

Supplementary Information

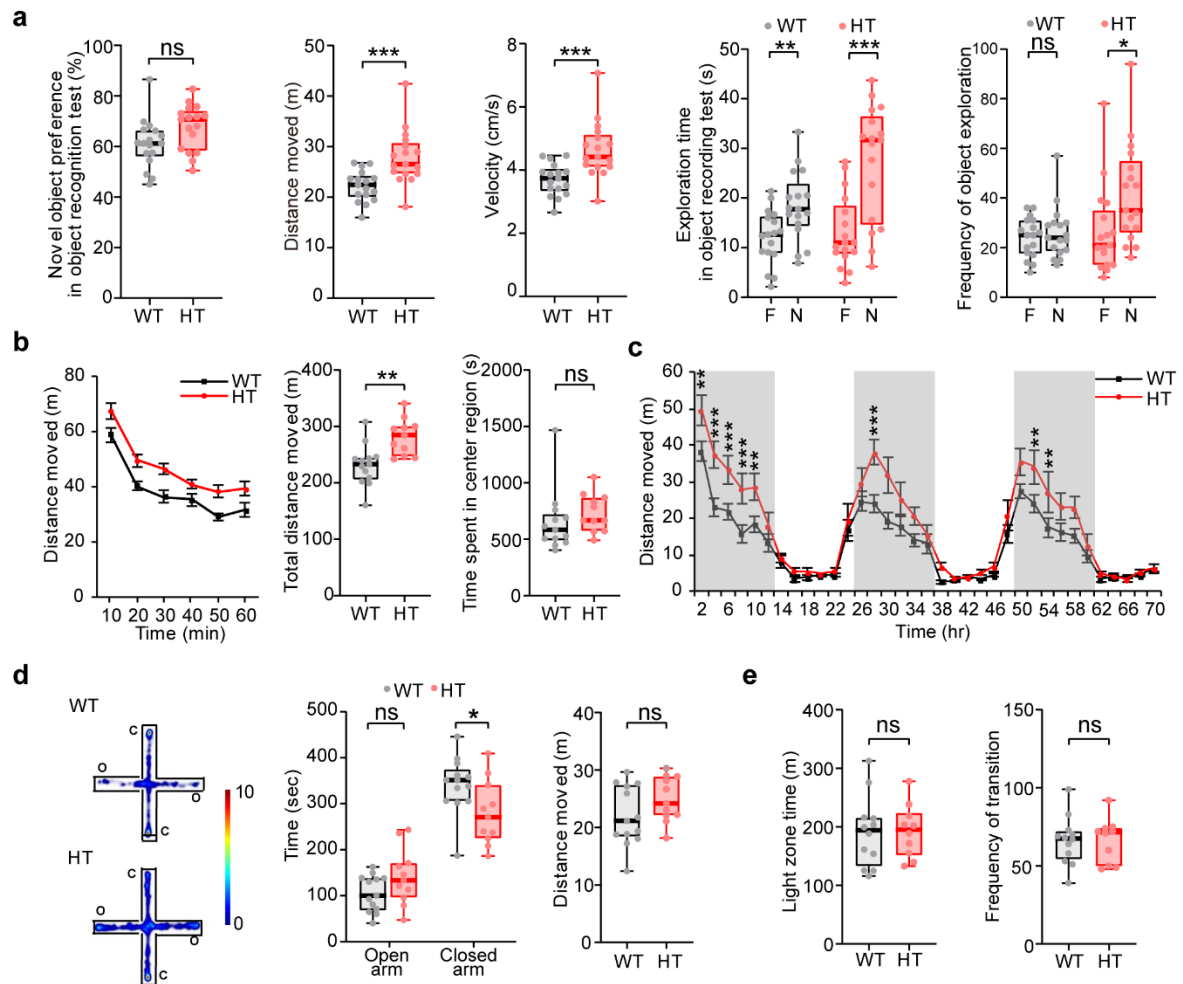
Tanc2-mediated mTOR inhibition balances mTORC1/2 signaling in the developing mouse brain and human neurons

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Supplementary figures and legends

Supplementary Fig. 1



Supplementary Fig. 1. Hyperactivity and moderate anxiolytic-like behavior in male *Tanc2*^{+/-} mice.

(a) Normal novel object-recognition memory in *Tanc2*^{+/-} mice (2–5 months; male) in the novel object-recognition test, as shown by novel object preference (% time spent in exploring a novel object relative to the total time spent exploring both familiar and novel objects). F, familiar object; N, new object. Note that these mice also showed hyperactivity in the novel object arena, as supported by the distance moved and movement velocity. The increased frequency of novel object exploration in the mutant mice might be attributable to the increased hyperactivity. Data: minimal, maximal, median, 25%, and 75% values. (n = 17 [WT, HT], *P < 0.05, **P < 0.01, ***P < 0.001, ns, not significant, Student's t-test).

(b) Hyperactivity of *Tanc2*^{+/-} mice (2–5 months; male) in the open-field test, as shown by distance moved over 60 minutes and total distance moved. Note also that the

time spent in the center region of the open field arena was normal in *Tanc2*^{+/-} mice, indicative of normal anxiety-like behavior. Data: mean ± SEM (line graphs), minimal, maximal, median, 25%, and 75% values. (n = 13 [WT], 11 [HT], **P < 0.01, ns, not significant, two-way RM ANOVA and Student's t-test).

(c) Hyperactivity of *Tanc2*^{+/-} mice (2–5 months; male) in the Laboras test, as shown by distance moved over 72 hours. The shaded areas represent light-off periods. Data: mean ± SEM. (n = 16 [WT, HT], **P < 0.01, ***P < 0.001, two-way RM ANOVA [genotype effect P < 0.05]).

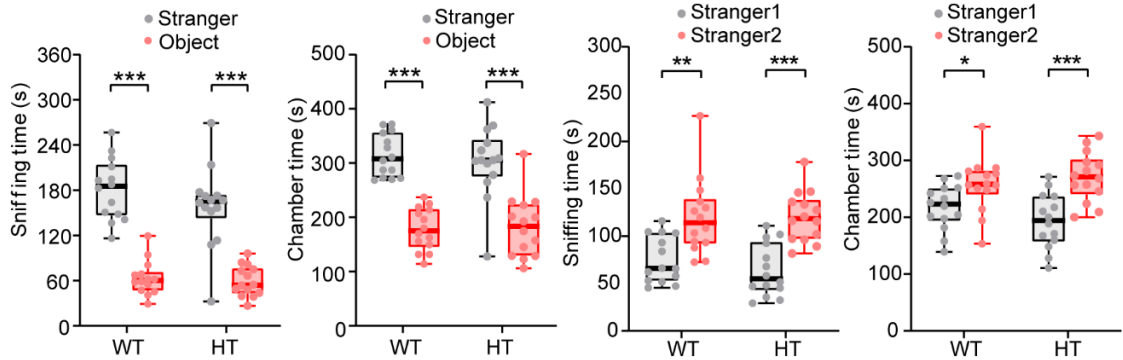
(d) Moderate anxiolytic-like behavior of *Tanc2*^{+/-} mice (2–5 months; male) in the elevated plus-maze test, as shown by time spent in open and closed arms. There was no genotype difference in the total distance moved. Data: minimal, maximal, median, 25%, and 75% values. (n = 13 [WT], 11 [HT], *P < 0.05, ns, not significant, two-way RM ANOVA with Bonferroni test and Student's t-test).

(e) Normal anxiety-like behavior of *Tanc2*^{+/-} mice (2–5 months; male) in the light-dark test, as shown by time spent in the light chamber/zone and frequency of transitions between light and dark chambers. Data: minimal, maximal, median, 25%, and 75% values. (n = 13 [WT], 10 [HT], ns, not significant, Student's t-test).

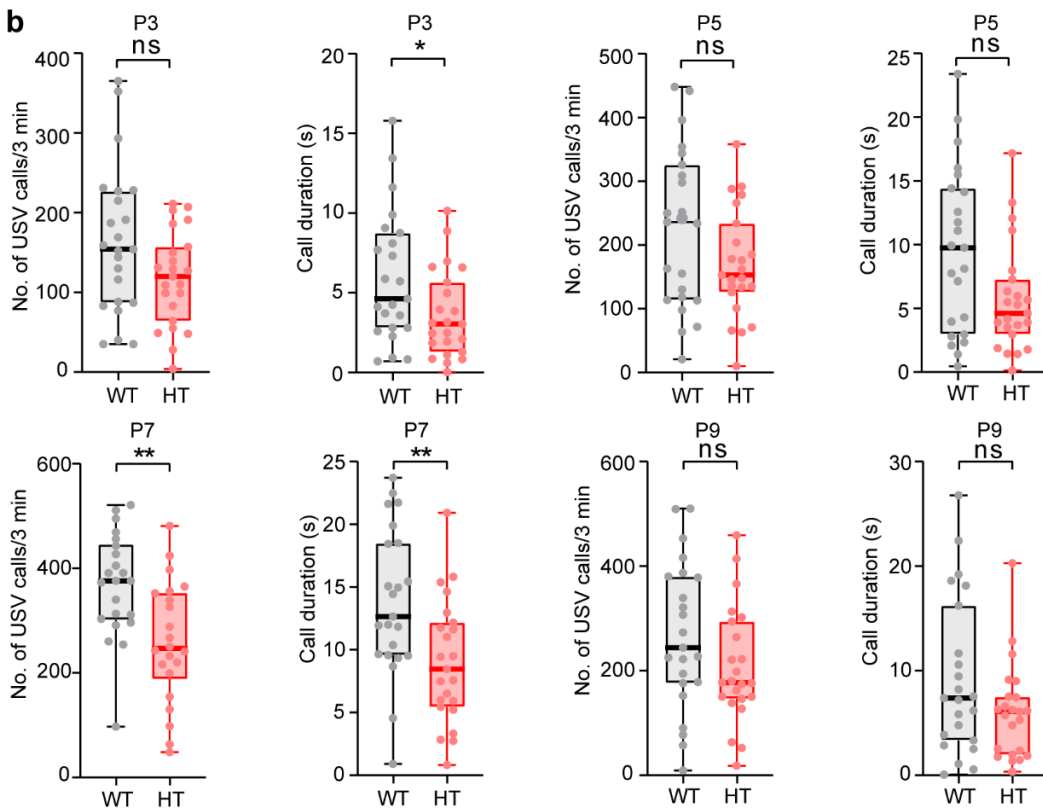
See Source Data 1 for raw data values and Supplementary Table 1 for statistical details.

Supplementary Fig. 2

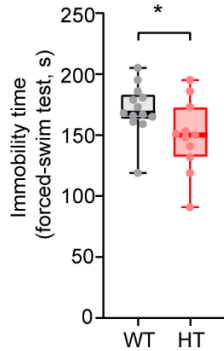
a



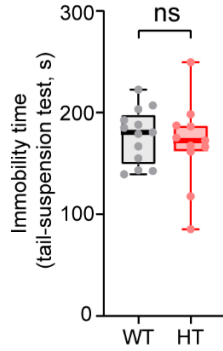
b



c



d



Supplementary Fig. 2. Suppressed pup ultrasonic vocalization but normal social interaction, social novelty recognition, and depression-like behavior in male *Tanc2*^{+/-} mice.

(a) Normal social interaction and social-novelty recognition in *Tanc2*^{+/-} mice (2–5 months; male) in the three-chamber social-interaction test, as shown by time spent in the chamber and in sniffing the target (stranger/object). Stranger 1, social stranger mouse; Object, inanimate object; Stranger 2, new social stranger mouse. Data: minimal, maximal, median, 25%, and 75% values. (n = 14 mice [WT, HT], ***P < 0.001, Student's t-test).

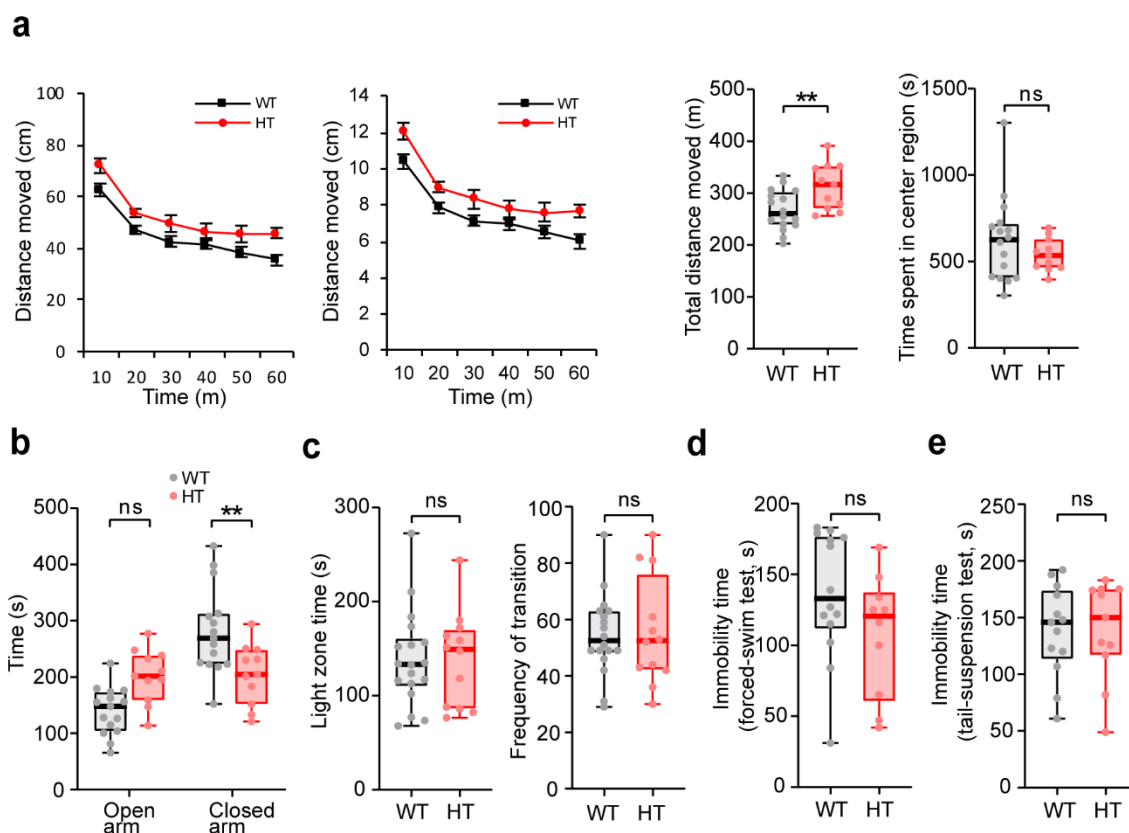
(b) Modestly suppressed ultrasonic vocalizations (USVs) in *Tanc2*^{+/-} pups (P3–9; male) separated from their mothers, as shown by the total number of USV calls and the duration of each call. Data: minimal, maximal, median, 25%, and 75% values. n = 23 pups [WT, HT], *P < 0.05, **P < 0.01, ns, not significant, Student's t-test).

(c) Anti-depression-like behavior of *Tanc2*^{+/-} mice (2–5 months; male) in the forced-swim test, as shown by immobility time. Data: minimal, maximal, median, 25%, and 75% values. (n = 13 [WT], 11 [HT], *P < 0.05, Student's t-test).

(d) Normal depression-like behavior of *Tanc2*^{+/-} mice (2–5 months; male) in the tail-suspension test, as shown by immobility time. Data: minimal, maximal, median, 25%, and 75% values. (n = 13 [WT], 11 [HT], ns, not significant, Student's t-test).

See Source Data 1 for raw data values and Supplementary Table 1 for statistical details.

Supplementary Fig. 3



Supplementary Fig. 3. Female *Tanc2*^{+/-} mice display hyperactivity and anxiolytic-like behavior, but normal depression-like behavior.

(a) Hyperactivity of female *Tanc2*^{+/-} mice (2–5 months) in the open-field test, as shown by distance moved over 60 minutes and total distance moved. Note also that the time spent in the center region of the open field arena was normal in female *Tanc2*^{+/-} mice, indicative of normal anxiety-like behavior. Data: mean ± SEM (line graphs), minimal, maximal, median, 25%, and 75% values. (n = 16 mice [WT], and 11 mice [HT], **P < 0.01, ns, not significant, two-way RM ANOVA and Student's t-test).

(b) Moderate anxiolytic-like behavior of female *Tanc2*^{+/-} mice (2–5 months) in the elevated plus-maze test, as shown by time spent in open and closed arms. Data: minimal, maximal, median, 25%, and 75% values. (n = 15 mice [WT], and 11 mice [HT], **P < 0.01, ns, not significant, two-way RM ANOVA with Bonferroni test).

(c) Normal anxiety-like behavior of female *Tanc2*^{+/-} mice (2–5 months) in the light-dark test, as shown by time spent in the light chamber and frequency of transitions between light and dark chambers. Data: minimal, maximal, median, 25%, and 75% values. (n = 18 mice [WT], and 12 mice [HT], ns, not significant, Student's t-test).

(d) Normal depression-like behavior of female *Tanc2*^{+/-} mice (2–5 months) in the

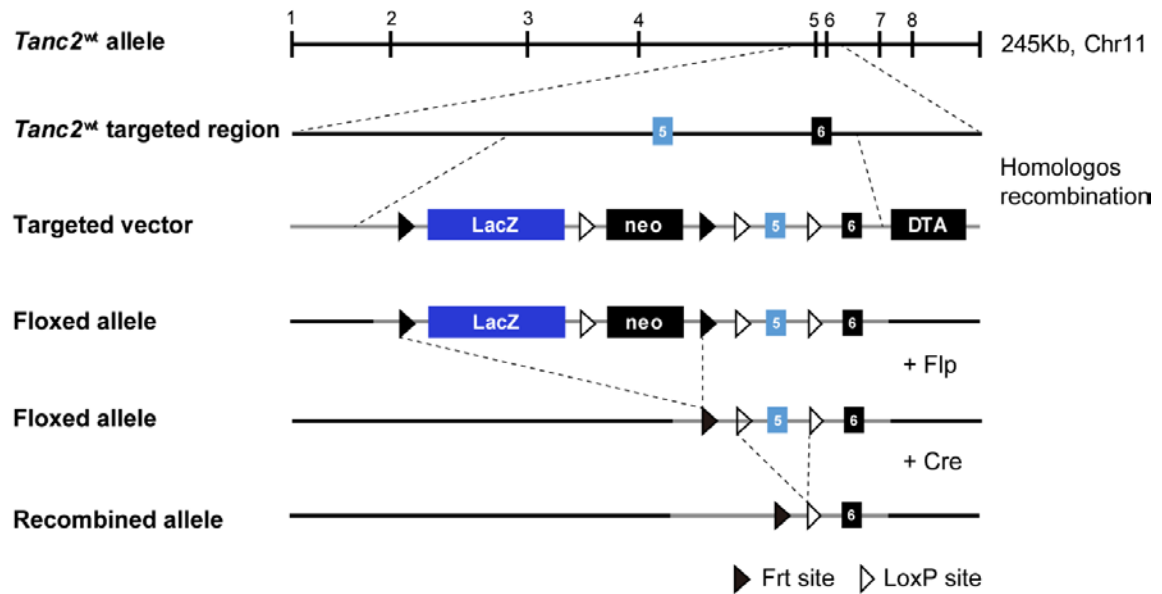
forced-swim test, as shown by immobility time. Data: minimal, maximal, median, 25%, and 75% values. (n = 14 mice [WT], and 10 mice [HT], ns, not significant, Student's t-test).

(e) Normal depression-like behavior of *Tanc2*^{+/-} mice (2–5 months; female) in the tail-suspension test, as shown by immobility time. Data: minimal, maximal, median, 25%, and 75% values. (n = 13 mice [WT], and 10 mice [HT], ns, not significant, Student's t-test).

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Supplementary Fig. 4

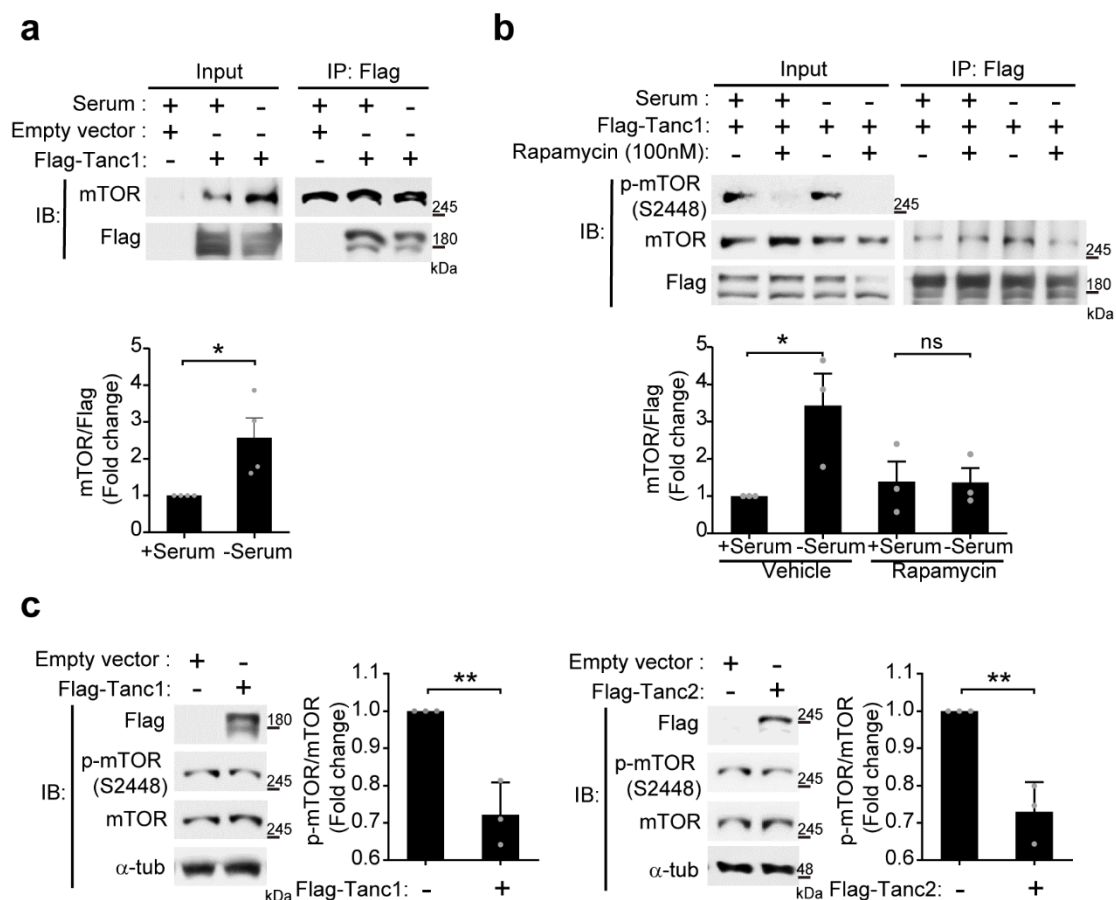
a



Supplementary Fig. 4. Conditional *Tanc2* KO strategy.

(a) Scheme for the generation of *Tanc2*^{fl/fl} mice carrying floxed exon 5 of the *Tanc2* gene. LacZ, β -galactosidase gene; neo, neomycin resistance cassette; DTA, diphtheria toxin gene; Flp, flippase; Cre, Cre recombinase; Frt site, site for flippase; LoxP site, cleavage site for cre recombinase action.

Supplementary Fig. 5



Supplementary Fig. 5. Tanc1 interacts with mTOR in a serum- and rapamycin-dependent manner and inhibits mTOR activity in heterologous Cells.

(a) Serum starvation (4 hours) promotes the interaction between Tanc1 and mTOR in heterologous cells, as demonstrated by coimmunoprecipitation. HEK293T cells expressing Flag-Tanc1 (human) were incubated with serum (or no serum; control) for 4 hours prior to coimmunoprecipitation and immunoblot experiments. mTOR signals were normalized to Tanc1 signals for quantification. Data: mean \pm SD. (n = 4 independent experiments, *P < 0.05, Student's t-test).

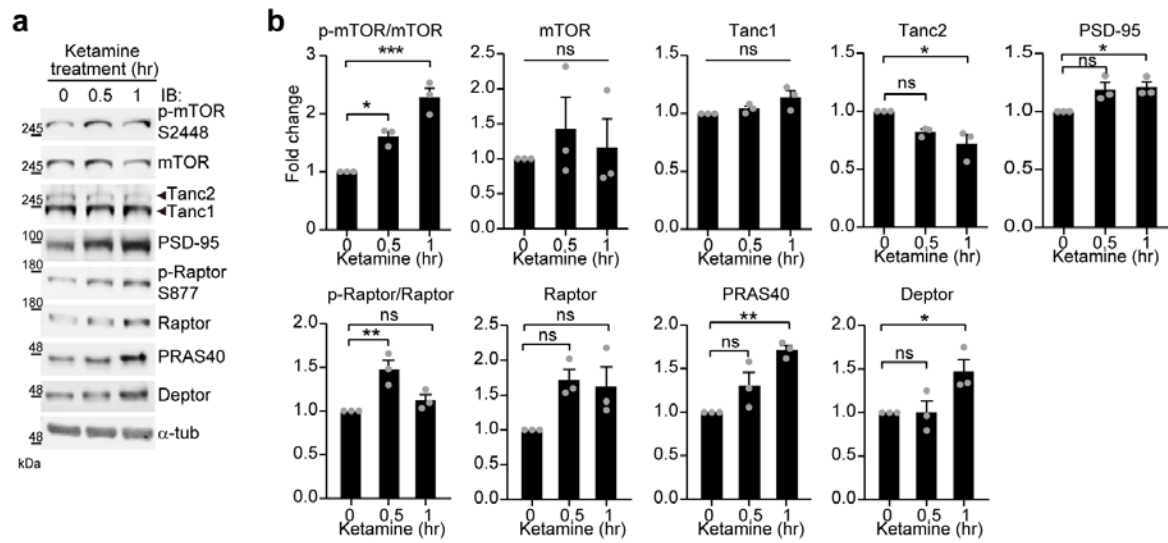
(b) Rapamycin inhibits serum starvation-induced increases in the interaction between Tanc1 and mTOR in heterologous cells. Flag-Tanc1 was expressed in HEK293T cells in the presence of serum. Pretreatment of cells with rapamycin for 2 hours was followed by serum starvation for 4 hours; cell lysates were subjected to coIP and immunoblot experiments. mTOR signals were normalized to Tanc1 signals for quantification. Data: mean \pm SD. (n = 3 independent experiments, ***P < 0.05, ns, not significant, Student's t-test).

(c) Overexpression of Tanc1 in HEK293T cells suppresses mTOR activity, as shown

by the ratio of total to phosphorylated (active) mTOR. Tanc2 (human) was also used as a control. Flag-tagged Tanc1/Tanc2 were overexpressed in HEK293T cells, followed by immunoblotting for total and phosphorylated (Ser-2448) mTOR. mTOR signals were normalized to α -tubulin signals. Data: mean \pm SD. (n = 3 independent experiments, ** $P < 0.01$, Student's t-test).

See Source Data 1 for raw data values and Supplementary Table 1 for statistical details.

Supplementary Fig. 6

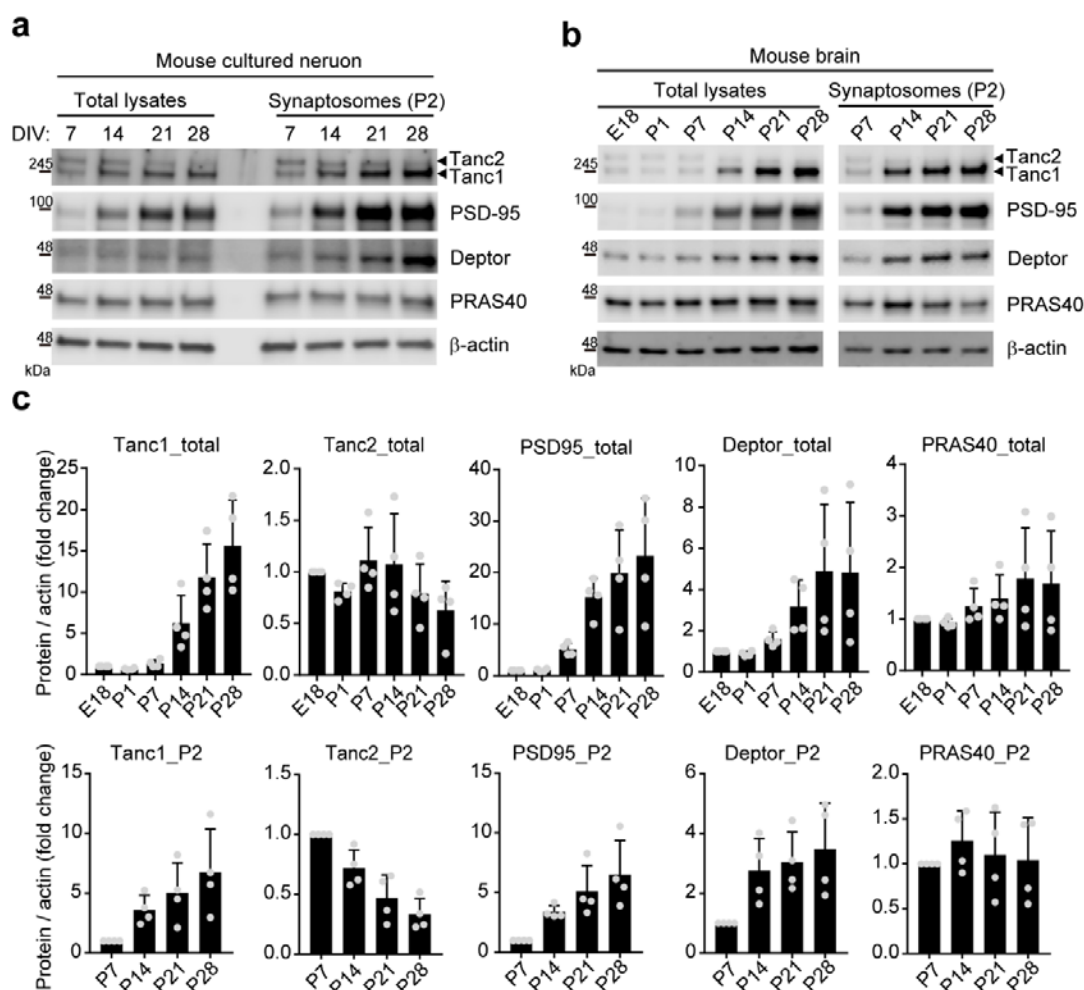


Supplementary Fig. 6. Ketamine induces changes in mTOR activity and synaptic levels of Tanc2-associated and mTORC proteins in the mouse brain.

(a and b) Ketamine induces mTOR activation in the mouse brain (P13–14; 10 mg/kg; i.p.), as shown by the time-dependent increases in the phosphorylation of mTOR and the mTORC1 protein Raptor in crude synaptosomes. Note that synaptic levels of other mTORC1/2 proteins (PRAS40 and Deptor) and PSD-95 also increased, whereas those of Tanc2 (but not Tanc1) were modestly decreased. Data: mean \pm S D. ($n = 3$ independent experiments, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, ns, not significant, one-way ANOVA with Bonferroni test).

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Supplementary Fig. 7



Supplementary Fig. 7. Temporal changes in total levels and synaptic enrichment of Tanc2, Deptor, and Tanc1 in cultured mouse hippocampal neurons and the mouse brain.

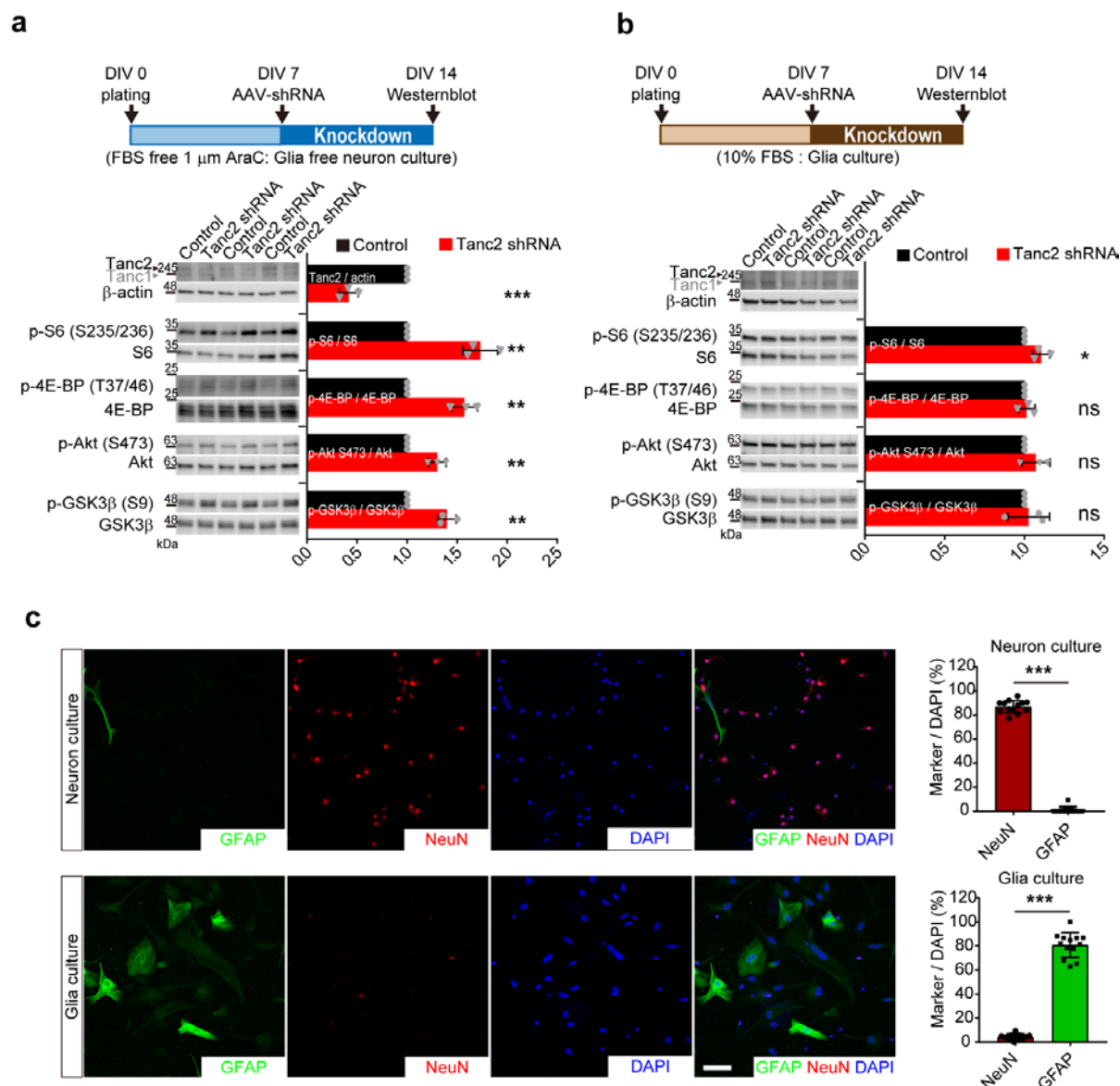
(a) Western blot analyses of whole lysates from cultured mouse hippocampal neurons show Tanc2 protein levels that are largely stable across developmental stages (DIV/days in vitro 7–28) and stronger relative to Tanc1 at early stages, while Deptor and Tanc1 protein levels gradually increase. Western blot analyses of crude synaptosomes from cultured mouse hippocampal neurons show largely unchanged synaptic levels of Tanc2 proteins across the developmental stages, which contrasts with the increasing synaptic levels of Deptor and Tanc1 proteins. Total and synaptic levels of PRAS40 (a component of mTORC1) were largely unchanged across developmental stages. PSD-95 was used as a positive control for stage-dependent increases in total expression and synaptic enrichment, and β -actin was used as a

loading control.

(b and c) Western blot analyses of whole lysates from the mouse brain (in vivo results) show Tanc2 protein levels that are largely stable across developmental stages (E/embryonic day 18–P/postnatal day 28) and higher relative to Tanc1 at early stages, while Deptor and Tanc1 protein levels gradually increase. Western blot analyses of crude synaptosomes from the mouse brain show decreasing synaptic levels of Tanc2 proteins across the developmental stages, which contrasts with the increasing synaptic levels of Deptor and Tanc1 proteins. Total and synaptic levels of PRAS40 were largely unchanged across developmental stages. PSD-95 was used as a positive control for stage-dependent increases in total expression and synaptic enrichment, and β -actin was used as a loading control. Data: mean \pm SD. (n = 4 independent experiments).

See Source Data 1 for raw data values and Supplementary Table 1 for statistical details.

Supplementary Fig. 8



Supplementary Fig. 8. Acute Tanc2 knockdown in cultured mouse neurons, but not glial cells, leads to mTORC1/2 hyperactivity.

(a) Acute knockdown of Tanc2 in cultured mouse neurons by infection with AAV-shRNA (DIV7–14) leads to increased phosphorylation levels of S6 (S235/236), 4E-BP (T37/46), Akt (S473), and GSK3 β (S9), indicative of mTORC1 and mTORC2 hyperactivity. Data: mean \pm SD. (n = 3 independent experiments, **P < 0.01, ***P < 0.001, Student's t-test).

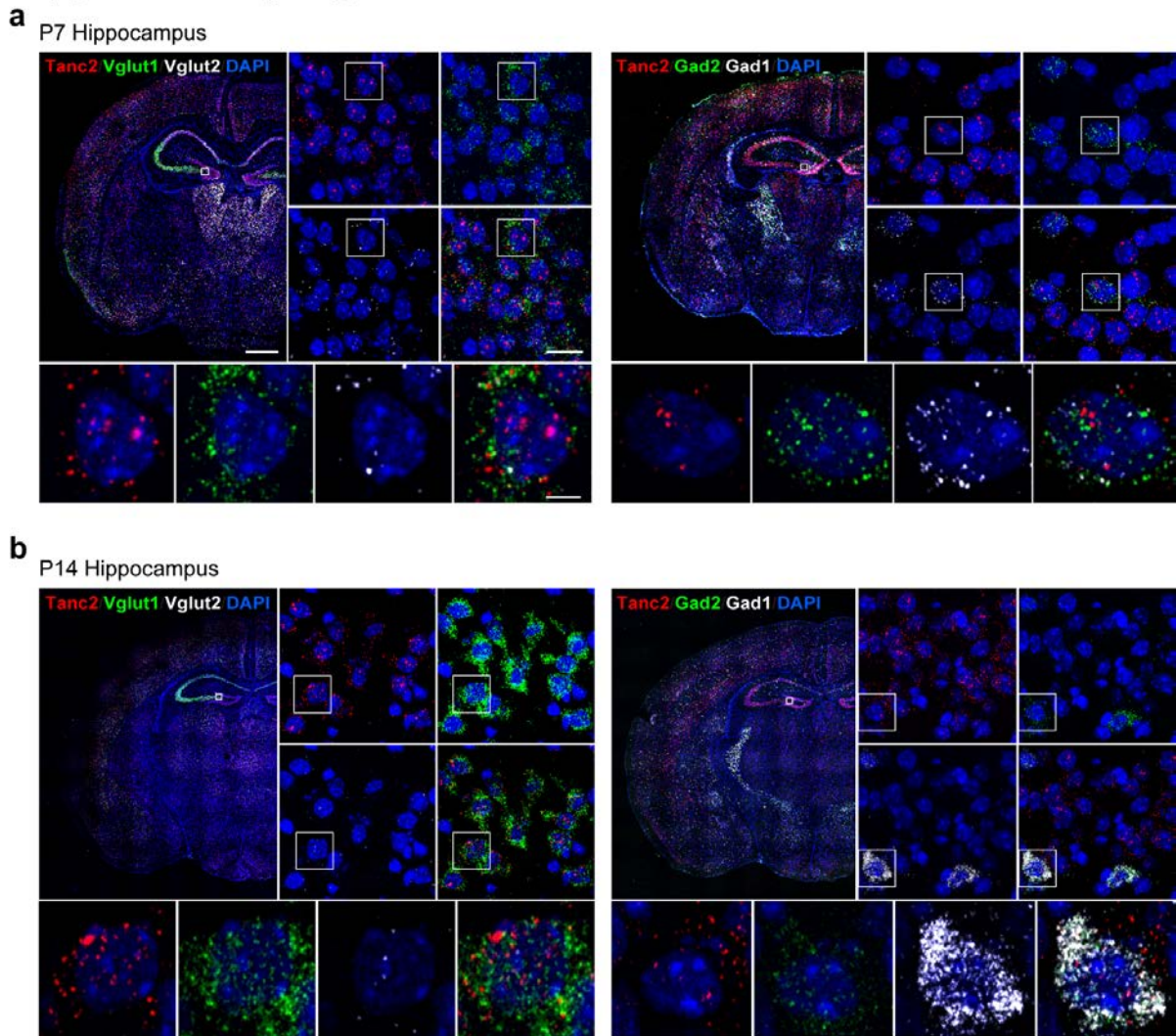
(b) Acute knockdown of Tanc2 in cultured mouse glial cells by infection with AAV-shRNA (DIV7–14) does not affect the phosphorylation levels of S6 (S235/236), 4E-

BP (T37/46), Akt (S473) or GSK3 β (S9), except for a small increase in p-S6, suggesting that Tanc2 mainly suppresses mTORC1/2 signaling in neurons rather than glial cells. Note that Tanc2 expression is much weaker relative to Tanc1. Data: mean \pm SD. (n = 3 independent experiments, *P < 0.05, ns, not significant, Student's t-test).

(c) Control co-immunofluorescence staining showing that cultured neurons are positive for NeuN (neuronal marker) but not for GFAP (astrocytic marker) and that cultured glia are positive for GFAP but not for NeuN. DAPI was used for nuclear staining. Scale bar, 50 μ m. Data: mean \pm SEM. (n = 13 images for neuron and glial cultures, ***P < 0.001, Student's t-test).

See Source Data 1 for raw data values and Supplementary Table 1 for statistical details.

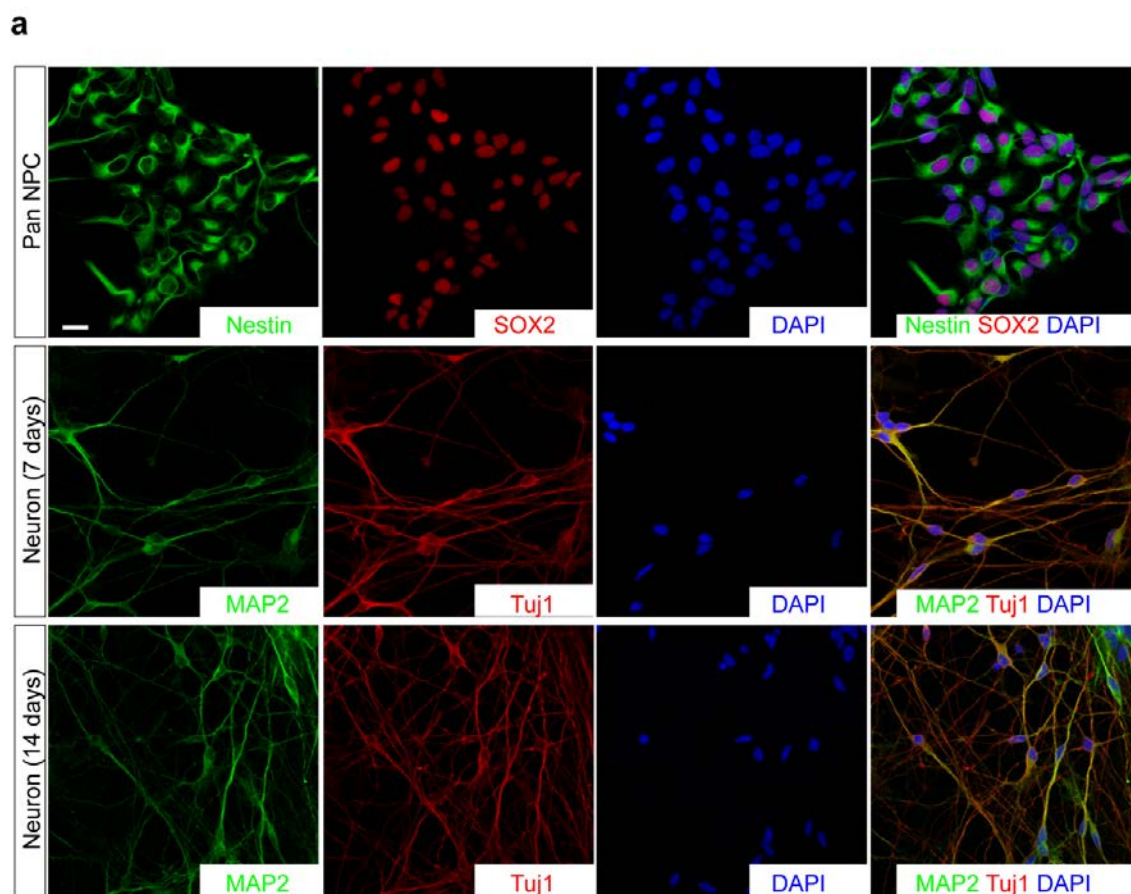
Supplementary Fig. 9



Supplementary Fig. 9. Tanc2 mRNA expression in both glutamatergic and GABAergic neurons in the mouse brain.

(a and b) Tanc2 expression is observed in both Vglut1/2-positive glutamatergic neurons and Gad1/2-positive GABAergic neurons in the cortical and hippocampal regions of the mouse brain at P7 and P14 (hippocampal examples are shown), as revealed by fluorescent in situ hybridization (FISH). Images in the bottom lines of each figure panel indicate enlarged single cells to clearly show the co-expression of Tanc2 with the indicated markers in the same cell. Scale bars, 500 μm (left), 20 μm (right), and 5 μm (bottom). Three independent experiments yielded similar results.

Supplementary Fig. 10



Supplementary Fig. 10. Characterization of human NPCs and neurons by immunofluorescence staining.

(a) Morphological characterization of pan-NPCs by immunofluorescence staining for Nestin and SOX2 (NPC markers) and pan-neurons at DIV7 and DIV14 by staining for MAP2 and Tuj1 (neuronal markers). Scale bar, 20 μ m. Three independent experiments yielded similar results.

						HT = 204.60 ± 16.17		p = 0.0001, ***																											
c	Light dark box test	3.4 months	WT = 18 HT = 12	Light zone (s)	WT = 138.50 ± 11.30 HT = 139.20 ± 14.56	Student's t-test	p = 0.9790, ns	(28) = 0.0266	Two-tailed																										
			WT = 14 HT = 10	Frequency of transition (count)	WT = 54.38 ± 3.35 HT = 55.83 ± 5.56	Student's t-test	p = 0.8151, ns	(28) = 0.2361	Two-tailed																										
			WT = 13 HT = 10	Immobility time (s)	WT = 107.1 ± 8.5 HT = 138.69 ± 11.13	Student's t-test	p = 0.1276, ns	(22) = 1.5840	Two-tailed																										
d	Forced swimming test		Immobility time (s)	WT = 107.1 ± 8.5 HT = 138.69 ± 11.13	Student's t-test	p = 0.3658, ns	(22) = 0.0180	Two-tailed																											
e	Tail suspension test		Immobility time (s)	WT = 107.1 ± 8.5 HT = 138.69 ± 11.13	Student's t-test	p = 0.3658, ns	(22) = 0.0180	Two-tailed																											
Supplement Fig 5	a	Coimmunoprecipitation (Western blotting)	n = 4	mTOR/Torc1 ratio in HEK293T cells (Fold change)	-siem + -siem 4 hr	1.00 ± 0.00 (SD) 2.57 ± 0.53 (SD)	Student's t-test	p = 0.0258, *	(8) = 2.945	Two-tailed																									
				n = 3	Vehicle	-siem -siem 4 hr	1.00 ± 0.00 (SD) 3.42 ± 0.85 (SD)	Student's t-test	p = 0.0487, *	(4) = 2.843	Two-tailed																								
						Rapamycin	-siem -siem 4 hr	1.36 ± 0.52 (SD) 1.35 ± 0.38 (SD)	Student's t-test	p = 0.9735, ns	(4) = 0.0353	Two-tailed																							
b	Western blotting	Torc1 overexpression (Empty vs Torc1)	p-mTOR/mTOR	Empty = 1.00 ± 0.00 (SD) Torc1 = 0.72 ± 0.05 (SD)	Student's t-test	p = 0.0051, **	(4) = 5.562	Two-tailed																											
				p-mTOR/mTOR	Empty = 1.00 ± 0.00 (SD) Torc2 = 0.72 ± 0.04 (SD)	Student's t-test	p = 0.0041, **	(4) = 5.507	Two-tailed																										
Supplement Fig 6	b	Western blotting (P2 fraction)	n=3	Protein levels normalized to tubulin (Fold change)	p-mTOR/mTOR	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.0130, *	F(2, 6) = 36.97, p = 0.0004																									
						0.5 hr	1.68 ± 0.25 (SD)																												
					mTOR	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.8546, ns	F(2, 6) = 0.3685, p = 0.7064																									
						0.5 hr	1.42 ± 0.45 (SD)																												
					Torc1	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.6933, ns	F(2, 6) = 3.735, p = 0.0884																									
						0.5 hr	1.04 ± 0.02 (SD)																												
					Torc2	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.9820, ns	F(2, 6) = 9.172, p = 0.0150																									
						0.5 hr	1.02 ± 0.6 (SD)																												
					PSD-95	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.0649, ns	F(2, 6) = 7.057, p = 0.0265																									
						0.5 hr	1.19 ± 0.59 (SD)																												
					p-Raptor/Raptor	0 hr	1.25 ± 0.45 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.0434, *	F(2, 6) = 12.54, p = 0.0072																									
						0.5 hr	1.47 ± 0.10 (SD)																												
					Raptor	0 hr	1.12 ± 0.06 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.5771, ns	F(2, 6) = 0.9666, p = 0.4326																									
						0.5 hr	1.17 ± 0.11 (SD)																												
					PRAS40	0 hr	1.42 ± 0.17 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.6489, ns	F(2, 6) = 14.92, p = 0.0047																									
						0.5 hr	1.30 ± 0.15 (SD)																												
					Deptor	0 hr	1.00 ± 0.00 (SD)	One-way ANOVA Bonferroni's multiple comparison test	p = 0.1072, ns	F(2, 6) = 5.93, p = 0.0379																									
						0.5 hr	1.71 ± 0.65 (SD)																												
Supplement Fig 7	c	Western blotting	n=3	Protein levels normalized to β-actin (Fold change)	Torc1_total	E18	1.00 ± 0.00 (SD)																												
						P1	0.68 ± 0.09 (SD)																												
					Torc2_total	P7	1.33 ± 0.49 (SD)																												
						P14	0.22 ± 0.34 (SD)																												
					PSD-95_total	P21	11.8 ± 4.05 (SD)																												
						P28	15.62 ± 5.54 (SD)																												
					Deptor_total	E18	1.00 ± 0.00 (SD)																												
						P1	0.81 ± 0.11 (SD)																												
					PRAS40_total	P7	1.11 ± 0.32 (SD)																												
						P14	1.07 ± 0.49 (SD)																												
					Torc1_P2	P21	0.78 ± 0.28 (SD)																												
						P28	0.62 ± 0.26 (SD)																												
					Torc2_P2	E18	1.00 ± 0.00 (SD)																												
						P1	1.1 ± 0.22 (SD)																												
					PSD-95_P2	P7	5.10 ± 1.16 (SD)																												
						P14	16.2 ± 3.72 (SD)																												
					Deptor_P2	P21	19.83 ± 8.48 (SD)																												
						P28	23.25 ± 11.21 (SD)																												
PRAS40_P2	E18	1.00 ± 0.00 (SD)																																	
	P1	0.92 ± 0.08 (SD)																																	
Torc1_P1	P7	1.25 ± 0.33 (SD)																																	
	P14	1.39 ± 0.48 (SD)																																	
Torc2_P1	P21	1.75 ± 0.58 (SD)																																	
	P28	1.68 ± 1.01 (SD)																																	
PSD-95_P1	P7	1.00 ± 0.00 (SD)																																	
	P14	3.90 ± 1.22 (SD)																																	
Deptor_P1	P21	5.00 ± 2.50 (SD)																																	
	P28	6.75 ± 3.59 (SD)																																	
PRAS40_P1	P7	1.00 ± 0.00 (SD)																																	
	P14	0.71 ± 0.19 (SD)																																	
Torc1_P2	P21	0.46 ± 0.10 (SD)																																	
	P28	0.33 ± 0.13 (SD)																																	
Torc2_P2	P7	1.00 ± 0.00 (SD)																																	
	P14	3.38 ± 0.93 (SD)																																	
PSD-95_P2	P21	5.08 ± 2.17 (SD)																																	
	P28	6.51 ± 2.84 (SD)																																	
Deptor_P2	P7	1.00 ± 0.00 (SD)																																	
	P14	2.75 ± 1.07 (SD)																																	
PRAS40_P2	P21	3.03 ± 1.00 (SD)																																	
	P28	3.48 ± 1.52 (SD)																																	
Supplement Fig 8	a	Western blotting	Control vs Torc2 shRNA	n=3	Protein levels normalized to β-actin (Fold change)	Torc2/actin	Control	1.00 ± 0.00 (SD)	Student's t-test	p = 0.0004, ***	(4) = 10.66																								
						Torc2 shRNA	0.41 ± 0.08 (SD)																												
						p-S6/S6	Control	1.00 ± 0.00 (SD)				Student's t-test	p = 0.0019, **	(4) = 7.27																					
						Torc2 shRNA	1.73 ± 0.17 (SD)																												
						p-4E-BP4E-BP	Control	1.00 ± 0.00 (SD)							Student's t-test	p = 0.0018, **	(4) = 7.4																		
						Torc2 shRNA	1.57 ± 0.12 (SD)																												
						p-Akt/Akt	Control	1.00 ± 0.00 (SD)										Student's t-test	p = 0.0045, **	(4) = 5.761															
						Torc2 shRNA	1.30 ± 0.09 (SD)																												
						p-GSK3/GSK3β	Control	1.00 ± 0.00 (SD)													Student's t-test	p = 0.0017, **	(4) = 7.456												
						Torc2 shRNA	1.39 ± 0.19 (SD)																												
						p-S6/S6	Control	1.00 ± 0.00 (SD)																Student's t-test	p = 0.0273, *	(4) = 3.397									
						Torc2 shRNA	1.10 ± 0.05 (SD)																												
						p-4E-BP4E-BP	Control	1.00 ± 0.00 (SD)																			Student's t-test	p = 0.6689, ns	(4) = 0.4607						
						Torc2 shRNA	1.01 ± 0.05 (SD)																												
						p-Akt/Akt	Control	1.00 ± 0.00 (SD)																						Student's t-test	p = 0.2364, ns	(4) = 1.392			
						Torc2 shRNA	1.07 ± 0.08 (SD)																												
						p-GSK3/GSK3β	Control	1.00 ± 0.00 (SD)																									Student's t-test	p = 0.7421, ns	(4) = 0.3528
						Torc2 shRNA	1.02 ± 0.13 (SD)																												
b	Immunocytochemistry	NeuN vs GFAP	n=13	Mikar / DAPI (%)	Neuron culture	NeuN	86.81 ± 5.09 (SD)	Student's t-test	p < 0.0001, ***	(24) = 54.13																									
					Glia culture	GFAP	1.03 ± 2.6 (SD)																												
					Glia culture	NeuN	4.01 ± 2.4 (SD)																												
c	Immunocytochemistry	NeuN vs GFAP	n=13	Mikar / DAPI (%)	Neuron culture	GFAP	80.77 ± 10.34 (SD)	Student's t-test	p < 0.0001, ***	(24) = 26.06																									
					Glia culture	NeuN	4.01 ± 2.4 (SD)																												
					Glia culture	GFAP	80.77 ± 10.34 (SD)																												

Supplementary table 2. Primers used in this study

Mouse line	Primer name	Sequence (5'-)
<i>Tanc2</i> ^{+/-}	<i>Tanc2</i> ^{+/-} -F	GGTTAGATAGGAAATAACCATCA
	WT-B	ATGTGGCATATAAAGTTATACCTC
	<i>Tanc2</i> ^{+/-} -B	TCCCCACAACGGGTTCTTCTG
<i>Tanc2</i> ^{fl/fl}	<i>Tanc2</i> -fl-F	CTGTGCATGAAAGCTTCTGG
	<i>Tanc2</i> -fl-B	TCTCCACCTCCCTGGTACTG

