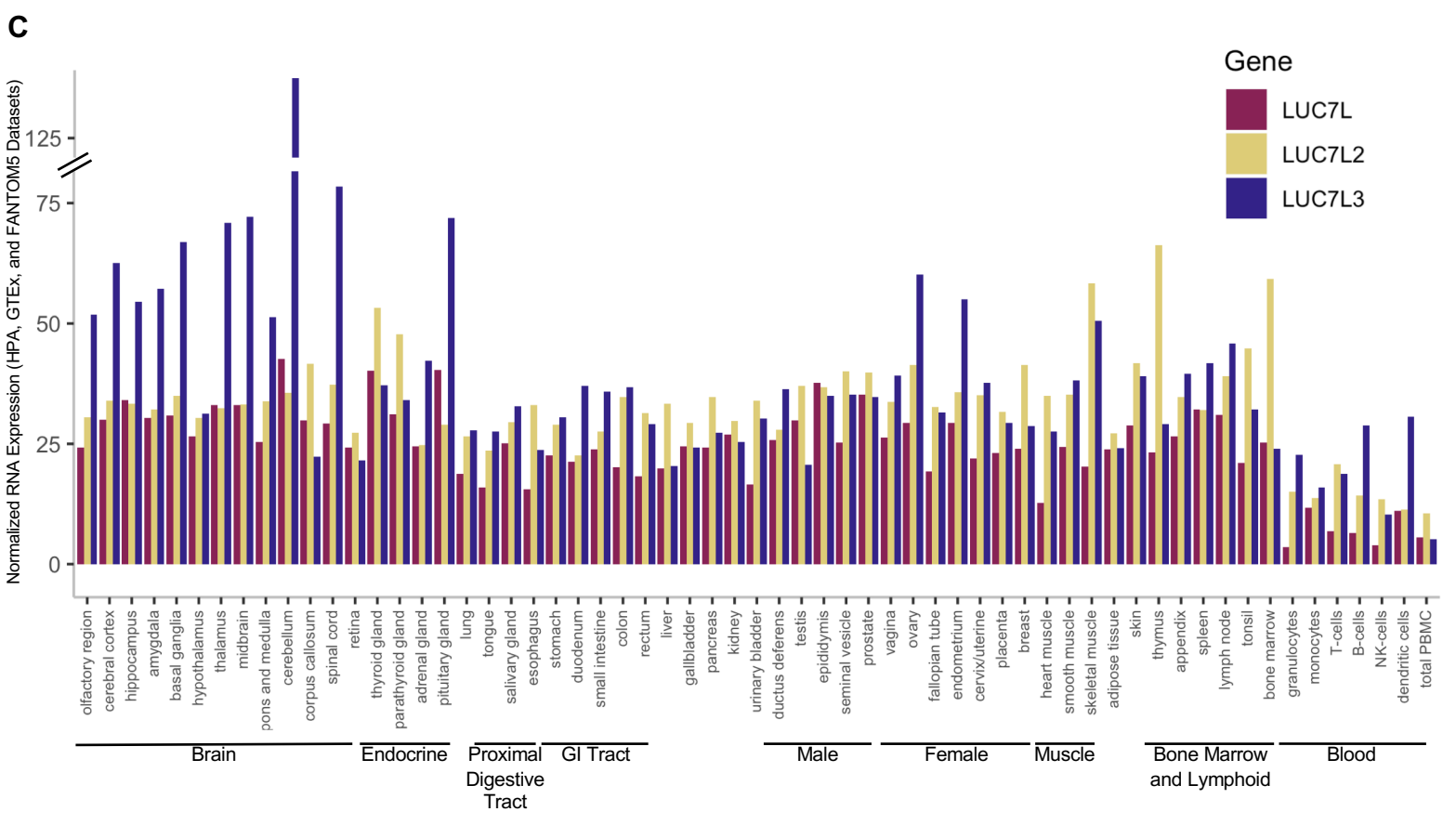
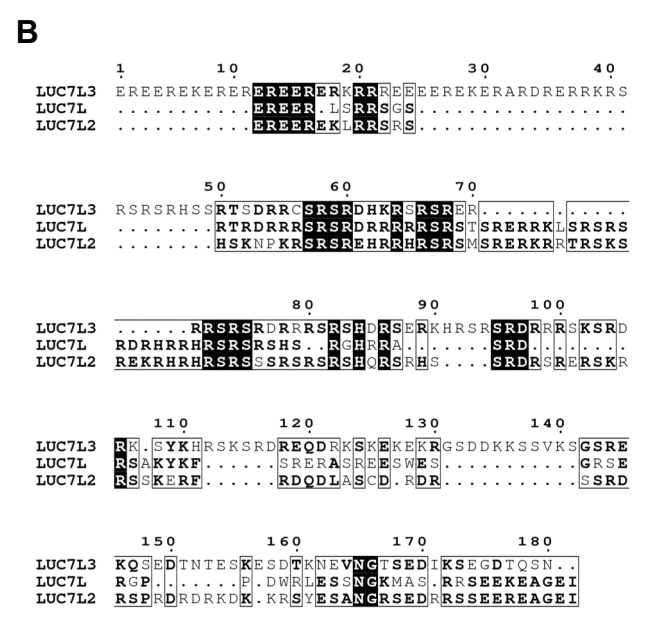
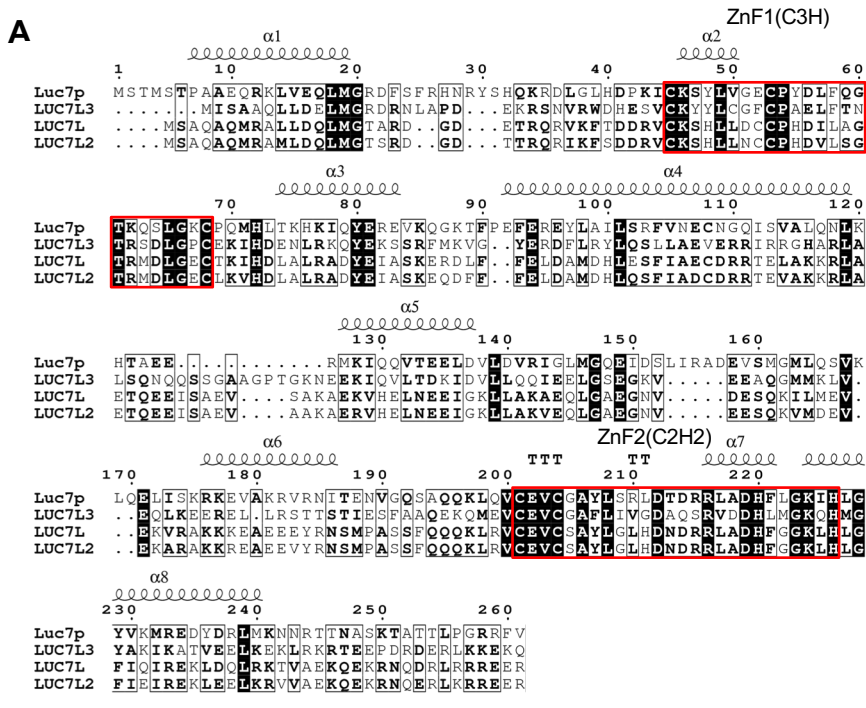


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**Supplemental information**

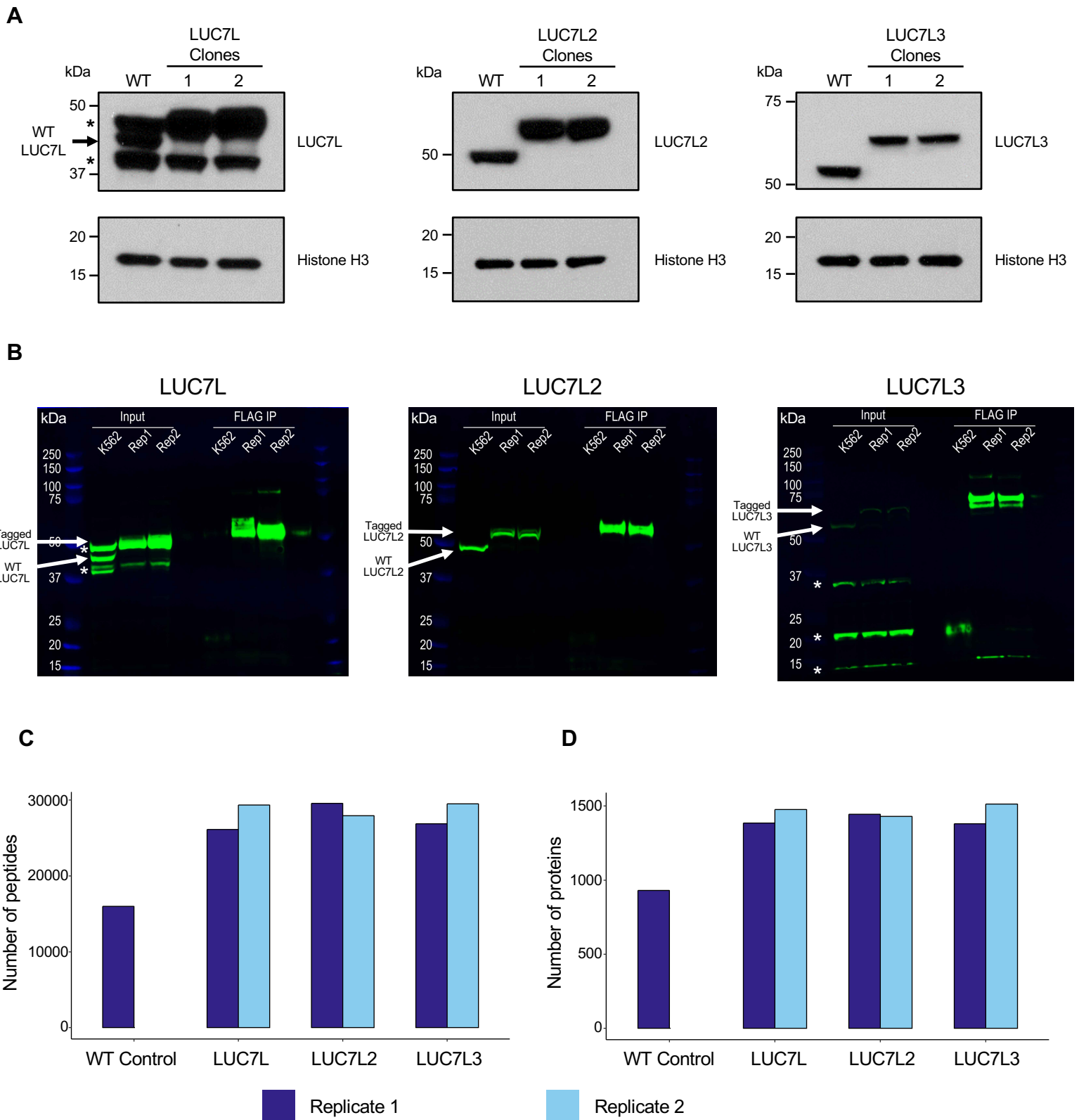
**Functional analyses of human LUC7-like proteins  
involved in splicing regulation  
and myeloid neoplasms**

**Noah J. Daniels, Courtney E. Hershberger, Xiaorong Gu, Caroline Schueger, William M. DiPasquale, Jonathan Brick, Yogen Sauntharajah, Jaroslaw P. Maciejewski, and Richard A. Padgett**



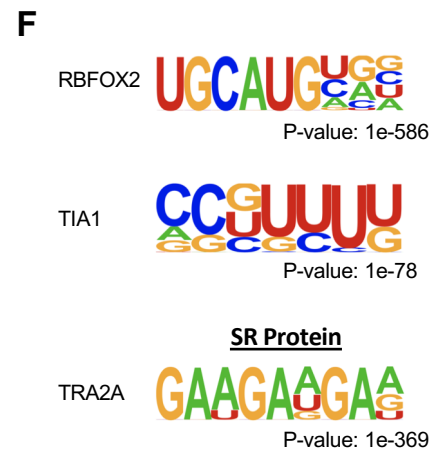
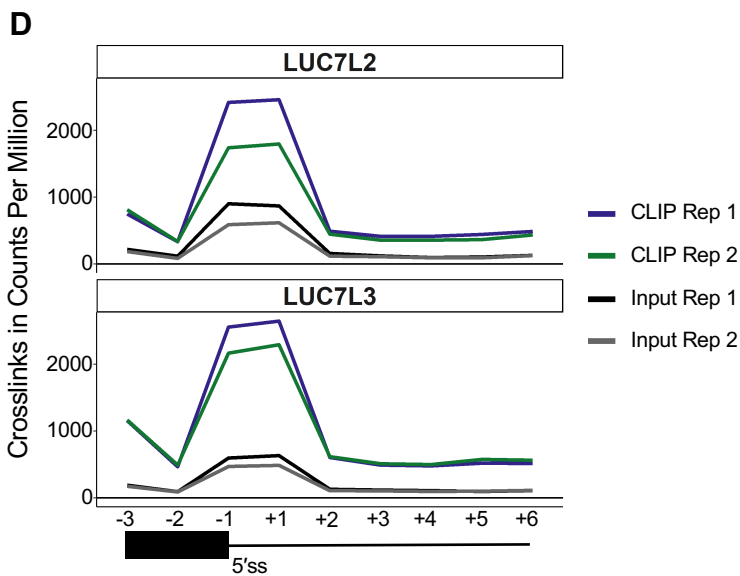
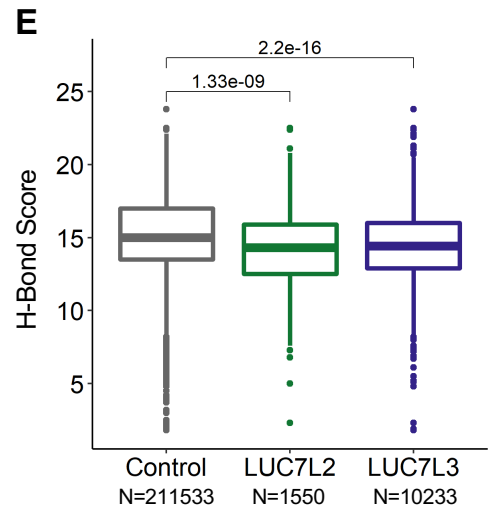
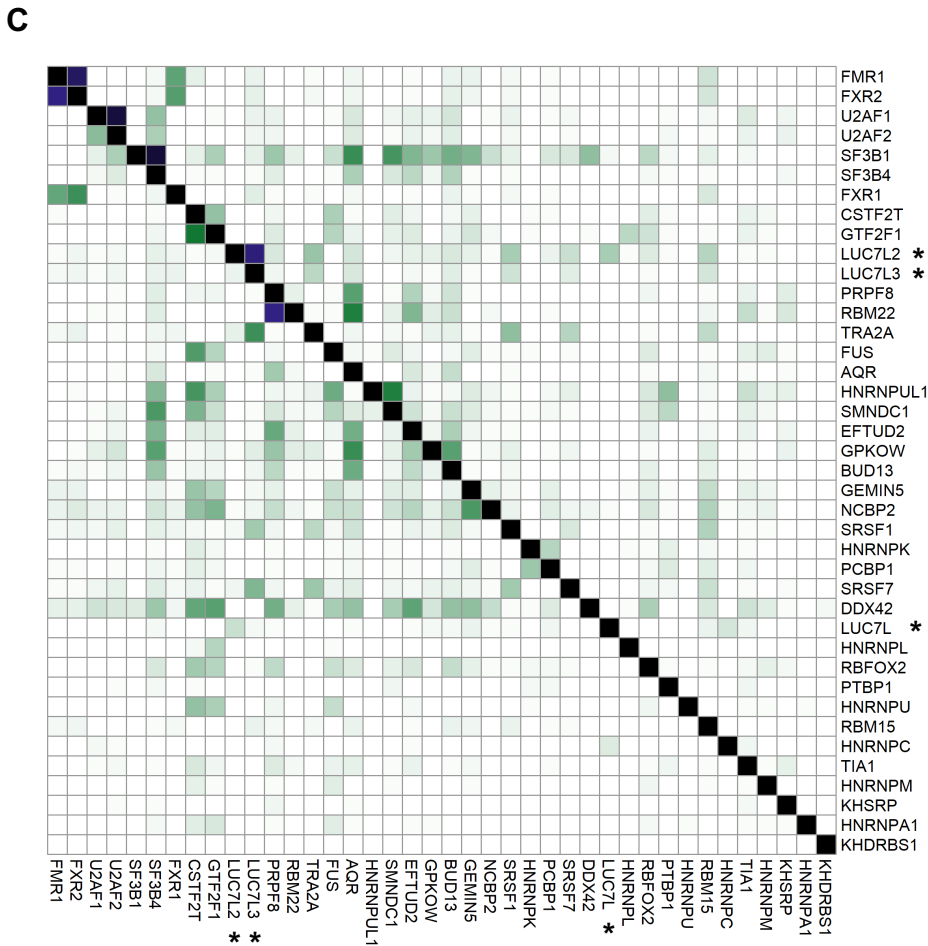
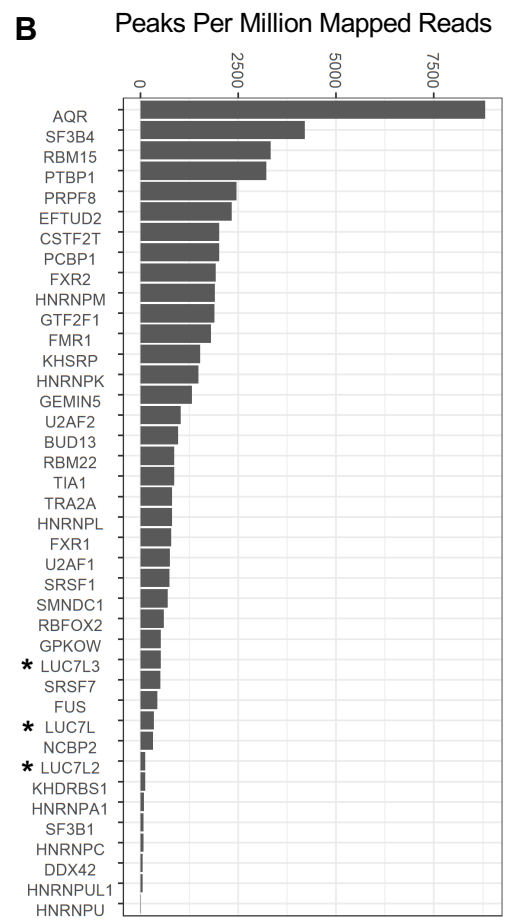
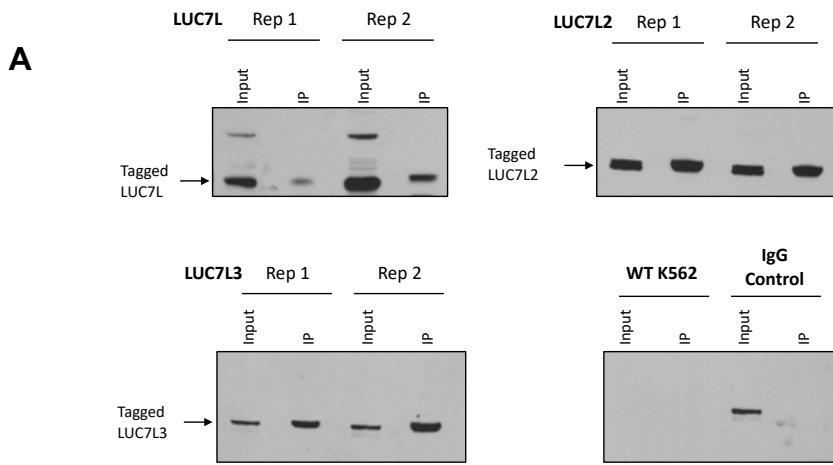
**Figure S1: Amino acid conservation and tissue specific expression of the LUC7-like proteins. Related to Figure 1.**

**(a)** Amino acid alignment of the N-terminal domains of the mammalian LUC7-like proteins along with the paralogous yeast protein Luc7p using CLUSTAL OMEGA 1.2.4 multiple sequence alignment tool and ESPrpt 3. Depicted are the conserved N-terminal  $\alpha$ -helix, ZnF1 (CH3 type containing three cysteines, one histidine) and ZnF2 (C2H2 type containing two cysteines, two histidines). The structure of the coiled-coil domain is located between the two ZnFs. White letters with black background represent 100% conservation among the four proteins. Black letters with black frame represent conservation among three proteins.  $\alpha$  and  $\beta$ -turns are depicted as TT and TTT. **(b)** Amino acid alignment of the arginine-glutamic acid rich (RE), arginine-serine rich (SR), and arginine rich (R) domains of the mammalian LUC7-like proteins following the second zinc finger. White letters with black background represent 100% conservation among the three proteins. Black letters with black frame represent conservation between two proteins. **(c)** RNA Expression (TMP) of the *LUC7*-like family across tissues from HPA, GTEX, and FANTOM databases [Uhlen et al., 2015].



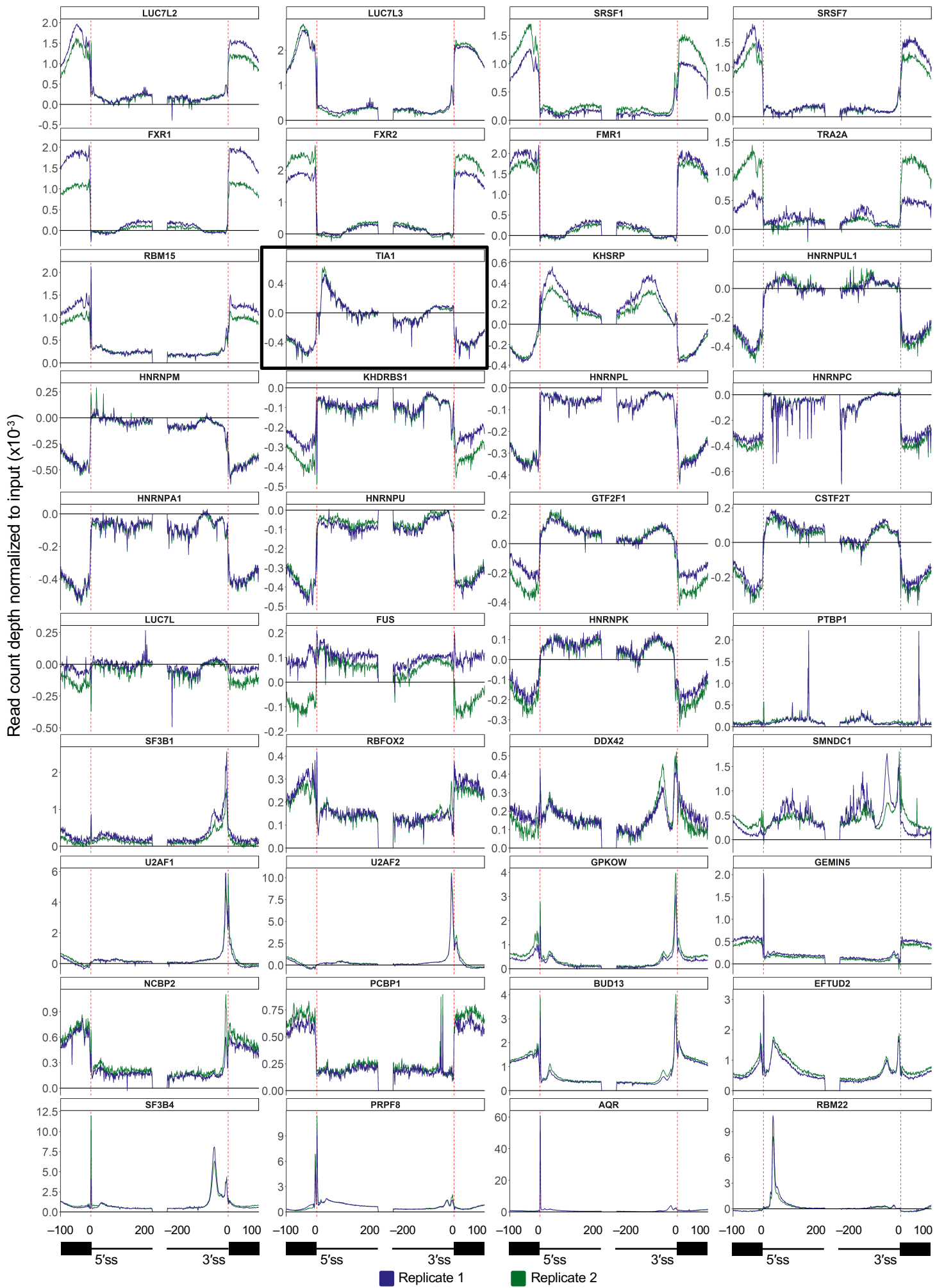
**Figure S2: Co-Immunoprecipitation of the LUC7-like proteins. Related to Figure 1.**

(a) LUC7-like genes homozygously CRISPR-tagged with V5, FLAG, HA in individual K562 clones shown by western blot (WB) using LUC7L, LUC7L2, and LUC7L3 antibodies, respectively. Epitope tagged LUC7L protein runs at the same size as the upper nonspecific band. Asterisks depict nonspecific bands. (b) Co-IP'd CRISPR-tagged LUC7-like cell lines using LUC7L, LUC7L2, and LUC7L3 antibodies respectively. WT and tagged proteins are depicted by arrows. Asterisks depict nonspecific bands. (c) Number of peptides analyzed in the Co-IP mass spectrometry experiments. (d) Number of proteins with at least one unique peptide identified from the Co-IP mass spectrometry experiments.



**Figure S3: seCLIP-Seq on CRISPR-tagged LUC7-like family members. Related to Figure 2.**

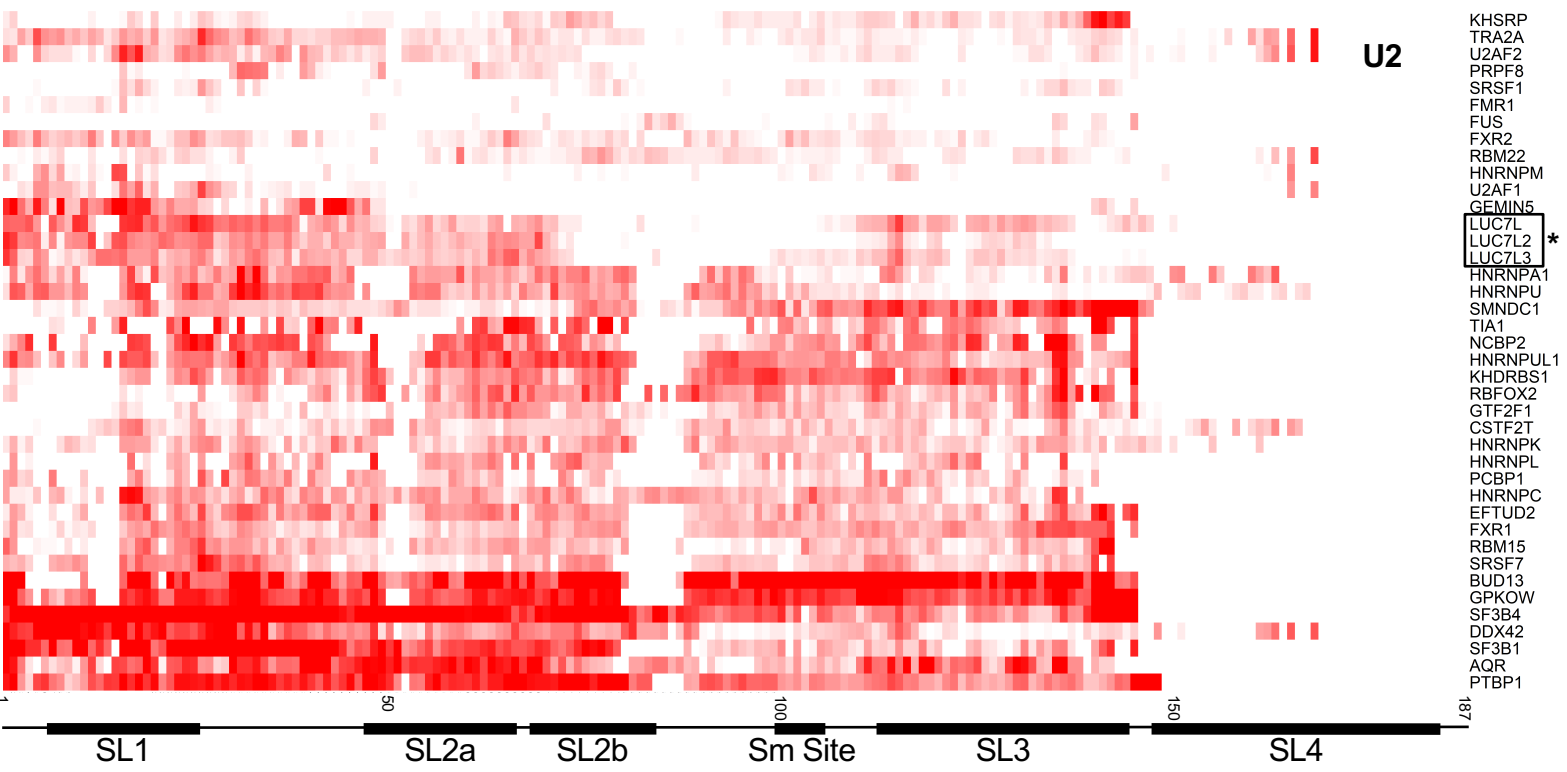
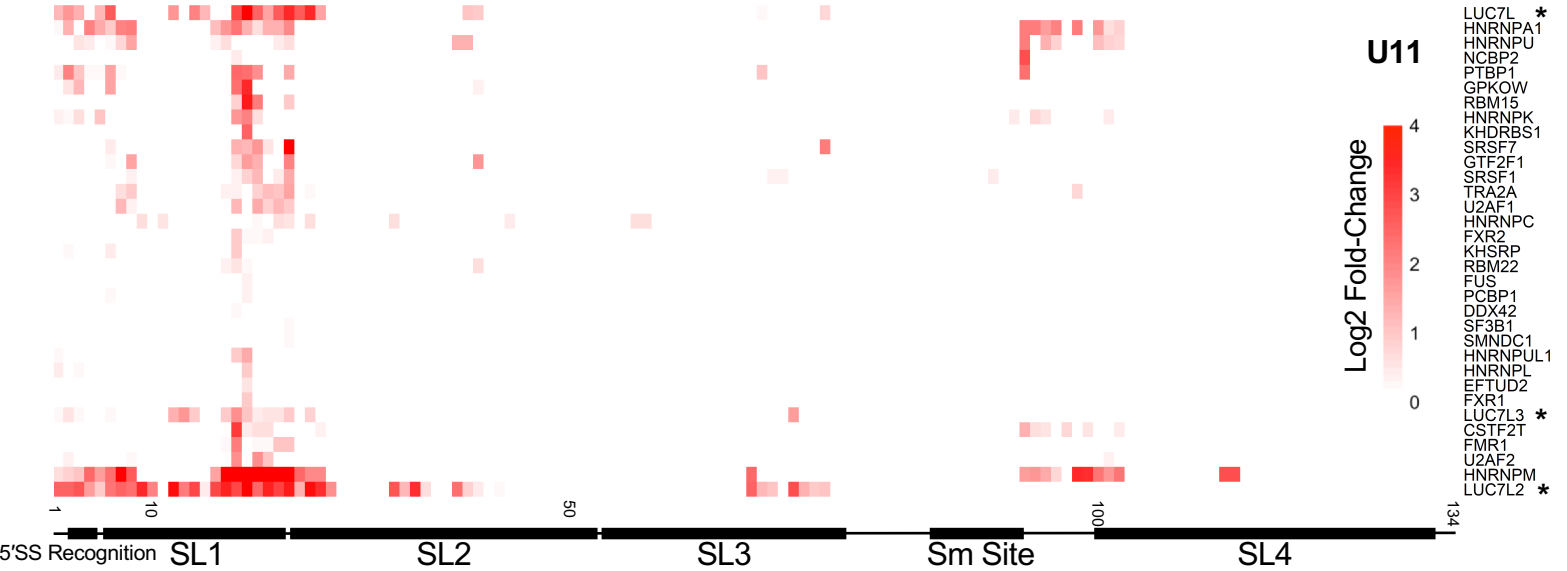
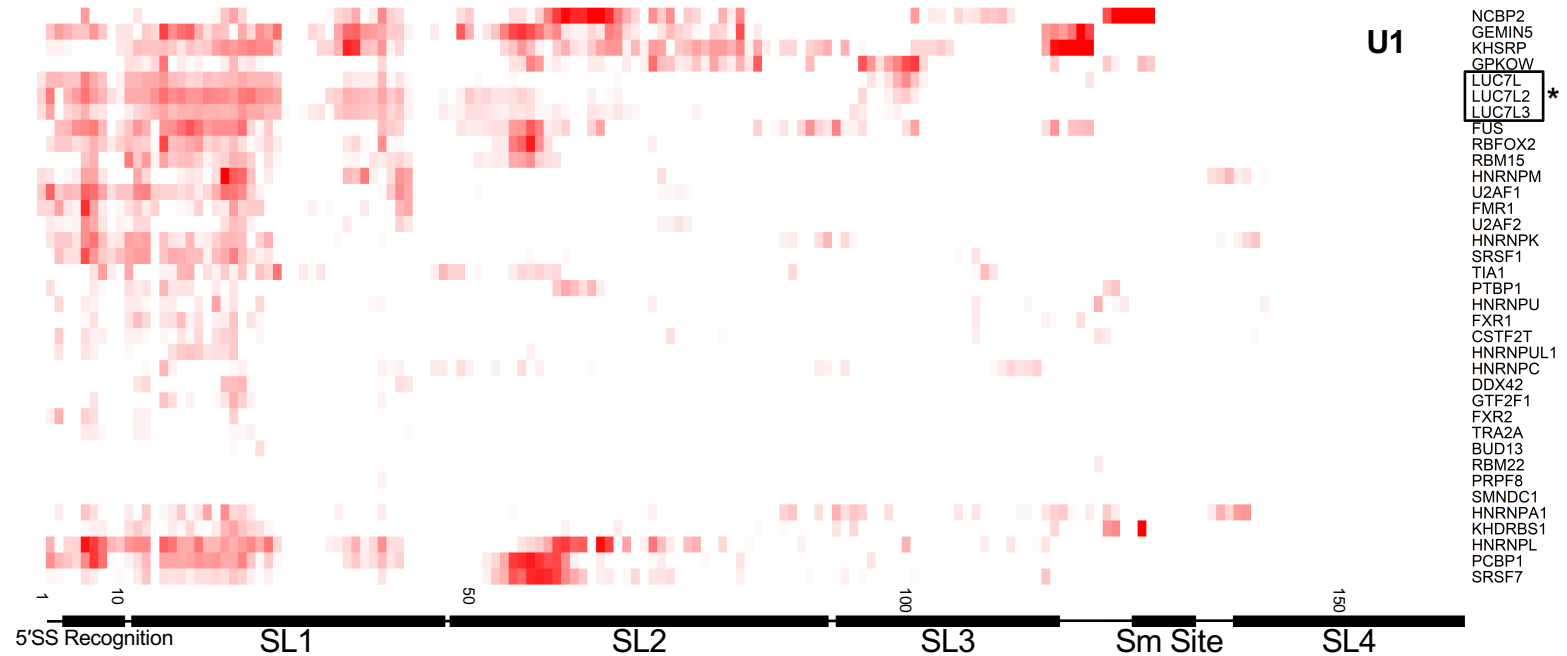
**(a)** CLIP-Seq: Input and V5-IP'd lysates in CRISPR-tagged cell lines and controls. **(b)** The number of reproducible and significantly enriched peaks per million mapped reads using the biological replicate containing the fewest uniquely mapped reads ( $\log_2$  fold-change  $\geq 3$ ,  $-\log_{10}$  p-value  $\geq 3$ , IDR  $\leq 0.01$ ) in CLIP-Seq experiments. **(c)** Proportion of significantly enriched peaks that overlap between the CLIP experiments. The LUC7-like experiments are depicted with a black asterisk. **(d)** CLIP crosslinking sites normalized to mapped library size in counts per million at a meta-5' splice site containing all 5'SS in the human genome. 5' splice site nucleotides are depicted as -3 – +8. Shown are CLIP and input replicates for LUC7L2 and LUC7L3. **(e)** U1 snRNA/5' splice site hydrogen bonding score using H-Bond tool. Control group contains all 5' splice sites used in CLIP-Seq mapping. LUC7L2 and LUC7L3 groups contain enriched 5' splice sites where there is an enriched crosslinking site at either position -1 or +1. A Wilcoxon rank-sum test was performed to determine significance. **(f)** Binding motifs enriched in significant CLIP-peaks identified in RBFOX3, TIA1, and the SR protein TRA2A.



**Figure S4: RBP binding profiles on generic 5'SS and 3'SS metagenes. Related to Figure 2.**

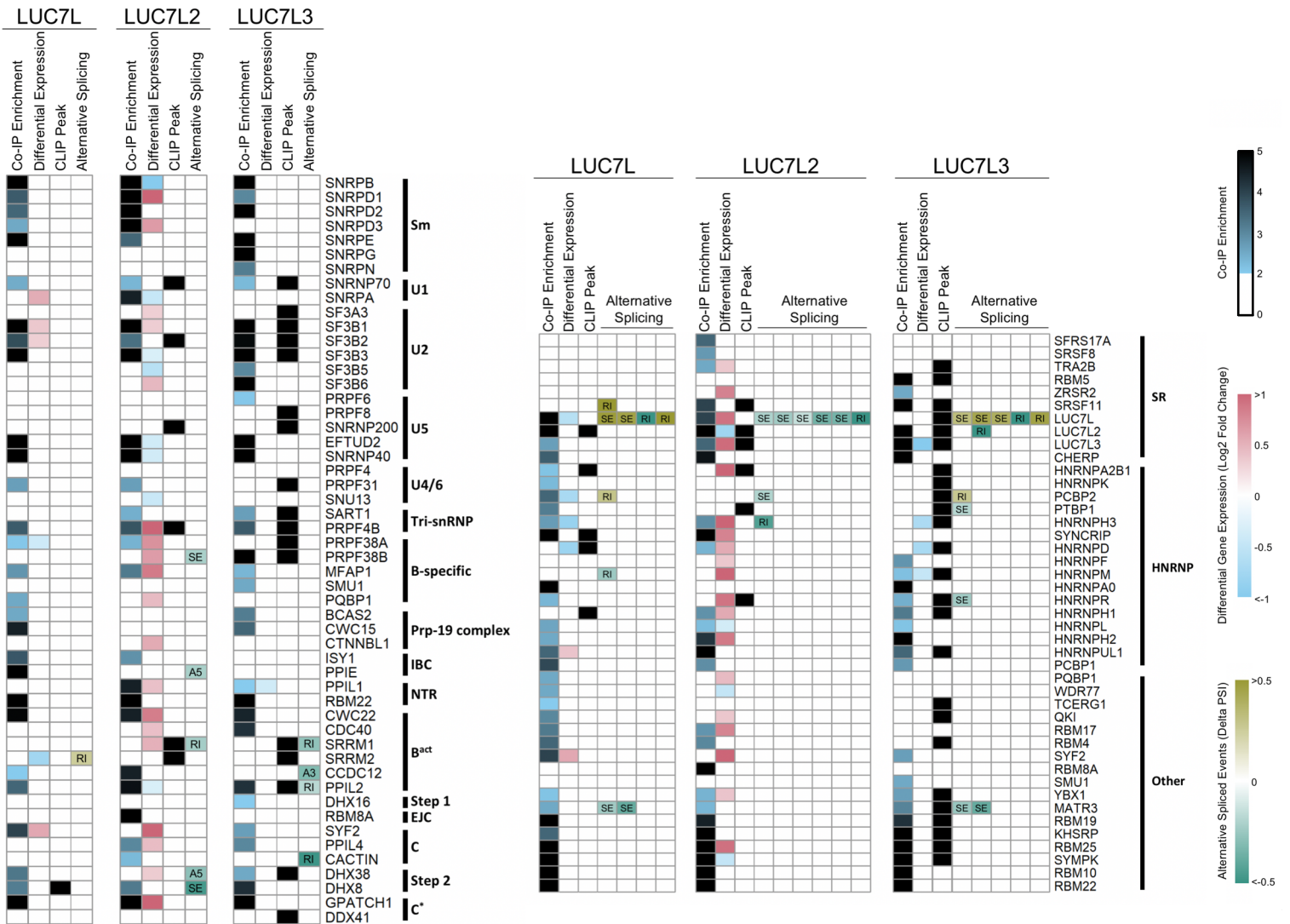
The number of crosslink sites, normalized to input crosslink sites at each base pair of all annotated human splice junctions. Depicted are binding data for 100 nucleotides into the exon and 200 nucleotides into the intron downstream and upstream of the 5'SS and 3'SS, respectively. Anything above the 0-y-axis threshold depicted by a bold black line is enriched binding over the input control.





**Figure S5: RBP crosslinking sites on snRNAs. Related to Figure 3.**

Single nucleotide resolution crosslinking maps on U1, U11, and U2 snRNAs shown as the averaged replicate log<sub>2</sub> fold-change enrichment over the input control.



**Figure S6: LUC7L2 knockdown reveals a complex interplay with its protein interactors. Related to Figures 1 and 6.**

Extension of Figure 1g that includes the significant Co-IP enrichment, differential expression (FDR  $\leq 0.05$ ), RNA binding, and alternative splicing events ( $\Delta$ PSI 10%, q-value  $\leq 0.05$ ) of SFs ordered by their appearance in sub-spliceosomal complexes (left) as well as factors involved in alternative splicing (right) split by each LUC7-like protein. A black cell in the CLIP-Peak column depicts whether there is at least 1 significant CLIP-Peak (log2 fold-change  $\geq 3$ , -log10 p-value  $\geq 3$ , IDR  $\leq 0.01$ ) on the gene in question.