Co-evolution of tumor and immune cells during progression of multiple myeloma Liu et al.



Supplementary Figure 1. (a) Ancestry analysis of multiple myeloma cases. Predicted ancestry for 14 cases, along with 856 Multiple Myeloma Research Foundation cases (including 31 multiple time point cases). Analysis indicated each patient in our cohort (triangle shape) is of European ancestry, consistent with clinical records. (b) Variant allele fraction (VAF) Pearson correlations of 24 germline variants showed samples came from same patients. (c) The landscape of copy number variation (CNV), structural variation (SV), and driver mutations across from 14 patients. Copy number amplification/gain, copy number deletion/loss and SV are shown in red, blue and purple respectively with darkness of color indicating the number of techniques supporting the event. Techniques for copy number events: FISH, 10xWGS, regular WGS, WES, scRNA-seq; for SV: FISH, 10xWGS, Bulk RNA-seq, scRNA-seq. Variant allele fraction for driver mutations are shown in yellow corresponding to different sequencing techniques. Plasma cells percentage indicates the percentage of plasma cells in scRNA-seq data.



b 27522-Primary

C 58408-Primary



d



Supplementary Figure 2. (a) Mapping reference and variant alleles to individual cell for one relapse tumor (27522 Relapse-2) and two primary tumors (b) 27522 Primary and (c) 58408 Primary. (d) Single cell CNV landscape from 20 samples.





d



Supplementary Figure 3. (a) t-SNE plot for non-malignant cells shows cell type dominates clustering. (b) t-SNE plot for macrophages, monocytes and CD4+ T cells, respectively. Colors indicate cells from particular samples. (c) Expression for *CTSS*, in monocytes and macrophages. (d) Expression changes over disease progression for *CD69*, *NFKBIA* in CD4+ T cells and *IL1R2*, *IL1B* in monocytes, respectively.









Supplementary Figure 4. (a) t-SNE plots showing malignant cells from eleven patients reveal tumor-specific clusters, colored by patient (left) and stage (right). (b) Heatmap showing genes with individual-specific expression pattern (c) Violin plot showing the expression pattern of plasma cells for samples potentially within CD1/CD2 groups.



Supplementary Figure 5. CyTOF profiling shows HLA-DQA1 is differentially expressed between primitive and mature B cells.



Supplementary Figure 6. (a) Plasma subpopulations identified in Patient 47491 SMM (47491_1) and Primary (47491_2). (b) Plasma subpopulations identified in Patient 37692 SMM (37692_1) and Primary (37692_2). (c) Copy number alterations in different plasma cell populations in Patient 47491. (d) Copy number alterations in different plasma cell populations in Patient 58408. (e) Mapping of plasma cell subpopulations from Patient 47491 SMM and Primary samples onto the integrated B cell and plasma cell t-SNE plot from all samples. (f) Mapping of plasma cell subpopulations from Patient 37692 SMM and Primary samples to their integrated t-SNE. (g) *JUN* and *FOS* expression in Patient 47491. (h) *JUN* and *FOS* expression in Patient 37692.





d

Ν

R.4

R.3

R.2

R.1

P.2

P.1

Supplementary Figure 7. (a) Expression of heat shock genes in different subclusters of plasma cells in Patient 58408 (SMM and Primary). (b) Quality control parameters in subclusters of plasma cells in Patient 58408. (c) Violin Plot showing expression of *FOS* and *JUN* in in different types of cells for Patient 58408. Cells are separated by nGene>=1000 (red) and nGene < 1000 (blue). (d) Dot Plot showing the expression of genes specifically upregulated in 81012 R.3 and R.4. (e) Pathway enrichment analysis for genes upregulated in case 81012 R.3. The x axis indicates the ratio of genes present in the corresponding pathway gene list. Dot size indicate the number of genes present in the corresponding pathway is the adjusted p value. Enrichment analysis is carried out by performing the hypergeometric test, p value is then adjusted by Benjamini-Hochberg (BH) correction.



Supplementary Figure 8. (a) Patient 56203 from the primary stage, with three plasma cell subpopulations (P.1, P.2, and P.3), to the relapse stage, with two plasma cell subpopulations (R.1 and R.2). (b) t-SNE plot showing the expression of *WHSC1/ FGFR3* and copy number profile in terms of chromosome 3 gain and chromosome 13 loss, in each subcluster of plasma cells in the relapse-2 sample of case 27522. (c) Expression of B cell markers in plasma cells from Patient 27522 Relapse-2.



Supplementary Figure 9. (a) Trajectory analysis for B and Plasma cells in Patient 47491; cells are colored by cell type (B/plasma) and subcluster information (corresponding to subcluster information from extended data figure 6). Left: cells from SMM; right: cells from primary diagnosis. (b) Normalized proportion for plasma cell subclusters within each state for Patient 47491. (c) Violin plot showing pseudotime distribution across states for plasma cell subclusters in Patient 47491. (d) Similar as (a) except that the case shown is from Patient 81012 and two time points are from primary diagnosis and relapse, respectively. (e-f) Normalized plasma cell subcluster proportion and pseudotime distribution for Patient 81012, similar to (b)-(c).



G1 G2 G3 Average Normalized Expression 0 0.5 1 1.5 2 2.5

group

Supplementary Figure 10. (a) Unsupervised clustering of sub-populations of plasma cells from 16 MM patients (x-axis and y-axis) based on correlation with significant pathways. Three major groups emerged (G1, G2, G3) that contained significantly correlated sub-populations based on shared pathways. The highly significant pathways from three groups displayed in the figure (Translation, Cellular Metabolism, Cell Cycle). The highly significant pathways from three groups displayed in the figure (Translation, Cellular Metabolism, Cellular Metabolism, Cell Cycle) contained significantly correlated sub-populations based on shared figure (Translation, Cellular Metabolism, Cellular Metabolism, Cell Cycle) contained significantly correlated sub-populations based on shared pathways. (b) Expression of two proliferation markers *MKI67* and *TOP2A* in different subclusters of plasma cells belong to three major groups. Each dot shows the normalized expression for individual cells; heatmap shows the average expression for each subcluster.



Supplementary Figure 11. (a) Correlation of single cell average expression and bulk RNA-Seq expression for each sample for FOS and JUN. Solid dot indicates unsorted samples. Blank dot indicates samples for bulk RNA-Seq is positively sorted for CD138, and single cell average expression in this scenario is calculated with plasma cells only. (b) CyTOF profiling of genes of interest in two manually-gated populations of plasma cells for three samples with good viability.