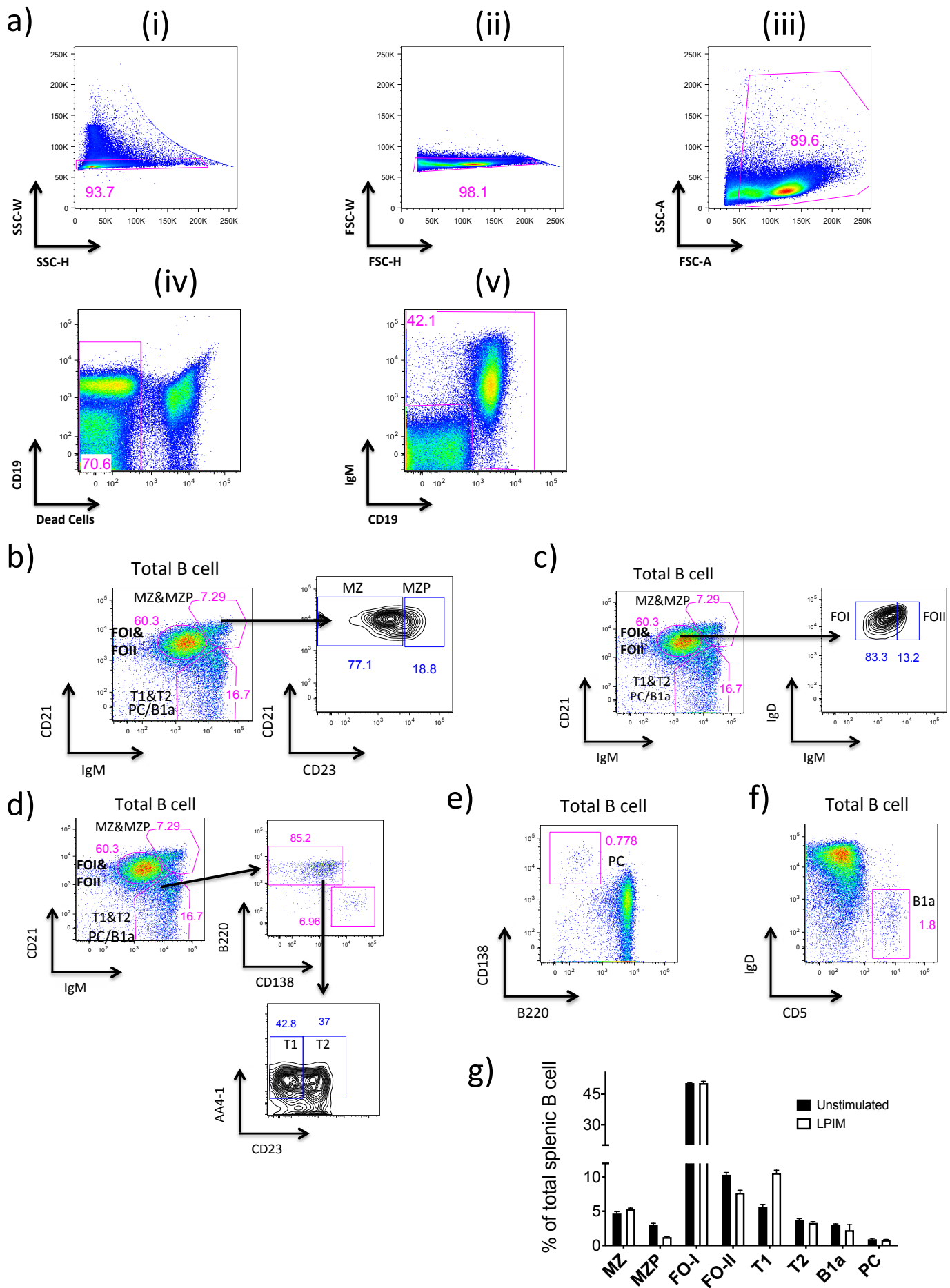


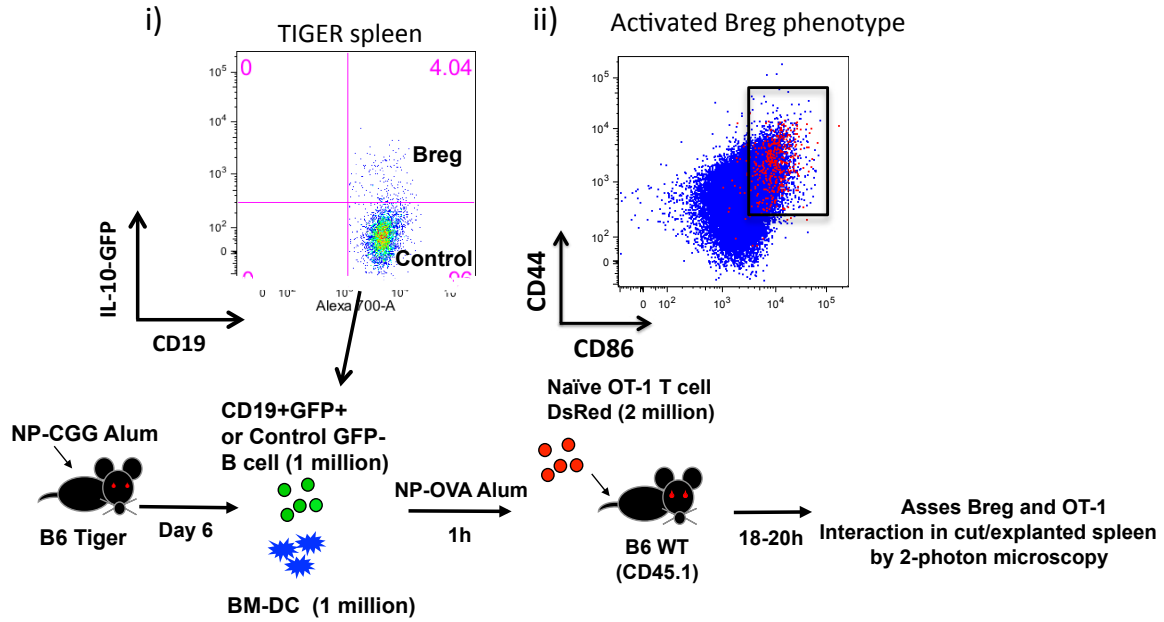
Supplemental Fig. 1



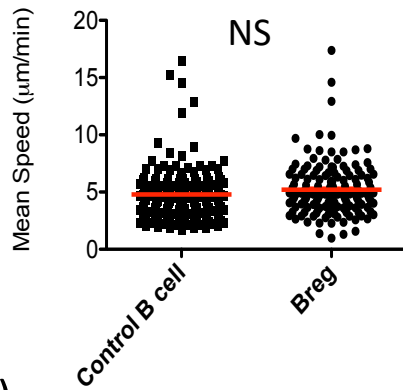
Supplemental Figure 1: Gating strategy to examine IL-10 expression in various B cell subsets. **a)** Gating strategy for total B cells. (i) SSC-W/SSC-H and (ii) FSC-W/FSC-H doublet exclusion, (iii) splenocyte gating, (iv) dead cell exclusion gate, (v) gating on B cells using CD19 and IgM. **b)** Gating shown to examine marginal zone (MZ: IgM^{hi}, CD21^{hi}, CD23^{lo/-}) and marginal zone precursor (MZP: IgM^{hi}, CD21^{hi}, CD23^{hi}) B cell subsets. **c)** Gating shown to examine follicular I (FOI: IgD^{hi}, IgM^{lo}, CD23^{hi}) and follicular II (FOII: IgD^{hi}, IgM^{int}, CD23^{hi}) B cell subsets. **d)** Gating shown to examine transitional 1 (T1: IgM^{hi}, IgD⁻, CD21⁻, CD23⁻, B220⁺, AA4.1⁺) and Transitional 2 (T2: IgM^{hi}, IgD⁻, CD21⁻, CD23⁺, B220⁺, AA4.1⁺) B cell subsets. **e)** Gating shown to examine plasma cell/plasmablast (PC: IgM^{hi}, IgD⁻, CD21⁻, B220⁻, CD138⁺) B cell subset. **f)** Gating shown to examine B1a B cell subset. **g)** Summary graphing showing the frequency of each subset from total splenic B cells before and after stimulation with LPIM.

Supplemental Fig. 2

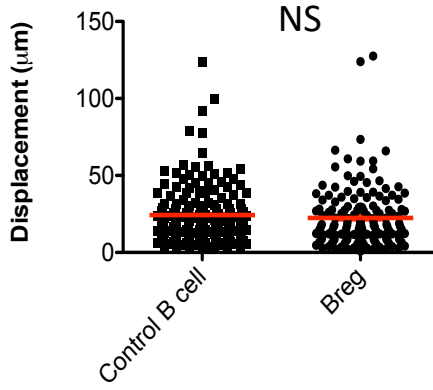
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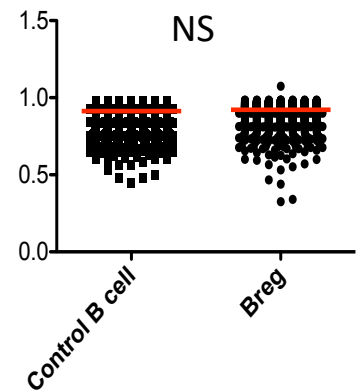
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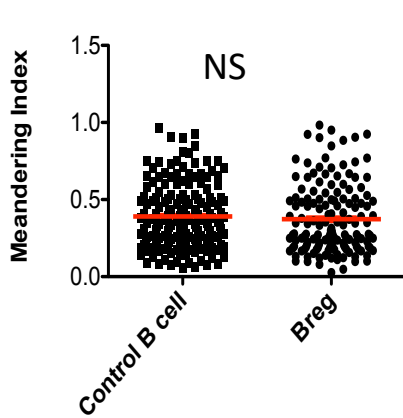
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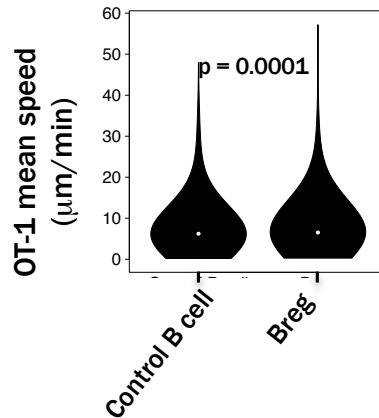
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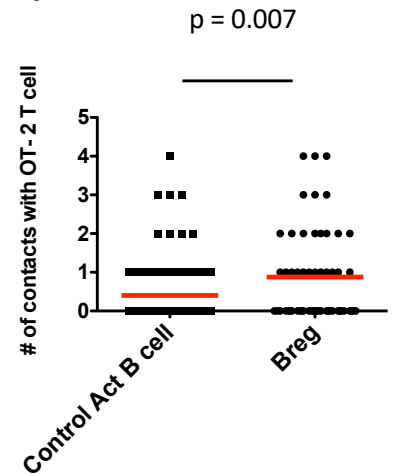
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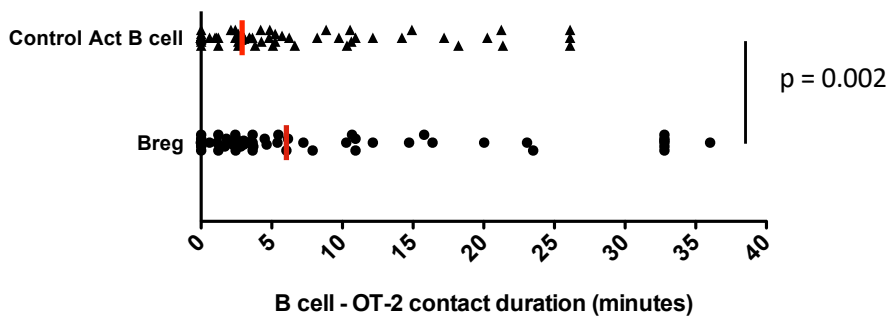
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g)

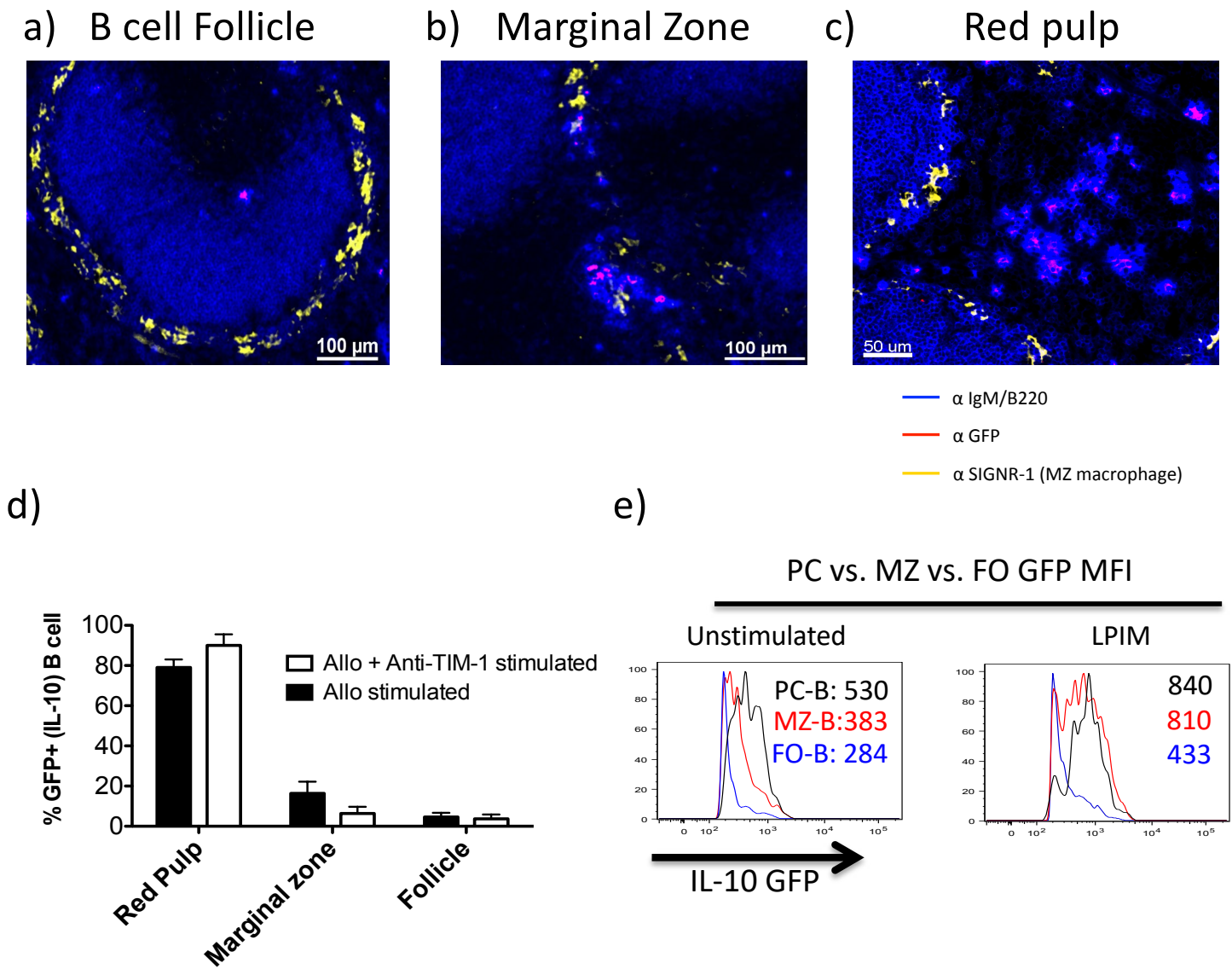


h)



Supplemental Figure 2: Experimental procedure and motility parameters derived from 2 photon imaging of Bregs vs. Control B cells. **a)** Breg (GFP+) (i), Control B cells (GFP-) (i) or Control Activated B cells (ii) were sorted from NP-CGG immunized Tiger mice with BD FACSAria on day 6-7 after immunization. Gate shows CD44 and CD86 expression level of Breg (pink) over-layed on total B cells (blue). Control activated B cells (GFP-) were sorted from within this gate. The sorted cells were stained briefly with 1.5mM CFSE and pulsed with 10 μ g/ml of NP-OVA for one hour. Bone marrow derived DC's (BMDC), generated using GM-CSF and IL-4 using a standard 7 day culture system, were treated with 100ng/ml of LPS for 5-6 hours and pulsed with 10 μ g/ml of NP-OVA for 1 hour. One million of both B cells and DCs were adoptively transferred into WT CD45.1 mice that had received 2 million naïve DsRed OT-1 the day before. After 18-20h, spleens from the recipient mice were removed and cut with a vibratome in order to facilitate imaging of the white pulp. Unused cut pieces of the spleen were snap frozen in OCT for further analysis by immunofluorescence staining. **b-g):** Graphs comparing motility parameters of Control B cells vs. Bregs as analyzed using Bitplane Imaris software. (Each dot represents one cell and red horizontal lines indicate mean for each graph). **b)** Mean speed of Control B cell vs. Bregs. **c)** Displacement of Control B cell vs. Bregs. **d)** Sphericity of Control B cell vs. Bregs. **e)** Meandering index of Control B cell vs. Bregs. **f)** Violin plot showing mean speed of OT-1 T cells obtained from all movies where recipient mice received either Control B cell or Bregs. **g)** The number of contacts individual (NP-OVA pulsed) Control Activated B cells or Bregs made with OT-2 T cells. **h)** Duration (minutes) of each contact made by Control Activated B cell or Bregs with OT-2 T cells. n = 2-5 mice for each group.

Supplemental Fig. 3



Supplemental Figure 3: Immuno-fluorescent images showing Breg localization in immunized Tiger spleens. **a)** Immuno-fluorescent image of a IL-10-GFP reporter mouse spleen immunized with allo-antigen for 72 hours and stained for B cells (anti-IgM/B220 - blue), marginal zone macrophage (anti- SIGNR-1- yellow) and IL-10-GFP (anti-GFP – red) showing a B cell follicle with Breg localized to T:B border, **b)** Breg localized to MZ, **c)** Breg localized to red pulp . **d)** Graph showing cumulative frequency of Breg in immunized IL-10-GFP reporter mice spleens with and without anti-TIM-1 treatment. n = 3-5 mice per group.