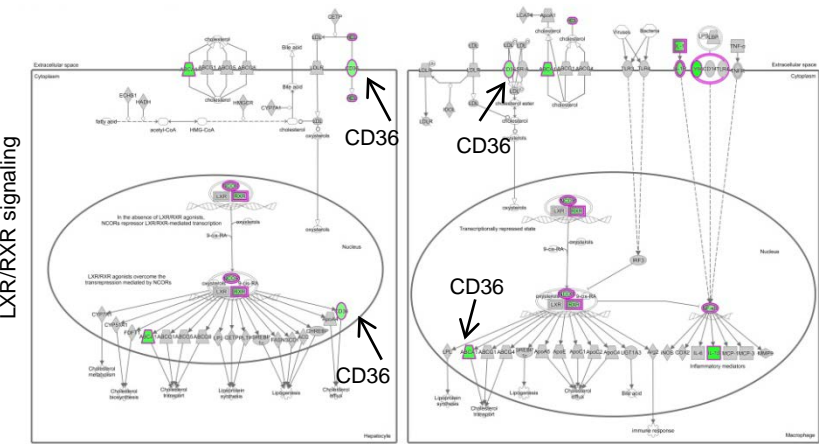
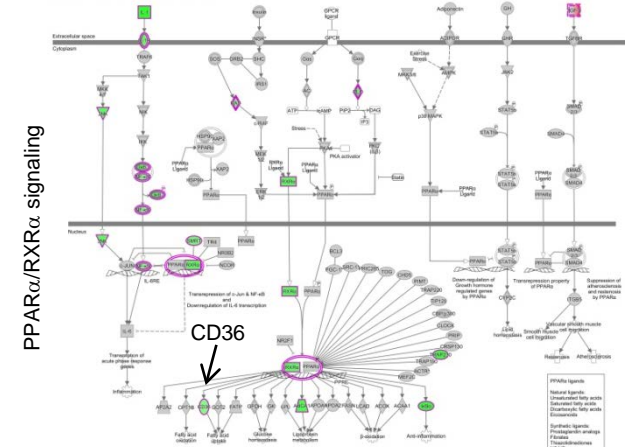


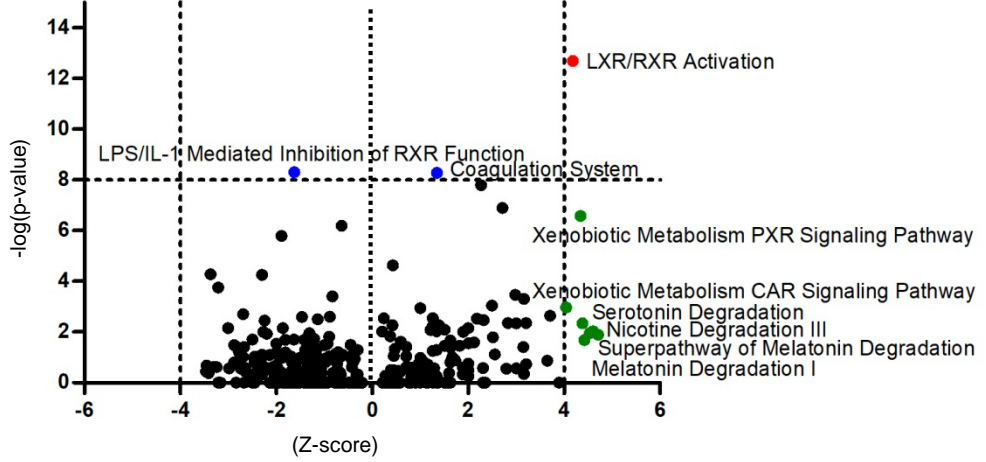
**A Long survival vs short survival**



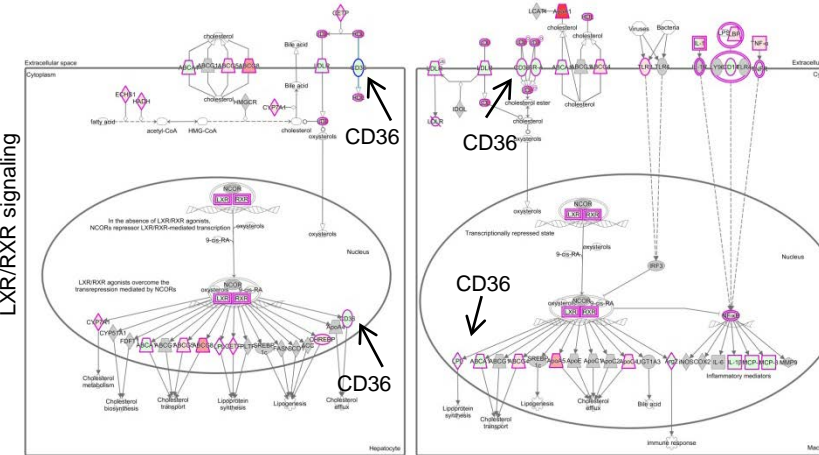
**B Long survival vs short survival**



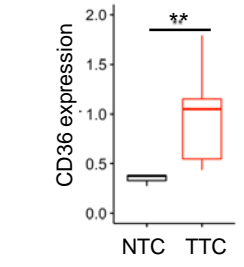
**C PD-1 response vs PD-1 non-response**



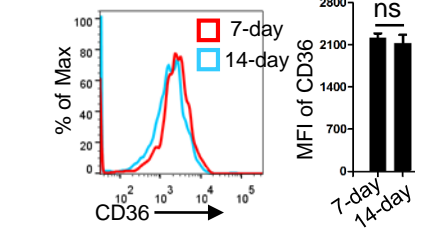
**D PD-1 response vs PD-1 non-response**



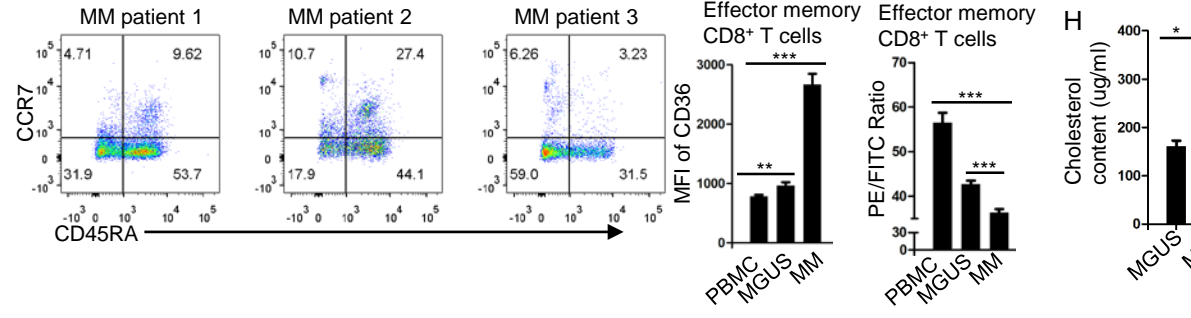
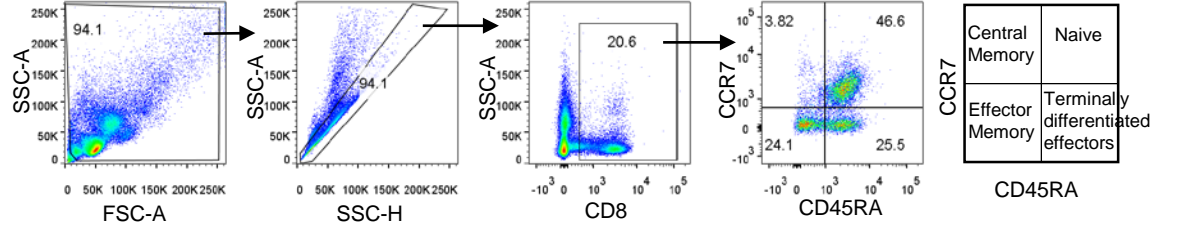
**E NSCLC tumor infiltrating CD8+ T cells**



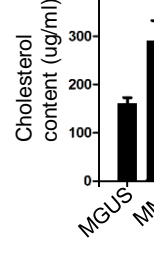
**G s.c. B16 tumor infiltrating CD4+ T cells**



**F PBMC**

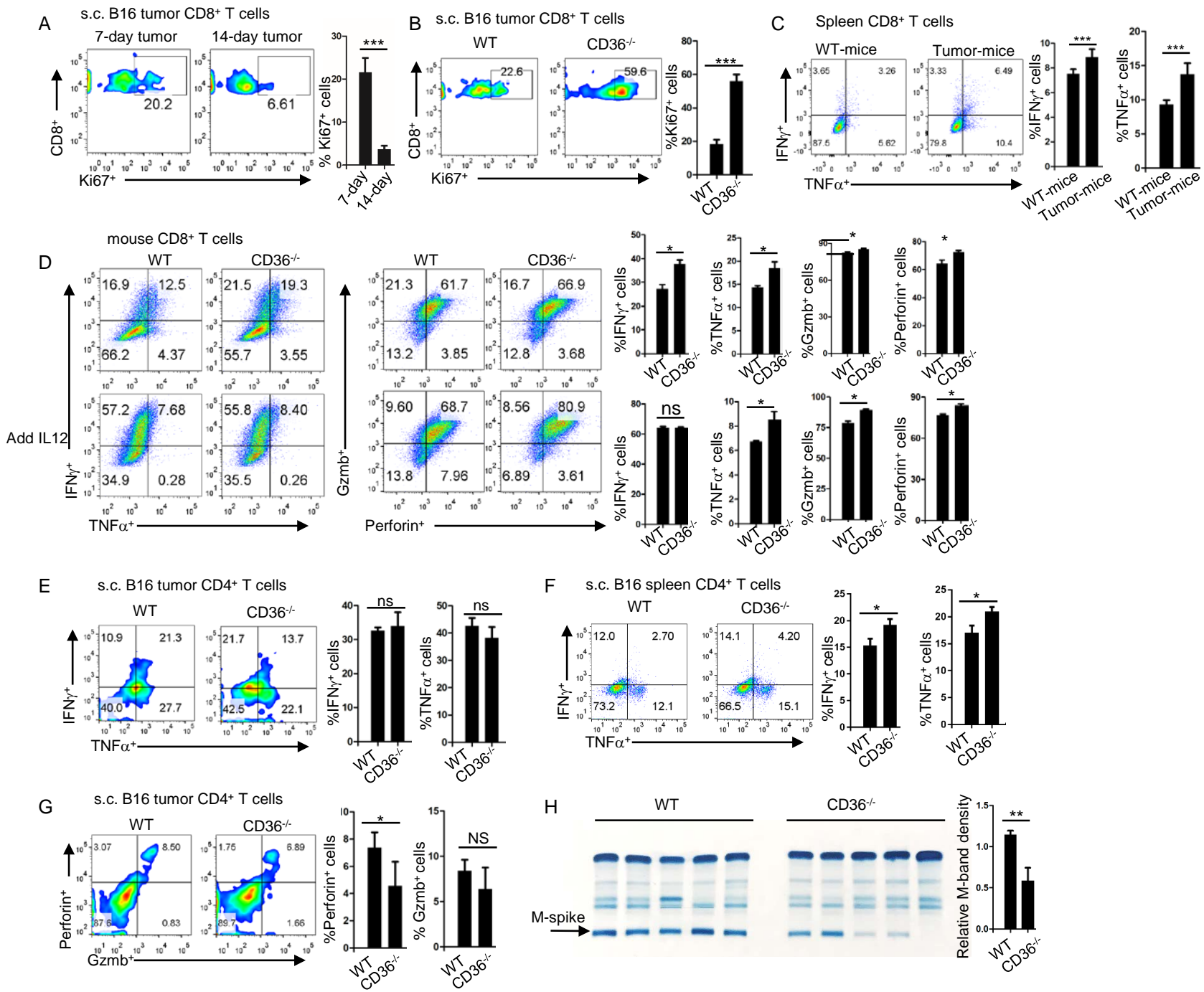


**H**



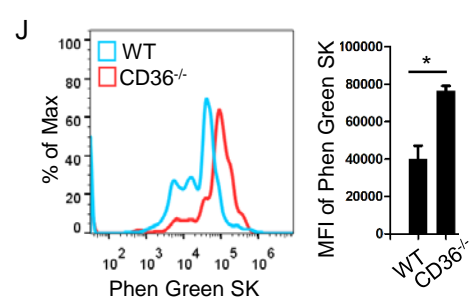
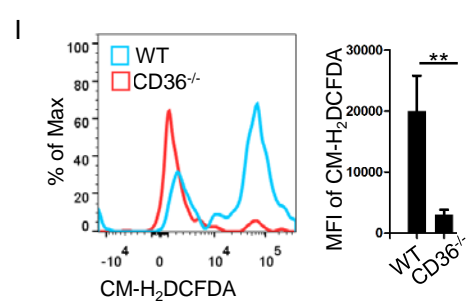
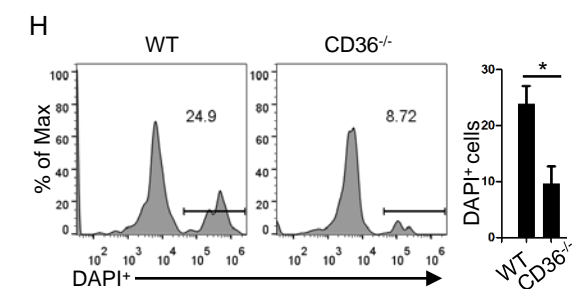
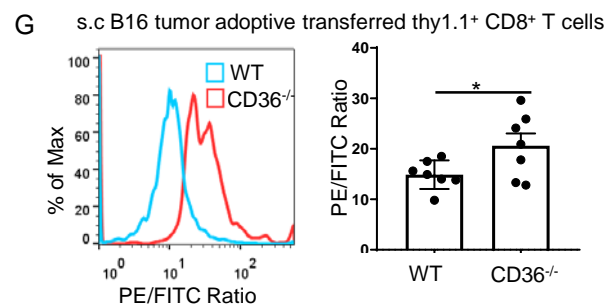
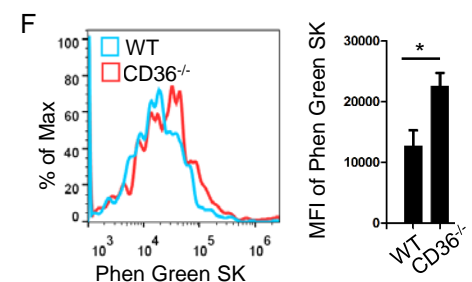
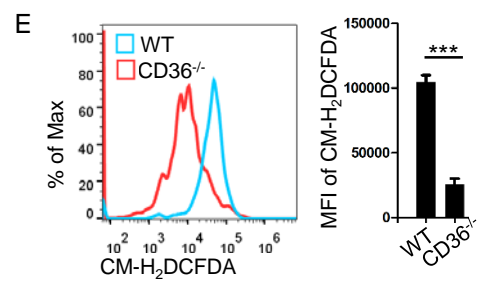
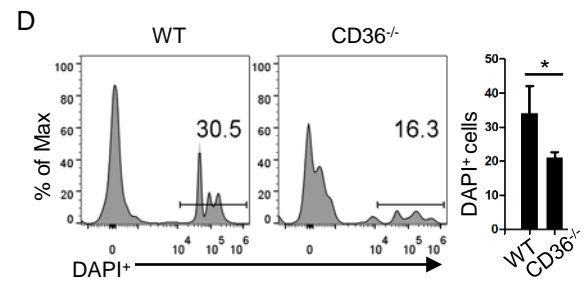
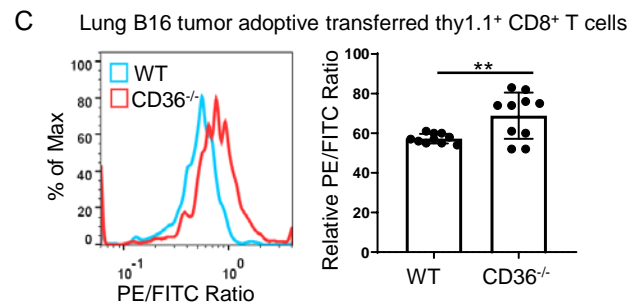
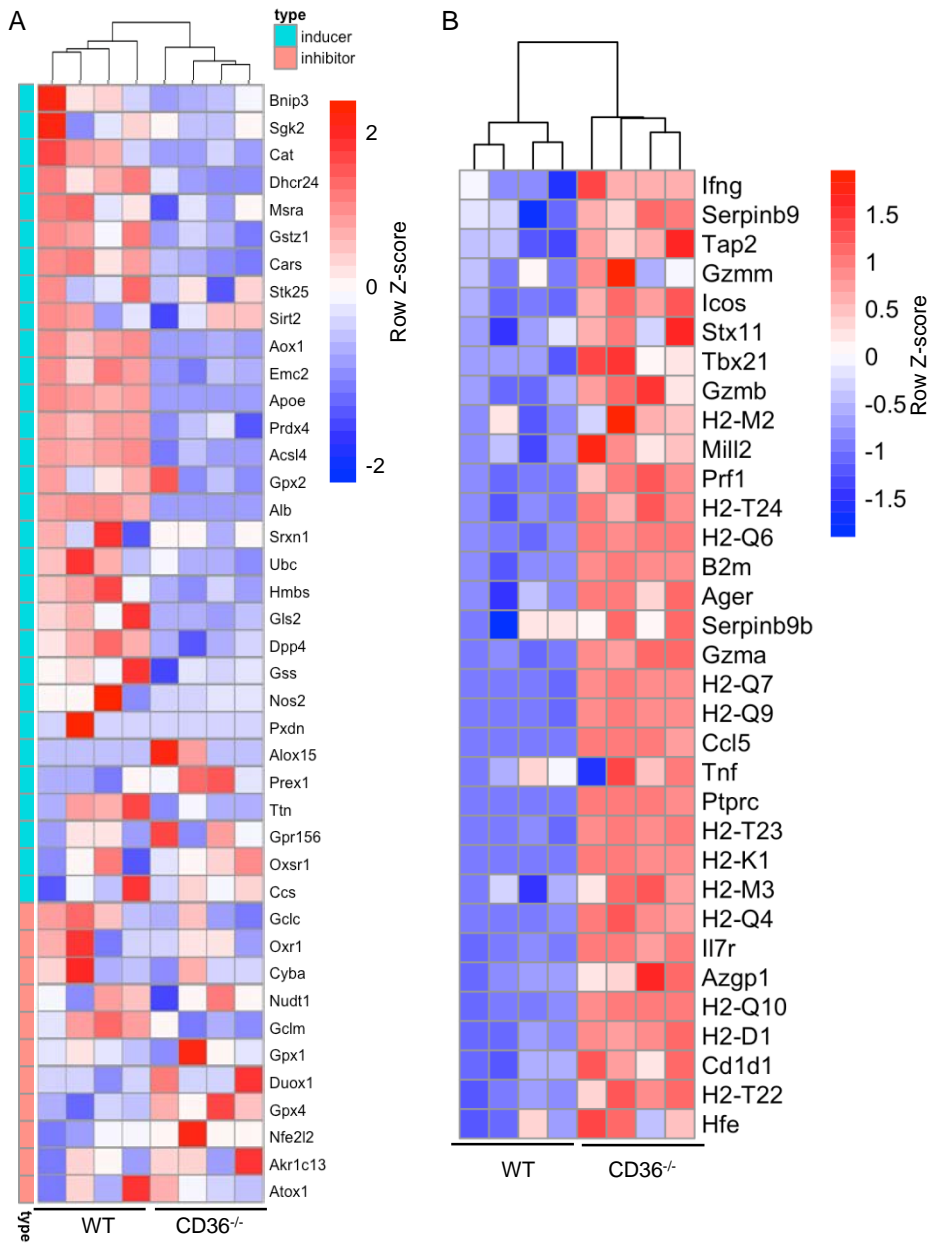
**Supplementary Figure 1. Increased CD36 expression on tumor-infiltrating CD8<sup>+</sup> T cells is associated with tumor progression and poor survival in human and murine cancers, Related to Figure 1.**

**(A and B)** IPA analysis of canonical pathway changes in tumor-infiltrating CD8<sup>+</sup> T cells between long- and short-survival melanoma patients. Shown are LXR/RXR signaling (A) and PPAR $\alpha$ /RXR $\alpha$  signaling (B). Red: upregulated; Green: downregulated. **(C and D)** IPA analysis of canonical pathway changes between PD-1 treatment responder and non-responder melanoma patients (C). Shown is LXR/RXR signaling (D). Red: upregulated; Green: downregulated. **(E)** CD36 expression of CD8<sup>+</sup> T cells from non-small-cell lung cancer (NSCLC) tumor tissues and normal tissues. NTC: normal adjacent tissue cells; TTC: tumor tissue cells. Data are presented as mean  $\pm$  SEM. \*\*p < 0.01. **(F)** CD8<sup>+</sup> T cells from MM or MGUS patient's bone marrow and blood were analyzed for the expression of CCR7, CD45RA, CD36 and lipid peroxidation. **(G)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Tumor-infiltrating CD4<sup>+</sup> T cells were analyzed for the expression of CD36 at days 7 and 14 after tumor injection. **(H)** Cholesterol content in bone marrow plasma of MGUS and MM patients. Data are presented as mean  $\pm$  SEM. \*\*p < 0.01; \*\*\*p < 0.001.



**Supplementary Figure 2. Expression of CD36 reduces cytotoxic cytokine production in murine CD8<sup>+</sup> T cells but not CD4<sup>+</sup> T cells, Related to Figure 2.**

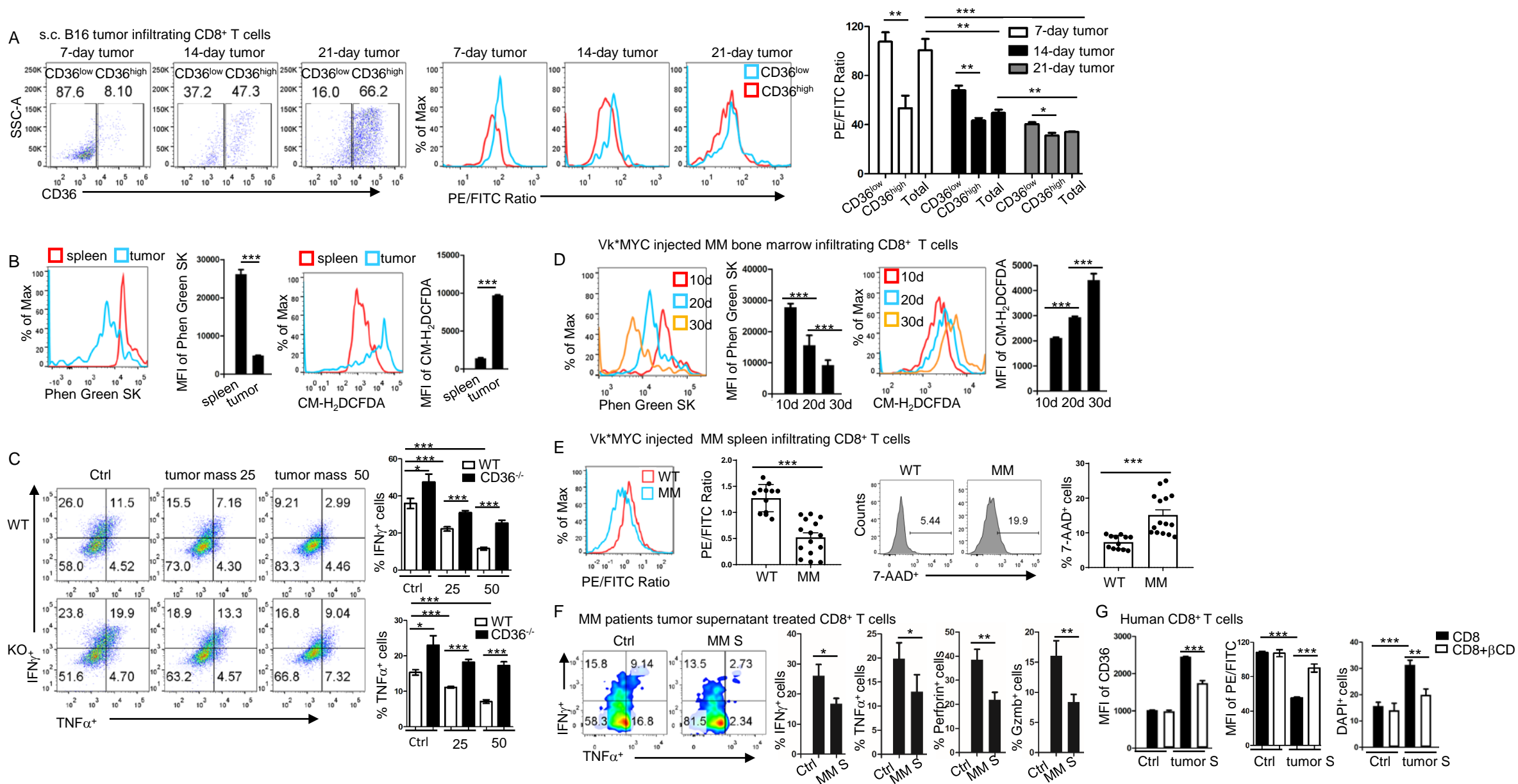
**(A)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Tumor-infiltrating CD8<sup>+</sup> T cells were analyzed for the expression of Ki67 at day 7 and 14 after tumor injection. **(B)** WT and CD36<sup>-/-</sup> B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Tumor-infiltrating CD8<sup>+</sup> T cells were analyzed for the expression of Ki67 at day 14 after tumor injection. **(C)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells and maintained for 10 days. Spleen CD8<sup>+</sup> T cells from tumor free or tumor bearing mice were analyzed for the expression of IFN $\gamma$  and TNF $\alpha$ . **(D)** Splenocytes from WT Pmel-1 and CD36<sup>-/-</sup> Pmel-1 mice were differentiated in vitro for 3 days in the presence of hgp100<sub>25-33</sub> peptide (1  $\mu$ g/ml) and IL-2 (10 ng/ml). Some of the cells were added with IL-12 as indicated. T cells were examined for the production of IFN $\gamma$ , TNF $\alpha$ , Gzmb and perforin on day 3. **(E-G)** WT and CD36<sup>-/-</sup> B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Tumor-infiltrating (E and G) and spleen (F) CD4<sup>+</sup> T cells were analyzed for the expression of IFN $\gamma$ , TNF $\alpha$ , Gzmb or perforin at day 14 after tumor injection. **(H)** WT and CD36<sup>-/-</sup> B6 mice were injected i.v. with  $1.5 \times 10^6$  Vk\*MYC cells. Tumor burden is shown as bands of M-spike at day 28. Data are presented as mean  $\pm$  SEM. \*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001.



**Supplementary Figure 3. CD36 regulates transcriptional and metabolic programs including lipid peroxidation in tumor-infiltrating murine CD8<sup>+</sup> T cells, Related to Figure 3.**

WT and CD36<sup>-/-</sup> B6 mice were injected i.v. with  $2 \times 10^5$  B16 cells, and 16 days later, spleen (A) and lung (B) of tumor-bearing mice were collected, sorted for CD8<sup>+</sup> T cells, and performed RNAseq on T cells. **(A)** Heatmap of lipid peroxidation- and ferroptosis-related genes. **(B)** Heatmap of T cell cytotoxicity-related genes. **(C-J)** Splenocytes from WT Pmel-1 and CD36<sup>-/-</sup> Pmel-1 mice were differentiated in vitro for 5 days in the presence of hgp100<sub>25-33</sub> peptide (1  $\mu$ g/ml) and IL-2 (10 ng/ml) and then adoptively transferred into tumor-bearing mice. **(C-F)** B6 mice were injected i.v. with  $2 \times 10^5$  B16 cells. At day 12 after tumor inoculation,  $2 \times 10^6$  WT or CD36<sup>-/-</sup> CD8<sup>+</sup> Pmel-1 T cells were i.v. injected into tumor-bearing mice. At day 16, WT or CD36<sup>-/-</sup> Pmel-1 CD8<sup>+</sup> T cells in tumor were analyzed for lipid peroxidation, cell death, cytosolic ROS and iron. **(G-J)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Two weeks after tumor inoculation,  $2 \times 10^6$  WT or CD36<sup>-/-</sup> Pmel-1 CD8<sup>+</sup> T cells were i.v. injected into tumor-bearing mice. Three days after T cell transfer, WT or CD36<sup>-/-</sup> Pmel-1 CD8<sup>+</sup> T cells in tumor were analyzed for lipid peroxidation, cell death, cytosolic ROS and iron. Data are presented as mean  $\pm$  SEM. \* $p < 0.05$ ; \*\* $p < 0.01$ ; \*\*\* $p < 0.001$ .

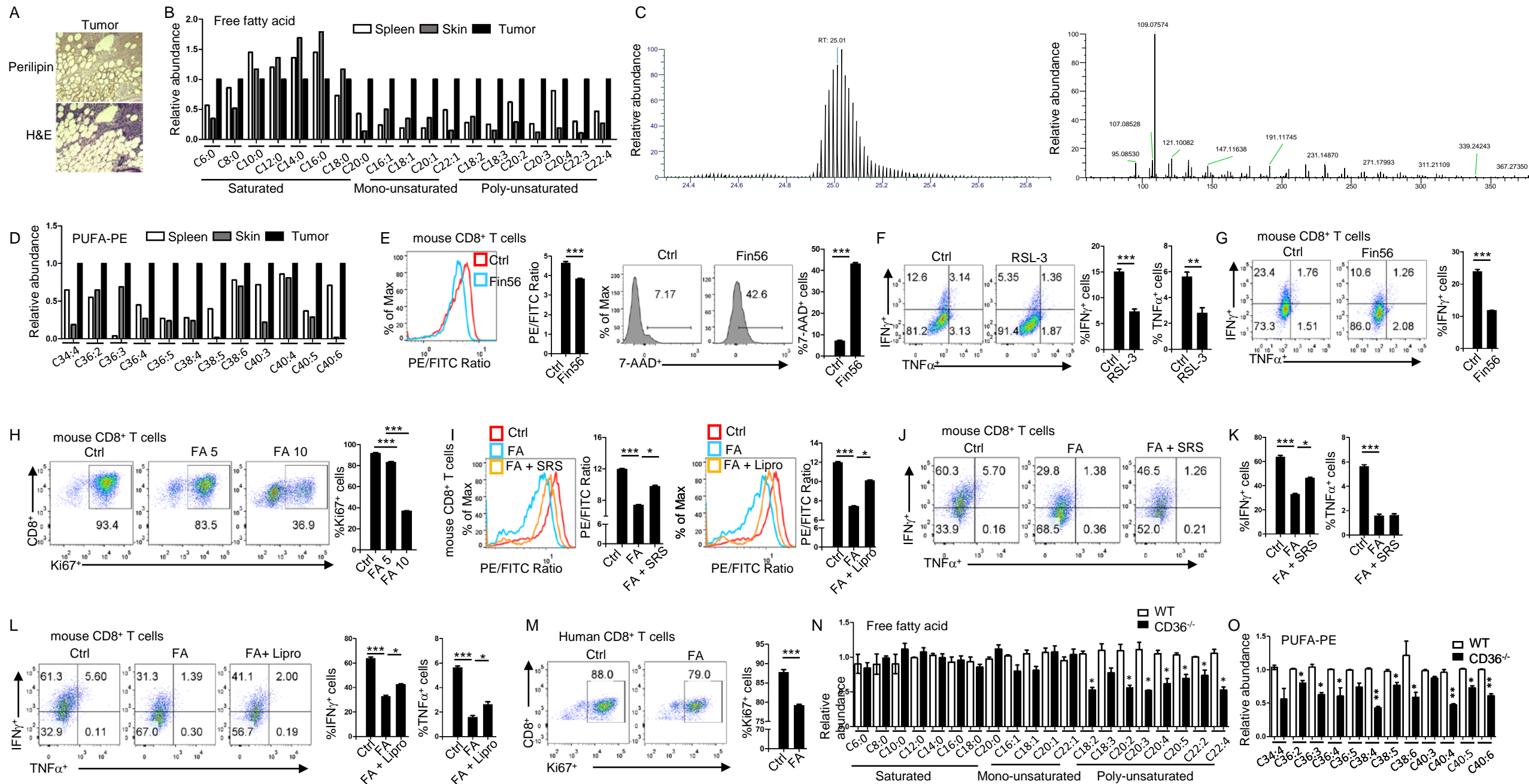




**Supplementary Figure 4. CD36 mediates ferroptosis and reduces cytotoxic cytokine production in human and murine CD8<sup>+</sup> T cells, Related to Figure 4.**

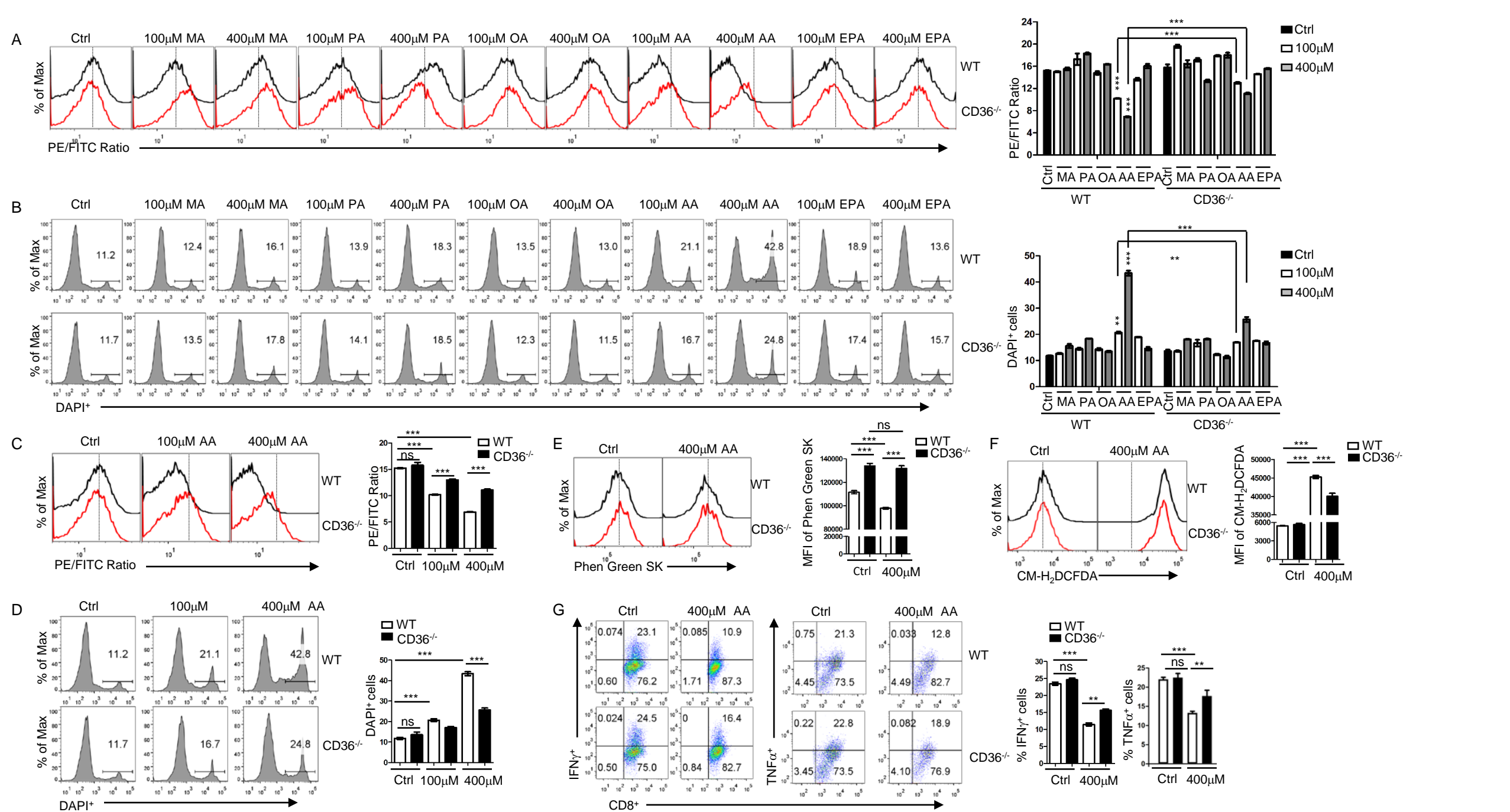
**(A)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. At days 7, 14 and 21 after tumor inoculation, tumor-infiltrating CD8<sup>+</sup> T cells were analyzed for CD36 expression and lipid peroxidation. **(B)** B6 mice were injected s.c. with  $1 \times 10^6$  B16 cells. Spleen and tumor-infiltrating CD8<sup>+</sup> T cells were analyzed for iron and ROS at day 14 after tumor injection. **(C)** WT Pmel-1 or CD36<sup>-/-</sup> Pmel-1 CD8<sup>+</sup> T cells were isolated and in vitro stimulated with CD3/CD28 antibodies in the presence of IL-2. Tumor mass (25 or 50  $\mu$ l at 1 mg/ml) was added to the culture of T cells for 1 day as indicated. Cultured T cells were analyzed for IFN $\gamma$  and TNF $\alpha$  expression. **(D)** B6 mice were injected i.v. with  $1.5 \times 10^6$  Vk\*MYC cells, and tumor-infiltrating CD8<sup>+</sup> T cells from bone marrow were analyzed for iron and cytosolic ROS at days 10, 20 and 30 after tumor injection. **(E)** B6 mice were injected i.v. with  $1.5 \times 10^6$  Vk\*MYC cells. Lipid peroxidation and 7-AAD level of spleen CD8<sup>+</sup> T cells from 4-week Vk\*MYC MM-bearing mice or tumor-free mice were examined. **(F)** Human CD8<sup>+</sup> T cells were isolated from blood and in vitro stimulated with CD3/CD28 beads in the presence of IL-2. MM patient bone marrow plasma (100  $\mu$ l) was added to the culture of T cells during T cell differentiation as indicated. Cultured human CD8<sup>+</sup> T cells were analyzed for Perforin, Gzmb, IFN $\gamma$ , or TNF $\alpha$  production. **(G)** Human CD8<sup>+</sup> T cells were isolated from blood and in vitro stimulated with CD3/CD28 beads in the presence of IL-2. MM patient bone marrow plasma (100  $\mu$ l) and  $\beta$ -CD (0.5 mM) were added to the culture of T cells during T cell differentiation as indicated. Cultured human CD8<sup>+</sup> T cells were analyzed for CD36, lipid peroxidation and cell death. Data are presented as mean  $\pm$  SEM. S: supernatant. \*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001.





**Supplementary Figure 5. CD36 mediates ferroptosis and reduces cytotoxic cytokine production in CD8<sup>+</sup> T cells through uptake of fatty acid, Related to Figure 5.**

**(A)** Immunohistochemistry showing the staining of perilipin (top) and H&E (bottom) on s.c. B16 tumors. **(B)** LC-MS analysis of indicated free fatty acid contents in tumor, adjacent skin, and spleen. **(C)** Representative normal-phase LS-MS chromatogram and mass spectra for arachidonic acid (AA). **(D)** LC-MS analysis of indicated poly unsaturated fatty acyl-phosphatidylethanolamines (PUFA-PE) contents in tumor, adjacent skin, and spleen. **(E-L)** Pmel-1 CD8<sup>+</sup> T cells were isolated and in vitro stimulated with CD3/CD28 antibodies in the presence of IL-2. **(E-G)** Fin56 (10 μM) or RSL-3 (10 μM) was added during T-cell differentiation. Lipid peroxidation, 7-AAD and cell viability (E), and IFN $\gamma$  or TNF $\alpha$  expression (F and G) of the cells were measured. **(H)** Fatty acid (5 or 10 μl) was added during T-cell differentiation as indicated. Ki67 ratio of the cells was measured. **(I-L)** Fatty acid (5 μl), SRS (10 μM), or Lipro (2 μM) was added to the culture of T cells during T-cell differentiation as indicated. Cultured T cells were analyzed for lipid peroxidation (I) and IFN $\gamma$  or TNF $\alpha$  expression (J-L). **(M)** Human CD8<sup>+</sup> T cells were isolated from blood and in vitro stimulated with CD3/CD28 beads in the presence of IL-2. Fatty acid (5 μl) was added during T-cell differentiation as indicated. Ki67 expression of the cells was measured. **(N and O)** LC-MS analysis of indicated free fatty acid contents (N) and PUFA-PE contents (O) in tumor-treated WT and CD36<sup>-/-</sup> CD8<sup>+</sup> T cells. Lipro: liproxstatin-1. SRS: SRS16-86. Data are presented as mean  $\pm$  SEM. \*p < 0.05; \*\*p < 0.01; \*\*\*p < 0.001.



**Supplementary Figure 6. AA is an effective component in fatty acids that induces CD36-mediated ferroptosis and reduces cytotoxic cytokine production in CD8<sup>+</sup> T cells, Related to Figure 5.**

WT Pmel-1 or CD36<sup>-/-</sup> Pmel-1 CD8<sup>+</sup> T cells were isolated and in vitro stimulated with CD3/CD28 antibodies in the presence of IL-2. Different free fatty acids were added during T cell differentiation as indicated. Lipid peroxidation (**A and C**), cell death (**B and D**), iron (**E**), cytosolic ROS (**F**), and IFN $\gamma$  or TNF $\alpha$  expression (**G**) of the cells were measured. Data are presented as mean  $\pm$  SEM. \*\*p < 0.01; \*\*\*p < 0.001.