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Supplementary Fig. 1 RCT-aAPCs and murine surrogate mRBC-aAPCs. a, Generation of

RCT-aAPCs that express peptide-MHC class I complex fused to an antigenic peptide as signal 1, 4-1BBL as signal 2, and a cytokine as signal 3. **b**, Generation of mRBC-aAPCs using a click chemistry reaction to conjugate on the surface of mRBC peptide-MHC class I complex as signal 1, 4-1BBL as signal 2, and a cytokine as signal 3. **c-d**, CellTrace Violet dye-labeled OT-1 cells (1*x*10⁵ per well, n=3) were co-cultured with RCT-OVA-4-1BBL-IL-12, RCT-CTRL, mRBC-CTRL, or mRBC-OVA-4-1BBL-IL-12 at a 1:9, 1:3 or 1:1 ratio. % proliferation in OT-1 cells (**c**), and IFNγ concentration in supernatant (**d**) on day 4. Data are depicted as mean ± s.d and are representative of two independent experiments. Source data are provided as a Source Data file.



Supplementary Fig. 2 Antigen-specificity of RCT-aAPCs. a, EL4 percent killing by RCT-activated OT-1 CD8⁺ T cells (n=3). **b**, OT-1 or pmel-1 numbers after 3 days co-incubation with a dose titration $(1x10^{6}, 3x10^{5}, \text{ or } 1x10^{5})$ of either RCT-CTRL or RCT-OVA-4-1BBL (n=3) with $1x10^{5}$ T cells, or $1x10^{5}$ anti-CD3/CD28 beads with $1x10^{5}$ T cells. Data are depicted as mean \pm s.d and are representative of two (**a**) or one (**b**) independent experiments. Source data are provided as a Source Data file.



Supplementary Fig. 3 mRBC-OVA-4-1BBL-IL-12 promotes tumor regressions in EG7.OVA tumor-bearing mice. CD45.1 Pep Boy mice were injected subcutaneously with $2x10^6$ EG7.OVA cells. When the tumors reached a volume of ~150 mm³, the animals were randomized (n=8) and dosed on day 1 post randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on days 1, 4 and 7 with mRBC. **a**–**c**, Tumor regression (TR, reduction of tumor volume on day 24 compared to tumor volume at randomization) in $2.5x10^8$ mRBC-OVA-4-1BBL-IL-12 (**a**), $1x10^9$ mRBC-OVA-4-1BBL (**b**), or $2.5x10^8$ mRBC-OVA-4-1BBL (**c**) treated mice. In a separate study, CD45.1 Pep Boy mice were injected subcutaneously with $2x10^6$ EG7.OVA cells. When the tumors reached a volume of ~230 mm³, the animals were randomized (n=8) and dosed on day 1 post-randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on day 1 post-randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on day 1 post-randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on day 1 post-randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on day 1 post-randomization with $1x10^6$ naïve OT-1 cells. The animals were then dosed on days 1, 4 and 7 with mRBC. **d-e**, TR of $2.5x10^8$ mRBC-OVA-4-1BBL-IL-12 (**d**) or $2.5x10^8$ mRBC-4-1BBL-IL-12 (**e**) treated mice. Results are representative of five (**a,d**), three (**b**), or two (**c**, **e**) independent experiments. Source data are provided as a Source Data file.



a Ki67 CD8 in tumor

TNF α CD8 in tumor

IL-2 CD8 in tumor

Supplementary Fig. 4 mRBC-OVA-4-1BBL-IL-12 promoted general anti-tumor immune effects.

CD45.1 Pep Boy mice were inoculated subcutaneously with $2x10^6$ EG7.OVA cells. When the tumors reached a volume of ~175 mm³, the animals were randomized (n=5) and treated with $1x10^6$ naïve OT-1 cells. After, $1x10^9$ mRBC-CTRL or a dose titration of mRBC-OVA-4-1BBL-IL-12 ($1x10^9$, $2.5x10^8$) was administered on days 0 and 3. Mice were sacrificed on day 7. **a**, Ki67, TNF α , and IL-2 per cell expression in tumor infiltrating endogenous CD8⁺ T cells; **b**, Ki67, and granzyme B per cell expression in tumor-infiltrating NK cells; **c**, Treg% and IFN γ ⁺Ki67⁺% in tumor infiltrating CD4⁺ T cells; and **d**, M1 macrophage % in tumor infiltrating leukocytes. Data are depicted as mean ± s.d. and are representative of two independent experiments. One-way ANOVA compared to mRBC-CTRL. Source data are provided as a Source Data file.



Supplementary Fig. 5 mRBC-OVA-4-1BBL-IL-12 was well tolerated. CD45.1 Pep Boy mice (n=10) were treated with $1x10^6$ naïve OT-1 cells or left untreated, and dosed with either $1x10^9$ mRBC-CTRL or a dose titration of mRBC-OVA-4-1BBL-IL-12 ($1x10^9$ or $3x10^8$) on days 0, 4, 7 and 11. n=5 were sacrificed on Day 12 with n=5 measured for time points post day 12 for all groups except for n=4 for mRBC-OVA-4-1BBL-IL-12+OT-1. a, Spleen weight; b, liver weight; c, serum aspartate aminotransferase (AST) levels; d, serum alkaline phosphatase (ALP) levels; e, liver perivascular inflammation score; f, liver macrophage infiltration; g, hematocrit levels; h, hemoglobin levels; i, platelet levels on days 12 and 25 (a-i: n=5 for all groups except for n=4 for mRBC-OVA-4-1BBL-IL-12+OT-1 on day 25). j, Plasma IL-10 levels; k, plasma TNF α levels; I, plasma IL-6 levels; m, plasma IL-1 β levels over time (j-m: n=10 for days 3 and 7, n=5 for days 10, 12, 17 and 25 except for n=4 for mRBC-OVA-4-1BBL-IL-12+OT-1 on days 17 and 25). One-way ANOVA (a, b, c, d, f, g, h, i), one-way ANOVA nonparametric Kruskal-Wallis test with Dunn's multiple comparisons test (e), two-way ANOVA (j, k, I, m) compared to mRBC-CTRL. Data are depicted as mean ± s.d. and are representative of two independent experiments. Source data are provided as a Source Data file.











Supplementary Fig. 6 mRBC-OVA-4-1BBL-IL-12 interacts with OT-1 cells in the spleen. CD45.1 Pep Boy mice (n=5) were transferred with $2x10^6$ naïve CellTrace Yellow dye-labeled OT-1 cells before dosing with $1x10^9$ CellTrace Far Red dye-labeled mRBC-CTRL, or mRBC-OVA-41BBL-IL12. One hour or 17 hours post mRBC injection, mice were sacrificed. **a-b**, immunofluorescence analyses of spleen. Representative confocal image of OT-1 and mRBC-OVA-4-1BBL-IL12 interaction 1-hour post dose (**a**) and quantification of % OT-1 co-localized with mRBCs (**b**). One-way ANOVA compared to mRBC-CTRL and time points; 1-hour mRBC-CTRL vs mRBC-OVA-4-1BBL-IL-12 *P* = 0.036. **c-d**, Flow cytometry analyses of (**c**) CellTrace Far Red (CTFR) dye⁺ mRBC signal associated with OT-1 cells (**c**) and % CD44⁺ OT-1 cells (**d**). One-way ANOVA compared to mRBC-CTRL vs mRBC-OVA-4-1BBL-IL-12 *P* = 0.0063; 1 hour % CD44⁺ OT-1 mRBC-CTRL vs mRBC-OVA-4-1BBL-IL-12 *P* = 0.0063; 1 hour % CD44⁺ OT-1 mRBC-CTRL vs mRBC-OVA-4-1BBL-IL-12 *P* = 0.0063; 1 hour % CD44⁺ OT-1 mRBC-CTRL vs mRBC-OVA-4-1BBL-IL-12 *P* < 0.0001; % CD44⁺ OT-1 mRBC-CTRL vs independent experiments. Source data are provided as a Source Data file.



Supplementary Fig. 7 Flow cytometry gating scheme for in vitro assays. a, Flow cytometry gating scheme for tumor killing assay in Fig. 1a and Supplementary Fig. 2a. b, Flow cytometry gating scheme for aAPC and OT-1 or pmel-1 co-culture assays in Supplementary Fig. 1c and 2b. c, Flow cytometry gating scheme for assays shown in Fig. 6. L/D, LIVE/DEAD[™] stain; CTFR, CellTrace[™] Far Red; CTV, CellTrace[™] Violet; T_{em}, effector memory T cells; T_{cm}, central memory T cells.



Supplementary Fig. 8 Flow cytometry gating scheme for in vivo studies. a, Flow cytometry

gating scheme for OT-1 T cell analyses in the blood in Fig. 1c, d, j, I (analysis I), Fig. 3d, e (analysis II), and Fig. 4c (analysis III). **b**, Flow cytometry gating scheme for OT-1 T cell analyses in the tumor in Fig. I, n (analysis I), Supplementary Fig. 4a-c (analysis II), and Supplementary Fig. 4d (analysis III). **c**, Flow cytometry gating scheme for pmel-1 T cell analyses in Fig. 5. **d**, Flow cytometry gating scheme for OT-1 T cell analyses in the spleen in Supplementary Fig. 6c, d. L/D, LIVE/DEAD[™] stain; CTV, CellTrace[™] Violet; CTFR, CellTrace[™] Far Red; CTY, CellTrace[™] Yellow; T_{em}, effector memory T cells; T_{cm}, central memory T cells; T_{scm}, stem memory T cells.