

## SYMPOSIUM: Oxidative Stress in Neurological Disease

# Oxidative Alterations in Alzheimer's Disease

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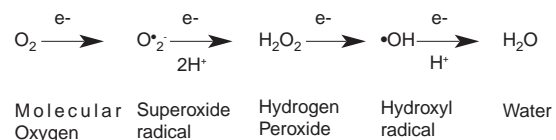
There is increasing evidence that free radical damage to brain lipids, carbohydrates, proteins, and DNA is involved in neuron death in neurodegenerative disorders. The largest number of studies have been performed in Alzheimer's disease (AD) where there is considerable support for the oxidative stress hypothesis in the pathogenesis of neuron degeneration. In autopsied brain there is an increase in lipid peroxidation, a decline in polyunsaturated fatty acids (PUFA) and an increase in 4-hydroxynonenal (HNE), a neurotoxic aldehyde product of PUFA oxidation. Increased protein oxidation and a marked decline in oxidative-sensitive enzymes, glutamine synthetase and creatinine kinase, are found in the brain in AD. Increased DNA oxidation, especially 8-hydroxy-2'-deoxyguanosine (8-OHdG) is present in the brain in AD. Immunohistochemical studies show the presence of oxidative stress products in neurofibrillary tangles and senile plaques in AD. Markers of lipid peroxidation (HNE, isoprostanes) and DNA (8-OHdG) are increased in CSF in AD. In addition, inflammatory response markers (the complement cascade, cytokines, acute phase reactants and proteases) are present in the brain in AD. These findings, coupled with epidemiologic studies showing that anti-inflammatory agents slow the progression or delay the onset of AD, suggest that inflammation plays a role in AD. Overall these studies indicate that oxidative stress and the inflammatory cascade, working in concert, are important in the pathogenet-

ic cascade of neurodegeneration in AD, suggesting that therapeutic efforts aimed at both of these mechanisms may be beneficial.

### Introduction

A growing body of evidence indicates that increased oxidative stress resulting from free radical damage to cellular function, is associated with the aging process and a number of age-related disorders including atherosclerosis and arthritis. In recent years, considerable data have emerged indicating that free radicals play a significant role in the pathogenesis of neurodegenerative disorders. This evidence has been most clearly demonstrated for Alzheimer's disease (AD), where multiple studies show increased oxidation of brain lipids, carbohydrates, proteins and DNA, and the presence of oxidative stress products in neurofibrillary tangles (NFT) and senile plaques (SP) (Tables 1 and 2). These oxidative modifications not only decrease or eliminate the normal function of these macromolecules, but also may activate an inflammatory process in the brains of AD patients. This report reviews the evidence for oxidative stress in the brain in AD and attempts to show a relationship between oxidative change and inflammatory responses in the pathogenesis of neuron degeneration.

Free radicals are defined as any atom or molecule with one or more unpaired electrons in its outer shell. Numerous radicals exist but some of the most potent are formed in the reduction of molecular oxygen to water as follows:



Reduction of molecular oxygen by one electron yields the superoxide radical ( $\text{O}_2^{\bullet-}$ ) which has limited reactivity with some proteins, but is not reactive with lipids or DNA. Under the influence of superoxide dis-

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1.	4-Hydroxynonenal (81,83,106)
2.	Protein carbonyls (117)
3.	Nitrotyrosine (37,119)
4.	Advanced glycation end products (133)
5.	Cu/Zn-superoxide dismutase (92)
6.	Hemeoxygenase 1 (115)

**Table 1.** Oxidative stress products in neurofibrillary tangles in AD.

1.	Advanced glycation end products (114,129)
2.	Cu/Zn-superoxide dismutase (92)
3.	Hemeoxygenase - 1 (115)
4.	Catalase (92)

**Table 2.** Oxidative stress products in senile plaques in AD.

mutase, hydrogen peroxide ( $H_2O_2$ ) is formed by addition of an electron and  $2H^+$ . Although  $H_2O_2$  does not have an unpaired electron and is not a free radical, it is an effective oxidant for many biological molecules. Reduction of  $H_2O_2$  yields the hydroxyl radical ( $\bullet OH$ ), which is the most reactive oxygen radical and capable of oxidizing lipids, carbohydrates, protein and DNA. These oxygen free radicals plus,  $H_2O_2$ , singlet oxygen, and hypochloric acid are spoken of as reactive oxygen species (ROS) (42).

Iron (Fe) and copper (Cu) are important reactive elements that catalyze oxidative reactions and the generation of oxygen radicals. Iron is the most important element in radical generation and may play a role in AD (see below). Iron contains a loosely bound electron and has the ability to exist in more than one valence state. The stable redox state of Fe is  $Fe^{3+}$ , but its bivalent form,  $Fe^{2+}$ , is capable of transferring one electron and facilitating free radical generation. The reaction of  $Fe^{2+}$  with  $H_2O_2$  produces  $\bullet OH$  and is termed the Fenton reaction.

Nitric oxide (NO) is synthesized in a variety of cells and tissues by the enzymatic oxidation of L-arginine to form citrulline through the action of calcium activated, calmodulin-dependent nitric oxide synthase (71). Nitric oxide is an important mediator of several physiologic processes. Excess NO production is found in excitotoxicity, inflammation and ischemia-reperfusion injury (11). Nitric oxide is a free radical with limited potential reactivity, but in biological systems, it reacts with oxygen,  $O_2^{\bullet -}$  and transition metals. Nitric oxide combines rapidly with  $O_2^{\bullet -}$  to form peroxynitrite ( $ONOO^-$ ), a powerful oxidant. Peroxynitrite can oxidize carbohydrates,

membrane lipids, proteins and DNA. It serves as a nitrating agent, promoting the addition of nitrogroups to aromatic and indolic groups in proteins containing tyrosine, phenylalanine, and tryptophan. In addition,  $ONOO^-$  can generate the highly reactive  $OH^{\bullet}$ .

To defend against free radicals, living organisms have learned over time to generate antioxidants and repair enzymes to remove and/or repair molecules that are oxidized. A few enzymatic antioxidants are synthesized by cells. These include Cu/Zn- and Mn-superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), glutathione reductase (GSSG-R), catalase (CAT) and methionine sulfoxide reductase. Other nonenzymatic antioxidants and metal chelators include vitamin E, vitamin C, beta carotene, selenium, ubiquinone, ferritin, ceruloplasmin, and uric acid. Antioxidant defense mechanisms can be upregulated in response to increased free radical or peroxide production (19). Although upregulating antioxidant defense systems may confer protection against free radicals, they are not completely effective in preventing oxidative damage. Also with aging, the efficiency of gene expression may decline or become defective as oxidative damage to the genome increases.

The term oxidative stress is used when free radicals and their products are in excess of the antioxidant defense mechanisms. This may occur as a result of increased radical production or a decrease in antioxidant defenses. If the increased demand on the cell's capacity to detoxify free radicals is not met, alterations occur in cells. Accumulation of oxidized products, such as aldehydes or isoprostanes from lipid peroxidation, protein carbonyls from protein oxidation, and oxidized base adducts from DNA oxidation, serve as markers of excess oxidative stress.

The brain is especially vulnerable to free radical damage because of its high oxygen consumption rate, abundant lipid content, and relative paucity of antioxidant enzymes compared with other organs (26). A role for oxidative stress in the pathogenetic cascade of events in AD and other neurodegenerative disorders is appealing because neurons are post-mitotic cells and gradually accumulate oxidative damage over time, which would account for the late life onset and slowly progressive nature of these disorders (26).

Mitochondrial dysfunction and consequent impairment of ATP production and free radical generation lead to excitotoxicity and neuron degeneration and have been strongly implicated in the pathogenesis of neurodegenerative disorders, especially in AD (7). Mitochondria generate ATP through the reduction of  $O_2$  by the sequential addition of electrons and  $H^+$ . The mitochondrial

electron transport system is the major intracellular source of oxygen radicals and  $H_2O_2$ . Paradoxically, mitochondria are one of the major targets of ROS. Considerable data show a number of age-related alterations in mitochondria including loss of membrane fluidity, increased proton leakage, and decreased levels of cardiolipin (3). Also there is an increase in  $O_2^{\bullet-}$  and  $H_2O_2$  generated in mitochondria with age (120).

It is well recognized that energy metabolism is impaired in the brain in AD (7). Positron emission tomography studies demonstrate a decline in cerebral metabolic rate in parietal and temporal lobes in AD subjects (reviewed in 51). These metabolic defects are present in those at high risk for the disease before symptoms develop (99, 112). Parker *et al.* (93) described a mild generalized reduction of the activity of the electron transport chain complexes (I-IV), but a more marked reduction of cytochrome oxidase (complex IV) in the brains of autopsied AD patients. Several other groups found a reduction of cytochrome oxidase activity in the cerebral cortex of AD subjects (17, 57, 86). Cytochrome oxidase is unique from the other respiratory complexes in that it requires cardiolipin for activity. Oxidative modification of this and other lipids may further damage this complex. Cytochrome oxidase mRNA levels are reduced in hippocampus in AD (109, 110). Parker and Parks (94) purified cytochrome oxidase from AD and control brains and found that control brain cytochrome oxidase had two  $K_m$  values, but the AD brain had only a single value, suggesting that AD brain cytochrome oxidase may be structurally abnormal. The decrease in cytochrome oxidase function could depress ATP synthesis and divert electrons from the normal pathway into increased  $O_2^{\bullet-}$  generation. Partridge *et al.* (95) found  $\bullet OH$  in mitochondria after cytochrome oxidase inhibition by sodium azide. Diminished cytochrome oxidase activity could lead to increased ROS generation, oxidative damage to mitochondrial membranes, and increased vulnerability to excitotoxins, and may play a role in the pathogenesis of AD.

### Lipid peroxidation

Increased lipid peroxidation has been described in several neurodegenerative diseases including Parkinson's disease and AD, and in ischemic and traumatic brain injury (10, 32, 41). Lipid peroxidation is assessed by measuring a) thiobarbituric acid reactive substances (TBARS), b) alterations in polyunsaturated fatty acids (PUFA), and c) the breakdown products of PUFA, such as aldehydes and isoprostanes.

Subbarao *et al.* (125) found an increase in TBARS in

frontal lobe specimens, but not cerebellum, in AD compared with control subjects. Others described increased TBARS in the temporal lobe but not in other neocortical areas in AD (4, 40, 66, 75). Our study of 13 AD and 10 prospectively evaluated control subjects, all from short postmortem interval autopsies, found elevated TBARS in all AD brain regions compared with controls, except the middle frontal gyrus (60). These elevations were statistically significant in the hippocampus and pyriform cortex, and marginally significant in the amygdala. This study demonstrated that lipid peroxidation is most pronounced in medial temporal lobe structures where histopathologic changes are most pronounced. Although TBARS measure the major lipid peroxidation burden, they also measure a variety of products including non-lipid derived malondialdehyde,  $C_3$ - $C_{10}$  aldehydes, and species resulting from chemical interaction among non-lipid molecules during the assay. Thus, it is important to determine other markers of lipid peroxidation to be certain of its presence.

Polyunsaturated fatty acids, which make up the brain's membrane phospholipids, are especially vulnerable to free radical attack because their double bonds allow easy removal of  $H^+$ . Because lipid peroxidation is increased in AD, it follows that PUFA would be expected to be diminished, especially arachidonic and docosahexenoic acids, which are especially vulnerable to attack by ROS. *In-vivo* phosphorus nuclear magnetic resonance (PNMR) studies demonstrated that the ratio of glycerophosphorylcholine to glycerophosphorylethanolamine was increased and levels of glycerophosphodiester and phosphomonoesters were elevated in AD compared with age-matched controls (79, 96). Another PNMR study showed a significant decrease in phosphatidylethanolamine (PE) and the phospholipid precursors—choline and ethanolamine—in AD (90). It was suggested that these alterations were specific for the pathogenesis of AD. Svennerholm and Gottfries (126) showed decreased levels of brain cortical membrane phospholipids in early-onset AD patients but not late-onset AD patients. Our study of membrane phospholipid levels in various brain regions in late-onset AD patients and age-matched controls found that PE and phosphatidylinositol (PI)-derived total fatty acids were significantly decreased in hippocampus of AD subjects (97). In PE-derived fatty acids, stearic, oleic, arachidonic and docosahexenoic acids were significantly decreased, and in PI-derived phospholipids, oleic and arachidonic were significantly decreased. In the inferior parietal lobule, significant decreases were found in total PE-derived fatty acids and in stearic,

oleic, and arachidonic acids. The decreases in PE and PI, which are rich in oxidizable arachidonic and docosahexenoic acids, but no change in the phosphatidylcholine pool, which contains lesser amounts of these fatty acids, suggest that free radicals are responsible for the alteration in membrane phospholipids.

Oxidation of PUFA results in the production of multiple aldehydes with different carbon chain lengths including propanal, butanal, pentanal, hexanal and 4-hydroxynonenal (HNE) (31). Aldehydes have half-lives of minutes to hours and can diffuse from their site of origin to more distant sites, much different from the highly reactive free radicals. HNE is a highly reactive  $\alpha$ ,  $\beta$ -aldehyde responsible for cytotoxicity in conjunction with oxidative stress and is capable of inhibiting DNA, RNA, protein synthesis, and glycolysis, and degrading proteins (31). HNE forms adducts with proteins by covalent bonding to histidine, lysine and cysteine residues through Michael addition or by Schiff base reactions (31, 127). We demonstrated elevations of free HNE in multiple brain regions in AD compared with age-matched controls (69). These elevations were statistically significant in the amygdala, hippocampus, and parahippocampal gyrus, regions showing the most striking histopathologic alterations in AD. We also demonstrated a highly significant elevation of free HNE in ventricular cerebrospinal fluid (CSF) in 19 AD patients compared with 13 control subjects (61).

In immunocytochemical studies, Montine *et al.* (81) demonstrated that HNE pyrrole adducts are present in NFT in AD and are significantly associated with the inheritance of the APOE  $\epsilon$ 4 alleles, a major risk factor for AD (103). Borohydride-reducible HNE adducts were increased in pyramidal neuron cytoplasm in the hippocampus, entorhinal cortex, and temporal neocortex in patients in AD who were homozygous for APOE  $\epsilon$ 4, and pyramidal neurons and astrocytes in AD patients who were homozygous for APOE  $\epsilon$ 3 (83). These studies suggest that APOE, the major lipoprotein trafficking molecule in the brain, might influence HNE accumulation in AD. Sayre *et al.* (106) demonstrated HNE-pyrrole immunoreactivity in NFT and in neurons lacking NFT in AD, but not in control brains. They did not find a correlation of HNE-pyrrole immunostaining with any particular APOE allele.

HNE is toxic to neurons and astrocytes in P19 neuroglial cultures and caused crosslinking of tau into high molecular weight species that are conjugated with ubiquitin (80). Mark *et al.* (67) demonstrated that HNE caused a time- and concentration-dependent decrease in rat hippocampal neurons in cultures by impairing

Na<sup>+</sup>K<sup>+</sup>-ATPase activity and disrupting calcium homeostasis which led to neuron death. Exposure of cultured rat hippocampal neurons to A $\beta$  induced a significant increase in free and protein-bound HNE (67). HNE caused impaired glucose transport in cultured rat hippocampal neurons and impaired glutamate transport in rat neocortical synaptosomes (54, 68). HNE also impaired coupling of metabotropic acetylcholine and glutamate receptor G<sub>q11</sub> in cortical neuron cultures (9). HNE is capable of inducing apoptosis in PC12 cells and cultured rat hippocampal neurons suggesting that it is a mediator of oxidative stress-induced apoptosis (59). HNE administered into the basal forebrain of rats, damaged cholinergic neurons, diminished ChAT activity, and impaired visuospatial memory (13). Taken together, these studies suggest that in addition to direct ROS damage to phospholipid membranes, there is an indirect mechanism involving HNE, which also may be involved in neuron death. These studies demonstrate that HNE may be an important molecule in the pathogenetic cascade of neuron degeneration in AD.

Glutathione transferases are a group of enzymes that function to inactivate the toxic products of oxygen metabolism including 4-hydroxyalkenals, such as HNE (27). Recently, we described decreased glutathione transferase activity in eight brain regions in AD including statistically significant reductions in the amygdala, hippocampus, and inferior parietal lobule compared with normal age-matched controls (63). In addition, a statistically significant decrease in glutathione transferase activity was found in postmortem ventricular CSF in AD compared with age-matched controls. This study suggests a loss of protection against HNE in AD that could be important in the pathogenesis of neuron degeneration.

Isoprostanes are prostaglandin-like compounds that are formed nonenzymatically by free radical-induced oxidation of arachidonic acid (84). Because they contain F-type prostane rings, they are referred to as F<sub>2</sub>-isoprostanes (F<sub>2</sub>-IsoP). Considerable evidence exists to show that F<sub>2</sub>-IsoP concentration is a reproducible quantitative index of lipid peroxidation *in vivo* (84). Oxidation of docosahexenoic acid leads to the formation of F<sub>2</sub>-IsoP-like compounds which have been called F<sub>4</sub>-neuroprostanes (F<sub>4</sub>-NP) (101). *In vitro* oxidation of docosahexenoic acid yielded higher levels of F<sub>4</sub>-NP than F<sub>2</sub>-IsoP by 3.4 fold (101). Concentrations of F<sub>2</sub>-IsoP in the CSF of AD patients were significantly elevated (72  $\pm$  pg/ml) compared with controls (46  $\pm$  4 pg/ml) ( $p$  < 0.05) (82). Linear regression analysis showed a signif-



icant correlation between F<sub>2</sub>-IsoP levels and brain weight in AD. Study of F<sub>4</sub>-NP in CSF of a small number of AD and control patients yielded a significantly higher level in AD (110 ± 12 pg/ml), compared with controls (64 ± 8 pg/ml) (101).

These studies indicate that a potential quantifiable marker of brain lipid peroxidation may be present in CSF in AD (Table 3). One or more of these potential biomarkers could be of considerable value in a) quantifying the response to therapeutic interventions to decrease lipid peroxidation, and b) aiding in the diagnosis of AD. Of these, F<sub>2</sub>-IsoP and F<sub>4</sub>-NP may be the most valuable. Multiple further studies of subjects in various clinical stages of AD and in other slowly progressive neurodegenerative diseases are required to define the value and specificity of lipid peroxidation breakdown products in CSF in AD. It is likely that products of lipid peroxidation are present in CSF in other chronic progressive neurodegenerative disorders such as motor neuron disease, Parkinson's disease, Huntington's disease, corticobasal degeneration and others with focal degenerative changes. However, it may be that the presence of more widespread alterations in AD increases the overall burden of lipid peroxidation and resultant breakdown products.

### Protein oxidation

Hydrazide reactive protein carbonyl (protein carbonyl) analysis is used as a general assay of oxidative damage to proteins (122). Smith *et al.* (113) demonstrated a significant increase in protein carbonyls in the frontal and occipital poles in normal aged subjects compared with young controls. This study also demonstrated that protein oxidation was significantly increased in the frontal pole (where numerous degenerative changes are found) of AD patients compared with normal aged control subjects, although no differences were found in the occipital pole (where few degenerative changes are present). When the level of protein carbonyl was compared to the age of the control subjects, it was found that the frontal pole accumulated carbonyl-bearing protein at twice the rate of the occipital pole. Hensley *et al.* (45) showed that protein carbonyl content was increased in the AD hippocampus and inferior parietal lobule compared with the cerebellum in AD. Control hippocampus and inferior parietal lobule levels were similar to control cerebellum levels. Others have shown a significant elevation of protein-carbonyl levels in the parietal lobe and a trend for elevations in the frontal, temporal and occipital lobes and hippocampus in AD (65). Smith *et al.* (117), using immunocytochemical techniques with *in*

- |    |  |
|----|--|
| 1. | Increased free 4-hydroxynonenal  |
| 2. | Increased glutathione transferases   |
| 3. | Increased isoprostanes   |
| 4. | Increased 8-hydroxy-2-deoxyguanosine (OHdG) in intact DNA and decreased free •OHdG |

**Table 3.** Oxidative stress products in CSF in AD.

*situ* 2, 4 dinitrophenylhydrazine (DNP) labeling linked to an antibody system against DNP, demonstrated the presence of protein carbonyls in NFT and glia but not in nonNFT-bearing neurons in AD. These changes were not found in control brains. In addition increased oxidation of synaptosomes in AD hippocampus and parietal lobe was shown using electron paramagnetic resonance (EPR) (44, 45).

Peroxonitrite causes nitration of tyrosine residues yielding nitrotyrosine which is used as an indicator of ONOO<sup>-</sup> activity. Several investigators found nitrotyrosine in NFT in the hippocampus in AD (37, 119). In addition, Smith *et al.* (119) found nitrotyrosine immunoreactivity in non-NFT bearing neurons and in nuclei of glia in AD. These studies demonstrate that ONOO<sup>-</sup> is likely involved in protein oxidation in AD.

The oxidation of proteins by free radicals may be responsible for damaging enzymes critical to neuron function (122). As demonstrated by Stadtman *et al.* (122), this is a two-step process in which there is oxidation of enzyme amino acids by the free radicals yielding carbonyl derivatives. The second step involves further degradation of the enzyme by proteases to amino acids and peptides. Two enzymes that are especially sensitive to oxidative modification are glutamine synthetase (GS) and creatinine kinase (CK) (122). A significant decline in glia-specific GS activity has been found in the hippocampus and neocortex in AD compared with age-matched controls (45, 113). Decreased levels of GS could result in decreased glutamate turnover causing prolonged NMDA receptor activation and neuron injury in brain areas susceptible to glutamate toxicity. In addition, because glutamate is converted to glutamine, less of the enzyme could alter nitrogen balance, pH and glutathione synthesis in astrocytes. Several studies have demonstrated a significant decline in CK activity in the frontal and temporal lobes in AD (1, 14). One of these studies demonstrated that the decline in CK levels was due to a decrease in brain CK and not ubiquitous mitochondrial CK (1). These studies demonstrate that the increase in protein oxidation in the brain in AD may be at the expense of enzymes critical to neuron energy

metabolism and function.

Yatin *et al.* (136) demonstrated that protein carbonyls develop immediately following oxidative insults in cultured neurons. They showed that ROS generated by A $\beta$  immediately attack amino acid residues causing carbonyl formation. Protein oxidation starts as soon as cells are exposed to A $\beta$ , and protein carbonyl levels are significantly elevated after one hour of A $\beta$  treatment. These changes occur before neuron death and parallel the EPR detection of free radicals. They also demonstrated that Mn-SOD and Cu/Zn-SOD increased rapidly in response to oxidative stress.

### DNA oxidation

Oxidation of DNA can cause several damage products including strand breaks, sister chromatid exchange, DNA-DNA and DNA-protein crosslinking, and base modifications (28). Damage to DNA occurs as a consequence of the generation of O $_2^{\bullet-}$ , H $_2$ O $_2$ ,  $\bullet$ OH, OONO $\cdot$ , and singlet oxygen. It has long been suggested that DNA damage accumulating in nondividing mammalian cells, may play a major role in aging changes. The ability to repair DNA damage is a critical factor in the function and longevity of cells and defects in DNA repair may be important in aging and age-associated disorders.

DNA degeneration occurs in apoptotic and necrotic cell death. It has been suggested that neuron death in AD is through an apoptotic mechanism (111, 123), although necrotic cell death also has been proposed (121). A two-fold increase in DNA strand breaks has been described in the brain in AD (85). DNA strand breaks can be detected at the single cell level by terminal deoxynucleotidyl transferase (TdT)-mediated *in situ* end labeling. Su *et al.* (124) showed robust TdT labeling in neurons lacking NFT in the occipital lobe in AD. Nitrotyrosine immunoreactivity was prominent in TdT-labeled neurons. These authors suggested that neurons with DNA damage without NFT may undergo degeneration caused by oxidative mechanisms involving OONO $\cdot$ .

Several biomarkers of oxidative DNA damage have been quantified but the most useful has been the adduct, 8-hydroxy-2'-deoxyguanosine (8-OHdG). Mecocci *et al.* (77) found an increase in 8-OHdG in nuclear and mitochondrial brain fractions in aging. Using HPLC, they demonstrated a 10-fold increase in mitochondrial DNA oxidation compared with nuclear DNA, and a 15-fold increase in DNA oxidation in subjects older than 70 years of age. These same investigators showed a three-fold increase in mitochondrial DNA oxidation in the parietal lobe in AD subjects compared with normal con-

trols (78). Their study demonstrated a small but significant increase in oxidative damage to nuclear DNA and a highly significant increase in mitochondrial DNA oxidation in AD samples compared with age-matched control subjects.

Gas chromatography with mass spectroscopy (GC-MS) is a sensitive method used to identify oxidative adducts from DNA bases. Using GC-MS in combined nuclear and mitochondrial DNA specimens, Lyras *et al.* (65) found various bases increased or decreased in different brain regions in AD. The most consistent elevations were in 8-hydroxyadenine, 5-hydroxycytosine, and 8-OHdG in the parietal lobe in AD. We used GC-MS with stable-labeled oxidized base analog standards to study nuclear DNA from four brain regions in AD patients and prospectively evaluated control subjects, all with short postmortem intervals (34). Our studies showed statistically significant elevations of 5-hydroxyuracil, 8-hydroxyadenine, and 8-OHdG in frontal, parietal, and temporal lobes, and 5-hydroxycytosine in the parietal and temporal lobes in AD. The increases in mean 8-OHdG were the largest elevations of all the base adducts analyzed indicating that guanine is the most vulnerable base to oxidation. These studies suggest that the pattern of damage to multiple bases in the brain is due to  $\bullet$ OH attack on DNA.

Recently, using GC-MS and stable isotope labeled 8-OHdG as the standard, we studied the levels of 8-OHdG in intact DNA and free 8-OHdG (representing the repair product) in ventricular CSF from AD and control subjects (64). We found significant elevation of 8-OHdG in intact DNA in AD compared with age-matched control subjects. In contrast, levels of free 8-OHdG were significantly decreased in AD samples. This suggests that in AD there may be a double insult of increased oxidative DNA damage and a deficiency of repair mechanisms responsible for removal of oxidized bases. Hermon *et al.* (46) described increased protein levels of two excision repair cross-complementing genes for nucleotide excision repair in AD brain, suggesting ongoing oxidative DNA damage.

Wade *et al.* (130) used immunocytochemistry to show that 8-OHdG was increased in mitochondria in neurons in AD compared with controls. The same investigators demonstrated that the 5Kb deletion, the most common alteration in human mitochondria, was prominent in large hippocampal pyramidal neurons in AD (47). This deletion was present in neurons immunostaining with 8-OHdG. They concluded that oxidative modifications of mitochondrial DNA are an early event in the neuropathologic changes in AD, and that accumu-

lation of deleted mitochondrial DNA may potentiate oxidative damage in vulnerable neurons.

These descriptive studies indicate significant oxidative damage to DNA in the brain in AD. The consequences of the oxidative DNA damage in the pathogenesis of AD will require further study as will the role of nucleotide and base excision-repair pathways in the brain in AD.

### **Glycooxidation**

There are two mechanisms by which glucose can induce changes in proteins (49, 50). Monosaccharides can oxidize when catalyzed by transition metals generating free radicals, H<sub>2</sub>O<sub>2</sub>, and reactive dicarbonyls (48). Proteins are then damaged by free radicals and by covalent bonding of the carbonyl products to protein components. The other reaction is the nonenzymatic glycation of proteins through the Maillard reaction. Advanced glycation end products (AGE) are post-translational modifications of proteins that are formed when the amino group of proteins, especially N-terminal amino groups and side chains of lysine and arginine, react nonenzymatically with monosaccharides (reviewed in 87). This leads by way of Schiff base products to protein-bound Amadori products. The Amadori products, through oxidation and dehydration including free radical intermediates, form AGE. This reaction, which is catalyzed by transition metals such as Fe and Cu, also yields oxygen free radicals. AGE-modified proteins can also produce free radicals through interaction with microglia (76). Modification of proteins by oxidation, glycooxidation, and products of lipid peroxidation can occur in an additive and synergistic manner (116).

Recent studies indicate a role for AGE in AD (116). AGE have been found in diffuse and neuritic SP in AD (114, 129). AGE-modified A $\beta$  accelerates aggregation of soluble nonfibrillar A $\beta$  *in vitro* suggesting that AGE may enhance SP formation *in vivo* (129). Hyperphosphorylated tau is a major component of NFT. Tau and AGE antigens are co-localized in NFT (114, 133). Glycated tau added to cultured neuroblastoma cells induces lipid peroxidation (133). These studies suggest that AGE may play a role in AD by oxidative modifications of A $\beta$  and tau, the two major proteins involved in this disorder.

Another possible way that AGE may play a role in AD has been described. One of the cell receptors for AGE, termed RAGE, is a member of the immunoglobulin superfamily of cell surface molecules (89,108) that is expressed on endothelium, neurons, smooth muscle cells, and phagocytes (12). RAGE expression is

increased in neurons, microglia and blood vessels in AD (134). Yan *et al.* (135) demonstrated that A $\beta$  binds to RAGE and generates oxidant stress, activating nuclear factor  $\kappa$ B(NF- $\kappa$ B), and inducing expression of macrophage-colony stimulating factor. Macrophage-colony stimulating factor enhances proliferation and migration of microglia. This study suggests that a free radical-dependent inflammatory pathway, triggered by interaction of A $\beta$  on RAGE, may be present in AD.

### **Antioxidant enzymes**

The increase in brain oxidative damage described above might be facilitated by a failure of antioxidant defense mechanisms. Antioxidant enzymes that are important in preventing an excessive accumulation of ROS include Cu/Zn- and Mn-SOD, GSH-Px, GSSG-R and CAT.

We studied a number of antioxidant enzymes in the brains of AD patients and control subjects with short postmortem intervals and found significant elevations of GSH-Px activity in hippocampus, GSSG-R activity in hippocampus and amygdala, and CAT activity in hippocampus and temporal neocortex in AD compared with normal subjects (60). These changes correlated with elevation of lipid peroxidation in the same regions. In that study and subsequent studies, we did not find significant elevations of Cu/Zn- or Mn-SOD activity in AD. In a separate series of AD and control subjects, we studied mRNA expression of oxidative stress-handling genes (2). We found significant elevations of GSH-Px, GSSG-R and CAT mRNA in hippocampus and inferior parietal lobules, but not in cerebellum in AD. An increase in Cu/Zn-SOD mRNA was present in the inferior parietal lobule. Importantly, no deficiencies of antioxidant enzyme expression at the transcription level were observed. The elevations observed in both these studies correlate with increased oxidative damage and may reflect a compensatory rise in antioxidant activity in response to increased ROS generation.

Other studies of antioxidant levels in the brain in AD found variable results. Two studies found no significant difference in GSH-Px in AD and controls (56, 66). Catalase was elevated in the amygdala (4) and in the temporal lobe in AD (66), but in another study it was reduced in several brain regions (39). Several studies reported no significant difference in Cu/Zn- or Mn-SOD activity in the brain in AD compared with controls (39, 53). One study described a significant reduction of SOD in frontal cortex, hippocampus, and cerebellum in AD (100). Other investigators reported a decrease in SOD activity in the caudate nucleus (70), and frontal and tem-

poral lobes in AD (66). In contrast, Kato *et al.* (53) found a significant elevation of Cu/Zn-SOD in AD brains.

Immunohistochemical studies (Table 1 and 2) show an increase in Cu/Zn-SOD and CAT in a subgroup of NFT and SP in the hippocampus in AD (92). Hemeoxygenase -1, an antioxidant, is present in NFT, SP and neuropil threads in AD (115), as well as in neurons and astrocytes in AD (107). Increased expression of hemeoxygenase-1 mRNA and protein content has been described in the frontal, temporal, and occipital lobes in AD (98).

Taken as a whole, studies of antioxidant enzyme activities and expression in the brain in AD do not show a consistent trend. Importantly, they do not demonstrate meaningful deficiencies of any enzyme activity, suggesting that the oxidative stress in the brain is not related to a failure of these defense mechanisms.

### Role of iron in oxidative stress

As noted in the Introduction, Fe has a potential major role as a catalyst for free radical generation. It is unique in that it has a loosely bound electron in the Fe<sup>2+</sup> oxidation state and the ability to exist in two oxidation states. In its bivalent Fe<sup>2+</sup> form, it is capable of transferring an electron and facilitating free radical generation.

We reported an increase in Fe in the brain in bulk specimens from different brain regions in AD compared with age-matched control subjects in several separate instrumental neutron activation analysis studies (25, 29, 30, 104). Our initial study of separated cerebral gray and white matter showed a statistically significant elevation of Fe in AD gray matter compared with control subjects (30). In our recent study of 58 AD and 21 prospectively evaluated normal control subjects, Fe was significantly elevated in the frontal, temporal, and parietal neocortex, and the hippocampus and amygdala (25).

Good *et al.* (36), using laser microprobe analysis demonstrated a significant elevation of Fe in NFT-bearing neurons in the hippocampus in AD. Using microparticle-induced x-ray emission, we found a significant elevation of Fe in the cores and rims of SP in the amygdala of AD subjects compared with neuropil of AD and control subjects (62).

Several studies found increased Fe and ferritin in SP in AD (22, 24) and ferritin immunoreactivity in microglia in SP in AD (38). One study showed that ferritin from the brains of AD patients contained more Fe than brains of age-matched controls (33). The frequency of the C<sub>2</sub> allele of transferrin was significantly higher in late-onset AD patients than age-matched controls, and

twice as high in AD patients homozygous for apolipoprotein E  $\epsilon$ 4 alleles compared with AD patients with one or no copies of the  $\epsilon$ 4 allele (88). Another study showed that transferrin immunoreactivity is homogeneously present around SP in AD (23). The Fe binding protein, P97 or melanotransferrin, is elevated in the serum, CSF, and brains of AD patients compared with control subjects (52, 55). Iron regulatory protein-2 is present in NFT, SP, and neuropil threads, whereas Fe regulatory protein 1 has similar expression in AD and control brains (16). Smith *et al.* (118) showed that redox-active Fe is associated with SP, NFT and neuropil threads in AD and catalyzes a H<sub>2</sub>O<sub>2</sub> dependent oxidation. Overall, these studies indicate an increase in Fe in the brain in AD suggesting an abnormality in Fe metabolism. This increase in Fe may contribute to enhancement of oxidative stress in the disease.

### Inflammatory alterations in AD

Numerous epidemiologic studies indicate that use of nonsteroidal anti-inflammatory drugs (NSAID) or corticosteroids is associated with slowing of the progression or delaying the onset of AD. Meta-analysis of 17 epidemiologic studies from nine countries demonstrated that the use of corticosteroids or NSAID, was associated with a low risk of AD (74). The use of NSAID had a stronger level of significance than use of corticosteroids. A six-month trial on indomethacin showed a slowing of the progression of AD, although the number of patients used in the study was small (102). The major effect of NSAID is through decreasing the activity of cyclooxygenase which catalyzes the first steps in the biosynthesis of prostaglandins from arachidonic acid. Cyclooxygenase-2 (COX-2) is induced by stimuli associated with inflammation, such as the cytokines, interleukin I (IL-1), interleukin 2 (IL-2), and TNF- $\alpha$  (128). Thus, the ability of NSAID to inhibit COX-2 makes them effective as anti-inflammatory agents in a number of different disorders.

Cultured murine and rat astrocytes express COX-2 and it is upregulated by IL-1 $\beta$  and TNF- $\alpha$  (91). Rat microglia stimulated with lipopolysaccharide express COX-2 (6). Expression of COX-2 mRNA is decreased in the brain in AD (18). One explanation for the lower COX-2 activity in the AD brain is that in end-stage disease the loss of neurons and decreased synthesis of COX-2 are greater than the increase of COX-2 in activated glia (128). Another explanation might be that the efficacy of NSAID in AD is through mechanisms that do not involve COX-2.

The above studies suggesting that anti-inflammatory



drugs may play a protective role against AD have stimulated an increased interest in the role of inflammation in the brain in AD. Support for the presence of an inflammatory response in the brain in AD has been gained from numerous autopsy studies. A number of markers of inflammation are present in the brain in AD and some are related to the morphologic changes of AD (reviewed in 73). These include the classic-complement cascade and their regulating cytokines, acute phase reactants, proteases, and protease inhibitors. Immunohistochemical studies have shown the presence of C1q, C4d, C3c, and C3d in amyloid in SP. Elements of the membrane attack complex, C7 and C9, are found in A $\beta$ -deposits in AD, and other components of the membrane-attack complex have been found in neurites in SP and in some NFT. The acute phase proteins,  $\alpha$ -1 antichymotrypsin,  $\alpha$ -antitrypsin, C-reactive protein, and  $\alpha$ -2 macroglobulin are upregulated in the AD brain and found around SP. Mutations in the  $\alpha$ -2 macroglobulin gene have been found in AD and appear to be a major risk factor for the disease (8).

Microglia cells are the immunological scavengers of the brain and may be the key factor in the inflammatory response in AD. Activated microglia cells are markedly increased in the brain in AD (15). Microglia cells are capable of releasing a number of interleukins, especially IL-1 and IL-6 and TNF- $\alpha$  (131, 132). Astrocytes also have the ability to produce interleukins, prostaglandins, complements, coagulation factors, proteases, and protease inhibitors (73). The relationship between reactive astrocytes and reactive microglia in the inflammatory cascades may be important in propagating the response.

The relationship between the inflammatory response and free radical generation, although not clearly worked out in AD, is of considerable theoretical and therapeutic interest. Activated microglia demonstrate respiratory burst activity which is derived from the hexose monophosphate shunt (20). Respiratory burst activity can be initiated by fatty acids, high extracellular calcium, eicosanoid derivatives and various synthetic peptides including A $\beta$  (58). Activated microglia are capable of generating O $_2^{\bullet-}$  which could lead to H $_2$ O $_2$  and  $\bullet$ OH formation (5, 20). Activated microglia are also capable of generating NO (21) which, as shown above, can generate the potent radical OONO $\bullet$ . Thus, the activated microglia in AD may be part of a neurotoxic mechanism through generation of free radicals.

Another possible pathway of neurodegeneration in AD is through the ability of IL-1 to cause an upregulation of APP expression (35). This could lead to increased A $\beta$  which has been shown to cause H $_2$ O $_2$  accu-

mulation in cultured hippocampal neurons (72). Hensley *et al.* (44), using EPR analysis, demonstrated that aggregated A $\beta$  is capable of generating free radicals. A $\beta$  is capable of inactivating GS and CK in cell free incubates and in cell cultures (43). Thus, microglia releasing IL-1 could set in motion a self-propagating series of events that lead to more free radical generation and neuron destruction.

It is most probable that AD is associated with multiple etiologies and pathophysiologic mechanisms. The above review clearly demonstrates that oxidative stress and the inflammatory cascade are, at least, a part of a pathophysiologic mechanism in AD. While they may be thought of as separate events, it appears that they work in concert. Although we do not know the initiating events, these mechanisms appear to be involved in the pathogenesis cascade that leads to specific neuron destruction in AD and possibly other neurodegenerative diseases. Thus, it seems logical to direct therapeutic efforts at oxidative and inflammatory events in the pathway of neuron degeneration in AD. Preliminary evidence that antioxidant therapy (105) and anti-inflammatory therapy (74) slow or delay AD supports this concept. Because some of the risk factors for AD are known, treatment of individuals at high risk long before they develop symptoms, using antioxidants and anti-inflammatory agents, may delay or prevent the onset of AD.

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#### References

1. Aksenov MY, Aksenov MV, Payne RM, Smith CD, Markesbery WR, Carnay JM (1997) The expression of creatine kinase isoenzymes in neocortex of patients with neurodegenerative disorders: Alzheimer's and Pick's disease. *Exp Neurol* 146: 458-465
2. Aksenov MY, Tucker HM, Nair P, Aksenova MV, Butterfield DA, Estus S, Markesbery WR (1998) The expression of key oxidative stress handling genes in different brain regions in Alzheimer's disease. Accepted *J Mol Neurosci*
3. Ames BN, Shigenaga MK, Hagen TM (1995) Mitochondrial decay in aging. *Biochem Biophys Acta* 1271: 165-170
4. Balazs L, Leon M (1994) Evidence of an oxidative challenge in the Alzheimer's brain. *Neurochem Res* 19: 1131-1137

5. Banati RB, Gehrman J, Schubert P, Kreutzberg GW (1993) Cytotoxicity of microglia. *Glia* 7: 111-118
6. Bauer MKA, Lieb K, Schulze-Osthoff K, Berger, M, Gebicke-Haerter PJ, Bauer J, Fiebich BL (1997) Expression and regulation of cyclooxygenase-2 in rat microglia. *Eur J Biochem* 243: 726-731
7. Beal MF (1995) Aging, energy, and oxidative stress in neurodegenerative diseases. *Ann Neurol* 38: 357-366
8. Blacker D, Wilcox MA, Laird NM, Rodes L, Horvath SM, Go RC, Perry R, Watson BJ, Bassett SS, McInnis MG, Albert MS, Hyman BT, Tanzi RE (1998) Alpha-2 macroglobulin is genetically associated with Alzheimer disease. *Nat Genet* 19: 357-360
9. Blanc EM, Kelly JF, Mark RJ, Waeg G, Mattson MP (1997) 4-Hydroxynonenal, an aldehydic product of lipid peroxidation, impairs signal transduction associated with muscarinic acetylcholine and metabotropic glutamate receptors: possible action on G alpha (q/11). *J Neurochem* 69: 570-580
10. Braughler JM, Hall ED (1989) Central nervous system trauma and stroke. I. Biochemical considerations for oxygen radical formation and lipid peroxidation. *Free Radic Biol Med* 6: 289-301
11. Bredt DS, Snyder SH (1994) Nitric oxide: a physiologic messenger molecule. *Annu Rev Biochem* 63: 175-195
12. Brett J, Schmidt A, M., Yan SD, Zou Y-S, Weidman E, Pinsky D, Nowgrad R, Nepper M, Przysiecki C, Shaw AEA (1993) Survey of the distribution of a newly characterized receptor for advanced glycation end products in tissues. *Am J Pathol* 143: 1699-1712
13. Bruce-Keller AJ, Li YJ, Lovell MA, Kraemer PJ, Gary DS, Brown RR, Markesbery WR, Mattson MP (1998) 4-Hydroxynonenal, a product of lipid peroxidation, damages cholinergic neurons and impairs visuospatial memory in rats. *J Neuropathol Exp Neurol* 57: 257-267
14. Burbueva GS, Aksenova MV, Makarenko IG (1992) Decreased level of creatine kinases BB in the frontal cortex of Alzheimer patients. *Dementia* 3: 91-94
15. Carpenter AF, Carpenter PW, Markesbery WR (1993) Morphometric analysis of microglia in Alzheimer's disease. *J Neuropathol Exp Neurol* 52: 601-608
16. Castellani, Smith MA (1998) Iron regulatory proteins in Alzheimer's disease. *J Neuropathol Exp Neurol* 57: 511
17. Chagnon P, Betard C, Robitaille Y, Cholette A, Gauvreau D (1995) Distribution of brain cytochrome oxidase activity in various neurodegenerative diseases. *NeuroReport* 6: 711-715
18. Chang JW, Coleman PD, O'Banion MK (1996) Prostaglandin G/H synthase-2 (cyclooxygenase-2) mRNA expression is decreased in Alzheimer's disease. *Neurobiol Aging* 17: 801-808
19. Cohen G, Werner P (1994) Free radicals, oxidative stress and neurodegeneration, In: *Neurodegenerative Diseases*, Calne DB (ed.), Chapter 10, pp 139-161, W. B. Saunders Co., Philadelphia, PA, USA
20. Colton CA, Gilbert DL (1987) Production of superoxide anions by a CNS macrophage, the microglia. *FEBS Lett* 223: 284-288
21. Colton CA, Snell J, Chernyshev O, Gilbert DL (1994) Induction of superoxide anion and nitric oxide production in cultured microglia. *Ann. NY Acad. Sci.* 738: 54-63
22. Connor JR, Menzies SL, St. Martin SM, Mufson EJ (1992 a) A histochemical study of iron, transferrin and ferritin in Alzheimer's diseased brains. *J Neurosci Res* 31: 75-83
23. Connor JR, Snyder BS, Beard JL, Fine RE, Mufson EJ (1992 b) Regional distribution of iron and iron-regulatory proteins in the brain in aging and Alzheimer's disease. *J Neurosci Res* 31: 327-335
24. Connor JR, Menzies SL (1995) Cellular management of iron in the brain. *J Neurol Sci* 134: 33-44
25. Cornett CR, Markesbery WR, W.D. (1998) Imbalances of trace elements related to oxidative damage in Alzheimer's disease brain. *Neurotoxicology* 19: 339-346
26. Coyle JT, Puttfarcken P (1993) Oxidative stress, glutamate and neurodegenerative disorders. *Science* 262: 689-695
27. Danielson UH, Esterbauer H, Mannervik B (1987) Structure-activity relationships of 4-hydroxyalkenals in the conjugation catalysed by mammalian glutathione transferases. *Biochem J* 247: 707-713
28. Davies KJ (1995) Oxidative stress: the paradox of aerobic life. *Biochem Soc Symp* 61: 1-31
29. Deibel MA, Ehmann WD, Markesbery WR (1996) Copper, iron, and zinc imbalances in severely degenerated brain regions in Alzheimer's disease: possible relation to oxidative stress. *J. Neurol Sci* 143: 137-142
30. Ehmann WD, Markesbery WR, Alauddin M, Hossain TIM, Brubaker EH (1986) Brain trace elements in Alzheimer's disease. *Neurotoxicology* 7: 197-206,
31. Esterbauer H, Schaur RJ, Zollner H (1991) Chemistry and biochemistry of 4- hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 11: 81-128
32. Evans PH (1993) Free radicals in brain metabolism and pathology. *Br Med Bull* 49: 577-587
33. Fleming J, Joshi JG: Ferritin (1987) Isolation of aluminum-ferritin complex from brain. *Proc Natl Acad Sci USA* 84: 7866-7870
34. Gabbita SP, Lovell MA, Markesbery WR (1998) Increased nuclear DNA oxidation in the brain in Alzheimer's disease. *J Neurochem* 71: 2034-2040
35. Goldgaber D, Harris HW, Hla T, Maciag T, Donnelly RJ, Jacobsen JS, Vitek MP, Gajdusek DC (1989) Interleukin-1 regulates synthesis of amyloid beta-protein precursor mRNA in human endothelial cells. *Proc Natl Acad Sci USA* 86: 7606-7610
36. Good PF, Perl DP, Bierer LM, Schmeidler J (1992) Selective accumulation of aluminum and iron in the neurofibrillary tangles of Alzheimer's disease: a laser microprobe (LAMMA) study. *Ann. Neurol.* 31: 286-292
37. Good PF, Werner P, Hsu A, Olanow CW, Perl DP (1996) Evidence of neuronal oxidative damage in Alzheimer's disease. *Am J Pathol* 149: 21-28
38. Grundke-Iqbal I, Fleming J, Tung Y-C, Lassmann H, Iqbal K, Joshi JG (1990) Ferritin is a component of the neuritic (senile) plaque in Alzheimer dementia. *Acta Neuropathol* 81: 105-110

39. Gsell W, Conrad R, Hickethier M, Sofic E, Frolich L, Wichart I, Jellinger K, Moll G, Ransmayr G, Beckmann H, Riederer P (1995) Decreased catalase activity but unchanged superoxide dismutase activity in brains of patients with dementia of Alzheimer type. *J Neurochem* 64(3): 1216-1223
40. Hajimohammadreza I, Brammer M (1990) Brain membrane fluidity and lipid peroxidation in Alzheimer's disease. *Neurosci Lett* 112: 333-337
41. Hall ED, Braughler JM (1989) Central nervous system trauma and stroke. II. Physiological and pharmacological evidence for involvement of oxygen radicals and lipid peroxidation. *Free Radic Biol Med* 6: 303-313
42. Halliwell B, Gutteridge JMC (1989) *Free Radicals in Biology and Medicine*, 2nd Edition, Clarendon Press: Oxford University Press, New York, NY, USA
43. Harris ME, Hensley K, Butterfield DA, Leedle RA, Carney JM (1995) Direct evidence of oxidative injury produced by the Alzheimer's beta-amyloid peptide (1-40) in cultured hippocampal neurons. *Exp Neurol* 131: 193-202
44. Hensley K, Carney JM, Hall N, Shaus W, Butterfield DA (1994 a) Electron paramagnetic resonance investigations of free radical induced alterations in neocortical synaptosomal membrane protein infrastructure. *Free Radic. Biol. Med.* 17: 321-331
45. Hensley K, Hall N, Subramaniam R, Cole P, Harris M, Aksenov M, Aksenova M, Gabbita SP, Wu JF, Carney JM, Lovell M, Markesbery WR, Butterfield DA (1994 a) Brain regional correspondence between Alzheimer's disease histopathology and biomarkers of protein oxidation. *J Neurochem* 65: 2146-2156 1995
46. Hermon M, Cairns N, Egly JM, Fery A, Labudova O, Lubec G (1998) Expression of DNA excision-repair-cross-complementing proteins p80 and p89 in brain of patients with Down syndrome and Alzheimer's disease. *Neurosci Lett* 251: 45-48
47. Hirai K, Smith MA, Wade R, Perry G (1998) Vulnerable neurons in Alzheimer disease accumulate mitochondrial DNA with the common 5KB deletion. *J Neuropathol Exp Neurol* 57: 511
48. Hunt JV, Smith CC, Wolff SP (1990) Autooxidative glycosylation and possible involvement of peroxides and free radicals in LDL modification by glucose. *Diabetes* 39: 1420-1424
49. Hunt JV, Wolff SP (1991) Oxidative glycation and free radical production: a causal mechanism of diabetic complications. *Free Radic Res Commun* 12-13: 115-123
50. Hunt JV, Bottoms MA, Mitchinson MJ (1993) Oxidative alterations in the experimental glycation model of diabetes mellitus are due to protein-glucose adduct oxidation. Some fundamental differences in proposed mechanisms of glucose oxidation and oxidant production. *Biochem J* 291: 529-535
51. Jagust WJ (1998) Neuroimaging of dementing disorders. In: *Neuropathology of Dementing Disorders*, Markesbery WR (ed). Chapter 2, pp. 25-55, Arnold: London, UK
52. Jefferies WA, Food MR, Gabathuler R, Rothenberger S, Yamada T, Yasuhara O, McGeer PL (1996) Reactive microglia specifically associated with amyloid plaques in Alzheimer's disease brain tissue express melanotransferrin. *Brain Res* 712: 122-126
53. Kato K, Kurobe N, Suzuki R, Morishita R, Asano T, Sato T, Inagaki T (1991) Concentrations of several proteins characteristic of nervous tissue in cerebral cortex of patients with Alzheimer's disease. *J Mol Neurosci* 3: 95-99
54. Keller JN, Pang Z, Geddes JW, Begley JG, Germeyer A, Waeg G, Mattson MP (1997) Impairment of glucose and glutamate transport and induction of mitochondrial oxidative stress and dysfunction in synaptosomes by amyloid  $\beta$ - peptide: role of the lipid peroxidation product 4-hydroxynonenal. *J Neurochem* 69: 273-284
55. Kennard ML, Feldman H, Yamada T, Jefferies WA (1996) Serum levels of the iron binding protein p97 are elevated in Alzheimer's disease. *Nat Med* 2: 1230-1235
56. Kish SJ, Morito CL, Hornykiewicz O (1986) Brain glutathione peroxidase in neurodegenerative disorders. *Neurochem Pathol* 4: 23-28
57. Kish SJ, Bergeron C, Rajput A, Dozic S, Mastrogiacomo F, Chang LJ, Wilson JM, Distefano LM, Nobrega JN (1992) Brain cytochrome oxidase in Alzheimer's disease. *J Neurochem* 59: 776-779
58. Klegeris A, Walker DG, McGeer PL (1994) Activation of macrophages by Alzheimer -amyloid peptide. *Biochem Biophys Res Commun* 199: 984-991
59. Kruman I, Bruce-Keller AJ, Bredesen D, Waeg G, Mattson MP (1997) Evidence that 4-hydroxynonenal mediates oxidative stress- induced neuronal apoptosis. *J Neurosci* 17: 5089-5100
60. Lovell MA, Ehmann WD, Butler SM, Markesbery WR (1995) Elevated thiobarbituric acid-reactive substances and antioxidant enzyme activity in the brain in Alzheimer's disease. *Neurology* 45:1594-1601
61. Lovell MA, Ehmann WD, Mattson MP, Markesbery WR (1997) Elevated 4- hydroxynonenal in ventricular fluid in Alzheimer's disease. *Neurobiol Aging* 18: 457-461
62. Lovell MA, Robertson JD, Teesdale WJ, Campbell JL, Markesbery WR (1998 a) Copper, iron and zinc in Alzheimer's disease senile plaques. *J Neurol Sci* 158: 47-52
63. Lovell MA, Markesbery WR (1998 b) Decreased glutathione transferase in brain and ventricular fluid in Alzheimer's disease. *Neurology*, in press
64. Lovell MA, Gabbita SP, Markesbery WR (1998 c) Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF. *J Neurochem*, in press
65. Lyras L, Cairns NJ, Jenner A, Jenner P, Halliwell B (1997) An assessment of oxidative damage to proteins, lipids, and DNA in brains from patients with Alzheimer's disease. *J Neurochem* 68: 2061-2069
66. Marcus DL, Thomas C, Rodriguez C, Simberkoff K, Tsai JS, Strafaci JA, Freedman ML (1998) Increased peroxidation and reduced antioxidant enzyme activity in Alzheimer's disease. *Exp Neurol* 150: 40-44



67. Mark RJ, Lovell MA, Markesbery WR, Uchida K, Mattson MP (1997 a) A role for 4-hydroxynonenal, an aldehydic product of lipid peroxidation, in disruption of ion homeostasis and neuronal death induced by amyloid  $\beta$ -peptide. *J Neurochem* 68: 255-264
68. Mark RJ, Pang Z, Geddes JW, Uchida K, Mattson MP (1997 b) Amyloid  $\beta$ - peptide impairs glucose transport in hippocampal and cortical neurons: involvement of membrane lipid peroxidation. *J Neurosci* 17: 1046-1054
69. Markesbery WR, Lovell MA (1998) Four-hydroxynonenal, a product of lipid peroxidation, is increased in the brain in Alzheimer's disease. *Neurobiol Aging* 19: 33-36
70. Marklund SL, Adolfsson R, Gottfries CG, Winblad B (1985) Superoxide dismutase isoenzymes in normal brains and in brains from patients with dementia of Alzheimer type. *J Neurol Sci* 67: 319-325
71. Marletta MA (1993) Nitric oxide synthase structure and mechanism. *J Biol Chem* 268: 12231-12234
72. Mattson MP, Lovell MA, Furukawa K, Markesbery WR (1995) Neurotrophic factors attenuate glutamate-induced accumulation of peroxides, elevation of intracellular  $CA^{2+}$  concentration, and neurotoxicity and increase antioxidant enzyme activities in hippocampal neurons. *J Neurochem* 65: 1740-1751
73. McGeer PL, McGeer EG (1995) The inflammatory response system of brain: implications for therapy of Alzheimer and other neurodegenerative diseases. *Brain Res Brain Res Rev* 21(2): 195-218
74. McGeer PL, Schulzer M, McGeer EG (1996) Arthritis and anti-inflammatory agents as possible protective factors for Alzheimer's disease: a review of 17 epidemiologic studies. *Neurology* 47: 425-432
75. McIntosh LJ, Trush MA, Troncoso JC (1997) Increased susceptibility of Alzheimer's disease temporal cortex to oxygen free radical-mediated processes. *Free Radic Biol Med* 23: 183-190
76. McMillian M, Kong LY, Sawin SM, Wilson B, Das K, Hudson P, Hong JS, Bing G (1995) Selective killing of cholinergic neurons by microglial activation in basal forebrain mixed neuronal/glia cultures. *Biochem Biophys Res Commun* 215: 572-577
77. Mecocci P, MacGarvey U, Kaufman AE, Koontz D, Shoffner JM, Wallace DC, Beal MF (1993) Oxidative damage to mitochondrial DNA shows marked age- dependent increases in human brain. *Ann Neurol* 34: 609-616
78. Mecocci P, MacGarvey U, Beal MF (1994) Oxidative damage to mitochondrial DNA is increased in Alzheimer's disease. *Ann Neurol* 36: 747-751
79. Miatto O, Gonzalez RG, Buonanno F, Growdon JH (1986) In vitro  $^{31}P$  NMR spectroscopy detects altered phospholipid metabolism in Alzheimer's disease. *Can J Neurol Sci* 13: 535-539
80. Montine TJ, Amarnath V, Martin ME, Strittmatter WJ, Graham DG (1996 a) E-4- hydroxy-2-nonenal is cytotoxic and cross-links cytoskeletal proteins in P19 neuroglial cultures. *Am J Pathol* 148: 89-93
81. Montine KS, Kim PJ, Olson SJ, Markesbery WR, Montine TJ (1997) 4-hydroxy- 2-nonenal pyrrole adducts in human neurodegenerative disease. *J Neuropathol Exp Neurol* 56(8): 866-871
82. Montine TJ, Markesbery WR, Morrow JD, Roberts LJ (1998 a) Cerebrospinal fluid F2 isoprostane levels are increased in Alzheimer's disease. *Ann Neurol* 44: 410-413
83. Montine KS, Recch E, Neely MD, Sidell KR, Olson SJ, Markesbery WR, Montine TJ (1998 b) Distribution of reducible 4-hydroxynonenal adduct immunoreactivity in Alzheimer disease is associated with APOE genotype. *J Neuropathol Exp Neurol* 57: 415-425
84. Morrow JD, Roberts LJ (1997) The isoprostanes: unique bioactive products of lipid peroxidation. *Prog Lipid Res* 36: 1-21
85. Mullaart E, Boerrieger ME, Swabb RR, Vijg J (1990) Increased levels of DNA breaks in cerebral cortex of Alzheimer's disease patients. *Neurobiol Aging* 11: 169-173
86. Mutisya EM, Bowling AC, Beal MF (1994) Cortical cytochrome oxidase activity is reduced in Alzheimer's disease. *J Neurochem* 63: 2179-2184
87. Münch G, Thome J, Foley P, Schinzel R, Riederer P (1997) Advanced glycation endproducts in ageing and Alzheimer's disease. *Brain Res Rev* 23: 134-143
88. Namekata K, Imagawa M, Terashi A, Ohta S, Oyama F, Ihara Y (1997) Association of transferrin C2 allele with late-onset Alzheimer's disease. *Hum Genet* 101: 126-129
89. Neeper M, Schmidt AM, Brett J, Yan SD, Wang F, Pan YC, Elliston K, Stern D, Shaw A (1992) Cloning and expression of a cell surface receptor for advanced glycosylation end products of proteins. *J Biol Chem* 267:(21)14998-15004.
90. Nitsch RM, Blusztajn JK, Pittas AG, Slack BE, Growdon JH, Wurtman RJ (1992) Evidence for a membrane defect in Alzheimer disease brain. *Proc Natl Acad Sci USA* 89: 1671-1675
91. O'Banion MK, Miller JC, Chang JW, Kaplan MD, Coleman PD (1996) Interleukin 1-B induces prostaglandin G/H synthase-2 (cyclooxygenase-2) in primary murine astrocyte cultures. *J Neurochem* 66: 2532-2546
92. Pappolla MA, Omar RA, Kim KS, Robakis NK (1992) Immunohistochemical evidence of oxidative stress in Alzheimer's disease. *Am J Pathol* 140: 621-628
93. Parker WD, Parks JK, Filley CM, Kleinschmidt-DeMasters BK (1994) Electron transport chain defects in Alzheimer's disease brain. *Neurology* 44: 1090-1096
94. Parker WD, Parks JK (1995) Cytochrome c oxidase in Alzheimer's disease brain: purification and characterization. *Neurology* 45: 482-486
95. Partridge RS, Monroe SM, Parks JK, Johnson K, Parker WD, Eaton GR, Eaton SS (1994) Spin trapping of azidyl and hydroxyl radicals in azide-inhibited rat brain sub-mitochondrial particles. *Arch Biochem Biophys* 310: 210-217
96. Pettigrew JW, Moosy J, Withers G, McKeag D, Panchalingam K (1988)  $^{31}P$  nuclear magnetic resonance study of the brain in Alzheimer's disease. *J Neuropathol Exp Neurol* 47: 235-248



97. Prasad MR, Lovell MA, Yarkin M, Dhillon H, Markesbery WR (1998) Regional membrane phospholipid alterations in Alzheimer's disease. *Neurochem Res* 23: 81-88
98. Premkumar DRD, Smith MA, Richey PL, Petersen RB, Castellani R, Kutty RK, Wiggert B, Perry G, Kalaria RN (1995) Induction of heme oxygenase-1 mRNA and protein in neocortex and cerebral vessels in Alzheimer's disease. *J Neurochem* 65: 1399-1402
99. Reiman EM, Caselli RJ, Yun LS, Chen K, Bandy D, Minoshima S, Thibodeau SN, Osborne D (1996) Preclinical evidence of Alzheimer's disease in persons homozygous for the epsilon 4 allele for apolipoprotein E. *N Engl J Med* 334: 752-758
100. Richardson JS (1993) Free radicals in the genesis of Alzheimer's disease. *Ann NY Acad Sci* 695: 73-76
101. Roberts LJ, Montine TJ, Markesbery WR, Tapper AR, Hardy P, Chemtob S, Dettbarn WD, Morrow JD (1998) Formation of isoprostane-like compounds (neuroprostanes) in vivo from docosahexaenoic acid. *J Biol Chem* 273: 13605-13612
102. Rogers J, Kirby LC, Hempelman SR, Berry DL, McGeer PL, Kaszniak AW, Zalski J, Cofield M, Mansukhani L, Wilson P (1993) Clinical trial of indomethacin in Alzheimer's disease. *Neurology* 43: 1609-1611
103. Roses AD (1996) Apolipoprotein E alleles as risk factors in Alzheimer's disease. *Ann Rev Med* 47: 387-400
104. Samudralwar DL, DiPrete CC, Ni BF, Ehmann WD, Markesbery WR (1995) Elemental imbalances in the olfactory pathway in Alzheimer's disease. *J Neurol Sci* 130:139-145
105. Sano M, Ernesto C, Thomas RG, Klauber MR, Schafer K, Grundman M, Woodbury P, Growdon J, Cotman CW, Pfeiffer E, Schneider LS, Thal LJ (1997) A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease. The Alzheimer's Disease Cooperative Study. *N Engl J Med* 336: 1216-1222
106. Sayre LM, Zelasko DA, Harris PL, Perry G, Salomon RG, Smith MA (1997) 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J Neurochem* 68: 2092-2097
107. Schipper HM, Cisse S, Stopa EG (1995) Expression of heme oxygenase-1 in the senescent and Alzheimer-diseased brain. *Ann Neurol* 37: 758-768
108. Schmidt A, M., Vianna M, Gerlach M, Brett J, Ryan J, Kao J, Esposito C, Hegarty H, Hurley W, Clauss M (1992) Isolation and characterization of two binding proteins for advanced glycosylation end products from bovine lung which are present on the endothelial surface. *J Biol Chem* 267(21): 14987-14997
109. Simonian NA, Hyman BT (1993) Functional alterations in Alzheimer's disease: diminution of cytochrome oxidase in the hippocampal formation. *J Neuropathol Exp Neurol* 52: 580-585
110. Simonian NA, Hyman BT (1994) Functional alterations in Alzheimer's disease: selective loss of mitochondrial-encoded cytochrome oxidase mRNA in the hippocampal formation. *J Neuropathol Exp Neurol* 53:508-512
111. Smale G, Nichols NR, Brady DR, Finche CE, Horton WE (1995) Evidence for apoptotic cell death in Alzheimer's disease. *Exp Neurol* 133: 225-230
112. Small GW, Mazziotta JC, Collins MT, Baxter LR, Phelps ME, Mandelkern MA, Kaplan A, LaRue A, Adamson CF, Chang LEA (1995) Apolipoprotein E type 4 allele and cerebral glucose metabolism in relatives at risk for familial Alzheimer disease. *JAMA* 273: 942-947
113. Smith CD, Carney JM, Starke-Reed PE, Oliver CN, Stadtman ER, Floyd RA, Markesbery WR (1991) Excess brain protein oxidation and enzyme dysfunction in normal aging and Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 88: 10540-10543
114. Smith MA, Taneda S, Richey PL, Miyata S, Yan S-D, Stern D, Sayre LM, Monnier VM, Perry G (1994 a) Advanced Maillard reaction end products are associated with Alzheimer disease pathology. *Proc Natl Acad Sci USA* 91: 5710-5714
115. Smith MA, Kutty RK, Richey PL, Yan SD, Stern D, Chader GJ, Wiggert B, Petersen RB, Perry G (1994 b) Heme oxygenase-1 is associated with the neurofibrillary pathology of Alzheimer's disease. *Am J Pathol* 145: 42-47
116. Smith MA, Sayre LM, Monnier VM, Perry G (1995) Radical AGEing in Alzheimer's disease. *Trends Neurosci* 18: 172-176
117. Smith MA, Perry G, Richey PL, Sayre LM, Anderson VE, Beal MF, Kowall N (1996) Oxidative damage in Alzheimer's. *Nature* 382: 120-121
118. Smith MA, Harris PLR, Sayre LM, Perry G (1997 a) Iron accumulation in Alzheimer disease is a source of redox-generated free radicals. *Proc Natl Acad Sci USA* 94: 9866-9868
119. Smith MA, Richey-Harris P, Sayre LM, Beckman JS, Perry G (1997 b) Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J Neurosci* 17: 2653-2657
120. Sohal RS (1997) Role of mitochondria and oxidative stress in the aging process. In: *Mitochondria and Free Radicals in Neurodegeneration Diseases*, Beal MF, Howell N, Bodis-Wollner I (eds.), Chapter 5, pp 91-107, Wiley-Liss, Inc: New York, NY, USA
121. Stadelmann C, Bruck W, Bancher C, Jellinger K, Lassman H (1998) Alzheimer disease: DNA fragmentation indicates increased neuronal vulnerability, but not apoptosis. *J Neuropathol Exp Neurol* 57:456-464.
122. Stadtman ER (1992) Protein oxidation and aging. *Science* 257: 1220-1224
123. Su JH, Anderson AJ, Cummings BJ, Cotman CW (1994) Immunohistochemical evidence for apoptosis in Alzheimer's disease. *NeuroReport* 5: 2529-2533
124. Su JH, Deng G, Cotman CW (1997) Neuronal DNA damage precedes tangle formation and is associated with up-regulation of nitrotyrosine in Alzheimer's disease brain. *Brain Res* 774: 193-197
125. Subbarao KV, Richardson JS, Ang LC (1990) Autopsy samples of Alzheimer's cortex show increased peroxidation in vitro. *J Neurochem* 55: 342-345

126. Svennerholm L, Gottfries CG (1994) Membrane lipids, selectively diminished in Alzheimer brains, suggest synapse loss as a primary event in early-onset (type I) and demyelination in late-onset form (type II). *J Neurochem.* 62: 1039-1047
127. Uchida H, Stadtman ER (1992) Modification of histidine residues in proteins by reaction with 4-hydroxynonenal. *Proc Natl Acad Sci USA* 89: 4544-4589
128. Vane JR, Bakhle YS, Botting RM (1998) Cyclooxygenases 1 and 2. *Annu. Rev. Pharmacol Toxicol* 38: 97-120
129. Vitek MP, Bhattacharya K, Glendening JM, Stopa E, Vlassara H, Bucala R, Manogue K, Cerami A (1994) Advanced glycation end products contribute to amyloidosis in Alzheimer disease. *Proc. Natl. Acad. Sci. USA* 91: 4766-4770
130. Wade R, Hirai K, Perry G, Smith MA (1998) Accumulation of 8-hydroxyguanosine in neuronal cytoplasm indicates mitochondrial damage and radical production are early features of Alzheimer disease. *J Neuropathol Exp Neurol* 57: 511
131. Walker DG, Kim SU, McGeer PL (1995) Complement and cytokine gene expression in cultured microglia derived from postmortem human brains. *J Neurosci Res* 40: 478-493
132. Yamabe T, Dhir G, Cowan EP, Wolf AL, Bergey GK, Krumholz A, Barry E, Hoffman PM, Dhib-Jalbut S (1994) Cytokine-gene expression in measles- infected adult human glial cells. *J Neuroimmunol* 49: 171-179
133. Yan SD, Chen X, Schmidt AM, Brett J, Godman G, Zou YS, Scott CW, Caputo C, Frappier T, Smith MA, Perry G, Yen SH, Stern D (1994) Glycated tau protein in Alzheimer disease: a mechanism for induction of oxidant stress. *Proc Natl Acad Sci USA* 91: 7787-7791
134. Yan SD, Chen X, Fu J, Chen M, Zhu H, Roher A, Slattery T, Zhao L, Nagashima M, Morser J, Migheli A, Nawroth P, Stern D, Schmidt AM (1996) RAGE and amyloid-beta peptide neurotoxicity in Alzheimer's disease. *Nature* 382: 685-691
135. Yan SD, Zhu H, Fu J, Yan SF, Roher A, Tourtellotte WW, Rajavashisth T, Chen X, Godman GC, Stern D, Schmidt AM (1997) Amyloid-B peptide-receptor for advanced glycation endproduct interaction elicits neuronal expression of macrophage-colony stimulating factor: a proinflammatory pathway in Alzheimer disease. *Proc Natl Acad Sci, USA* 94: 5296-5301
136. Yatin SM, Aksenova M, Aksenova M, Markesbery WR, Aulick T, Butterfield DA (1998) Temporal relations between amyloid B-peptide-induced free radical oxidative stress and neuronal toxicity and neuronal defense responses. Submitted