# **Expanded View Figures**

Α						В	
BRAF	51%						
NRAS	30%		-	-			
NF1	13%		1 🚥			elR	10
SLC9A5	2,8%	10.0		1.		8M	1.0
SMAD3	2,4%					X	
BIRC3	5%	1.1				<u>ک</u> (5	
GAGE1	0,3%					nsit	0.5- **
DCAF17	0,7%	1 (C)				de	
ANGPT4	5%	0.0	111	0.1		Bell	
YAP1	2,8%					Ve	
YY2	1,4%	1				alti	DMSO Birinanai
SLC25A41	1,4%	1 I. I.		1		R	Billoo Billiapa
SLC22A18AS	0%						
GTF2A1	1,7%			1			
ACSM2B	15%	1 111 11110000	1 H H	18.18			
CDKN2C	1,4%			1.1			
KCNQ3	17%	1100 110 0		11		С	
VAV1	6%	1 0 1 1 1		0.1.1.1		-	
AFAP1	2,1%	0	1.1			5)	
PDE6A	6%	100 1	1.0	1.1		64	1.0 J
RRAD	2,4%	1	1 I.	1		Ve.	
TGIF1	1,7%		1 - C	T.		لک لک	**
TMEM59L	1,4%	1	1 I I			insi	0.5 -
PCOLCE	4%	10 I. I. I.	1 I.			lde	
GADL1	3%	- 1 (C)	1 - E	1.00		8	
FBXO38	5%	1 I I I I I I I I I I I I I I I I I I I	1	1.11		ive	
KLK8	5%	0.1.1	1.1	0 0 0		elat	
SREK1	2,1%	1	1	1.1		Ř	DMSO Birinapai
LCE1D	4%	the second second		1.00			
HEPACAM2	5%	<b>II</b> 1.111	1	1			
ARL13B	2,1%		1	1			
RAD18	2,4%	1 I. I.	1 - E				
CRYGB	2,1%	1	1	1.11			
SGMS2	1%	- 1					
ZNF211	3%	1.1.1.1		1.1			
KIF3B	5%	1 I.	1.1.1				
ITPRID2	4%	11-11-1 F	1 I	Î.			
AGAP9	1,4%						
FAM9A	3%	0	1 ( ) ( ) ( ) ( ) ( ) ( ) ( ) ( ) ( ) (				
Genetic Altera	ation	Inframe Mutation (putative d Missense Mutation (unknown Truncating Mutation (unknow	river) Missense Muta significance) Trunca m significance) Ampli	tion (putative drive ting Mutation (puta fication	r) tive driver)		

Figure EV1. Tumor-promoting genes are not frequently altered in cutaneous melanoma. Related to Fig 1.

A Genomic alterations and 36 tumor-promoting genes identified in Fig 1. *BRAF, NRAS,* and *NF1* have been added to the list of tumor-promoting genes according to melanoma classification (Akbani *et al*, 2015). Analyses were performed using the webtool available at www.cbioportal.org.

B Birinapant, an inhibitor of BIRC proteins (BIRC3 and BIRC2), decreased cell density (SKMel28R cells, Birinapant 100 nM, 96 h).

C Birinapant, an inhibitor of BIRC proteins (BIRC3 and BIRC2), decreased cell density (Me1402 cells, Birinapant 100 nM, 96 h).

Data information: Each histogram represents the mean  $\pm$  s.d. from three independent experiments. Bilateral Student test (with non-equivalent variances) \*\*P < 0.01. Source data are available online for this figure.

#### Figure EV2. BRAFi-resistance genes are associated with invasive phenotype and resistance to BRAFi in vitro and in vivo. Related to Fig 3.

- A *BIRC3* expression levels (fold change) in melanoma Rr and Ra cell lines. The fold change has been calculated by comparing *BIRC3* expression levels in resistant/parental cell lines (Song *et al*, 2017). Two groups have been made: Rr (n = 6) and Ra (n = 5) cell lines; Ra for resistance and MAPK reactivation and Rr for resistance and MAPK redundant. BRAFi resistance in Ra cell lines is due to additional genetic alterations such as BRAF splicing events for M395 SDR. BRAFi resistance in Rr cell lines is due to gene expression reprogramming. Two-tailed Mann–Whitney test, P = 0.0303. Vertical bars correspond to the medians.
- B Schematic representation of the workflow. As described in the original paper (Shaffer *et al*, 2017), fresh melanoma cells were obtained by patient tumor dissociation. Cells mix was treated with BRAFi. Cell sorting (EGFR positive and/or negative) was performed at two time points: before treatment (no drug) and after 4 weeks of treatment. Cells "mix" corresponds to the unsorted population. The EGFR-positive cells (exposed to BRAFi) are able to produce colonies.
- C Heat map depicting mRNA expression of BRAFi-resistance genes, using an RNA-Seq dataset obtained from BRAFi-treated melanoma cells (Shaffer *et al*, 2017). Scale corresponds to Z scores.
- D Heat map illustrating the expression levels of BRAFi-resistance genes in invasive (Inv.) and proliferative melanoma cell lines (n = 2 and n = 9, respectively) (Verfaillie *et al*, 2015). Scale corresponds to Z scores.
- E Genomic alterations in 18 BRAFi-resistance genes identified in Fig 3 (from www.cbioportal.org). BRAF, NRAS, and NF1 have been added according to melanoma classification (Akbani et al, 2015).
- F SMAD3 expression levels (RNA Seq V2 (log<sub>2</sub>) in the TCGA dataset. Picture was downloaded from www.cbioportal.org. Arrow indicates cutaneous melanoma.
- G TYRP1 expression levels (RNA Seq V2 (log<sub>2</sub>) in the TCGA data set. Pictures were downloaded from www.cbioportal.org. Arrow indicates cutaneous melanoma. TYRP1 is highly expressed in drug-naive tumors; cutaneous and uveal melanoma in contrast to SMAD3.
- H SMAD3 expression and cell types (https://www.proteinatlas.org/ENSG00000166949-SMAD3/celltype). The consensus normalized expression (NX) value for SMAD3 and organ/tissue represents the maximum NX value in the three data sources (HPA, GTEx, and FANTOM5). Black arrow highlights the melanocytes.

Source data are available online for this figure.



Figure EV2.

### Figure EV3. BIRC3 and EGFR are potent BRAFi-resistance genes. Related to Fig 5.

- A Pictures illustrating the ratio (invasive diameter/spheroid diameter; E/S) used in Fig 5E (Invasion assay, melanoma spheroids). Two cell lines are illustrated: CTR cells correspond to 501Mel cells expressing dCas9 and HSF1-p65-MS2 (named here 501Mel 2+) and the SMAD3 cells (overexpressing SMAD3).
- B SMAD3 Knock-down efficiency of two siRNAs targeting SMAD3 in SKMel28R (Fig 5G). SMAD3 protein expression levels have been quantified in three independent experiments by Western blots. Expression levels have been compared to HSC70. Values obtained with the control siRNA (CTR) have been set at 1. Bilateral Student test (with non-equivalent variances) \*\*\*P < 0.001.
- C PCA analysis of *BIRC3* expression in melanoma cell lines in function of their dedifferentiation states (generated by the webtool http://systems.crump.ucla.edu/dediff/ index.php). Scale: red color corresponds to a high *BIRC3* expression level (unit of the scale: Log<sub>2</sub> FPKM).
- D *BIRC3* expression in these four subtypes of melanoma cells (undifferentiated = 10; neural crest-like = 14; transitory = 12; melanocytic = 17) (Tsoi *et al*, 2018). Error bars represents the mean  $\pm$  s.d. Multiple comparisons have been done using ordinary one-way ANOVA, \**P* < 0.05, \*\*\**P* < 0.001.
- E PCA analysis of EGFR expression in melanoma cell lines in function of their dedifferentiation states (unit of the scale: Log<sub>2</sub> FPKM).
- F *EGFR* expression in these four subtypes of melanoma cells (undifferentiated = 10; neural crest-like = 14; transitory = 12; melanocytic = 17). Error bars represents the mean  $\pm$  s.d. Multiple comparisons have been done using ordinary one-way ANOVA, \*\*P < 0.01, \*\*\*P < 0.001.
- G BIRC3 depletion (siRNA#1 & #2) reduced cell density and restored BRAFi (vemurafenib) effect on BRAFi-resistant cells (SKMel28R). CTR for non-targeting siRNA. Vem for the BRAFi vemurafenib and DMSO for dimethylsulfoxide (solvent of Vem). n = 2 biologically independent experiments. Each histogram represents the mean  $\pm$  s.d.
- H *BIRC3* Knock-down efficiency of two siRNAs targeting *BIRC3* in SKMel28R (Fig EV3G). *BIRC3* mRNA expression levels have been quantified in three independent experiments by RT–qPCR. Values obtained with the control siRNA (CTR) have been set at 1. Each histogram represents the mean  $\pm$  s.d. n = 3 biologically independent experiments. Bilateral Student test (with non-equivalent variances) \*\*\*P < 0.001.
- 1 Knock-down of *EGFR*, *IL6*, *or AQP1* (siRNA#1, #2, and #3) restored BRAFi (vemurafenib) effect on BRAFi-resistant cells (SKMel28R). n = 3 biologically independent experiments are presented. Each histogram represents the mean  $\pm$  s.d.. Bilateral Student test (with non-equivalent variances) \*P < 0.05.
- J Knock-down efficiency of siRNAs targeting *EGFR*, *IL6*, or *AQP1* in SKMel28R (Fig EV3I). mRNA expression levels have been quantified in two or three independent experiments by RT–qPCR. Each histogram represents the mean  $\pm$  s.d. n = 3 biologically independent experiments for siCTR, siEGFR#1,2,4, IL6#3, and n = 2 for the other siRNAs. Values obtained with the control siRNA (CTR) have been set at 1. Bilateral Student test (with non-equivalent variances) \*\*P < 0.01, \*\*\*P < 0.001
- K Efficiency of BRAFi (Vem) and MEKi (Cobi) to inhibit the MAPK signaling pathway (P-ERK) (as used in Fig 50). Values obtained with the solvent (CTR, here DMSO) have been set at 100%.

Source data are available online for this figure.



Figure EV3.



## Figure EV4. SMAD3 signature is associated with mesenchymal phenotype in liver cancer cell lines. Related to Fig 7.

- A Plot showing correlation between SMAD3-signature and mesenchymal signature (Mak et al, 2016) in the TCGA dataset (SKCM). Pearson correlation test: P < 0.0001
- B Gene Set Enrichment Analysis (GSEA) of the SMAD3 signature in TGF-β treated versus non-treated Huh-28 cholangiocarcinoma cells (Merdrignac et al, 2018). C Gene Set Enrichment Analysis of the SMAD3 signature in hepatoma cell lines : mesenchymal (3sp cells) versus epithelial (3p cells) (van Zijl et al, 2011).



## Figure EV5. sgRNA targeting four isoforms of SMAD3 and BRAFi resistance.

- A SMAD3 mRNA isoforms according to NCBI. The longest isoform NM\_005902.4 encodes SMAD3 protein (NP\_005893.1). Three shorter isoforms of SMAD3 mRNA encode shorter SMAD3 proteins.
- B Snapshot of SMAD3 mRNA expression in 501Mel and SKMel28S cells using IGV tool. Note that exon 1 is detectable in these melanoma cell lines. Arrows indicate the binding sites of primers for RT–qPCR (exon 1 or 6 for NM\_005902.4).
- C Scheme to illustrate the binding site of sgRNAs targeting SMAD3 gene. These 12 sgRNAs are found in the CRISPR-SAM library (Konermann *et al*, 2015). Three sgRNAs per isoform. Only two sgRNA targeting the longest isoform of SMAD3 are associated with BRAFi resistance (Fig 2).
- D Ability of six sgRNAs to transactivate *SMAD3* expression have been compared. Among the five sgRNAs targeting the first exon, three are found in the library (g71, g92, and g116). We designed g50 and g155 to evaluate the impact of sgRNA position on exon 1. sgRNA gIntron3 has been designed as a control (unable to promote the longest *SMAD3* isoform but efficient to transactivate the smallest isoform). These two isoforms contain the exon 6 used for RT–qPCR.
- E *SMAD3* quantification by RT–qPCR (exon 1 or 6) in HEK293 cells transiently transfected with the three plasmids required for the CRISPR-SAM as previously described (Konermann *et al*, 2015). CTR for empty plasmid (no sgRNA cloned into lenti sgRNA(MS2)\_zeo backbone). RT–qPCR data are representative of two independent experiments. Source data are available in Fig EV5 source data. Each histogram represents the mean of three technical replicates from one representative experiment. No statistical analyses were thus performed. Dotted line highlights the value of 1 (corresponding to the effect obtained by the control sgRNA).

Source data are available online for this figure.