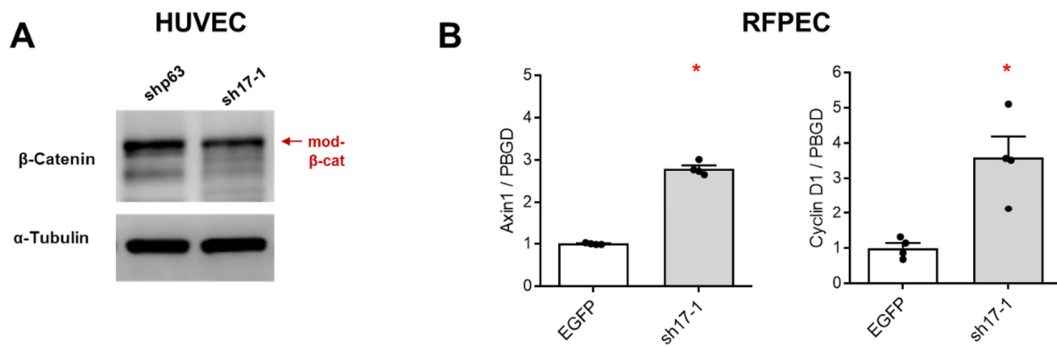


Suppl. Figure 1

Figure S1: RhoGEF17 is essential for cell-cell contacts and AJ protein regulation in EC. **A)** Shown is the schematic domain structure of RhoGEF17. The positions of the two shRNA target sequences in the PH domain are depicted. ABD = actin binding domain, DH = Dbl homology domain, PH = pleckstrin homology domain, WD40 = domain with seven WD40-related repeats. **B)** RFPEC were transduced for 48 h. Shown are representative immunoblots of RhoGEF17 and α -tubulin and the quantitative analysis. The values were normalized by α -tubulin and are given relative to the respective EGFP controls (not shown). Presented are the means+SEM and the single data points, $n=12-24$, $*p<0.05$ vs. the respective EGFP controls assessed by paired t-testing. **C)** Imaging of semi-efficiently transduced HUVEC was performed after 48 h. Depicted are bright field/EGFP overlay images. Scale bar = 100 μ m. In addition, an immunoblot of RhoGEF17 and α -tubulin is shown, demonstrating the knockdown efficiency of sh17-2. **D)** Imaging of semi-efficiently transduced RFPEC was performed. Depicted are bright field/EGFP overlay images of different time points.

Scale bar = 100 μm . **E)** RFPEC were transduced for 48 h and then used to generate spheroids. Bright field and fluorescence images are shown. Scale bar = 200 μm . **F)** Immunoblot analysis of N-cadherin, p120-catenin and α -tubulin was performed. Shown are representative immunoblots and the quantitative analysis. The values were normalized by α -tubulin and are given relative to the respective EGFP controls (not shown). Presented are the means+SEM and the single data points, n=6-21, *p<0.05 vs. the respective EGFP controls assessed by t-testing. **G)** Immunoblot analysis of RhoGEF17, pan-cadherin, p120-catenin, and β -actin was performed. Shown are representative immunoblots.



Suppl. Figure 2

Figure S2: RhoGEF17 regulates β -catenin. **A)** HUVEC were transduced for 48 h. β -catenin and α -tubulin were detected by immunoblot. **B)** RFPEC were transduced for 48 h and qPCR of axin1, cyclin D1, and the housekeeping gene PBGD was performed. Given are means +SEM of 4 independent experiments, * $p < 0.05$ vs. EGFP assessed by paired t-test.

Primer sequences:

axin 1

forward CCA GTG CCA ATG ACA GTG AG

reverse CCT TCG GTG CTG CTT ACG

cyclin D1

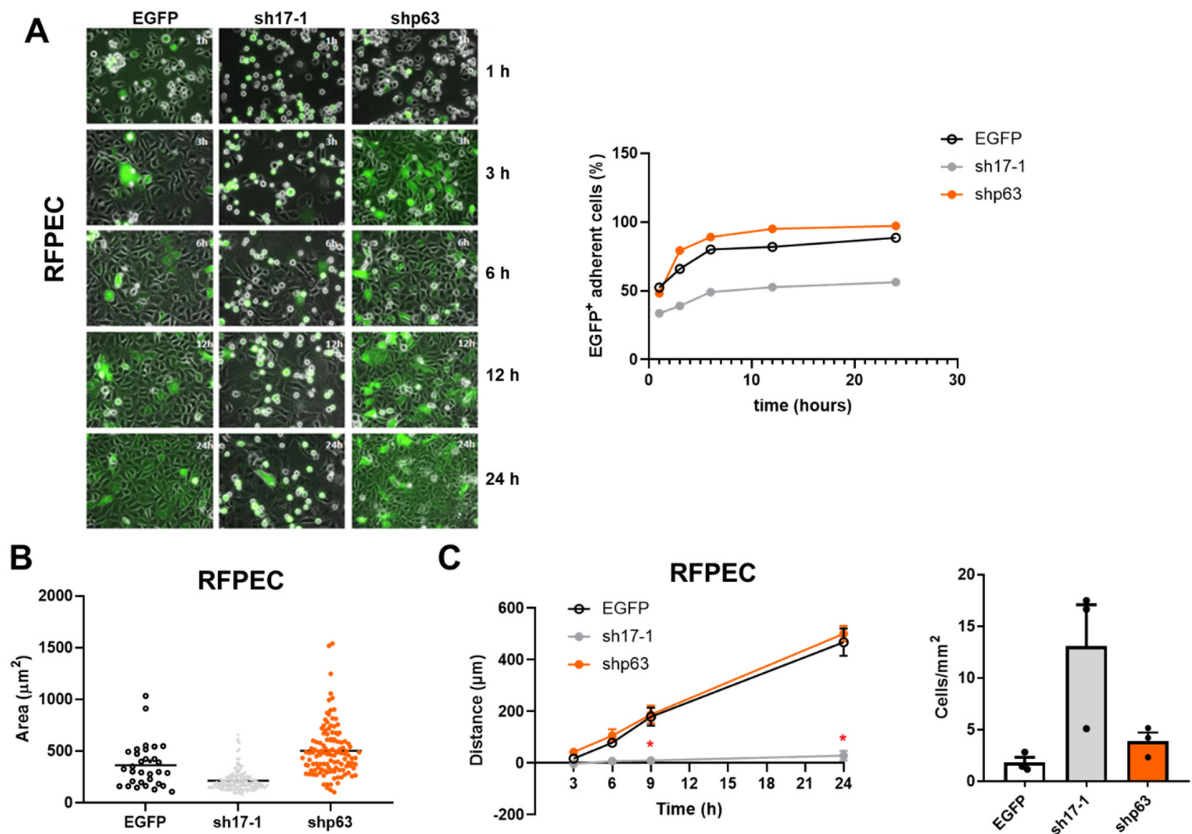
forward CAC CAA TCT CCT CAA CGA C

reverse CAC AGA CCT CCA GCA TCC

PBGD

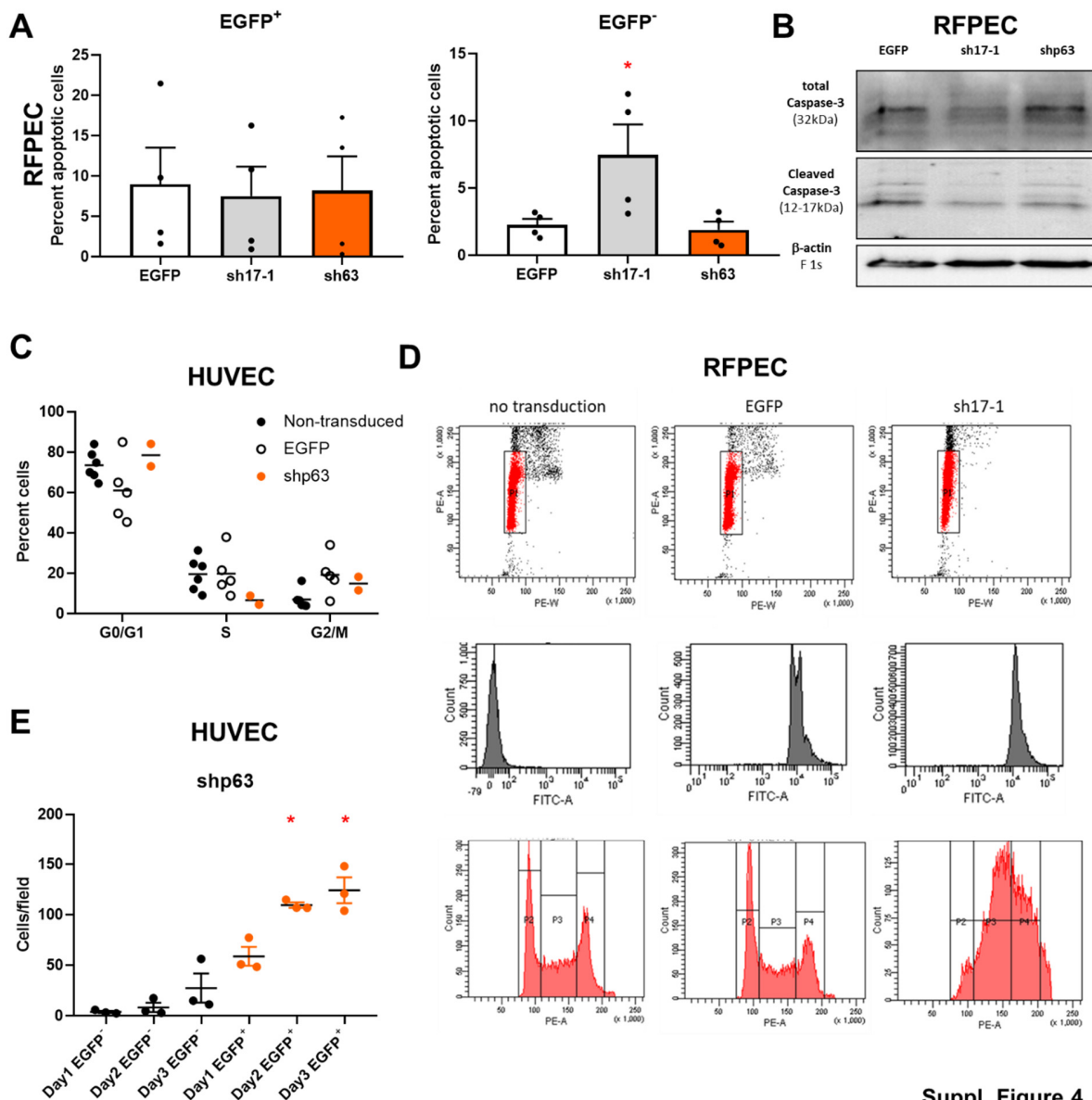
forward CCT GAAACTCTGCTTCGCTG

reverse CTGGACCATCTTCTTGCTGAA



Suppl. Figure 3

Figure S3: The shp63 adenovirus has no effect on RFPEC adhesion, cell size, and sheet migration. RFPEC were transduced for 48 h with the indicated adenoviruses. **A)** The cells were detached and reseeded. Adhesion was monitored by fluorescence microscopy over a time course of 24 h. Depicted are the percentages of transduced (EGFP⁺), adherent cells given as mean, n=1-2. **B)** The surface area of the cells was determined at the end (24 h) of the adhesion assay. Given are the sizes of the measured cells of 1-2 independent experiments. **C)** Confluent transduced cells were scratched and imaged at the indicated time points. The migration distance of the sheet was measured. Given are the quantified data as mean±SEM, n=3, *p<0.5 vs. EGFP assessed by 2-way ANOVA with Tukey's multiple comparison test. Single, EGFP⁺ cells in the wound were counted at the end of the assay. The number of cells per mm² are given as means+SEM, n=3.



Suppl. Figure 4

Figure S4: The shp63 adenovirus has no effect on EC apoptosis, caspase 3 expression, and cell cycle. RFPEC (A, B, D) or HUVEC (C, E) were transduced with the respective viruses for 48h or the indicated times (E). **A**) Annexin V staining was performed in semi-efficiently transduced cells and Annexin V positive cells were counted and are given in percent of EGFP⁺ and EGFP⁻ cells, n=4, *p<0.05 vs. EGFP assessed by 1-way ANOVA with Tukey's post test. **B**) A representative immunoblot of caspase 3 and cleaved caspase 3 together β -actin as loading control is shown. **C-D**) Cell cycle analysis was performed by FACS with PI-stained HUVEC (C) or RFPEC (D). **C**) Analysis of the FACS results for HUVEC, n=2-6. **D**) Cell gating strategy is shown. Upper row: Living single cells, middle row: EGFP-positive cells, bottom row: PI staining vs. cell count. **E**) A proliferation analysis of shp63RhoGEF-transduced HUVEC is presented. EGFP⁻ and EGFP⁺ cells were counted from microscope images starting one day after transduction, n=3, *p<0.05 vs. Day1 EGFP⁺ assessed by 1-way ANOVA with Tukey's post test.