

1 **Supplementary Materials**

2 **Dynamics of HEV Antibodies and Development of a Multi-factorial Model to**
3 **Improve the Diagnosis of Current HEV Infection in Resource-limited Settings**

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5 **Validation of the laboratory-developed test (LDT) of HEV RNA detection**

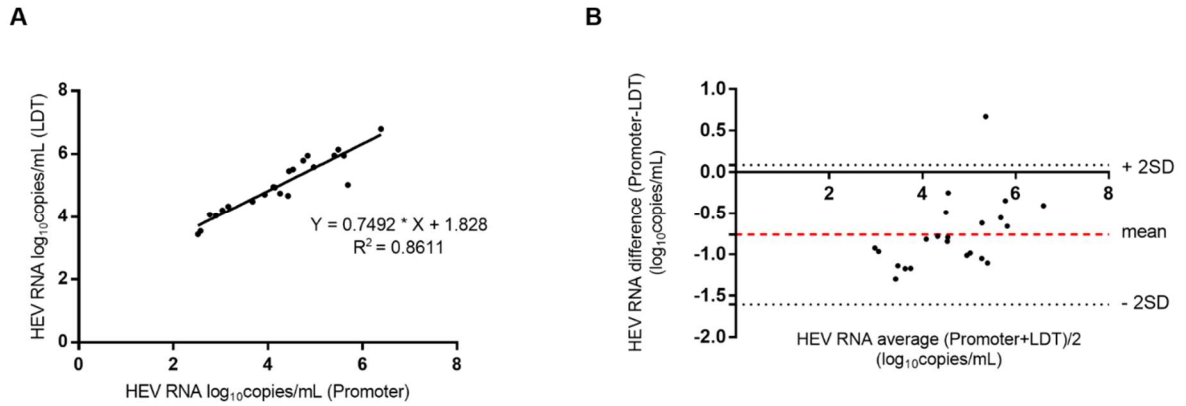
6 The in-house developed HEV RNA RT-PCR assay was validated against the
7 commercial Promotor® HEV RNA detection kit (ACON, Hangzhou, China), which
8 was approved by the National Medical Products Administration (NMPA) of China to
9 provide a qualitative dichotomous positive/negative diagnostic result. For each
10 method, the reproducibility was assessed by calculating the coefficient of variation
11 (CV) of the threshold cycle (C_t) obtained for each standard dilution tested in 5
12 replicates. The CV was found to be inferior to 2% (Table S1). The LDT yielded good
13 dilution linearity at HEV RNA levels within 3.73–7.73 \log_{10} copies/mL. The standard
14 curve of the LDT gave a slope of -3.2534, with Y-intercept of 48.083 and a R^2 value
15 of 0.9979.

16 The limit of detection (LOD) was determined using serial 3-fold dilutions of
17 HEV RNA standard in nuclease-free water to give 4.73, 4.25, 3.77, 3.30, 2.82 and
18 2.34 \log_{10} copies/mL (5 replicates tested by the LDT and 3 replicates tested by the
19 Promoter® assay of each concentration). Probit analysis predicted the 95% LOD of
20 1849 (95% CI: 258-13234) copies/mL for the Promoter® assay and 2295 (95% CI:
21 1.6-3265472) copies/mL for the LDT (Table S2).

22 HEV RNA presence was tested in 28 clinical samples in parallel by each assay
23 and 100% qualitative agreement (negative or positive) was reached across specimens.
24 The viral loads in 22 HEV RNA-positive samples measured by each assay are shown
25 in Fig. S1. The results by the Promoter® assay and the LDT were linearly associated

26 and correlated ($R^2=0.8611$, $p < 0.0001$) (Fig. S1A). Bland-Altman analysis indicated
27 that the LDT gave a slightly higher viral load than did the Promoter® assay. The mean
28 [Promoter®-LDT] difference was $-0.76 \log_{10}$ copies/mL (Fig. S1B). Notably, the
29 other 6 samples with positive anti-HEV IgM but negative HEV RNA determined by
30 the LDT were confirmed negative by the Promoter® assay.

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33 **Fig S1** Agreement between HEV RNA concentrations measured by the LDT and the

34 Promoter® assay. (A) Linear regression analysis of HEV RNA concentrations

35 determined by the LDT and the Promoter® assay. (B) Bland-Altman plot for bias

36 analysis between the LDT and the Promoter® assay. The red dashed line represents

37 the mean of the difference ($n = -0.76 \log_{10}$ copies/mL) and black dotted lines show

38 the mean ± 2 standard deviation.

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40 **Table S1** Detection results of 10-fold dilutions of HEV RNA standard

HEV RNA concentration		Promoter®				LDT			
copies/mL	log ₁₀ (copies/mL)	Detected/Tested	Mean <i>Ct</i>	SD	CV%	Detected/Tested	Mean <i>Ct</i>	SD	CV%
5.35E+07	7.73	5/5	18.95	0.24	1.27	5/5	22.80	0.13	0.57
5.35E+06	6.73	5/5	22.40	0.19	0.85	5/5	26.20	0.22	0.84
5.35E+05	5.73	5/5	25.75	0.14	0.54	5/5	29.55	0.25	0.85
5.35E+04	4.73	5/5	29.02	0.39	1.34	5/5	33.02	0.20	0.60
5.35E+03	3.73	5/5	32.51	0.50	1.54	5/5	35.66	0.54	1.51
5.35E+02	2.73	3/5	35.49	0.71	2.00	0/5	/	/	/

41 LDT: laboratory-developed test; *Ct*: cycle threshold; SD: standard deviation; CV: coefficient of variation

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45 **Table S2** Detection limit of the RT-PCR assays

HEV RNA concentration		Promoter®		LDT	
copies/mL	log ₁₀ copies/mL	Detected/Tested	%	Detected/Tested	%
			Detected		Detected
5.35E+04	4.73	3/3	100.0	5/5	100.0
1.78E+04	4.25	3/3	100.0	5/5	100.0
5.94E+03	3.77	3/3	100.0	5/5	100.0
1.98E+03	3.30	3/3	100.0	4/5	80.0
6.60E+02	2.82	2/3	67.7	0/5	0.0
2.20E+02	2.34	1/3	33.3	0/5	0.0
7.34E+01	1.87	0/3	0.0	0/5	0.0

46 Probit value (95% detection rate) for the Promoter® assay = 1849 copies/mL (95% CI: 258.4-
 47 13234).

48 Probit value (95% detection rate) for the LDT assay = 2295 copies/mL (95% CI: 1.612-
 49 3265472).

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52 **Table S3** HEV Ag and HEV RNA levels in sample A and sample B diluted with either
 53 positive or negative anti-HEV IgG serum

	IgG (COI)	IgM (COI)	HEV Ag (S/CO)	HEV RNA (log ₁₀ copies/mL)
IgG positive pooled sera	16.7	0.32	0.08	(-)
IgG negative pooled sera	0.02	0.05	0.08	(-)
Sample #A	7.81	42.29	37.11	7.44
#A_1/10 diluted by IgG_positive sera	/	/	0.07	6.20
#A_1/100 diluted by IgG_positive sera	/	/	0.09	5.23
#A_1/1000 diluted by IgG_positive sera	/	/	0.06	4.29
#A_1/10 diluted by IgG_negative sera	/	/	23.79	6.35
#A_1/100 diluted by IgG_negative sera	/	/	4.21	5.43
#A_1/1000 diluted by IgG_negative sera	/	/	0.41	3.96
Sample #B	4.47	12.06	34.81	7.91
#B_1/10 diluted by IgG_positive sera	/	/	0.06	7.32
#B_1/100 diluted by IgG_positive sera	/	/	0.08	6.50
#B_1/1000 diluted by IgG_positive sera	/	/	0.08	5.42
#B_1/10 diluted by IgG_negative sera	/	/	16	7.39
#B_1/100 diluted by IgG_negative sera	/	/	1.81	6.36
#B_1/1000 diluted by IgG_negative sera	/	/	0.15	5.28

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55 **Table S4** Multivariate stepwise logistic regression analysis of factors associated with HEV

56 RNA presence

Variables	Current HEV infection					
	univariate analysis			multivariate analysis		
	β coefficient	OR (95% CI)	<i>p</i> value	β coefficient	OR (95% CI)	<i>p</i> value
Age	0.017	1.02 (1.00–1.04)	0.098	–	–	–
Gender	0.276	1.32 (0.76–2.29)	0.328	–	–	–
Log ₁₀ (Ag)	3.04	20.88 (8.62-50.57)	< 0.001	3.08	25.52 (4.75-136.97)	< 0.001
Log ₁₀ (IgG)	-0.97	0.38 (0.11-1.34)	0.132	–	–	–
Log ₁₀ (IgM)	2.94	18.86 (8.47-41.96)	< 0.001	5.14	175.57 (19.78- > 999.99)	< 0.001
$\sqrt{ALT/ULN}$	1.61	5.00 (2.98-8.40)	< 0.001	2.64	12.28(2.09-72.01)	< 0.001

57 OR: odds ratio; CI: confidence interval; Ag: antigen; ALT: alanine aminotransferase; ULN:

58 upper limit of normal

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