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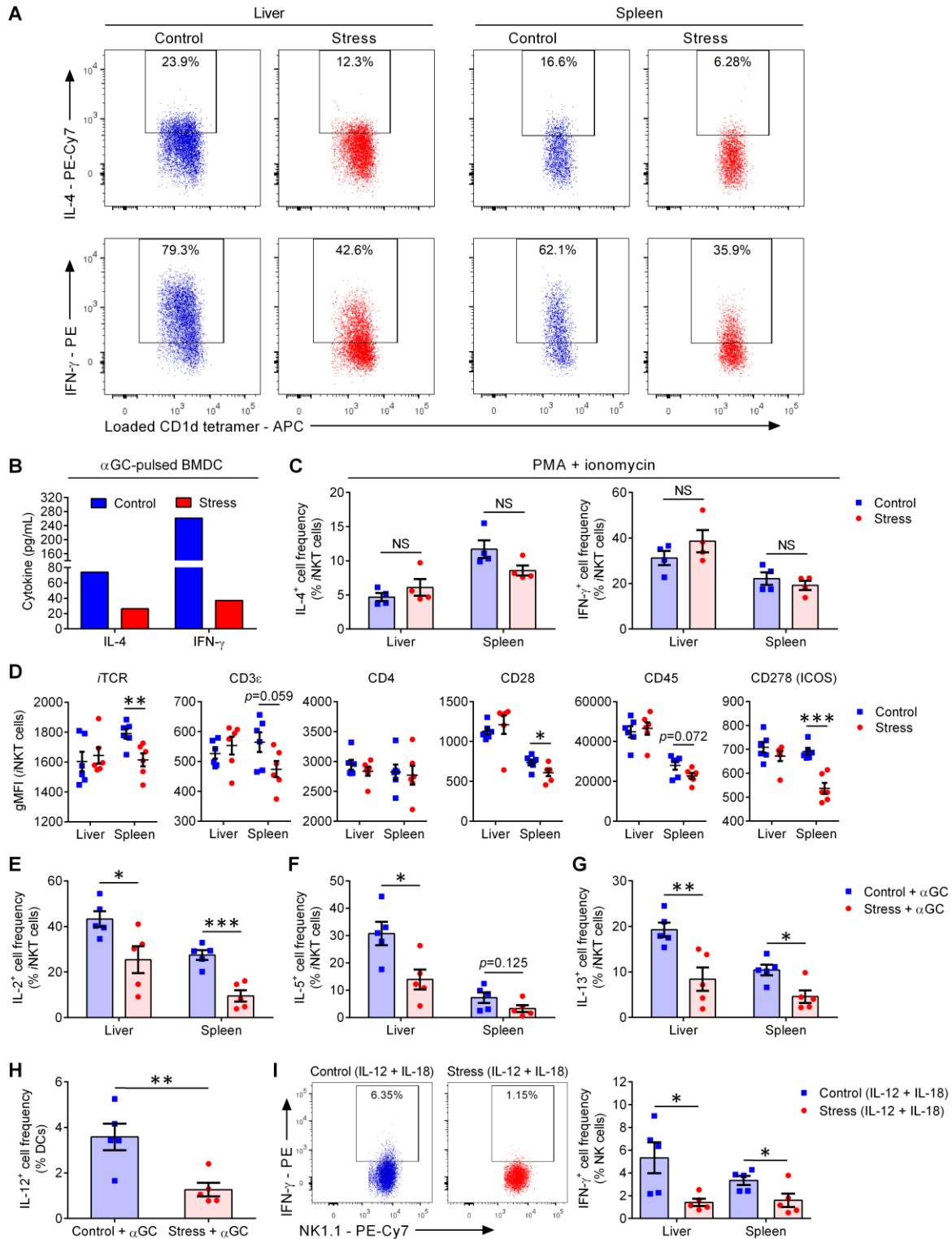
Supplemental information

Chronic stress physically spares

but functionally impairs

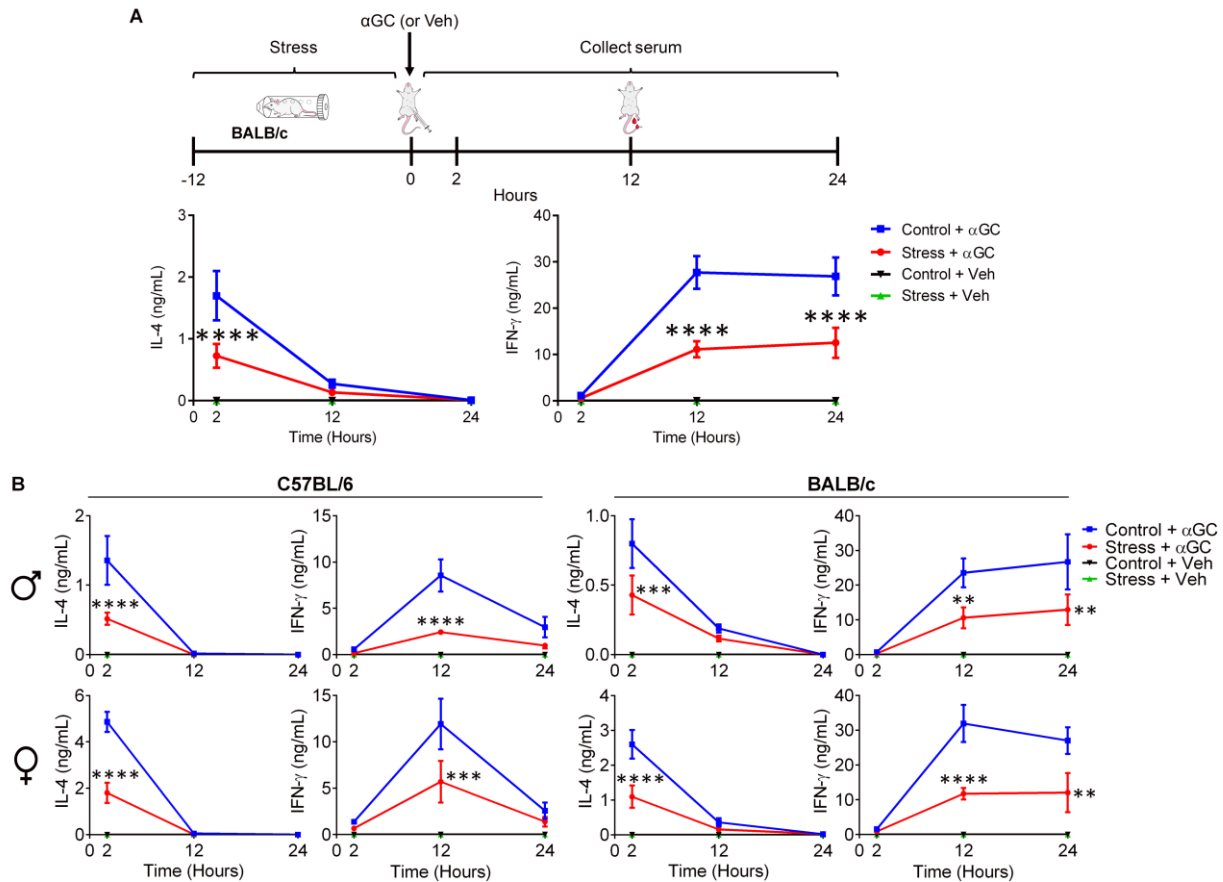
innate-like invariant T cells

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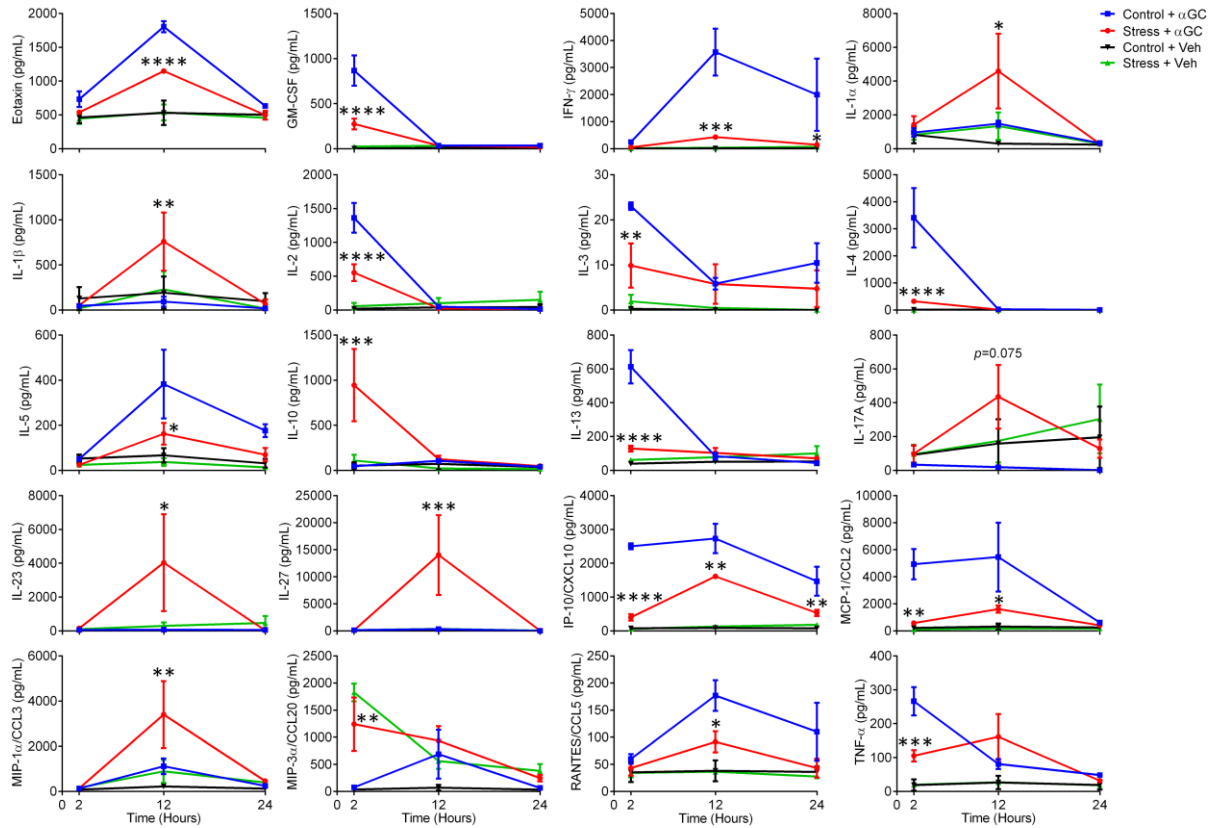


Supplemental Figure 1. Prolonged physical restraint impairs the ability of i NKT cells to elicit T_H1 and/or T_H2 -type cytokine responses to α GC or to a combination of IL-12 and IL-18, but not to a combination of PMA

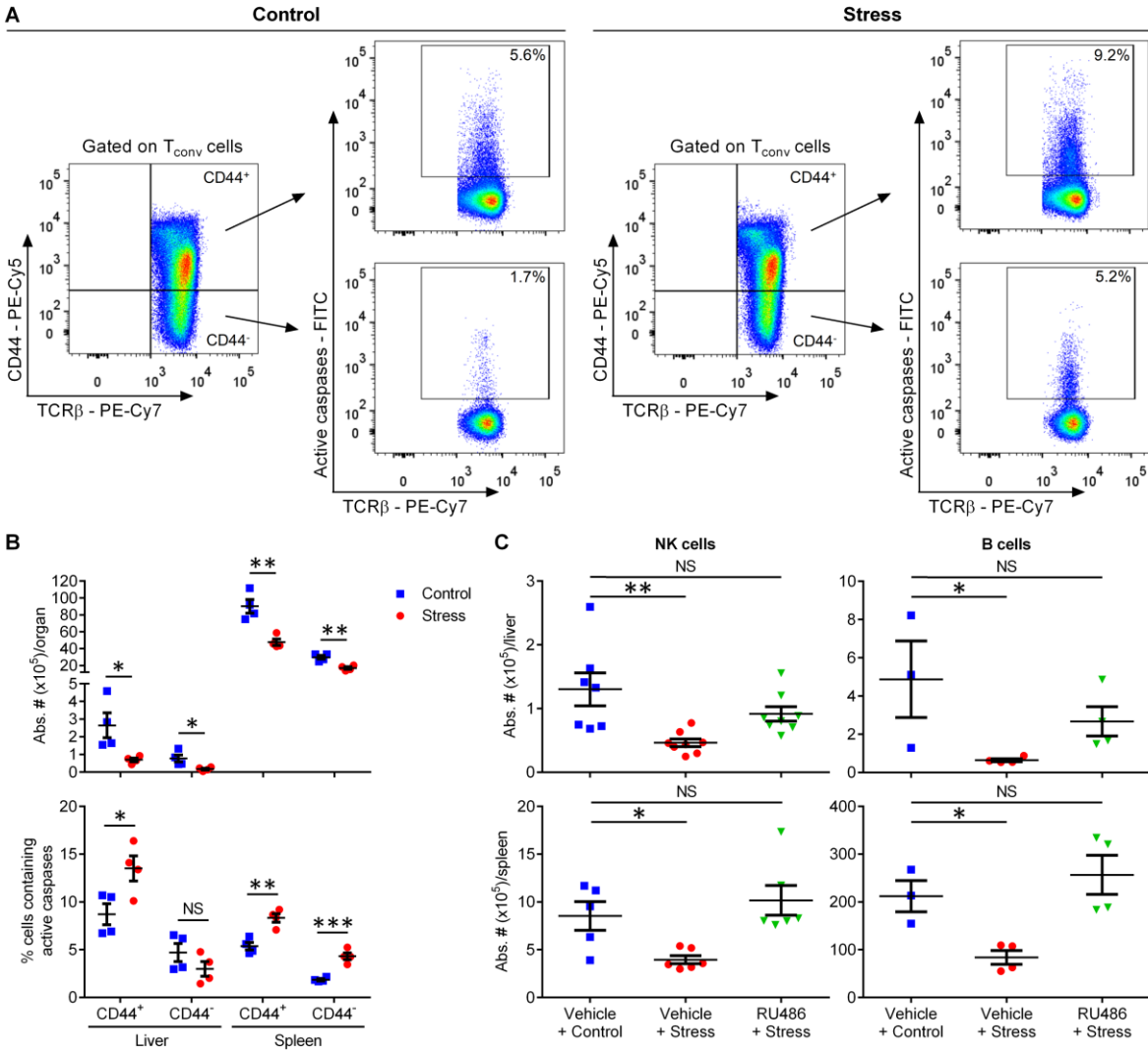
and ionomycin (related to Figure 1). (A) B6 mice were physically restrained (or not) for 12 h before they were given α GC. Two hours later, HMNCs and splenocytes were stained with mAbs against IL-4 and IFN- γ and analyzed by flow cytometry. Representative dot plots illustrate the frequencies of IL-4⁺ and IFN- γ ⁺ *i*NKT cells after gates were set based on isotype control staining. (B) Sorted hepatic *i*NKT cells pooled from ≥ 5 stressed or control B6 mice were stimulated *ex vivo* with 100 ng/mL of α GC in the presence of CD11c⁺ BMDCs. After 24 h, IL-4 and IFN- γ levels in culture supernatants were measured. (C) HMNCs and splenocytes from stressed and control B6 mice were stimulated for 2 h with 15 ng/mL of PMA and 500 ng/mL of ionomycin before intracellular levels of IL-4 and IFN- γ in *i*NKT cells were determined by flow cytometry. (D) HMNCs and splenocytes from restrained or control animals were stained with loaded CD1d tetramer or mAbs against indicated molecules. After gating on *i*NKT cells, the gMFI of staining for each molecule is depicted. (E-G) As in (A), but summary data indicates the frequencies of *i*NKT cells staining positively for mAbs against IL-2 (E), IL-5 (F) and IL-13 (G). (H) B6 mice were restrained (or not) for 12 h before they were given α GC. Six h later, HMNCs were stained with an anti-IL-12p35 mAb or a rat IgG2 κ isotype control. The frequency of IL-12⁺ DCs was determined after gating on TCR β ⁺CD11c⁺ events. (I) Mice that had been restrained or left undisturbed were injected with IL-12 and IL-18 one h before the percentages of IFN- γ ⁺ events among TCR β ⁺NK1.1⁺ NK cells were determined. Representative flow plots and summary data are shown. Each symbol represents an individual mouse, and error bars represent SEM. *, ** and *** denote significant differences with $p < 0.05$, $p < 0.01$ and $p < 0.001$, respectively, by unpaired Student's *t*-tests. NS = not significant



Supplemental Figure 2. Prolonged stress impairs cognate *i*NKT cell responses in male and female B6 and BALB/c mice (related to Figure 1). (A) WT BALB/c mice were subjected to physical restraint or left undisturbed for 12 h before they were injected i.p. with α GC or Veh. At indicated time points, IL-4 and IFN- γ in serum samples were quantified by ELISA ($n=8-9$ /group). (B) Data presented in Figure 1B-C and Figure S2A were segregated by sex. The kinetics of serum cytokine levels following α GC (or Veh) administration in male (B6: $n=5$; BALB/c: $n=5$) and female (B6: $n=5$; BALB/c: $n=3-4$) mice are depicted. Error bars represent SEM. **, * and **** denote differences with $p<0.01$, $p<0.001$ and $p<0.0001$, respectively, using two-way ANOVA with Dunnett's post-hoc analysis.**

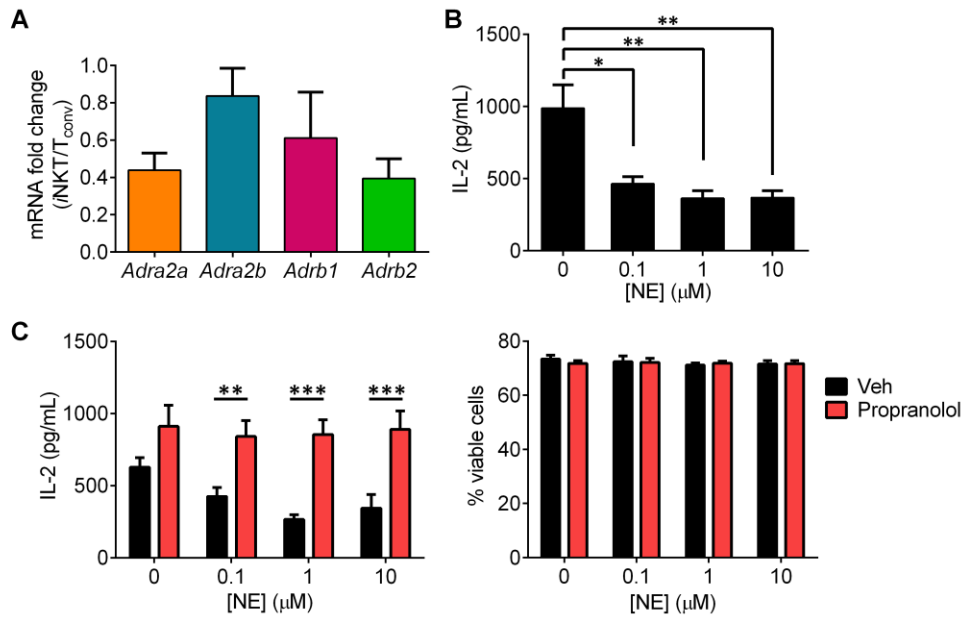


Supplemental Figure 3. Prolonged physical restraint before α GC administration alters the serum concentrations of a wide range of inflammatory mediators (related to Figure 1). WT B6 mice were restrained or left undisturbed for 12 h before they were injected with α GC or Veh (n=3/cohort). Two, 12 and 24 h later, mice were bled and serum levels of indicated mediators were quantified by cytokine/chemokine multiplexing. *, **, *** and **** denote significant differences between stressed and control animals receiving α GC with $p < 0.05$, $p < 0.01$, $p < 0.001$ and $p < 0.0001$, respectively, using two-way ANOVA with Tukey's post-hoc analysis.

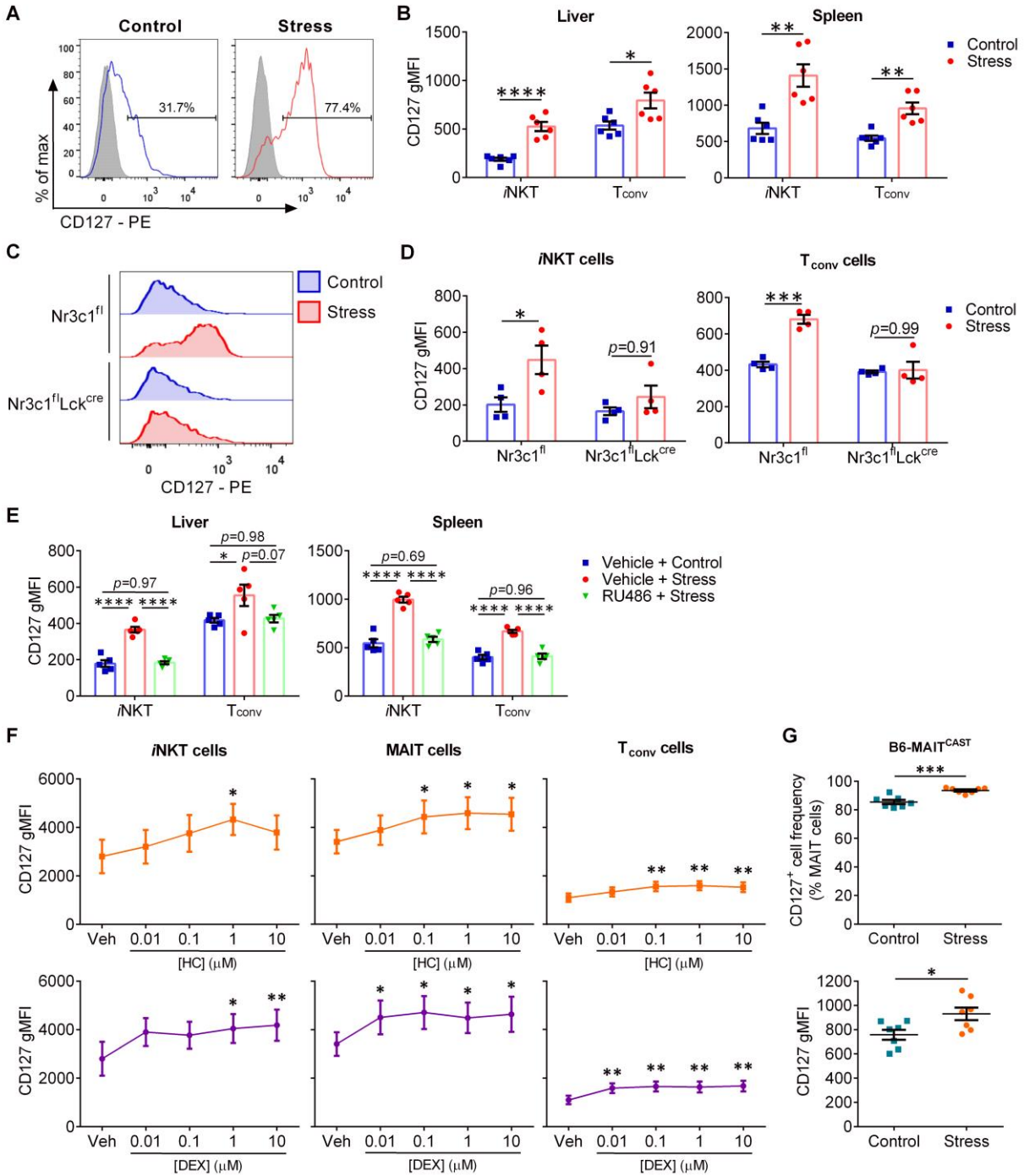


Supplemental Figure 4. Unlike *i*NKT cells, naïve and memory T_{conv} cells, NK cells and B cells are prone to stress-induced apoptosis (related to Figure 2). (A-B) Splenocytes and HMNCs from restrained and control B6 mice were stained for surface TCR β , surface CD44 and intracellular active caspases. A rat IgG2b κ isotype control was used to set the gate for CD44 staining. (A) Representative FACS plots illustrate our gating strategy to distinguish between splenic CD44 $^+$ and CD44 $^-$ populations among TCR β^+ CD1d tetramer $^-$ events, which correspond to memory and naïve T_{conv} cells, respectively. Gates containing T_{conv} cells with intracellular active caspases are also shown. (B) The absolute numbers of hepatic and splenic memory and naïve T_{conv} cells and the percentages of T_{conv} cells staining positively for active caspases are depicted. (C) Separate cohorts of B6 mice were treated i.p. with RU486 or Veh.

One hour later, animals were subjected to prolonged restraint stress or were left undisturbed for 12 h. HMNCs and splenocytes were prepared shortly afterwards and stained with a panel of mAbs against TCR β , NK1.1 and B220. TCR β NK1.1⁺ NK cells and TCR β B220⁺ B cells were identified by flow cytometry, and their absolute numbers were calculated. Each symbol represents an individual mouse, and error bars represent SEM. *, ** and *** denote differences with $p < 0.05$, $p < 0.01$ and $p < 0.001$, respectively, using unpaired Student's *t*-tests (**B**) or one-way ANOVA with Dunnett's post-hoc analysis (**C**). NS = not significant

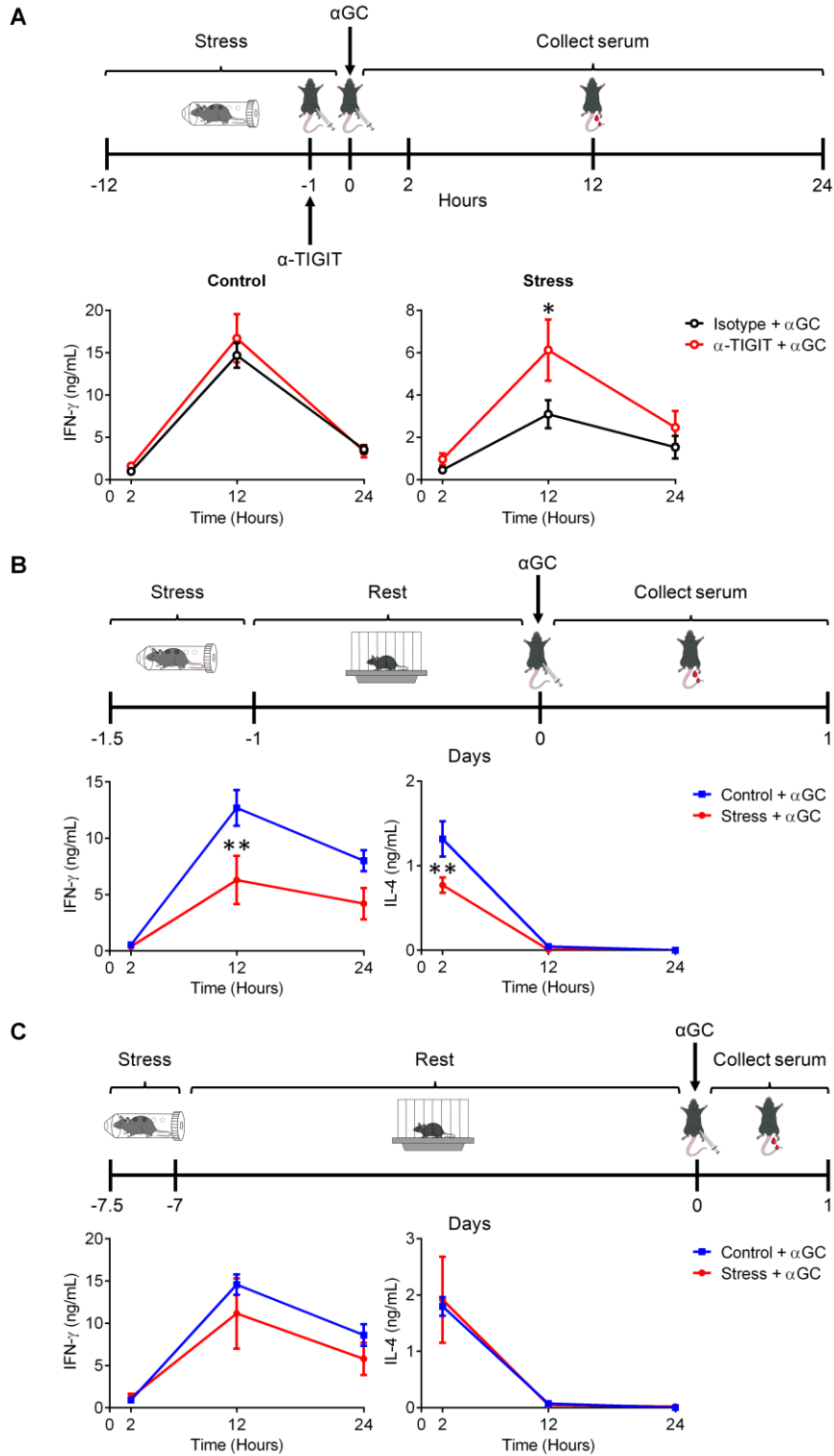


Supplemental Figure 5. *i*NKT cells express adrenergic receptors and respond weakly to α GC in the presence of NE in an *in vitro* setting (related to Figure 3). (A) HMNCs from 10 naïve B6 mice were pooled and co-stained with an anti-TCR β mAb and PBS-57-loaded CD1d tetramers. *i*NKT and T_{conv} cells were FACS-purified for cDNA synthesis and gene expression analysis by quantitative PCR. The expression of indicated genes by *i*NKT cells relative to T_{conv} cells was calculated using the $2^{-(\Delta\Delta C_t)}$ method. Data from 3 independent experiments were used to determine fold change values. (B) DN32.D3 cells were exposed for 20 minutes to indicated concentrations of norepinephrine (NE) before they were stimulated with 100 ng/mL of α GC. The IL-2 content of culture supernatants was quantified after 24 h by ELISA (n=3-5). (C) DN32.D3 cells were pretreated for 20 minutes with propranolol before they were exposed to NE and subsequently stimulated with α GC (n=4). After 24 h, IL-2 was measured in supernatants (n=4) and cellular viability was assessed by 7-AAD staining (n=3). Error bars represent SEM. *, ** and *** denote differences with $p < 0.05$, $p < 0.01$ and $p < 0.001$, respectively, by one-way ANOVA with Dunnett's post-hoc analysis (B) or by unpaired Student's *t*-tests (C).



Supplemental Figure 6. Mouse and human *i*NKT, MAIT and T_{conv} cells upregulate CD127 in response to GR signaling (related to Figure 7). B6 mice were subjected to 12 h of restraint stress (or not). Shortly afterwards, HMNCs and splenocytes were prepared and stained to detect CD127 expression on the surface of *i*NKT and T_{conv}

cells. Open and filled histograms correspond to staining with an anti-CD127 mAb and a rat IgG2 κ isotype control, respectively, after gating on hepatic *i*NKT cells (**A**). Cumulative data depicting the gMFI of CD127 staining in indicated populations are also shown (**B**). Nr3c1^{fl} and Nr3c1^{fl}Lck^{cre} mice were physically restrained (or not) and assessed for CD127 expression in their *i*NKT and T_{conv} cell compartments (**C-D**). Blue and red histograms correspond to CD127 expression on hepatic *i*NKT cells from control and stressed mice, respectively (**C**), and cumulative data for indicated cell populations are summarized using bar graphs (**D**). Separate cohorts of WT B6 mice received RU486 (or Veh) i.p. 1 h before they were physically restrained (or not). The gMFI of CD127 staining is shown (**E**). Human PBMCs were cultured for 24 h in the presence of hydrocortisone (HC), dexamethasone (DEX) or vehicle before they were stained with either an anti-CD127 mAb or a mouse IgG1 κ isotype control. The gMFI of CD127 staining in *i*NKT, MAIT and T_{conv} cells was assessed by flow cytometry (n=5) (**F**). B6-MAIT^{CAST} mice were physically restrained or left undisturbed for 12 h before they were sacrificed for their liver. HMNCs were stained with a mAb to CD127 or a rat IgG2 κ isotype control. After gating on TCR β ⁺B220⁻MR1 tetramer⁺ MAIT cells, the percentages of CD127⁺ cells and the gMFI of CD127 staining were determined (**G**). Each symbol in (**B**, **D-E** and **G**) represents an individual mouse. Error bars represent SEM. *, **, *** and **** denote differences with $p < 0.05$, $p < 0.01$, $p < 0.001$ and $p < 0.0001$, respectively, using unpaired Student's *t*-tests (**B**, **D** and **G**) or one-way ANOVA (**E-F**).



Supplemental Figure 7. Stress-induced *i*NKT cell impairments are partially mediated by TIGIT and do not last long after the stressor is removed (related to Figure 4). (A) B6 mice were restrained or left undisturbed

before they received a 200- μ g i.p. dose of an anti-mouse TIGIT mAb (or a mouse IgG1 κ isotype control) followed by α GC administration as schematically illustrated. Mice were bled at indicated time points, and serum IFN- γ levels were quantified (n=8-9/group). **(B-C)** Separate cohorts of B6 mice were physically restrained or left undisturbed for 12 h. Mice were then returned to standard housing conditions for one day **(B)** or seven days **(C)** before they were injected i.p. with α GC. Serum IFN- γ and IL-4 levels were measured at indicated time points (n=4/group). Error bars represent SEM. * and ** denotes differences with $p<0.05$ and $p<0.01$, respectively, using two-way ANOVA with Sidak's post-hoc analysis.

Supplemental Table 1: Taqman-based qPCR primer/probe sets used in this study (related to STAR Methods).

Target	Assay Identification
<i>Abl1</i>	Mm00802029_m1
<i>Actb</i>	Mm00607939_s1
<i>Adra1a</i>	Mm00442668_m1
<i>Adra1b</i>	Mm00431685_m1
<i>Adra1d</i>	Mm01328600_m1
<i>Adra2a</i>	Mm00845383_s1
<i>Adra2b</i>	Mm00477390_s1
<i>Adra2c</i>	Mm00431686_s1
<i>Adrb1</i>	Mm00431701_s1
<i>Adrb2</i>	Mm02524224_s1
<i>Adrb3</i>	Mm02601819_g1
<i>Aifm1</i>	Mm00442540_m1
<i>Anxa5</i>	Mm01293059_m1
<i>Apaf1</i>	Mm01223702_m1
<i>Api5</i>	Mm00500189_m1
<i>Atf5</i>	Mm04179654_m1
<i>Bad</i>	Mm00432042_m1
<i>Bag1</i>	Mm01208593_m1
<i>Bag3</i>	Mm00443474_m1
<i>Bak1</i>	Mm00432045_m1
<i>Bax</i>	Mm00432051_m1
<i>Bbc3</i>	Mm00519268_m1
<i>Bcl2</i>	Mm00477631_m1
<i>Bcl2a1a</i>	Mm03646861_mH
<i>Bcl2l1</i>	Mm00437783_m1
<i>Bcl2l11</i>	Mm00437796_m1
<i>Bid</i>	Mm00432073_m1
<i>Bik</i>	Mm00476123_m1
<i>Birc2</i>	Mm00431811_m1
<i>Birc3</i>	Mm01168413_m1
<i>Birc5</i>	Mm00599749_m1
<i>Bmf</i>	Mm00506773_m1
<i>Bnip2</i>	Mm00443990_m1
<i>Card10</i>	Mm00459941_m1
<i>Card6</i>	Mm01297056_m1
<i>Casp1</i>	Mm00438023_m1
<i>Casp2</i>	Mm00432314_m1
<i>Casp3</i>	Mm01195085_m1
<i>Casp4</i>	Mm00432307_m1
<i>Casp6</i>	Mm00438053_m1
<i>Casp7</i>	Mm00432324_m1
<i>Casp8</i>	Mm00802247_m1
<i>Casp9</i>	Mm00516563_m1
<i>Cblb</i>	Mm01343092_m1

<i>Cd27</i>	Mm01185212_g1
<i>Cd28</i>	Mm00483137_m1
<i>Cd40lg</i>	Mm00441911_m1
<i>Cd44</i>	Mm01277161_m1
<i>Cd69</i>	Mm01183378_m1
<i>Cd274</i>	Mm03048248_m1
<i>Cdk2</i>	Mm00443947_m1
<i>Cdk4</i>	Mm00726334_s1
<i>Cflar</i>	Mm01255578_m1
<i>Cradd</i>	Mm01226172_m1
<i>Csf1</i>	Mm00432686_m1
<i>Dad1</i>	Mm01319221_m1
<i>Dffa</i>	Mm00438410_m1
<i>Dffb</i>	Mm00432822_m1
<i>Diablo</i>	Mm01194441_m1
<i>Egr2</i>	Mm00456650_m1
<i>Egr3</i>	Mm00516979_m1
<i>Fadd</i>	Mm00438861_m1
<i>Fas</i>	Mm01204974_m1 Mm00433237_m1
<i>Fasl</i>	Mm00438864_m1
<i>Fos</i>	Mm00487425_m1
<i>Foxo3</i>	Mm01185722_m1
<i>Foxp1</i>	Mm00474848_m1
<i>Gata3</i>	Mm00484683_m1
<i>Gzma</i>	Mm01304452_m1
<i>Gzmb</i>	Mm00442834_m1
<i>Hells</i>	Mm00468580_m1
<i>Icam1</i>	Mm00516023_m1
<i>Icos</i>	Mm00497600_m1
<i>Ifng</i>	Mm01168134_m1
<i>Ifnar1</i>	Mm00439544_m1
<i>Ifnar2</i>	Mm00494916_m1
<i>Igf1r</i>	Mm00802831_m1
<i>Il10ra</i>	Mm00434151_m1
<i>Il12rb1</i>	Mm00434189_m1
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<i>Il2</i>	Mm00434256_m1
<i>Il2ra</i>	Mm01340213_m1
<i>Il2rb</i>	Mm00434268_m1
<i>Il4</i>	Mm00445259_m1
<i>Il7ra/Cd127</i>	Mm00434295_m1
<i>Irf4</i>	Mm00516431_m1
<i>Itch</i>	Mm01246513_m1
<i>Jak1</i>	Mm00600614_m1
<i>Jak3</i>	Mm00439962_m1
<i>Jun</i>	Mm00495062_s1

<i>Lat</i>	Mm00456761_m1
<i>Lta</i>	Mm00440228_gH
<i>Myb</i>	Mm00501741_m1
<i>Naip2</i>	Mm00440446_m1
<i>Nfatc1</i>	Mm00479445_m1
<i>Nfatc2</i>	Mm00477776_m1
<i>Nfatc3</i>	Mm01249200_m1
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<i>Npy6r</i>	Mm00627550_m1
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<i>Nr4a1</i>	Mm01300401_m1
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<i>Pmaip1</i>	Mm00451763_m1
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<i>Xiap</i>	Mm01311594_mH
<i>Zbtb16</i>	Mm01176868_m1
<i>Zc3hc1</i>	Mm01168068_m1