Mann & Whitney non parametric test). n=6

A**B**

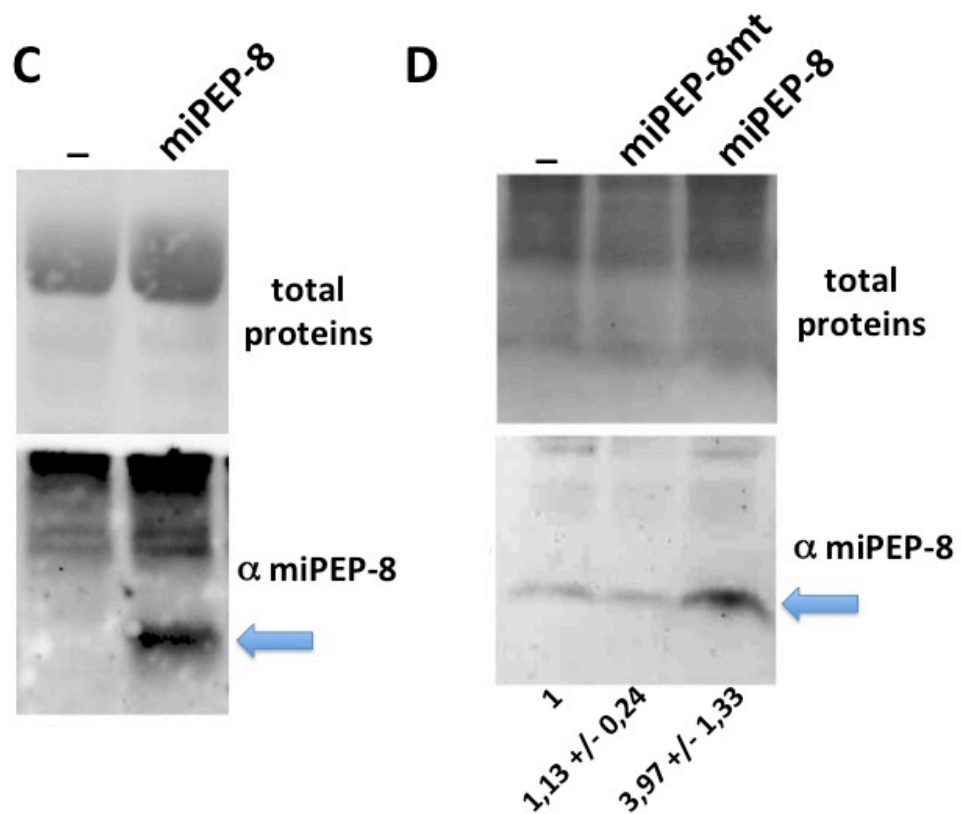


Figure S4 : anti miPEP-8 antibodies characterization. - indicates programmed lysate with empty vector. **A** : *In vitro* translated miPEP-8 HA were produced in insect cell extracts and subjected to western blot experiments. **B** : the miPEP-8 ORFs ATGs were placed in natural, kozak (K) or mutated (mt) translational context and were tested for their ability to be translated. **C** : *N.benthamiana* agroinfiltrated with pCambia expressing vectors, empty vector (ctrl) or miPEP-8 encoding vector (miPEP-8). Arrows indicate miPEP-8. Note the signal in miPEP-8 overexpressing plants. **D** : miR-8 GAL4 flies were crossed with the respective UAS constructs (ctrl is driver crossed with the recipient white flies). Young Adults (1-2 days old) were subjected to western blot using the purified anti miPEP8 antibody. Arrows indicate miPEP-8. Note the increased signal in miPEP-8 overexpressing flies for the wt construct but not the ATG mutated one.

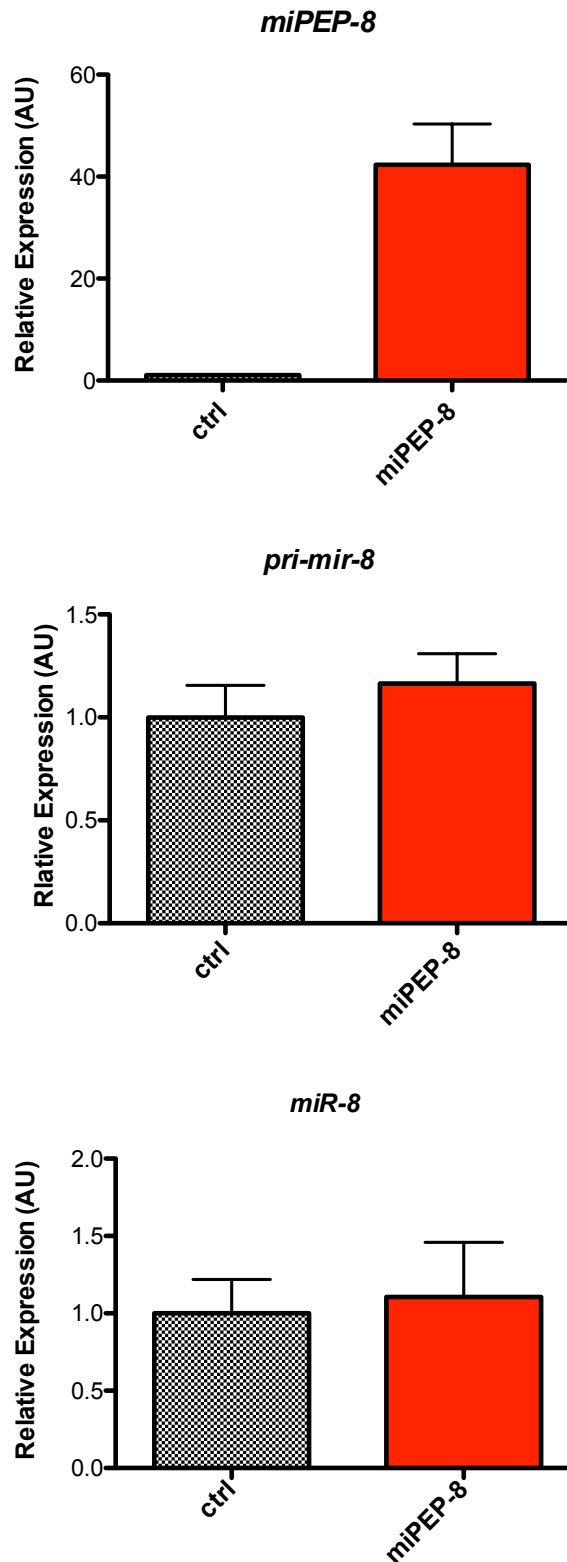


Figure S5 : qPCR on flies over-expressing miPEP-8 using the *miR-8* GAL4 line as driver. Top panel, the overexpression level is more than 45 fold times higher than the endogenous level (n=9). Middle and bottom panels, endogenous level of *pri-mir-8* (n= 17) and mature *miR-8* (n=10) were determined by qPCR. Non significant variation (Mann & Whitney non parametric test) of endogenous *pri-mir-8* or mature *miR-8* was observed upon miPEP-8 overexpression.

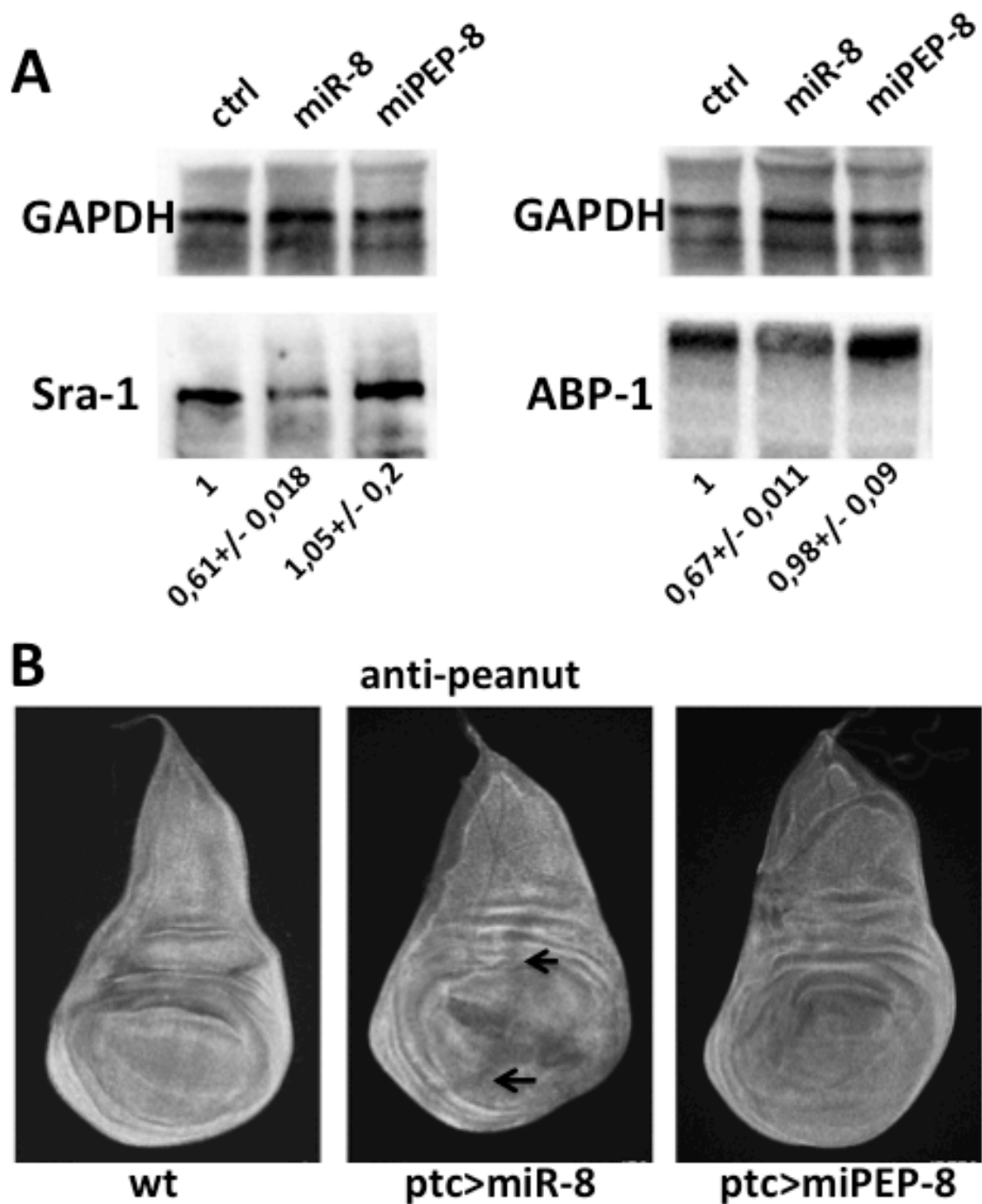


Figure S6 : Lack of miPEP-8 effect on miR-8 endogenous targets.

A : western blot experiment on S2 cells overexpressing *miR-8* or miPEP-8 as indicated.

Bottom : average value of the relative expression/GAPDH from two independent

experiments with SD indicated. **B** : expression of *miR-8* and miPEP-8 using *ptc*-GAL4 in

wing imaginal discs. Third instar discs are stained with an anti-peanut antibody. Arrows :

repression of this Peanut expression in the *ptc* domain with *miR-8* but not miPEP-8.

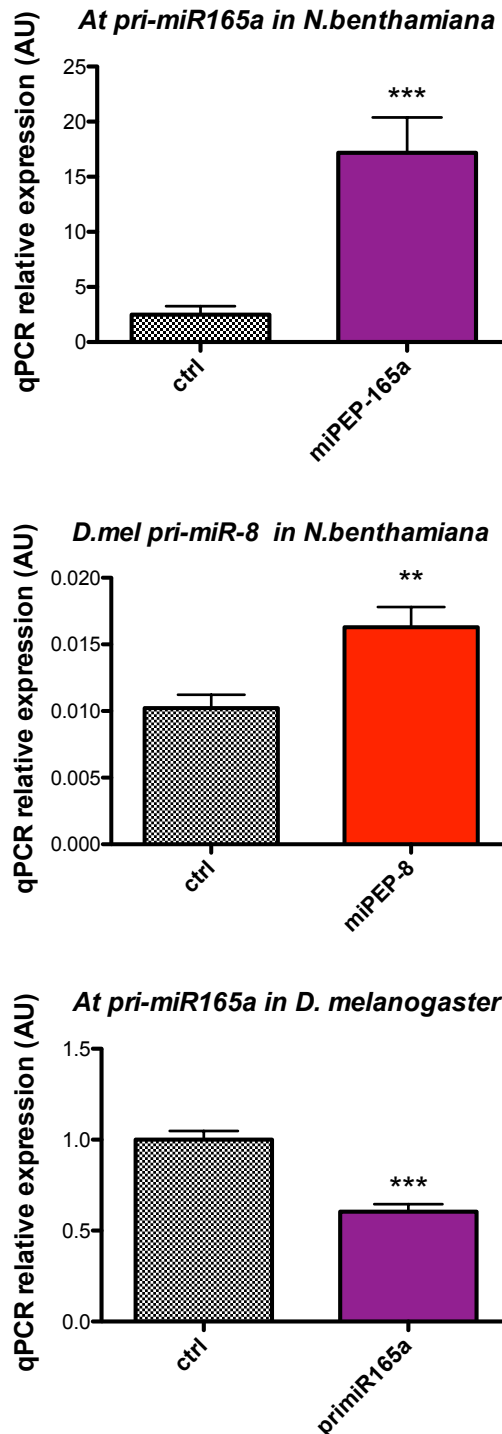


Figure S7 : qPCR on agroinfiltrated *N.benthamiana* leaves with either *A.thaliana pri-miR165a* (n= 8) or *Drosophila pri-miR-8* (n= 12) expressed from ubiquitous promoter (35S) together with either an empty expression vector (ctrl) or a vector expressing miPEP-165a or miPEP-8 respectively. Bottom transfected *drosophila* S2 cells (n= 13) expressing the *A.thaliana pri-miR165a* together with the p-actin empty vector (ctrl) or a p-actin vector expressing the miPEP-165a. * indicate that the difference is significant with a pvalue<0,01 (**), or 0,001 (***)

A

gene	symbol	logFC	pvalue (FDR)	updown	reference
FBgn0015838	Vang	-0,25	0,000100718	down	Bolin et al, 2016
FBgn0011225	jar	-0,41	3,07216E-11	down	Bolin et al, 2016
FBgn0005672	spi	-0,34	4,63246E-09	down	Morante et al, 2013
FBgn0036372	Abp1	-0,18	0,01777008	down	Bolin et al, 2016
FBgn0262716	Arp3	-0,22	0,002244126	down	Bolin et al, 2016
FBgn0013726	pnut	-0,25	6,25716E-05	down	Bolin et al, 2016; Eichenlaub et al, 2016
FBgn0003514	sqh	-0,26	9,0503E-05	down	Bolin et al, 2016
FBgn0038320	Sra-1	-0,28	2,44587E-05	down	Bolin et al, 2016
FBgn0034970	yki	-0,32	1,82097E-06	down	Umegawachiet al, 2017; Sander et al, 2018
FBgn0001257	ImpL2	-0,35	6,13257E-06	down	Lee et al, 2015
FBgn0036141	wls	-0,36	1,78976E-07	down	Kennel et al, 2008
FBgn0034709	Swim	0,82	3,1208E-15	up	Lucas et al, 2015
FBgn0034407	DptB	1,00	0,000544687	up	Choi & Hyun, 2012

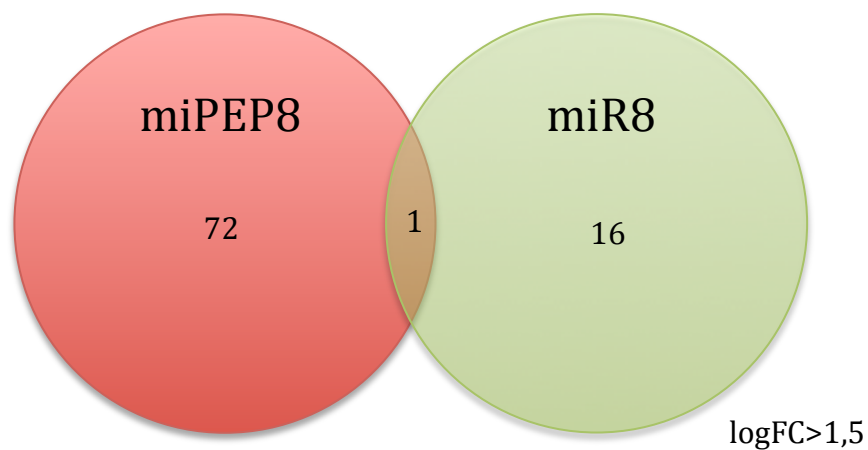
B

Figure S9 : miPEP-8 and *miR-8* deregulated genes in S2 cells overexpressing miPEP-8 or miR-8. **A** : published *miR-8* targets identified in this study that are highly significant (see FDR value) with a logFC > 0,25. **B** : venn diagram for significant deregulated genes (number indicated in the center of each circle) with a logFC > 1 ,5.

A

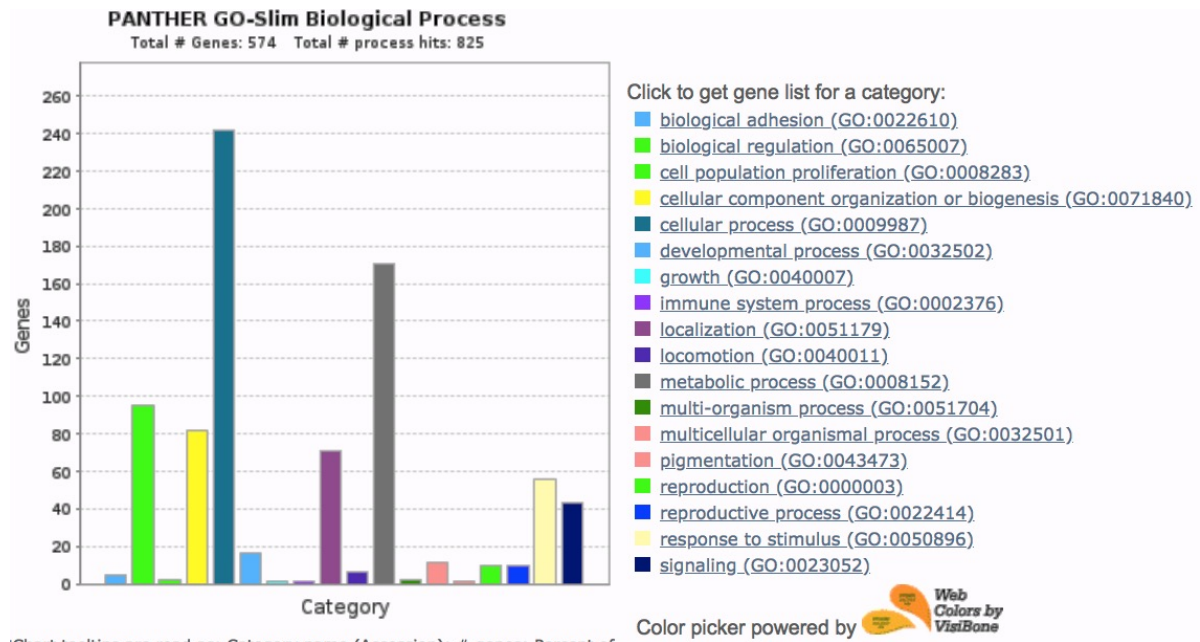


Chart tooltips are read as: Category name (Accession): # genes; Percent of the hit against total # genes; Percent of gene hit against total # Process hits

B

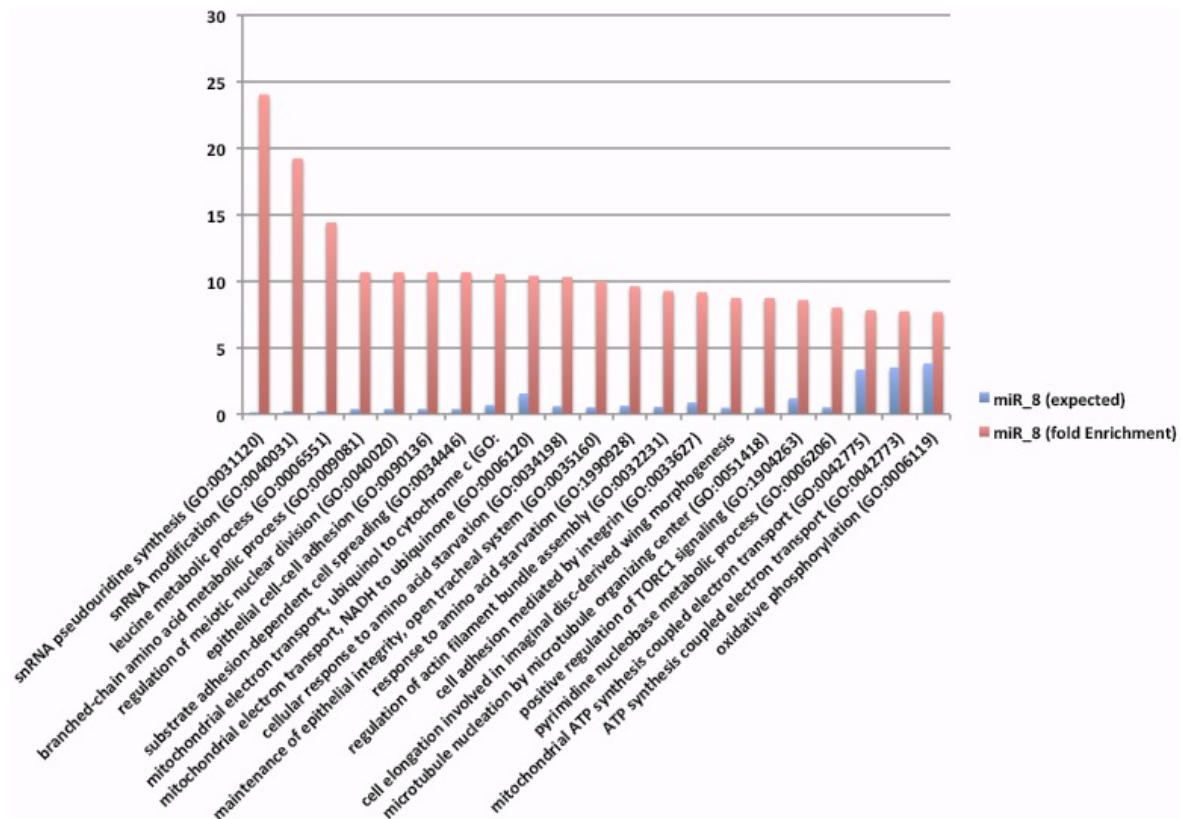


Figure S10 : GO of transcriptome of *miR-8* specific genes. **A** : Most significant regulated biological processes. **B** : Fold enrichment of the most representative biological activities controlled by *miR-8*. Note enrichment of epithelial cell adhesion, and integrity, regulation of actin filament assembly, wing imaginal disc morphogenesis.

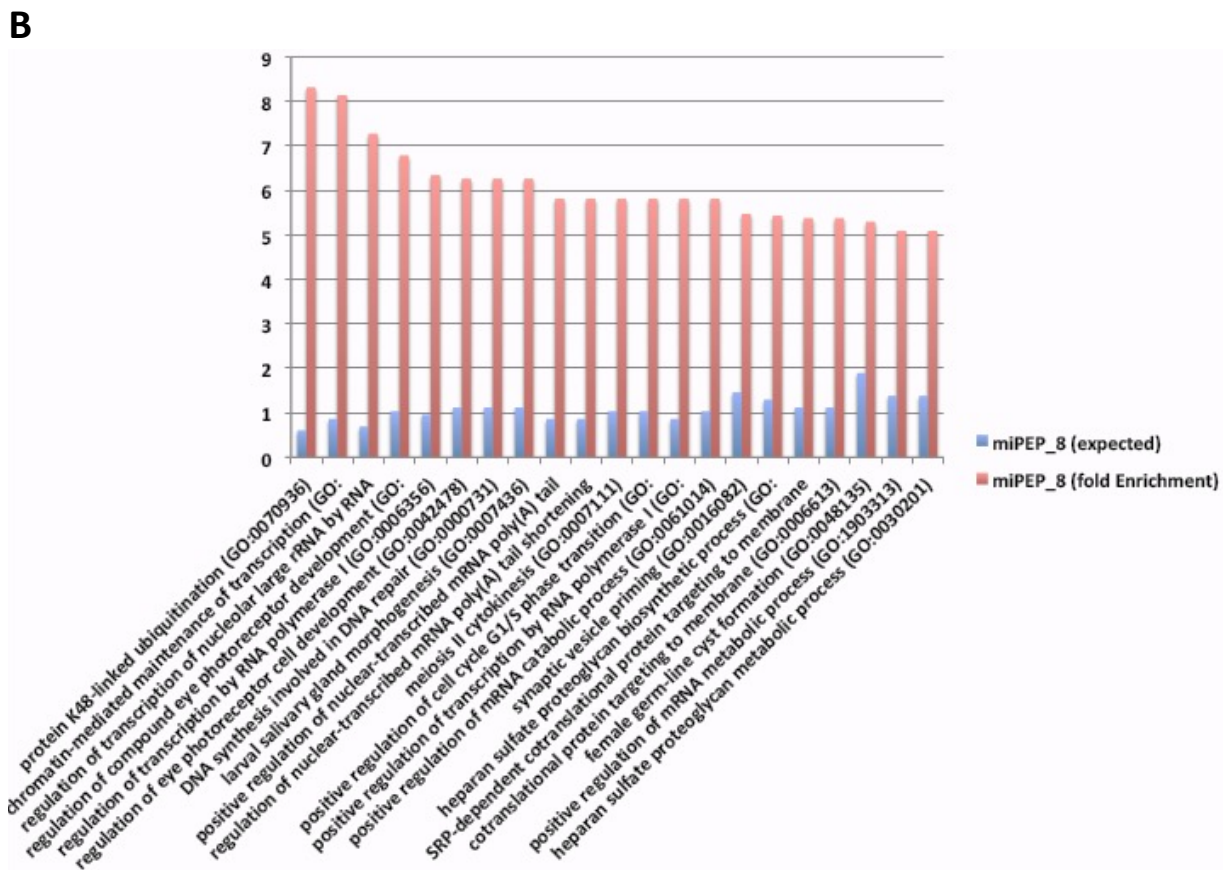
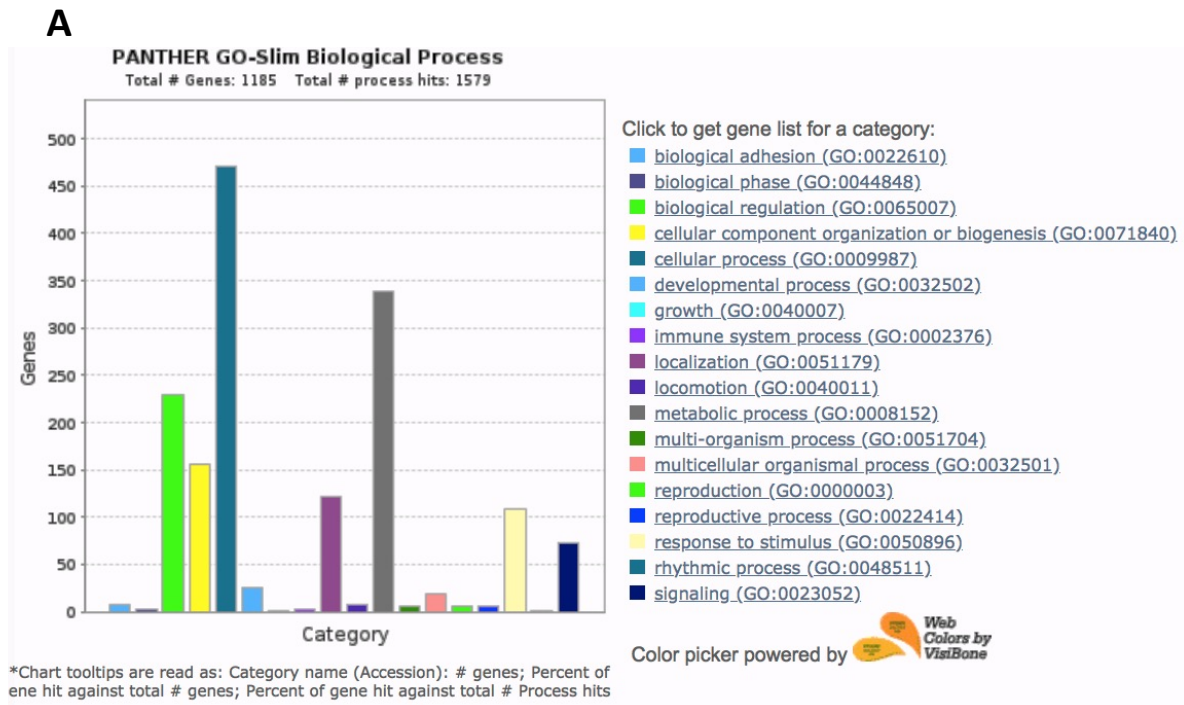


Figure S11 : Gene Ontology of transcriptome of miPEP-8 specific genes. **A** : Most significant regulated biological processes. **B** : Fold enrichment of the most representative biological activities.

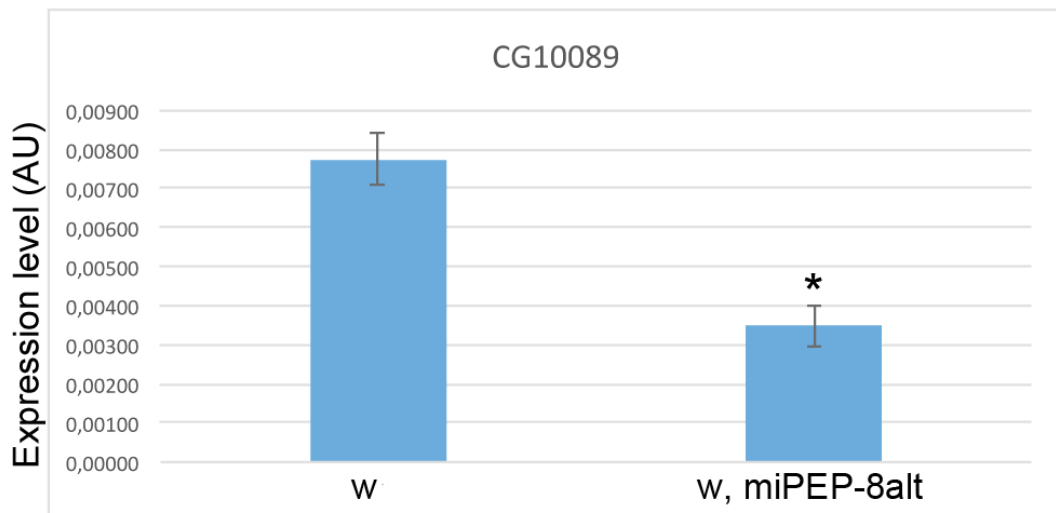
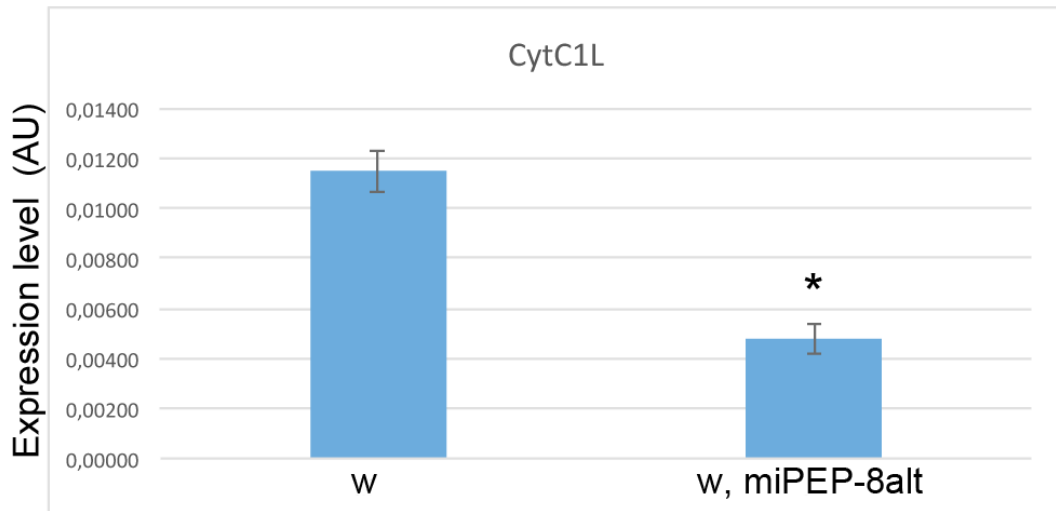


Figure S12 : Modulation of two miPEP-8 regulated genes analysed by qPCR in white (wt miPEP-8) and in w ; miPEP-8 mutated adult flies (miPEP-8alt) (N= 6 and 8 respectively). The differences are significant ($p < 0,05$).

AGATCGTGAAGAAGCGCACCAAGC
 GCACCAGGAACTTCTTGAATCCGG
 CGAGACCTACTGCATCGACA
 AGGTCACCGTATGTGGGTGT
 AGGACACGGACGACATCTTT
 TCTTACCGGGCAGCATTAGA
 TCGCCTTGAGAACTTTGAGC
 TGATTTGTTTGGATTTTACGC
 TTGTTTGTTCGCTTTGCTTTG
 GAGCCTGGCTTTGTTTTTGT
 GGGGGTAATACTGTCAGGTAAA
 CCAGTGCAGGGTCCGAGGTA
 GCTTCGGCTTAATGATGGTC
 GGGTGTGATTCTGCTTGCA
 GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACAGTCAG
 GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACGATCAG
 GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACGACATC
 CTCAGAACCGGAGACCGAC
 TCTTGCGAGACTTGAGCGTT
 GGAGCAACTGGATCGCACTA
 GCAGTTACCCTCGCAGATGT

RP49 q5
 RP49 q3
 Dm tub q5
 Dm tub q3
 premiR8 q3
 premiR8 q5
 primiR8 q5
 primiR8 q3OK
 primiR8 q3 quatuor
 PrimiR8q5quatuor
 New mir8fwqRT
 New universal rv stemloop
 U14fwqRT
 snoR442fwqRT
 stemloopRTU14
 stemloopRTsnoR442
 SL RT miR8
 Cyt C1L Fw
 Cyt C1L Rv
 CG10089 Fw
 CG10089 Rv

Figure S13 : sequences and list of primers used in real time quantitative PCR experiments.