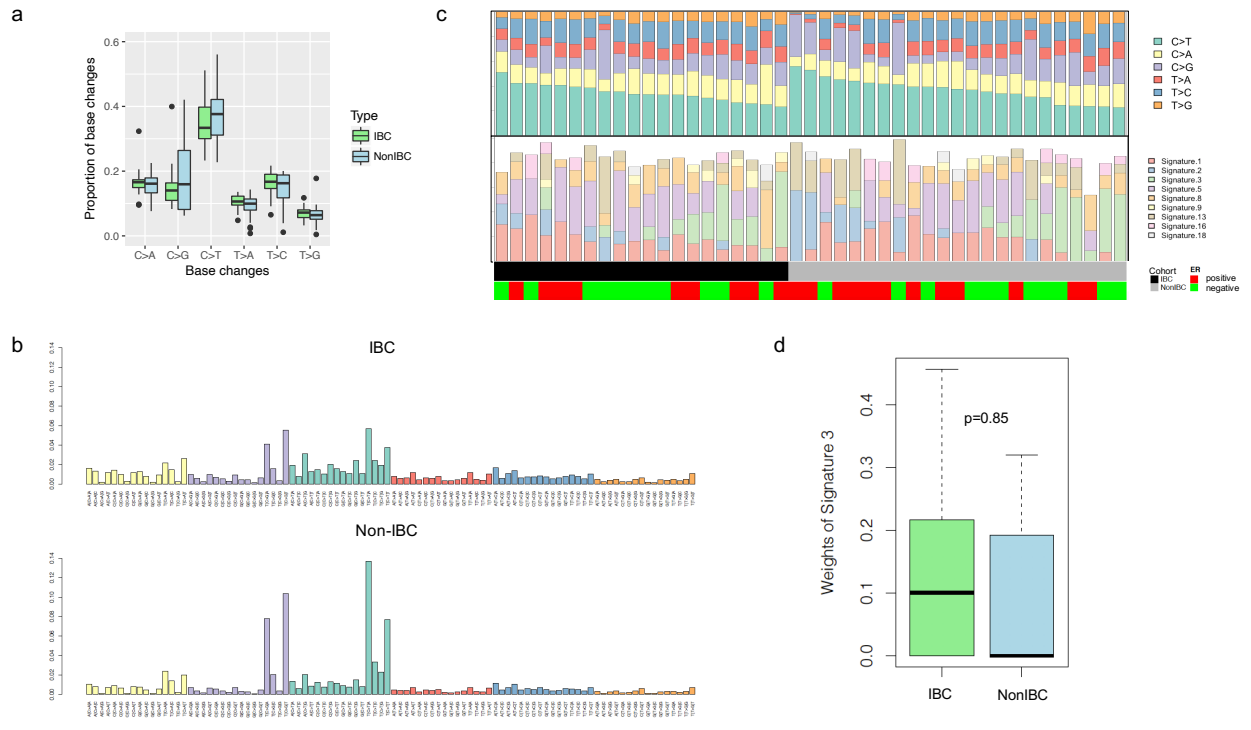


1 **Fig S1**  
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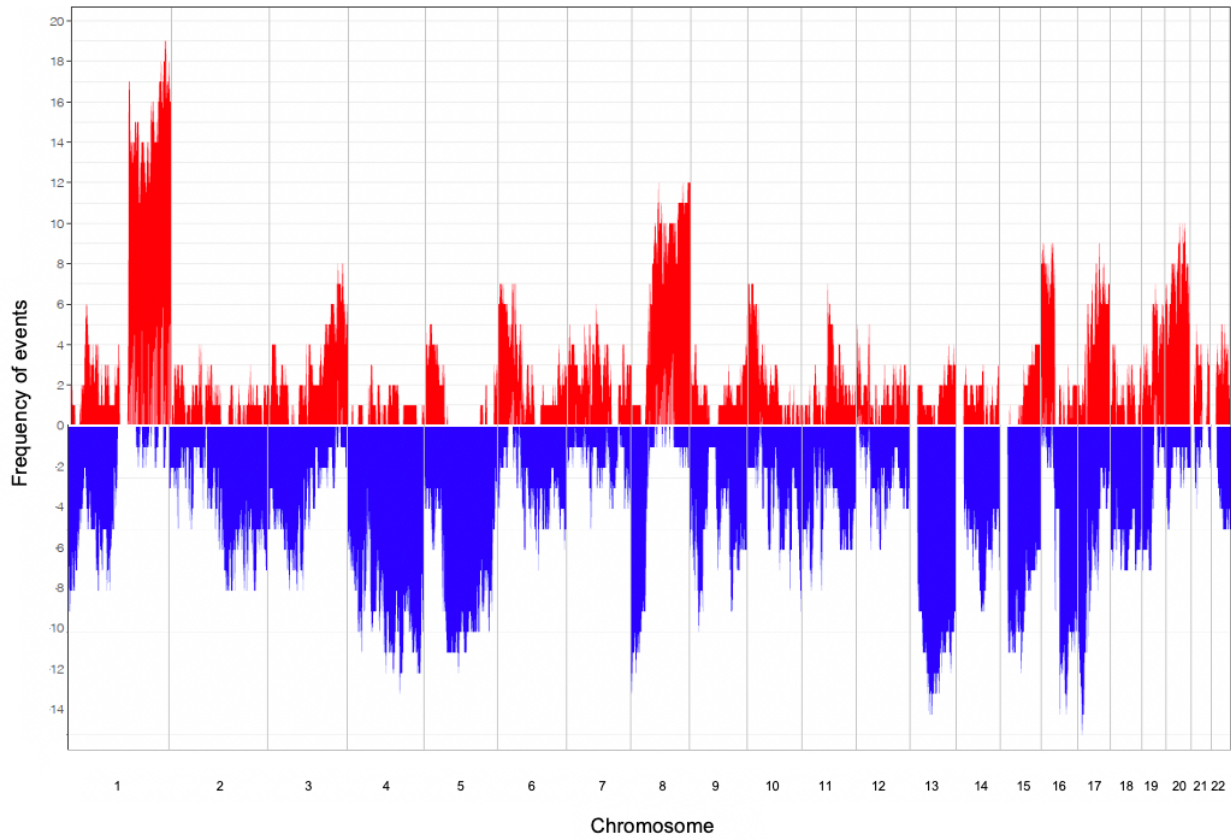
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 6 **Fig S1. Mutation spectra and mutational signatures**

7 **a.** Proportion of somatic base changes in IBC and non-IBC cohorts. **b.** Mutation spectra of IBC and non-IBC cohorts.  
 8 **c.** Proportion of somatic base changes and weights of nine reference mutational signatures, for individual IBC and  
 9 non-IBC cases, annotated with ER status. **d.** Comparison of weights of signature 3, which is associated with  
 10 homologous recombination defect, in IBC and non-IBC cohorts. For panel a (base change), c and d (weights of  
 11 signatures), two cohorts were compared by Wilcoxon rank sum test. Subsequent p-values were adjusted by  
 12 Bonferroni method. All adjusted p-values>0.05.

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1 **Fig S2**

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5 **Fig S2. Somatic copy number profile of the non-IBC cohort**

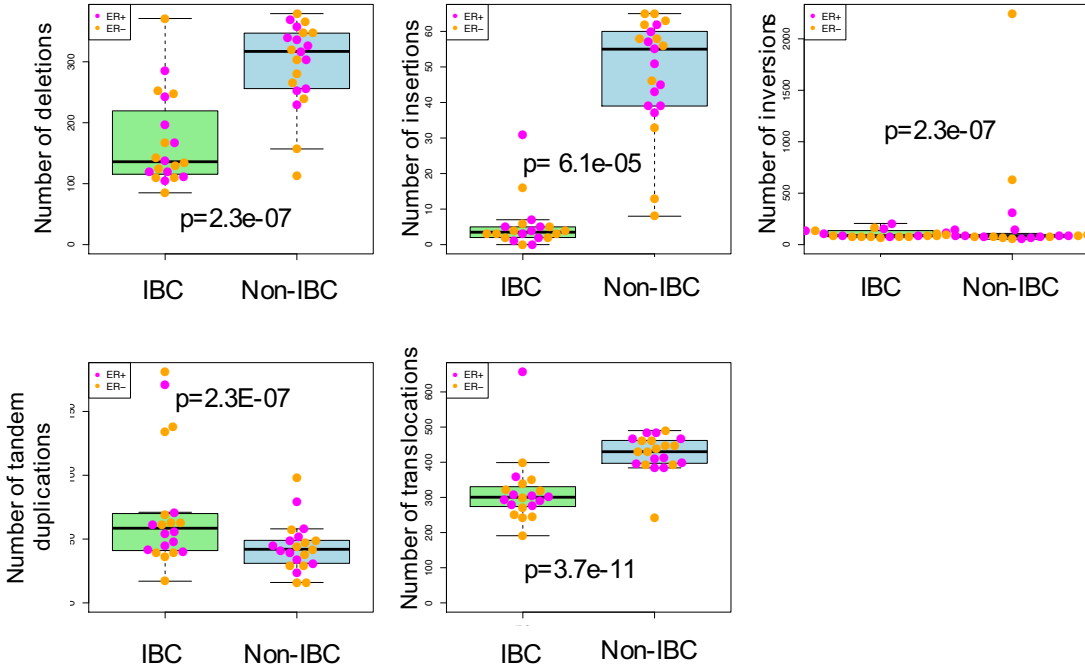
6 X-axis represents genome coordinates ordered by chromosomes. Y-axis shows the frequency of copy number gain

7 (red) and copy number loss (blue) in 1Mb-length bins across the genome in non-IBCs.

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1 **Fig S3**

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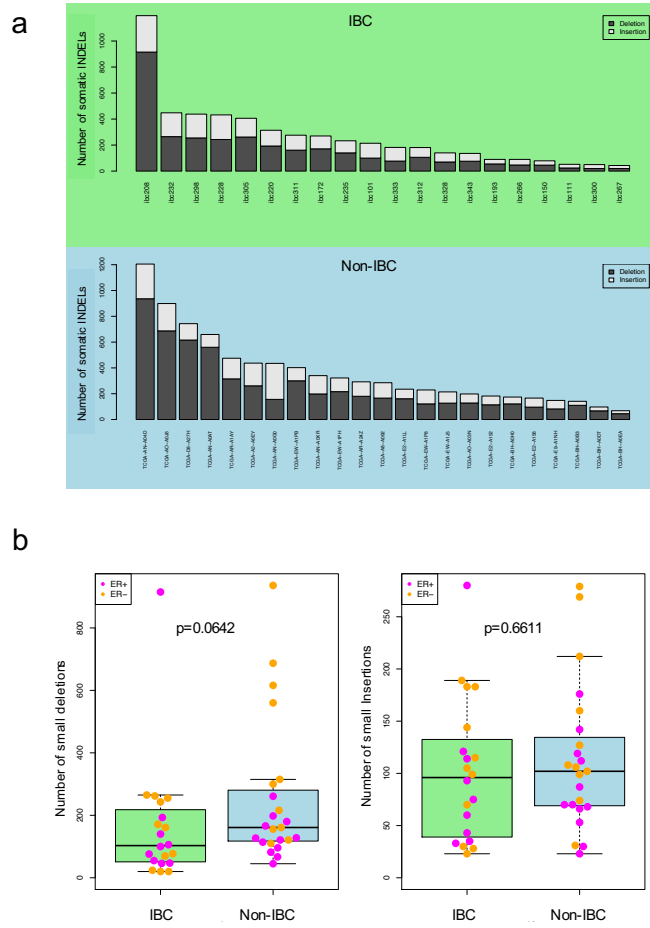
5 **Fig S3. Number of somatic structural variants in IBC and non-IBC cohorts**

6 Number of somatic SVs in each category in IBC and non-IBC cohorts. Each dot represents a sample color-coded by  
7 its ER status. P-values were calculated by Wilcoxon test and adjusted by Bonferroni method.

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1 **Fig S4**

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5 **Fig S4. Number of small insertions and deletions (INDELS) in IBC and non-IBC cohorts**

6 a. Number of small INDELS in individual IBC and non-IBC samples. Shades represent the types of small INDELS. **d.**

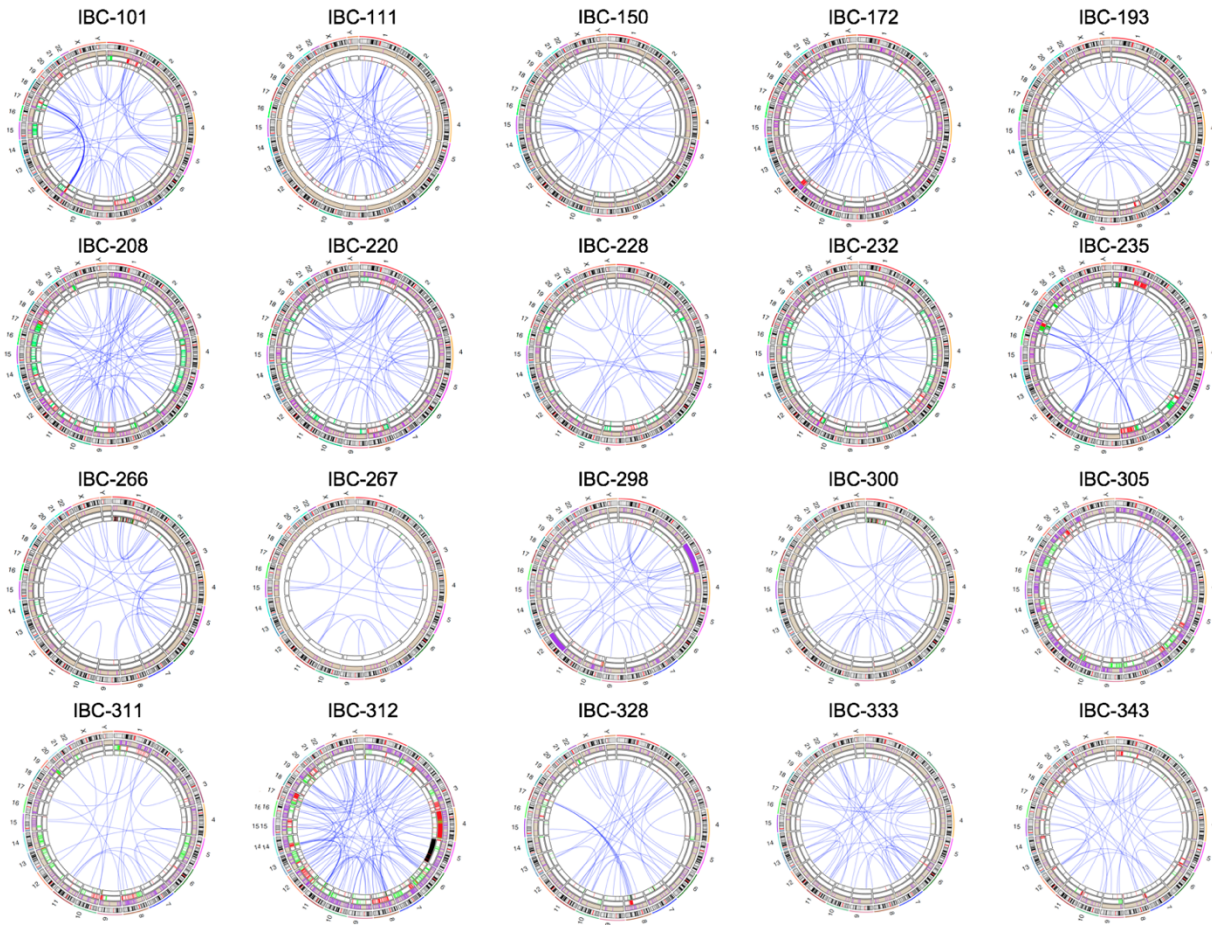
7 Number of small deletions and small insertions in IBC and non-IBC cohorts. Each dot represents a sample color-

8 coded by its ER status. P-values were calculated by Wilcoxon test.

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1 **Fig S5**

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6 **Fig S5. Circos plots of individual IBC cases**

7 The outer ring represents each chromosome ideogram, the next ring is the somatic SNV (purple), the next ring is

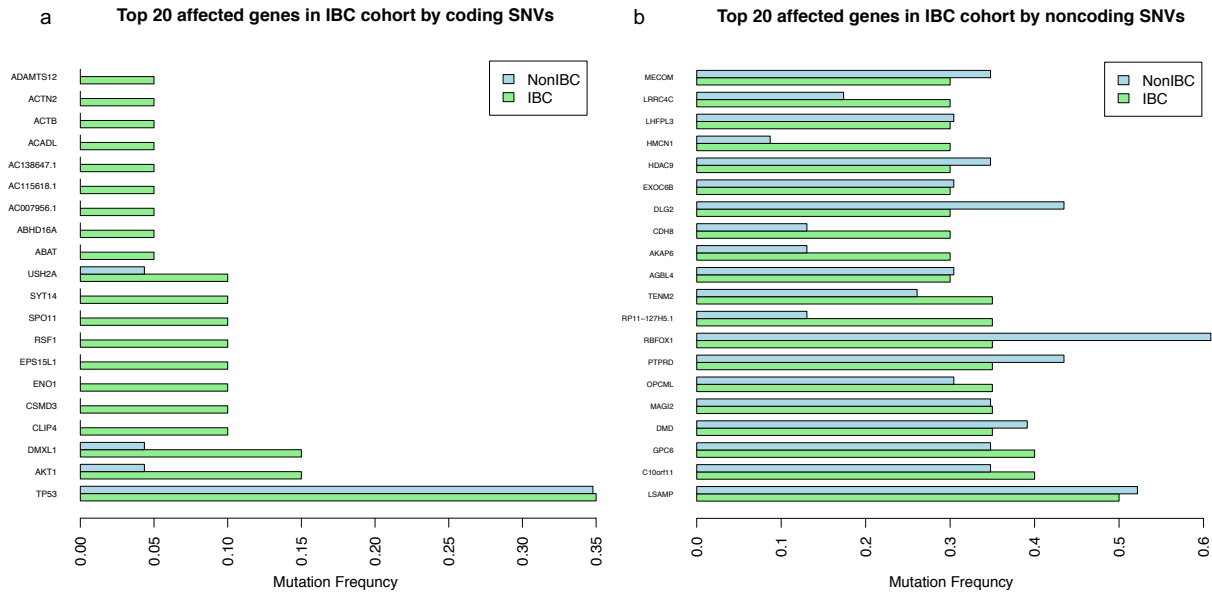
8 somatic CNV (red: gain; green: loss), the inner ring is somatic SV (red: tandem duplication; green: deletion; black:

9 inversion). The interchromosomal translocations are shown by the lines in the center of the plot.

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1 **Fig S6**

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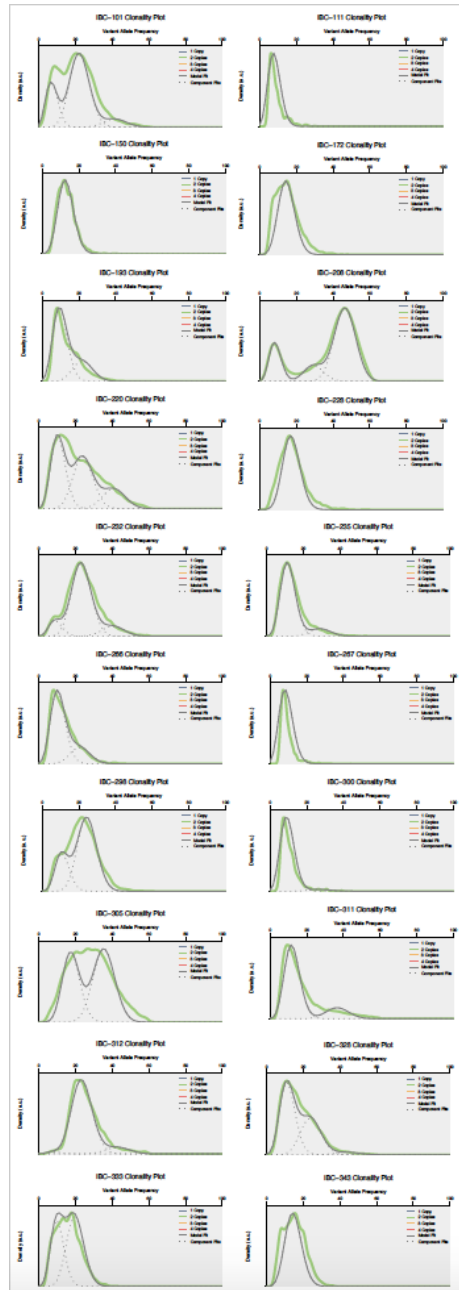
5 **Fig S6. Top 20 most frequently affected genes by coding and non-coding SNVs**

6 **a.** Top 20 most frequently affected genes by coding SNVs in IBC. Green and blue bars represented mutation  
 7 frequency of a given gene in IBC and non-IBC cohort, respectively. **b.** Top 20 most frequently affected genes by  
 8 noncoding SNVs in IBC. Green and blue bars represented mutation frequency of a given gene in IBC and non-IBC  
 9 cohort, respectively. For all genes in **a** and **b**, their mutation frequencies were not significantly different between IBC  
 10 and non-IBC cohorts (Adjusted  $p$ -values > 0.05, Fisher's exact test, Bonferroni correction).

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1 Fig S7

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6 **Fig S7. Clonality Plot for each individual IBC sample**

7 Clonality plot presented the distribution of variant allele frequencies for all input somatic SNVs for each sample (green  
8 line), as well as the model fit (grey solid line) and the component fits (grey dashed line) results from the SciClone  
9 method.

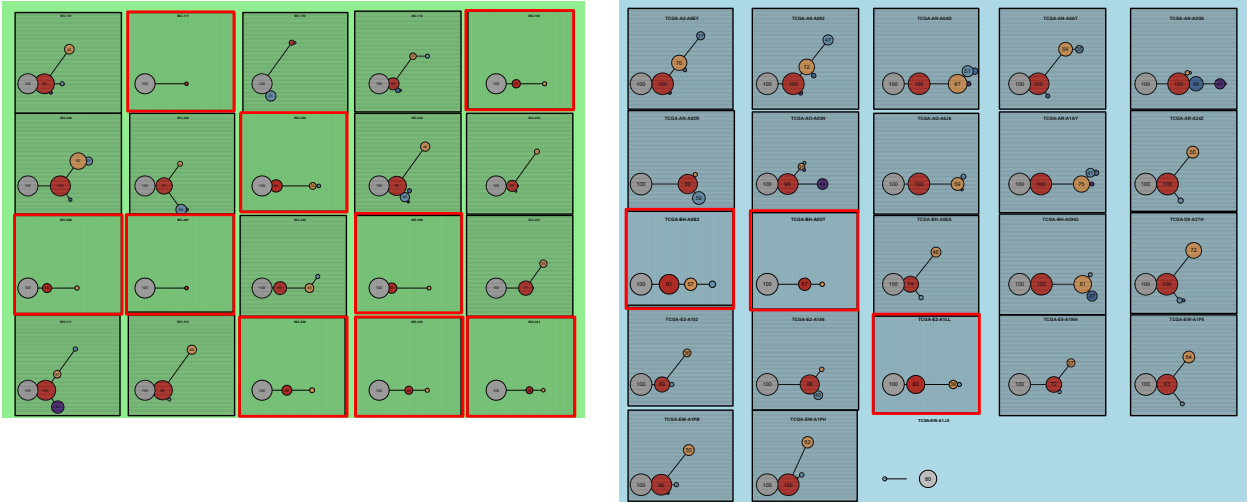
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1 **Fig S9**

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6 **Fig S9. Individual evolutionary trees of IBC and non-IBC cohorts**

7 Each sample is shaded with horizontal or vertical lines to indicate branching and linear pattern, respectively. Samples  
8 belonging to the linear group are also highlighted by red.

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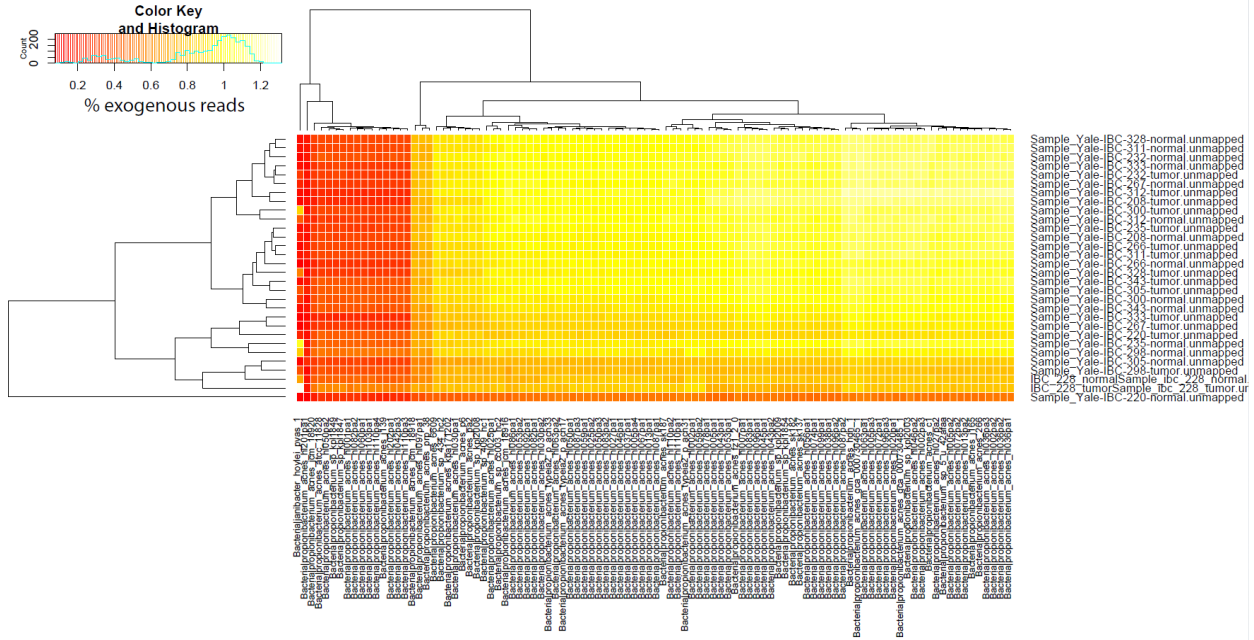
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1 **Fig S10**

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7 **Fig S10. Top 100 most frequent non-human sequences in the IBC cancer and normal DNA**

8 Each row is one IBC cancer or normal sample. Each column is one microorganism. Color scales correspond to the  
9 percentage of exogenous reads detected in a given sample, for the indicated microorganism.

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