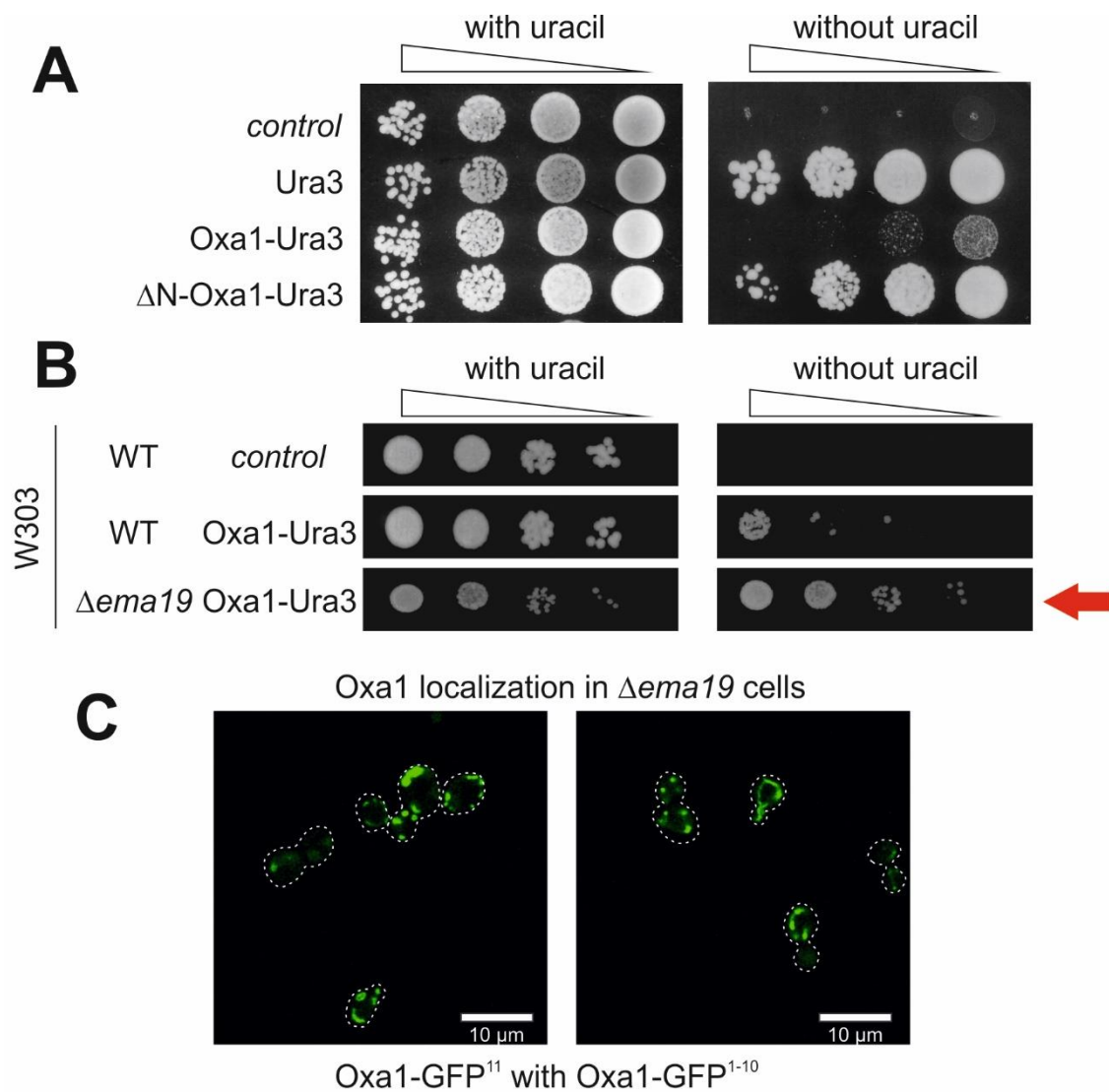


# Supplemental Materials

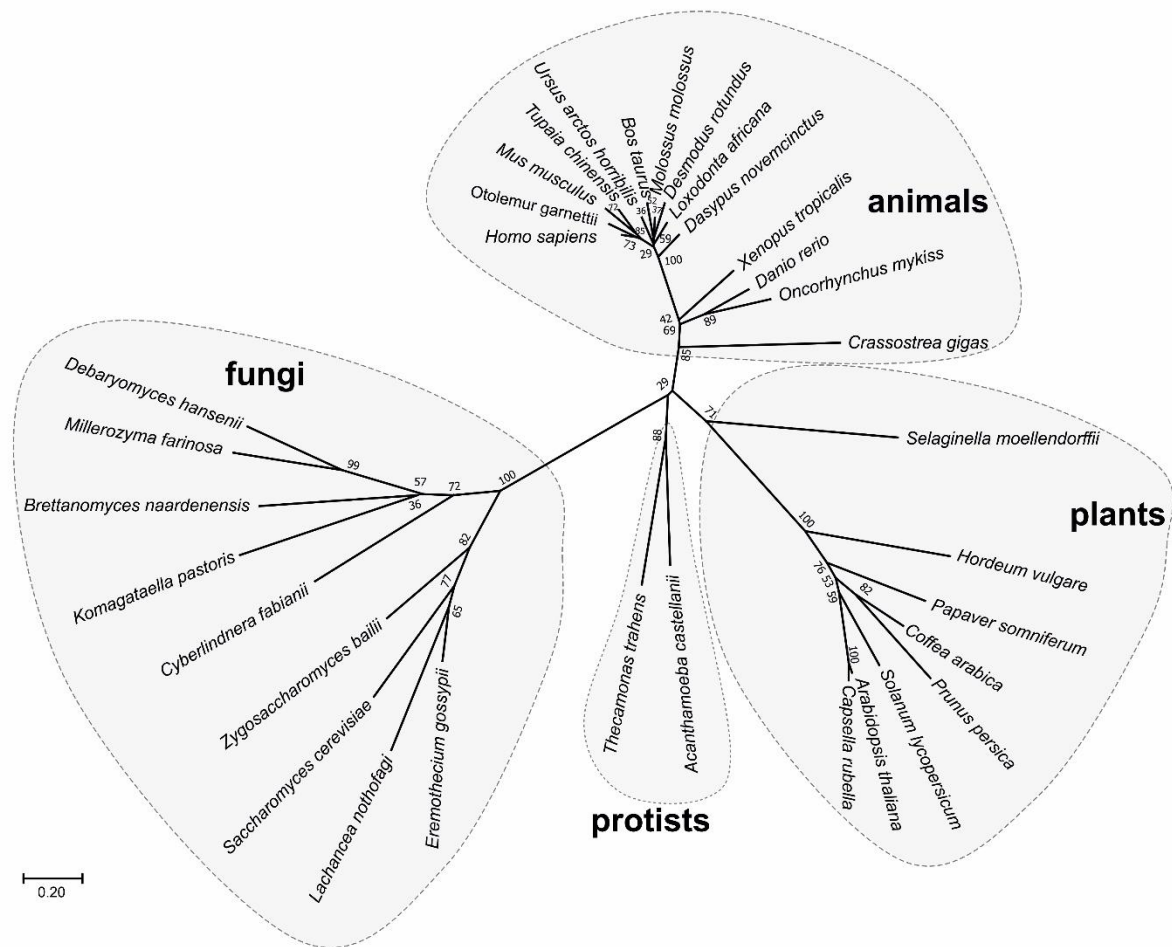
*Molecular Biology of the Cell*

Laborenz *et al.*

Supplemental Figures



**Figure S1. The uracil-independent growth of Oxa1-Ura3-expressing  $\Delta$ *ema19* cells is influenced by the strain background. A, B.** The indicated strains were generated in the wild type background YPH (A), W303 or BY4742 and analyzed as described for Fig. 1. **C.** Cells expressing Oxa1-GFP<sup>11</sup> and Sec63-GFP<sup>1-10</sup> were analyzed by fluorescence microscopy.



**Figure S2. Phylogenetic analysis of the members of Ema19/TMEM97/sigma 2 receptor family.** For the phylogenetic tree construction, pairwise sequence alignments were generated using ClustalW with default settings and subjected to the calculation of maximum likelihood distances implementing a JTT matrix (Jones *et al.*, 1992). This matrix was then used to calculate a neighbor-joining tree. Given bootstrap values are based on 1000 random reconstructed trees and indicate the confidence of the respective branches. All analyses were conducted in MegaX (Kumar *et al.*, 2018). Sequences and corresponding accession numbers are listed in the Supplementary Tables file.

Jones, D.T., Taylor, W.R., and Thornton, J.M. (1992). The rapid generation of mutation data matrices from protein sequences. *CABIOS* 8, 275-282.

Kumar, S., Stecher, G., Li, M., Knyaz, C., and Tamura, K. (2018). MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol Biol Evol* 35, 1547-1549.

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Sc -----
Cg -----
Ca -----
Zb -----
Kl -----
Dh -----
Hs -----
Mm -----
Dr MVVLRLPHEPMSWESAHSRNQPSRLTNWKTRIKKIRAKGRIRNRGLTRYSTLPTDANPP 60

Sc -----MKLGHREQQFYLWYFIVHIPITIFIDSSVVI 37
Cg -----M-ISSLSVFEEKFYYYYCLVHIPITVLDSSVAVPDKWVL 39
Ca -----MKLIDKYLWYFIIHIPITIFIDSSIVIPKQYQL 34
Zb -----MAGLSRLEISFFYFYFLLHIPITILIDS AVVIPPKFRI 38
Kl -----MSKEQRD-SRKTQLWKINLWKI WYAVNIPIITFLDSNAVLPPYLQY 45
Dh -----MPGANK-STKTMCGIDKFYVAYFLMHIPITLIIDSCIIIP EEQRF 44
Hs -----MGAPATRRCVEWLLGLYFLSHIPITL FMDLQAVLPRELYP 40
Mm -----MGALAARRCVEWLLGLYFVSHIPITL FIDLQAVLPPELYP 40
Dr LLNHPDGGTTCKRHLTRHLNQTIKMLFLVLEI IYFIYFASHIPITLLVDLQALLPEHVYP 120
      . : :*** ::* :*

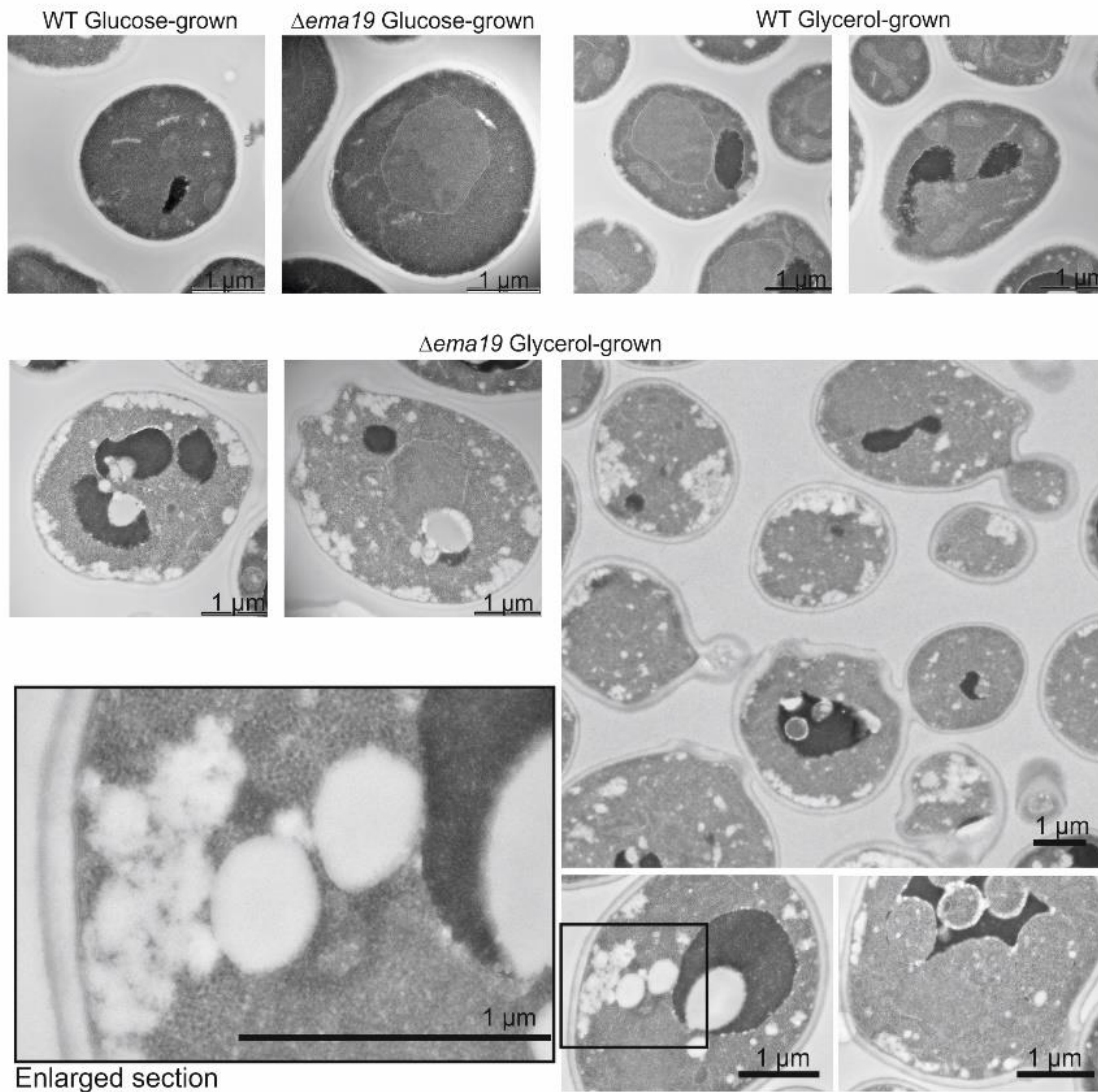
Sc GIAQKVSDHIAKQHDFFLLSEKPEWLYWV VLELVQLPLFVYFVNKFWNSSE----- 90
Cg L--PGLVQWHIRQNNDFLLYEKPMWLQLFV WVELVFLPLFFYFAHQFKKI WALRSKDTK 97
Ca PITKSILEFHISTNNDILLAYPQTFWKFIF GFIELIFQLPLFFYFYIKLLSSNR----- 87
Zb A--GWLLERHIAQNNDFLLCDRPMWLQVFVA LELTVQLPLFFQFAKQLRRNSV----- 89
Kl T--KGLLDGYIDNFNDFLTANPENWLNK YMTIIEILFQLPVAIYCLYQLLNLSKSKTIK-- 101
Dh QFQKQFLEFHISSNKDFLLVSLPLWLKVF GLFELFVQLPFFAIGAYMLVKQMK----- 97
Hs VEFRNLLKWYAKEFKDPLLQEPFAWFKSFL FCELVFQLPFFPIATYAFKLGSC----- 93
Mm QEFSNLLRWYSKEFKDPLMQEPPVWFKSFL LCELVFQLPFFPIAAYAFFKGC----- 93
Dr PELIKLLHWYAGEFKDPMMMDPPAWFKSF VVCEALVQLPFFPVAAAYAFKGGC----- 173
      .: : :* : * : * .***. :

Sc ----LQVNTNSRLKKWLR----IYGNASL TTLICIVVIFKRGYIPYDVLKT----SLS 137
Cg NAKAERASTKKSLEYLWLR----VYGLNAAL TTWICIVVILYRGYYPFTLDASRIAGTKLE 153
Ca -----RVLDVNYLWSI----IYGFNAGFT TFCVLIWLIIE-YKNF-----QLS 126
Zb -----PRRSLK----LYGLLASTTTLV CIGAILLEGHYPGTAIP-----MT 125
Kl -----RTLFTVDRLSK----CYAFNVIT TTSFCIWVWAYGYYPATMNTDH---VLSL 147
Dh -----QVYPYML----IYGFNASFTTL VCLVHIFCD-YERF-----GLT 131
Hs -----KWIRTPAI IYSVHTMTTLIPIL STFLFEDFS---KASGFKGQRPET 136
Mm -----RWIRIPAI IYAAHTITTLIPIL YTLLEDFES---KAVAFKGQRPES 136
Dr -----KWIRTPAI IYSVHATTLPILSHI LFHKFP---LSP---HPGPQT 213
      * . * : . :

Sc MTQKCQLASVYLPTFLIPLRLCFV*----- 162
Cg VRDTLALMGLYLPFTFLLPLRLCMLQQ----- 179
Ca DLQLINLLAIYIPYLLPLILLIHSFKQIQ-Q YN-N--NNHNKCLKQO- 169
Zb SLDKGLVCVYLPTFLLPFRLVLL----- 149
Kl NPKLALTMVYVPYVLI PALFFV----- 170
Dh TGESYKLAALYIPYLVIPVMLVDYSIRIN-K SI-K--AHIPPTKKN I 175
Hs LHERLTLVSVYAPYLLIPFILLIFMLRSP YKYEEKRKKK----- 176
Mm FRERLTLVGVYAPYLIIPILLLLFMLRN PYYKYEEKRKKK----- 176
Dr LNERLTLVSIYAPYLIIPIMILLTMLFSAT YNSPSLKGNAKAKKQR 261
      : * :* * .:* : .

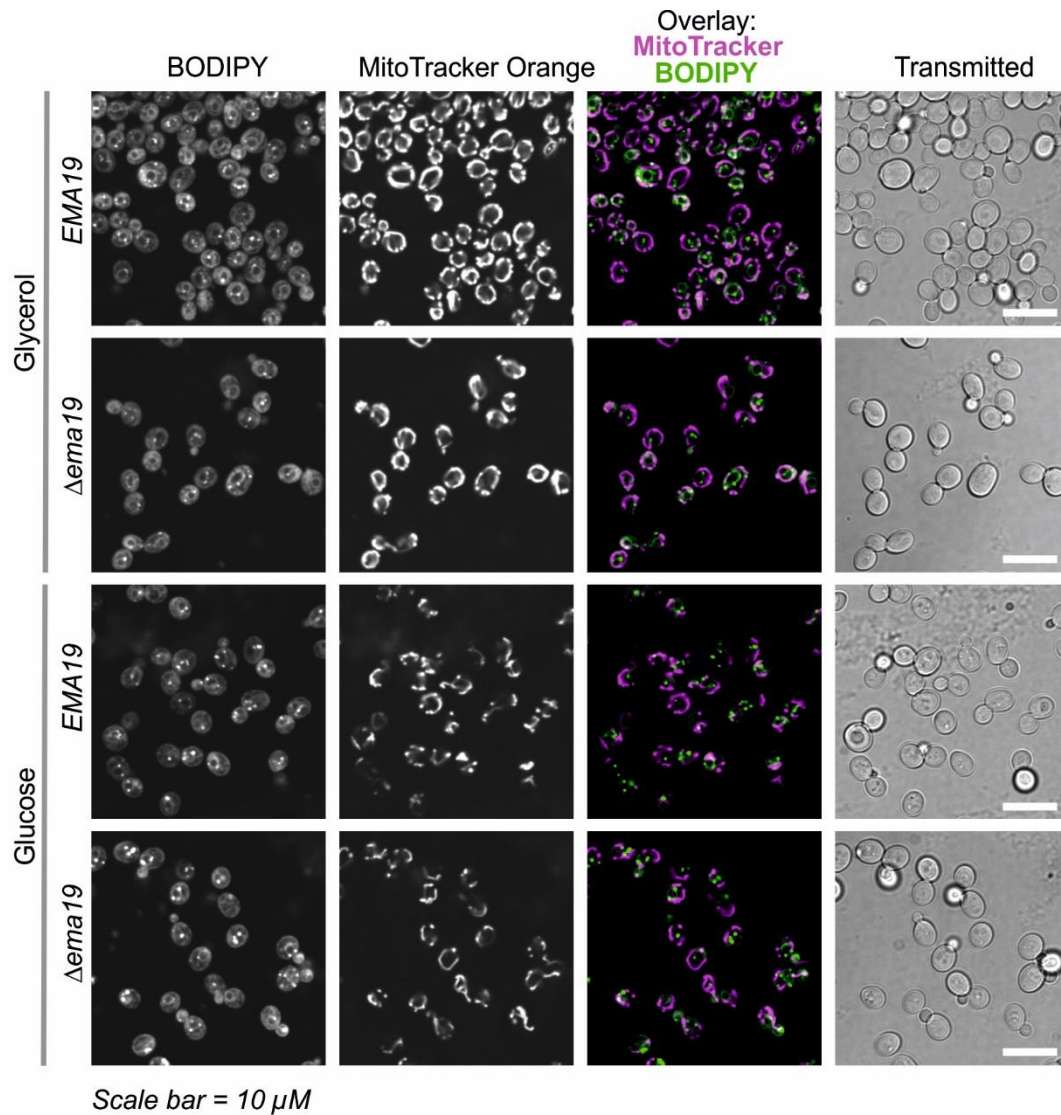
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**Figure S3. Alignment of sequences of members of the Ema19/TMEM97 protein family.** Sc, *Saccharomyces cerevisiae*; Cg, *Candida glabrata* KTA97029; Ca, *Candida albicans* EEQ46884; Zb, *Zygosaccharomyces bailii* AQZ15181; Kl, *Kluyvermyces lactis* CAH00503; Dh, *Debaromyces hansenii* CAG89731; Hs, *Homo sapiens* TMEM97; Mm, *Mus musculus* TMEM97; Dr, *Danio rerio* tmem97. Transmembrane domains are highlighted in blue.

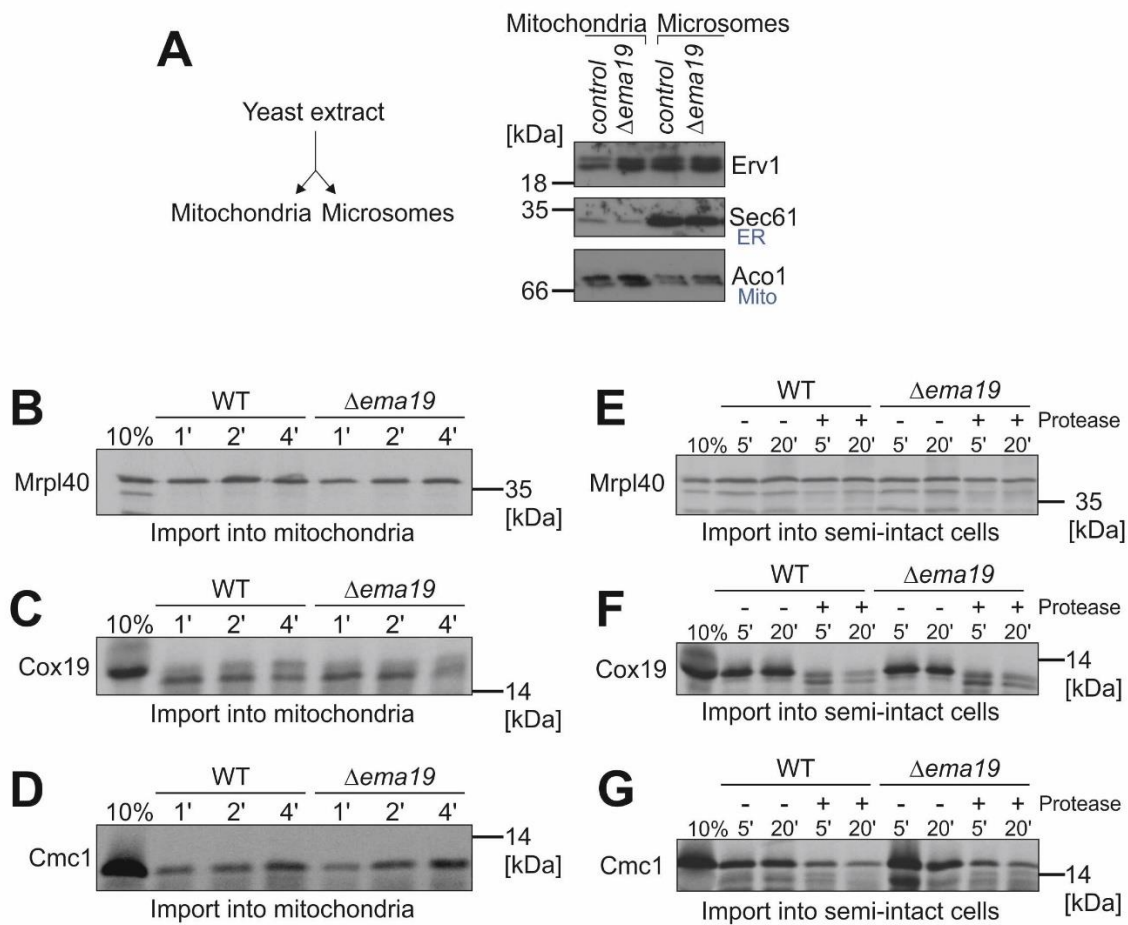


**Figure S4. *Δema19* cells show morphological changes upon growth on glycerol but staining with BODIPY does not show obvious changes.** Yeast cells were visualized by electron microscopy as described for Fig. 3A.

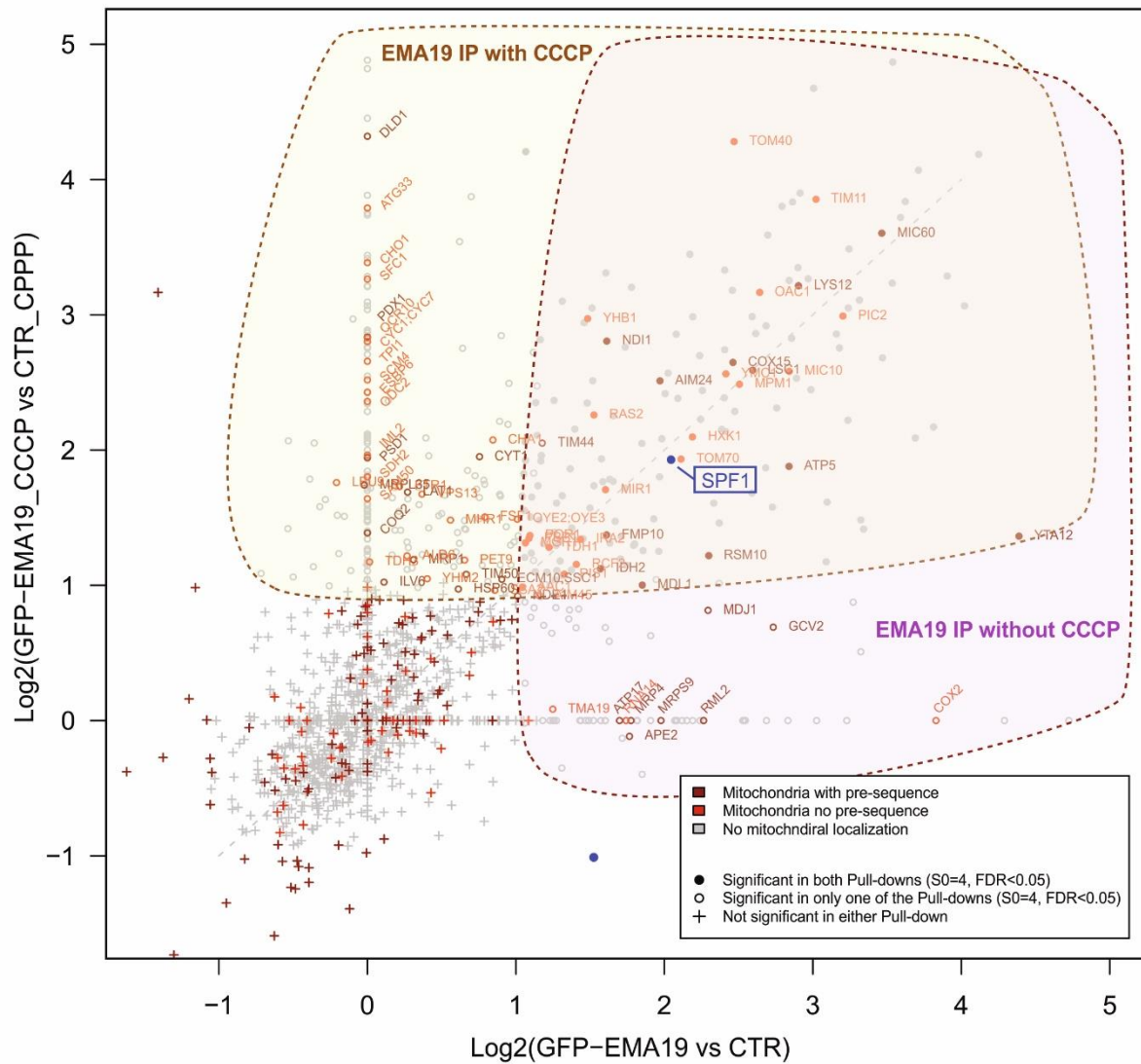




**Figure S5. BODIPY staining does not indicate the accumulation of lipid droplets in  $\Delta$ ema19 cells.** Cells were grown in glycerol medium before mitochondria and lipid bodies were visualized by fluorescence microscopy using MitoTracker Orange and BODIPY 493/503, respectively.



**Figure S6. Ema19 is not critical for the import of mitochondrial proteins.** **A.** Microsomes and mitochondrial fractions were isolated from cells expressing or lacking Ema19. The levels of the indicated proteins were analyzed by Western blotting. **B.-G.** *In vitro* import experiments with isolated mitochondria or semi-intact cells as described in the legend to Fig. 5.



**Figure S7. Pull-down experiments indicate that the mitochondrial membrane potential influences the interaction partners of Ema19.** The log<sub>2</sub>-fold change of proteins in Ema19-GFP pull-downs in the presence or absence of 200 μM CCCP was plotted against each other (see Table S5 for data). To visualize proteins that were identified only in one of the two pull-down experiments, missing values were replaced by 0 (indicated by rotated labels). Proteins with a predicted mitochondrial localization are shown in red, proteins with a predicted matrix targeting sequence are shown in dark red. Proteins with a significant enrichment in both pull-down experiments are shown as filled circles. Proteins with significant enrichment in only one of the two pull-down experiments are shown in non-filled circles (four replicates, FDR<0.05). Note that Ema19 (fold change = 9.14 / 9.22) has been omitted for clarity.



## Supplemental Table S1 listing the strains used in this study

Strain	Genotype	Reference
BY4742 WT	MAT $\alpha$ <i>his3 leu2 lys2 ura3</i>	
YPH499 WT	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2</i>	
W303 WT	MAT $\alpha$ <i>ura3-1 ade2-1 his3-11 leu2-3,112 trp1<math>\Delta</math>2</i>	
$\Delta$ <i>ema19</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2</i>	This study
$\Delta$ <i>ema19</i>	MAT $\alpha$ <i>ura3-1 ade2-1 his3-11 leu2-3,112 trp1<math>\Delta</math>2 EMA19::NatNT2</i>	This study
$\Delta$ <i>ema19</i>	MAT $\alpha$ <i>his3 leu2 lys2 ura3 EMA19::NatNT2</i>	This study
$\Delta$ <i>djp1</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 DJP1::KanMX4</i>	This study
$\Delta$ <i>ema19<math>\Delta</math><i>djp1</i></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 DJP1::KanMX4</i>	This study
$\Delta$ <i>oxa1</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 OXA1::KanMX4</i>	This study
$\Delta$ <i>ema19<math>\Delta</math><i>oxa1</i></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 OXA1::KanMX4</i>	This study
$\Delta$ <i>cox23</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 COX23::KanMX4</i>	This study
$\Delta$ <i>ema19<math>\Delta</math><i>cox23</i></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 COX23::KanMX4</i>	This study
$\Delta$ <i>qcr2</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 QCR2::KanMX4</i>	This study
$\Delta$ <i>ema19<math>\Delta</math><i>qcr2</i></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 QCR2::KanMX4</i>	This study
$\Delta$ <i>coa4</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 COA4::KanMX4</i>	This study
$\Delta$ <i>ema19<math>\Delta</math><i>coa4</i></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 COA4::KanMX4</i>	This study
$\Delta$ <i>ema19-<math>\Delta</math>N- Oxa1-HA</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 pYX223-<math>\Delta</math>N-Oxa1-HA</i>	This study
$\Delta$ N-Oxa1-HA	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 pYX223-<math>\Delta</math>N-Oxa1- HA</i>	This study
$\Delta$ <i>ema19-Erv1-HA</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 pYX232-Erv1-HA</i>	This study
Erv1-HA	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 pYX232-Erv1-HA</i>	This study
$\Delta$ <i>ema19-Oxa1- Ura3</i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 pRS313 Oxa1-Ura3</i>	This study
YPH499-Oxa1- Ura3	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 pRS313 Oxa1-Ura3</i>	This study
$\Delta$ <i>ema19-Oxa1- Ura3</i>	MAT $\alpha$ <i>his3 leu2 lys2 ura3 EMA19::NatNT2 pRS313 Oxa1-Ura3</i>	This study
BY4742-Oxa1- Ura3	MAT $\alpha$ <i>his3 leu2 lys2 ura3 pRS313 Oxa1-Ura3</i>	This study
$\Delta$ <i>ema19-Oxa1- Ura3</i>	MAT $\alpha$ <i>ura3-1 ade2-1 his3-11 leu2-3,112 trp1<math>\Delta</math>2 EMA19::NatNT2 pRS313 Oxa1-Ura3</i>	This study
W303-Oxa1-Ura3	MAT $\alpha$ <i>ura3-1 ade2-1 his3-11 leu2-3,112 trp1<math>\Delta</math>2 pRS313 Oxa1-Ura3</i>	This study
$\Delta$ <i>ema19-Oxa1- GFP<sup>11</sup> + Oxa1- GFP<sup>1-10</sup></i>	MAT $\alpha$ <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2 pYX142-Oxa1-GFP<sup>11</sup> + pYX122-Oxa1-GFP<sup>1-10</sup></i>	This study

<i>Δema19- ΔN-Oxa1-GFP<sup>11</sup> + Sec63-GFP<sup>1-10</sup></i>	MATa <i>ura3 lys2 ade2 trp1 his3 leu2 EMA19::NatNT2</i> pYX142-Oxa1-GFP <sup>11</sup> + pYX122-Sec63-GFP <sup>1-10</sup>	This study
YPH499- <i>ΔN-Oxa1-GFP<sup>11</sup> + Sec63-GFP<sup>1-10</sup></i>	MATa <i>ura3 lys2 ade2 trp1 his3 leu2</i> pYX142-Oxa1-GFP <sup>11</sup> + pYX122-Sec63-GFP <sup>1-10</sup>	This study
Ema19-GFP Sec63-mCherry	MATa <i>his3Δ1 leu2Δ0 met15Δ0 ura3Δ0 EMA19-GFP::HIS</i> <i>SEC63-mCherry::KanMX</i>	This study
GFP-Ema19 Sec63-mCherry	MATa <i>his3Δ1 leu2Δ0 met15Δ0 ura3Δ0</i> <i>hphΔn::URA3::SpNOP1pr-sfGFP-EMA19 SEC63-</i> <i>mCherry::KanMX</i>	This study
GFP-Ema19	MATa <i>his3Δ1 leu2Δ0 met15Δ0 ura3Δ0</i> <i>hphΔn::URA3::SpNOP1pr-sfGFP-EMA19</i>	Hansen et al., 2018

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**Supplemental Table S2 listing the plasmids used in this study**

<b>Plasmid</b>	<b>Description</b>	<b>Reference</b>
pYX223- $\Delta$ N-Oxa1-HA	2 $\mu$ , GAL- $\Delta$ N-Oxa1-HA	Hansen et al., 2018
pYX232-Erv1-HA	2 $\mu$ , TPI-Erv1-HA	Bien et al., 2010
pRS313	CEN, empty vector	Hansen et al., 2018
pRS313-Oxa1-Ura3	CEN, Oxa1-Ura3	Hansen et al., 2018
pGem4-Mrp140		Herrmann lab collection
pGem4-Cox19		Herrmann lab collection
pGem4-Cmc1		Herrmann lab collection
pGem4-Oxa1		Herrmann lab collection
pYX142-Oxa1-GFP <sup>11</sup>	CEN, TPI-Oxa1- GFP <sup>11</sup>	This study
pYX122-Oxa1-GFP <sup>1-10</sup>	CEN, TPI-Oxa1-GFP <sup>1-10</sup>	This study
pYX142- $\Delta$ N-Oxa1-GFP <sup>11</sup>	CEN, TPI- $\Delta$ N-Oxa1-GFP <sup>11</sup>	This study
pYX122-Sec63-GFP <sup>1-10</sup>	CEN, TPI-Sec63-GFP <sup>1-10</sup>	This study

**Supplemental Table S3 listing the antibodies used in this study**

<b>Antibodies</b>	<b>Source</b>	<b>Identifier</b>
anti-Ilv5	Herrmann lab collection	Peleh et al., 2017
anti-Oxa1	Herrmann lab collection	
anti-Atp23	Herrmann lab collection	
Anti-Cmc1	Herrmann lab collection	
anti-Erv1	Herrmann lab collection	
anti-Rpl6B	Herrmann lab collection	
anti-HA-Peroxidase	Roche	12 013 819 001
anti-rabbit secondary antibody	BioRad	172-1019

**Supplemental Table S4 listing organisms, accession numbers and protein names of the sequences used for the unrooted neighbor-joining tree shown in Fig. S2**

	<b>Organism</b>	<b>Protein</b>	<b>Accession Number</b>
fungi	<i>Brettanomyces naardenensis</i>	DEKNAAC105048	VEU23783.1
	<i>Cyberlindnera fabianii</i>	TMEM97	ONH65138.1
	<i>Debaryomyces hansenii</i>	DEHA2F22616p	XP_461326.1
	<i>Eremothecium gossypii</i>	AFR508Wp	NP_986055.1
	<i>Komagataella pastoris</i>	BA75_00687T0	ANZ74746.1
	<i>Lachancea nothofagi</i>	LANO_0B02652g1_1	SCU81319.1
	<i>Millerozyma farinosa</i>	Piso0_004871	CCE86385.1
	<i>Saccharomyces cerevisiae</i>	Ema19p	NP_013151.1
	<i>Zygosaccharomyces bailii</i>	ZYBA0S09-00276g1_1	CDF90900.1
protists	<i>Acanthamoeba castellanii</i>	TMEM97	XP_004367690.1
	<i>Thecamonas trahens</i>	TMEM97	XP_013756112.1
plants	<i>Arabidopsis thaliana</i>	TMEM97	NP_565742.1
	<i>Capsella rubella</i>	sigma intracellular receptor 2	XP_006295112.1
	<i>Coffea arabica</i>	sigma intracellular receptor 2	XP_027063739.1
	<i>Hordeum vulgare</i>	TMEM97	KAE8810566.1
	<i>Papaver somniferum</i>	sigma intracellular receptor 2	XP_026392546.1
	<i>Prunus persica</i>	TMEM97	XP_007216330.1
	<i>Selaginella moellendorffii</i>	sigma intracellular receptor 2	XP_002969527.1
	<i>Solanum lycopersicum</i>	sigma intracellular receptor 2	XP_004243464.1
animals	<i>Bos taurus</i>	sigma intracellular receptor 2	NP_001029575.1
	<i>Crassostrea gigas</i>	sigma intracellular receptor 2	XP_011421127.1
	<i>Danio rerio</i>	sigma intracellular receptor 2	NP_001313327.1
	<i>Dasypus novemcinctus</i>	sigma intracellular receptor 2	XP_004450017.2
	<i>Desmodus rotundus</i>	sigma intracellular receptor 2	XP_024420904.1
	<i>Homo sapiens</i>	TMEM97	AAH91504.1
	<i>Loxodonta africana</i>	sigma intracellular receptor 2	XP_003416883.2
	<i>Molossus molossus</i>	TMEM97	KAF6417492.1
	<i>Mus musculus</i>	sigma intracellular receptor 2	NP_598467.1
	<i>Oncorhynchus mykiss</i>	sigma intracellular receptor 2	XP_021444210.2
	<i>Otolemur garnettii</i>	sigma intracellular receptor 2	XP_012665224.1



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<i>Tupaia chinensis</i>	sigma intracellular receptor 2	XP_006165494.1
<i>Ursus arctos horribilis</i>	sigma intracellular receptor 2	XP_026373477.1
<i>Xenopus tropicalis</i>	sigma intracellular receptor 2	sp Q6DFQ5.1

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