Supplementary Information for

Neutralizing antibody responses to SARS-CoV-2 in symptomatic COVID-19 is persistent and critical for survival.

by Stefania Dispinseri et al.

Supplementary Figures 1 to 10

- Figure 1: Schematic representation of transfer vectors and plasmids.
- Figure 2: Gating strategy used for FACS analysis to detect ACE expression on VEROE6 cells.
- Figure 3: Correlation of anti-SARS-CoV2 spike neutralizing and RBD, S1+S2 antibodies during follow-up of the COVID-19 patients.
- Figure 4: Development and kinetics of SARS-CoV-2 antibody responses of non-hospitalized COVID-19 patients.
- Figure 5: Univariate Hazard Ratios (HR) for time to a negative SARS-CoV-2 viral RNA RT-PCR of the nasopharyngeal swab and to death of 134 hospitalized COVID-19 patients.
- Figure 6: Bivariate Hazard Ratios (HR) for time to death of COVID-19 patients.
- Figure 7: HCoV-HKU1 and -OC43 S2 IgG correlation with neutralization is temporary.
- Figure 8: Variations of the SARS-CoV-2 neutralizing titers between the 2nd and 3rd out-patient visit.
- Figure 9: Gating strategy used for FACS analysis to detect SARS-CoV-2 spike.
- Figure 10. Schematic representation of plasmids used for the LIPS.

Supplementary Tables 1 to 5

- Table 1: Laboratory values at first Biobank blood sampling of the COVID-19 study population.
- Table 2: SARS-CoV-2 Antibody responses according to week from symptoms onset. (provided as xls file)
- Table 3: Characteristics of non-hospitalized COVID-19 patients.
- Table 4: Characteristics of COVID-19 patients grouped according to their SARS-CoV-2 neutralizing antibody response. (provided as xls file)
- Table 5: COVID-BioB study team and collaborators.

Supplementary Figure 1



Footnote to Supplementary Figure 1: Schematic representation of transfer vectors and plasmids. (a) the lentiviral transfer vector plasmid expressing luciferase reporter gene under the control of CMV promoter (pGAE-LucW); (b) the pseudotyping plasmids expressing full-length Spike (pSpike), cytoplasmic tail truncated Spike (pSpike-C3) and VSV.G envelope (phCMV-VSV.G); (c) the packaging plasmid providing the proteins for producing the vector particles (pADSIV3+). The packaging signal (ψ), the primer binding site (PBS), the deleted packaging signal ($\Delta\psi$), the major splice donor (SD), the bovine growth hormone polyadenylation signal (polyA) and the central polypurine tract (cPPT) are indicated.



Footnote to Supplementary Figure 2: Gating strategy used for FACS analysis to detect ACE expression on cells. The first gate (G1) was set to include the singlets. Forward (FSC) and side scatter (SSC) gating (G2) was used to identify the cells of interest. The identification of the ACE2 positive cells was performed by using a primary mouse anti-human ACE2 antibody (Millipore, Catalog Number: MAB5676) followed by a secondary goat anti-mouse IgG-PE (SoutherBiotech; Catalog Number: 1030-09). Staining with secondary antibody only (G3) was used as negative control to set the gate of negative cells (G3) and quantify the percentage of positive cells expressing ACE2 (G4).



Footnote to Supplementary Figure 3: Correlation of anti-SARS-CoV2 spike neutralizing and RBD, S1+S2 antibodies during follow-up of the COVID-19 patients. Sera of 162 COVID-19 patients, collected at the indicated timepoints from symptoms onset, were assessed by LV-based neutralization assay (ID50) and the LIPS assay to the various antigens, as indicated in the panels. (a) Correlation matrix of the indicated variables at week 3-4. For each pair, the Pearson's correlation coefficient is shown as number and on a color scale. Statistically non-significant correlations are crossed. (b) Sera of COVID-19 patients, collected at the indicated timepoints from symptoms onset, were assessed by LV-based neutralization assay (ID50) and the LIPS assay, indicated in grey labels above each row/column. Boxes under the diagonal show each correlation plot of the ID50 reciprocal and arbitrary units after log10 conversion. Dots correspond to individual measurements; the black line represents the regression line and the grey area its 95%CI. Boxes on the diagonal show as histograms the distribution of values in each assay. Boxes above the diagonal show the corresponding Pearson correlation analysis coefficients. Asterisks correspond to the following p values: *** $p \le 0.001$; ** $p \le 0.01$; * $p \le 0.05$. Source data are provided as a Source Data file.





Footnote to Supplementary Figure 4: Development and kinetics of SARS-CoV-2 antibody responses of non-hospitalized COVID-19 patients. Each colored dot corresponds to the ID50 reciprocal of a given infected individual's serum of 26 patients. Shown is the moving average of data + SE (black curve line + grey band) as obtained by a LOESS curve fitting polynomial regression. Sampling occurred during hospital attendance (ER or ward), and at follow-up visits post discharge; colors define the number of the visits attended. In Table 1 the serum sample availability of the non-hospitalized COVID-19 patients is described. The clinical characteristics are described in Supplementary Table 3. Source data are provided as a Source Data file.



Footnote to Supplementary Figure 5: Univariate Hazard Ratios (HR) for time to a negative SARS-CoV-2 viral RNA RT-PCR of the nasopharyngeal swab and to death of 134 hospitalized COVID-19 patients. Forest plots of Hazard Ratios obtained by univariable Cox regression analysis of the shown variables at the time of the in-hospital serum sampling. The analysis was adjusted for sex and age. Dots represent the HR, filled dots stand for p<0.05 (two-sided). Wald statistics were used for comparison. Abbreviations are: Body Mass Index (BMI), Coronary Artery Diseases (CAD), Chronic obstructive pulmonary disease (COPD), Chronic Kidney Disease (CKD), Neurodegenerative disease (ND), White Blood cells (WBC), Lactate dehydrogenase (LDH), C-reactive protein (CRP). Antibody binding are to SARS-CoV-2, when not otherwise specified. Antibody binding is expressed in arbitrary units (AU). Source data are provided as a Source Data file.



Footnote to Supplementary Figure 6: Bivariate Hazard Ratios (HR) for time to death of COVID-19 patients. Forest plot of Hazard Ratios (HR) with lower and upper limit of the 95%CI for neutralizing antibodies at the time of in-hospital serum sampling and survival in COVID-19 patients calculated with multivariable Cox regression analysis. The analysis used a neutralization negative score, corrected for age and sex, and the shown variables measured at the time of in-hospital serum sampling. Dots represent the HR, filled dots stand for p<0.05 (two-sided). Wald statistics were used for comparison. Abbreviations are: Body Mass Index (BMI), Coronary Artery Diseases (CAD), Chronic obstructive pulmonary disease (COPD), Chronic Kidney Disease (CKD), Neurodegenerative disease (ND), Lactate dehydrogenase (LDH), C-reactive protein (CRP). Antibody binding (IgG, IgM and IgA) are to SARS-CoV-2, and expressed in arbitrary units (AU). Source data are provided as a Source Data file.



Footnote to Supplementary Figure 7: HCoV-HKU1 and -OC43 S2 IgG correlation with neutralization is temporary. Sera of COVID-19 patients, collected at the indicated timepoints from symptoms onset, were measured by LV-based neutralization assay and the LIPS assay indicated in grey labels above each row/column. Boxes under the diagonal show each correlation plot of the ID50 reciprocal and arbitrary units after log10 conversion. Dots correspond to individual measurements, the black line represents the regression line and the grey area its 95%CI. Boxes on the diagonal show as histograms the distribution of values in each assay. Boxes above the diagonal show the corresponding Pearson correlation analysis coefficients. Asterisks correspond to the following p values: *** $p \le 0.001$; ** $p \le 0.01$; ** $p \le 0.05$. Source data are provided as a Source Data file.









Range (min-max)	0,3-98,13	1,29-441,92	
Standard deviation	28,01	170,41	
No. with variation >30%	22	6	

Footnote to Supplementary Figure 8: Variations of the SARS-CoV-2 neutralizing titers between the 2nd and 3rd out-patient visit. (a) the inhibitory serum dilution (ID50) at the 2nd and 3rd visit (dark and light grey bars, respectively), and (b) the percent variation of the ID50 of each of the 43 patients tested for nAbs. The 3rd visit occurred between September 22nd and November 6th 2020, when the Rt in the same geographical area of the San Raffaele Hospital, the Lombardy region, ramped-up from 0.86 (CI 0.73-1.01) at the end of September to 1.61 (CI 1.08-2.33) in the first week of November. (http://www.salute.gov.it/portale/nuovocoronavirus/dettaglioNotizieNuovoCoronavirus.jsp?lingua=italia no&menu=notizie&p=dalministero&id=5093). Source data are provided as a Source Data file.



Footnote to Supplementary Figure 9: Gating strategy used for FACS analysis to detect SARS-CoV-2 spike. Forward (FSC) and side scatter (SSC) gating (G1) was used to identify the cells of interest, while removing debris. Lenti-X cells transfected with pSpike were gated and analyzed for Spike expression using a primary rabbit-anti-Spike S2 antibody (Sino Biological, Catalog Number: 40590-T62) followed by a secondary PE donkey anti-rabbit antibody (Biolegend, Catalog Number: 406421). Staining with secondary antibody only was used as negative control (CTR-, in panels a) to set the quadrant and quantify the percentage of Spike-expressing cells (in panels b).



Footnote to Supplementary Figure 10. Schematic representation of plasmids used for the LIPS. The recombinant nanoluciferase (sNanoLuc) tagged antigens used in this study are shown. Serial Cloner 2.6.1 for virtual design of recombinant antigens plasmid clones. See Methods for details on construction.

Supplementary Tables

Supplementary Table 1: Laboratory values at first Biobank blood sampling of the COVID-19 study population (No. of patients = 162).

	COVID-19 patients	Missing
		data
Median days (95%CI) from symptoms onset to first sampling for	11.5 (7-18)	0
Biobank		
Laboratory values at time of first blood sampling (Median, 95%CI):		
- White Blood Cells, x10 ⁹ /L	6.9 (5.2-9.8)	17
- Lymphocyte (Ly) count, x10 ⁹ /L	1.1 (0.7-1.5)	25
- Neutrophil (N) count, x10 ⁹ /L	4.8 (3.5-7.5)	25
- Monocyte count, x10 ⁹ /L	0.5 (0.4-0.7)	25
- N/Ly ratio	4.67 (2.55-8.75)	25
- Haemoglobin, g/dL	13.1 (11.6-14.6)	17
- Platelet count, x10 ⁹ /L	236 (182-323)	17
- Bilirubin total, mg/dL	0.56 (0.37-0.86)	45
- ALT, U/L	41 (23.5-66.5)	35
- AST, U/L	44 (31-60)	34
- Creatinine, mg/dL	0.97 (0.77-1.34)	23
- LDH, U/L	355 (273-428)	39
- CRP, mg/L	54.6 (18.75-118.1)	19
- D-Dimer, μg/mL	1.11 (0.53-2.45)	104
- IL-6, pg/mL	33 (16.7-81)	117
- Ferritin, ng/mL	1222 (597-1701)	103

Footnote to Supplementary Table 1: ALT: Alanine Amino Transferase; AST: Aspartate Amino Transaminase; LDH: Lactate dehydrogenase; CRP: C-reactive protein.

Supplementary Table 2: SARS-CoV-2 Antibody responses according to week from symptoms onset.

		N	/eeks after sympt	oms onset		
	1-2	3-4	5-8	9-16	17-36	Overall
Samples tested (No.)	101	43	76	76	66	362
Neutralization of SARS-CoV-2						
Mean (SD)	3500 (8600)	45300 (252000)	9950 (24400)	2160 (3910)	1460 (1910)	9190 (87900)
Median	238	2640	3500	869	660	1080
[Min, Max]	[10, 61500]	[10, 1660000]	[10, 161000]	[10, 28000]	[10, 10900]	[10, 1660000]
Antibody binding to SARS-CoV-2						
lgG_RBD						
Mean (SD)	13.0 (29.4)	165 (296)	2670 (5710)	4360 (7570)	6100 (6640)	2610 (5660)
Median	1,4	43	1060	2010	3830	333
[Min, Max]	[0.00296, 163]	[0.00859, 1130]	[0.116, 46500]	[0.0453, 36800]	[0.0171, 31500]	[0.00296, 46500]
IgG_S1S2		4070 (4040)	0050 (0000)	0000 (0000)	7040 (0070)	F000 (7F00)
Mean (SD)	37.5 (50.6)	1370 (4040)	8850 (8960)	8080 (8660)	7310 (6970)	5060 (7580)
	0,00	02,3 [0.370, 33400]	0420	5350	0000	1000
	[0.0228, 258]	[0.379, 23400]	[0.160, 46900]	[0.05, 33400]	[0.112, 33400]	[0.0226, 46900]
IgG_52 Mean (SD)	349 (1760)	818 (3030)	1550 (4050)	1640 (1670)	3550 (3930)	1510 (3140)
Median	7 81	159	429	1040 (1070)	2740	312
[Min_Max]	[0 00373 16000]	[0 00885 18800]	10 0289 310001	[0 0159 6400]	[0 0169 25600]	[0 00373 31000]
	[0.00010, 10000]	[0.00000, 10000]	[0.0200, 01000]	[0.0100, 0100]	[0.0100, 20000]	[0.00010, 01000]
Mean (SD)	20.3 (19.7)	31.0 (19.8)	49.3 (18.9)	NA	NA	28.3 (22.4)
Median	12,4	35,7	53,1			29,4
[Min, Max]	[0.237, 65.9]	0.313, 65.6]	[0.443, 75.4]			[0.237, 75.4]
missing	1 (1%)	4 (9.3%)	43 (56.6%)	100%	100%	190 (52.5%)
Antibody binding to other Viruses						
IgG_OC43 S2						
Mean (SD)	1430 (2040)	2310 (2370)	3490 (2430)	2610 (2070)	1630 (1190)	2290 (2220)
Median	492	1220	3410	2350	1320	1440
[Min, Max]	[10.2, 10200]	[0.307, 8410]	[60.1, 8780]	[29.8, 8340]	[57.6, 4420]	[0.307, 10200]
missing	0%	0%	0%	0%	18 (27.3%)	18 (5.0%)
IgG_HKU1 S2						
Mean (SD)	1760 (2550)	3600 (3030)	4650 (4060)	3580 (3020)	2120 (1780)	3080 (3210)
Median	587	3410	3240	2910	1590	1930
[Min, Max]	[21.4, 13500]	[92.8, 10500]	[56.8, 15100]	[2.73, 12800]	[108, 7310]	[2.73, 15100]
missing	0%	0%	0%	0%	18 (27.3%)	18 (5.0%)
IgG_FLU HA	40000 (05000)	40500 (00000)	40000 (45400)	N14	N14	40000 (00000)
Mean (SD)	78600 (25600)	12500 (20900)	13300 (15100)	NA	NA	16200 (23000)
	7040	4750	6940			6890
	[72.6, 106000]	[172, 117000] 4 (0.3%)	[311, 56200] 43 (56.6%)	100%	100%	[72.0, 117000] 100 (52.5%)
Initial Initian Initia	1 (176)	4 (9.3 %)	43 (30.0%)	100 /6	100 /6	190 (32.3 %)
Mean (SD)	9 34 (15 4)	11 0 (17 7)	14 0 (23 3)	NΔ	NΔ	10.6 (17.6)
Median	2 82	3 52	2 97			3 12
[Min_Max]	[0 328 92 5]	[0 159 90 8]	[0 377 91 4]			[0 159 92 5]
missing	1 (1%)	4 (9.3%)	44 (57.9%)	100%	100%	191 (52.8%)
Days post symptoms			(
Median [Min, Max]	8.00 [1, 14]	18.0 [15, 28]	39.0 [30, 54]	95.0 [57, 112]	204 [114, 250]	39.0 [1, 250]
Age years						
Median [Min, Max]	63.0 [34, 94]	63.0 [34, 88]	61.5 [19, 87]	59.5 [26, 87]	61.0 [37, 87]	62.0 [19, 94]
Sex Male (No.)	67 (66.3%)	31 (72.1%)	48 (63.2%)	49 (64.5%)	41 (62.1%)	236 (65.2%)
Sample Category No.	•				,	,
In-hospital visit	101 (100%)	39 (90.7%)	10 (13.2%)	0 (0%)	0 (0%)	150 (41.4%)
1st follow-up visit	0 (0%)	4 (9.3%)	66 (86.8%)	17 (22.4%)	0 (0%)	87 (24.0%)
2nd follow-up visit	0 (0%)	0 (0%)	0 (0%)	59 (77.6%)	18 (27.3%)	77 (21.3%)
3rd follow-up visit	0 (0%)	0 (0%)	0 (0%)	0 (0%)	43 (65.2%)	43 (11.9%)
4th follow-up visit	0 (0%)	0 (0%)	0 (0%)	0 (0%)	4 (6.1%)	4 (1.1%)
5th follow-up visit	0 (0%)	0 (0%)	0 (0%)	0 (0%)	1 (1.5%)	1 (0.3%)

Footnote to Supplementary Table 2: No. = number. Neutralization is expressed as the inverse of the serum dilution at which the ID50 was obtained in the LV-based SARS-CoV-2 neutralization assay. A value of 10 was ascribed to the serum that displayed absence of neutralization at the first dilution used (1/40) for the assay. Antibody binding is expressed as arbitrary units. Sample category shows the number and percent of patient's sera tested at each visit according to time (in week intervals) from symptoms onset as depicted in Figure 2. NA= not applicable.

	Non-hospitalized COVID-19 patients (N=26)
Age, years	49.5 (46-62)
Sex Male	38.5%
Ethnicity:	
 Caucasian 	84.6%
 Hispanic 	15.4%
 Asian 	0
 ○ African 	0
Co-morbidities:	
	11 5%
	7 7%
	7.7%
\circ COPD	0%
	3.8%
• Cancer	0%
	0%
Number of comercialities	070
	00.0%
	80.8%
	11.5%
0 2	3.8%
0 3	3.8%
04	0
Body Mass Index:	
o <25	31.8%
o 25-30	40.9%
o >30	27.3%
Median days (95%CI) from symptoms onset:	
- to admission at Emergency Department	5.5 (3-10)
- to first blood sampling for Biobank	21.5 (5-37)
Symptoms at disease onset:	
- general	
\sim fever	68%
	36%
 fatique/malaise 	60%
 mvalgie/malaise mvalgie/arthralgia 	36%
- respiratory	0070
\circ cough	60%
	28%
 ayophed sore throat 	20%
\circ chest pain	24%
- astrointestinal	21/0
o diarrhea	40%
	12%
\circ abdominal pain	12%
- others	,.
o conjunctivitis	12%
o hypo/anosmia	28%
o hypo/dysgeusia	40%
\circ skin rash	1.3%
Median days from symptoms to negative RT-PCR swab (95%CI)	33 (31-35)
Median follow up days (95%CI)	97 (89-104)

Supplementary Table 3: Characteristics of non-hospitalized COVID-19 patients.

Footnote to Supplementary Table 3: abbreviations are as in Table 2.

Supplementary Table 4: Characteristics of COVID-19 patients grouped according to their SARS-CoV-2 neutralizing antibody response.

Characteristics	SARS-CoV-2 neutralizing Ab score			Statistics	
	Negative (N=43 patients)	M	Positive (N=107 patients)	м	p Value
Median age, years (95%CI)	74 (59-79)	0	62 (51-70)	0	0,003
Co-morbidities:	· · · ·		· · · ·		
 Hypertension 	26 (60.5%)		45 (42.1%)		0.048
• CAD	8 (18.6%)		13 (12.1%)		0,308
 Diabetes 	10 (23.3%)	0	30 (28%)	0	0,684
 COPD 	5 (11.6%)	0	1 (0.9%)	0	0,008
 CKD 	15 (34.9%)		9 (8.4%)		<0.001
 Cancer 	10 (23.3%)		7 (6.5%)		0,008
 Neurodegenerative disease 	3 (7%)		1 (1.9%)		0,142
Number of co-morbidities					
 None 	10 (23.3%)		48 (44.9%)		
o 1	6 (14%)	0	22 (20.6%)	0	0.005
o 2	14 (32.6%)		28 (26.2%)		-,
o 3	9 (20.9%)		7 (6.5%)		
o 4	4 (9.3%)		2 (1.9%)		
Body Mass Index					0,012
<25	15 (44.1%)	۹	19 (18.6%)	5	
25-30	11 (32.4%)	0	45 (44.1%)	0	
>30	8 (23.5%)		38 (37.3%)		
Symptoms at disease onset:		1		4	
- general					
o fever	71,40%		91,30%		0,004
 headache 	21,40%		25,20%		0,541
 fatigue/malaise 	28,60%		67%		<0.001
 myalgia/arthralgia 	26,20%		35,90%		0,331
- respiratory					
∘ cough	35,70%		71,80%		<0.001
o dyspnea	47,60%		79,60%		<0.001
 sore throat 	11,90%		17,50%		0,464
o cnest pain	14,30%		26,20%		0,133
- gastrointestinai	16 70%		21 109/		0.000
	9.50%		14 60%		0,099
	11 90%		7 80%		0,503
- others	11,0070		1,0070		0,020
 conjunctivitis 	4.80%		18.40%		0.038
o hypo/anosmia	16,70%		46,60%		0,001
 hypo/dysgeusia 	21,50%		50,50%		0,002
○ skin rash	7,10%		3,90%		0,413
Median days (95%CI) from symptoms onset:					
- to admission	5 (3-12)	0	10 (7-12)	0	<0.001
- to blood sampling for Biobank	6 (4-12)	0	13 (9-16)	0	<0.001
Admitted to the bosnital	43/45 (95 5%)		107/117 (91.4%)		
	5/43 (11.6%)		11/107 (10.3%)		
- Hospitalized	5/45 (11.070)		11/10/ (10.5%)		
o ≤7davs	6/43 (14%)		19/107 (17.8%)		
o >7days	16/43 (37.2%)	0	49/107 (45.8%)	0	
 deceased 	15/43 (34.9%)	0	3/107 (2.8%)	0	<0.001
 in need of ICU, recovered 	1/43 (2.3%)		14/107 (13.1%)		
 in need of ICU, deceased 	0/43 (0%)		11/107 (7.3%)		
Median days of hospital stay for 134 patients	8 (4-24)		13 (6-21)		0.317
(95%CI) Median days from symptoms to pagetive DT	- ()				-,
PCR swab (95%CI)	46 (38-54)	2	40 (37-43)	5	0,041
Median follow up days (95%Cl)	194 (99-289)	0	203 (198-208)	0	0.521
Lehersten volues at time of first blood compliant	101 (00 200)	Ũ	200 (100 200)	Ū	0,021
Laboratory values at time of hist blood sampling:		-		40	0.000
- White Blood Cells, x10 [°] /L	6.2 (4.7-8.1)	5	7.2 (5.5-10.3)	12	0,028
- Lympnocyte (Ly) count, x10 ⁻ /L	U.90 (U.5-1.5)	9	1.∠ (U.8-1.5)	10	0,000
- Neutrophil (N) count, XTU /L Monopute count, X10 ⁹ //	4.3 (∠.0-C0./3)	9	4.9 (3./-/.8) 0 ∈ (0 /_0 ∘)	16	0,000
- N/Ly ratio	0.4 (0.3-0.0) 4 2 (2 AG_R 25)	9	0.0 (0.4-0.0) 4 92 (2 55-0 04)	16	0,009
- Haemoglobin g/dl	11 95 (9 72-14 05)	5	13.5 (12.1-14.8)	12	0,000
- Platelet count x10 ⁹ /l	187 (109-234)	5	256 (199-355)	12	<0.001
- Bilirubin total, mg/dL	0.44 (0.36-0.72)	13	0.65 (0.38-0.98)	32	0,053
- ALT, U/L	30 (18.5-34.5)	12	47 (26-75)	23	< 0.001
- AST, U/L	36 (26-53)	12	46 (33-64)	22	0,045
- Creatinine, mg/dL	1.22 (0.87-1.81)	7	0.95 (0.76-1.24)	16	0,002
- LDH, U/L	282 (232-409)	12	368.5 (294-441)	27	0,007
- CRP, mg/L	46.4 (20.4-99)	7	62.6 (18.1-128.1)	12	0,417

Footnote to Supplementary Table 4: The median BMI is 25.9 (IQR 23-29.7) for the neutralizing antibody negative group and 28.3 (IQR 25.4-32.3) for the neutralizing antibody positive group of patients (p= 0.018). Chi-square or Fischer's exact test were used to compare categorical variables. Wilcoxon rank sum test was used to compare continuous variables. A two-sided P Value was reported. Abbreviations used are: M = missing data. Intensive Care Unit (ICU) Coronary Artery Diseases (CAD), Chronic obstructive pulmonary disease (COPD), Chronic Kidney Disease (CKD), Neurodegenerative disease (ND); Lactate dehydrogenase (LDH), C-reactive protein (CRP), Alanine Amino Transferase (ALT), Aspartate Amino Transaminase (AST), Lactate dehydrogenase (LDH), C-reactive protein (CRP).

Supplementary Table 5: COVID-BioB study team and collaborators.

Affiliation is IRCCS Ospedale San Raffaele, Milan, Italy.

Name	Affiliation
Giovanni Landoni	Università Vita-Salute San Raffaele
Alberto Zangrillo	Università Vita-Salute San Raffaele
Angelo AM Manfredi	Autoimmunity and Vascular Inflammation Unit
Antonella Castagna	Department of Infectious Diseases
Paolo Scarpellini	Department of Infectious Diseases
Paola Cinque	Department of Infectious Diseases
Andrea Poli	Department of Infectious Diseases
Laura Galli	Department of Infectious Diseases
Roberta Caccia	Department of Infectious Diseases
Patrizia Rovere Querini	Department of Internal Medicine
Elena Cantarelli	Molecular Hematology Unit
Michela Grossi	Molecular Hematology Unit
Chiara Bonini	Division of Immunology, Transplantation and
	Infectious Diseases
Mauro Malnati	Division of Immunology, Transplantation and
	Infectious Diseases
Massimo Clementi	Laboratory of Microbiology and Virology
Nicasio Mancini	Laboratory of Microbiology and Virology
Elisa Vicenzi	Viral Pathogens and Biosafety Unit
Guido Poli	Viral Pathogens and Biosafety Unit