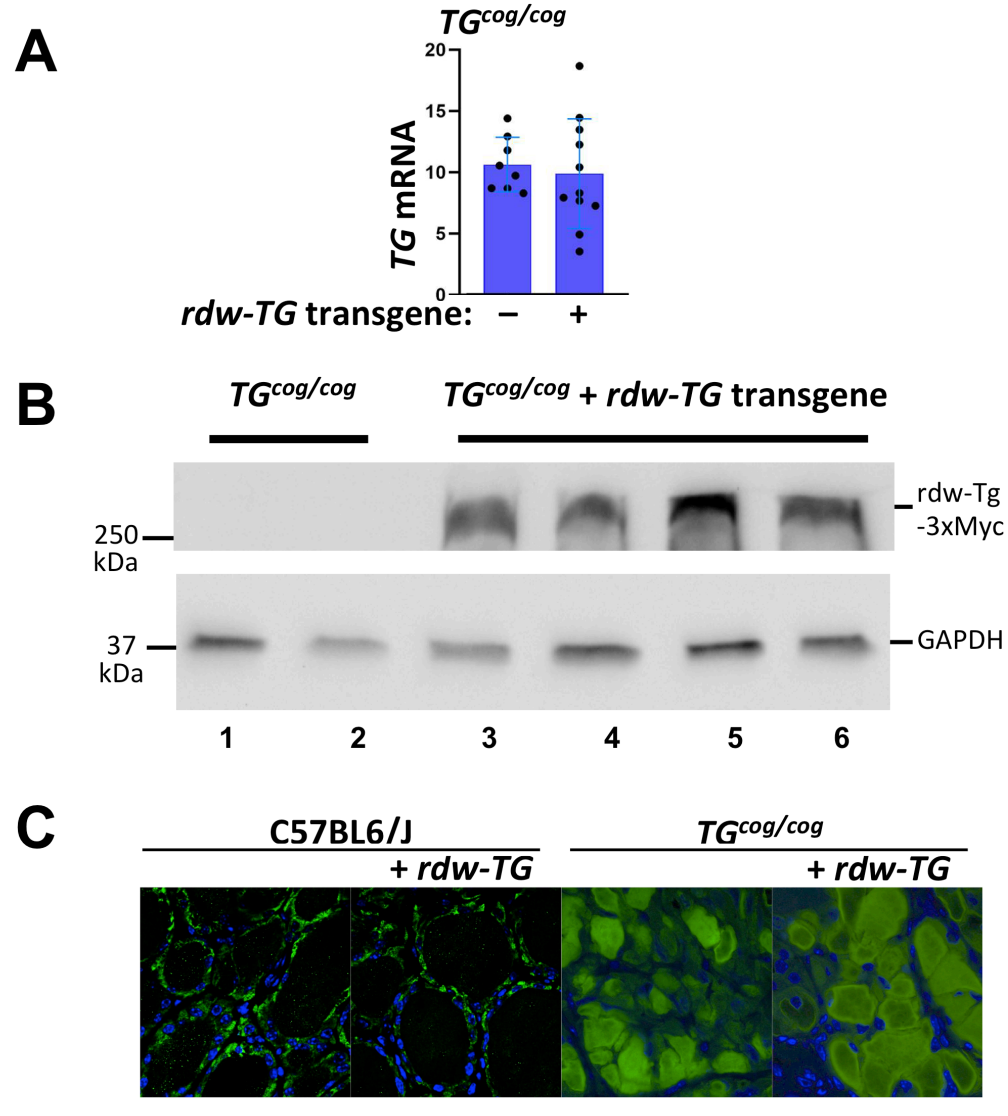


Morishita et al., SI Fig. S1

Figure S1. Generation of $TG^{cog/cog}$ mice bearing a TG^{rdw} transgene. **A.** Depiction of the Tg primary structure, indicating regions I, II, III, ChEL domain, and the location of a triple-myc epitope tag at the C-terminus. The positions of *cog* and *rdw* mutations within the Tg-ChEL domain are indicated. **B.** Construction of a plasmid bearing the bovine *TG* promoter followed by the ORF encoding *rdw*-Tg-3xMyc. **C.** PCR identification of the TG^{rdw} transgene specified by a 284 bp PCR fragment amplified from genomic DNA; a 500 bp beta-actin amplicon was included as an internal control (Con) for the PCR reaction.



Morishita et al., SI Fig. S2

Figure S2. Characterization of the thyroid gland of $TG^{cog/cog}$ mice bearing the TG^{rdw} transgene. **A.** Total TG mRNA in the thyroid gland of $TG^{cog/cog}$ mice lacking or bearing the TG^{rdw} transgene, normalized to housekeeping RNA \pm s.d.. **B.** Western blot detection of $rdw-Tg-3xMyc$ in $TG^{cog/cog}$ mice bearing the TG^{rdw} transgene, and nontransgenic controls. GAPDH is a loading control. **C.** BiP immunofluorescence in the thyroid glands of 2 month-old control mice lacking or bearing the TG^{rdw} transgene, or $TG^{cog/cog}$ mice lacking or bearing the TG^{rdw} transgene (magnification identical in all four panels; but the ER is vastly swollen in $TG^{cog/cog}$ mice).

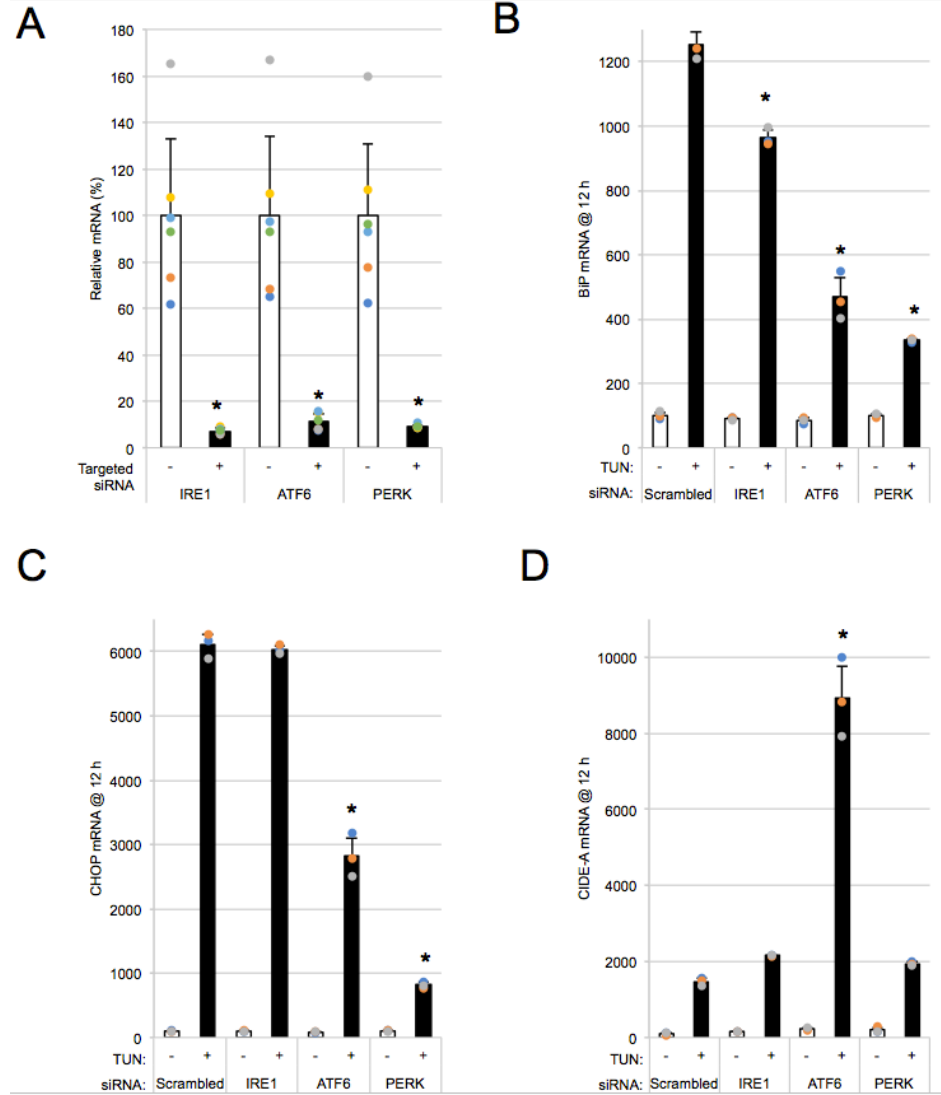


Figure S4. Acute ER stress response after siRNA-mediated knockdown of IRE1, ATF6, or PERK in PCCL3 cells. Replicate wells of PCCL3 were transfected with 30 nM siRNA duplexes for knockdown of IRE1, ATF6, or PERK, or scrambled/noncoding oligos. At 24 h after oligofection, the cells were treated \pm TUN (0.1 μ g/mL). After an additional 12 h, all cells were lysed, and mRNA levels were measured by q-rtPCR for IRE1, ATF6 and PERK (**panel A**; * $p < 0.05$ versus scrambled oligo), BiP (**panel B**), CHOP (**panel C**), and CIDE-A (**panel D**). The data show the mean (from three biological replicates) \pm s.d in a single experiment; * $p < 0.05$ versus TUN-treated cells transfected with scrambled oligo (*decreased* for BiP and CHOP; *increased* for CIDE-A).

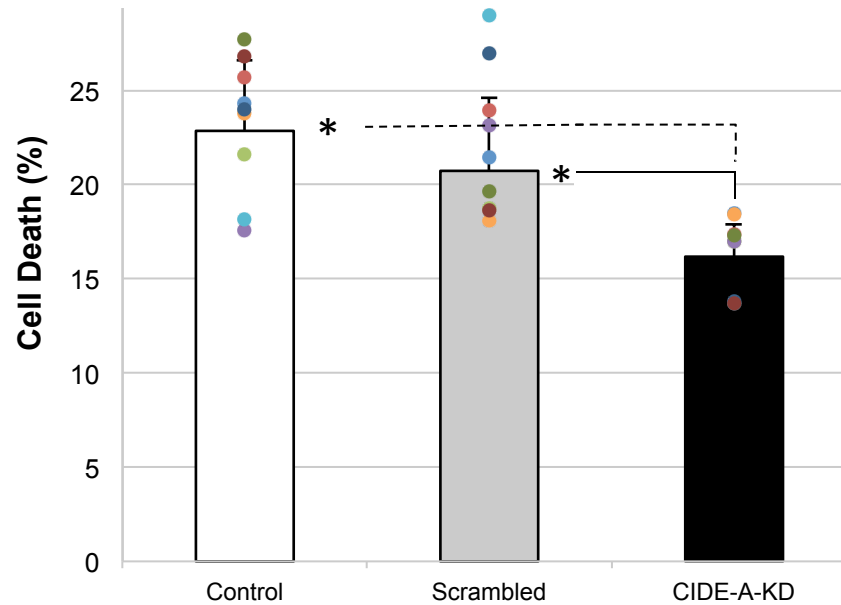
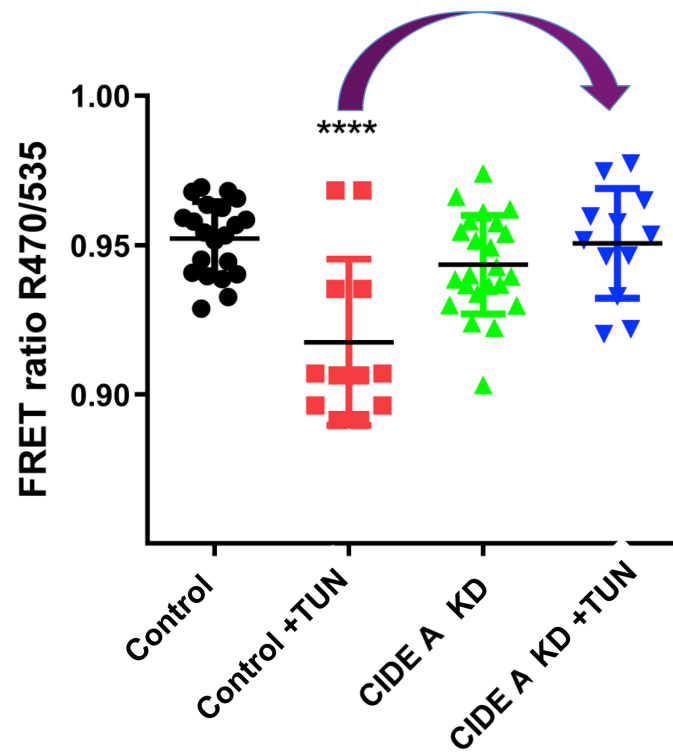


Figure S5. Cell death limited by siRNA-mediated knockdown of CIDE-A. Each set of cells were either untransfected (Control) or transfected with Scrambled or siRNA-mediated knockdown of endogenous CIDE-A (CIDE-A-KD). All cells were treated with DOX (for the full 3 d) to induce CIDE-A-myc (as in Fig. 7) plus TUN (0.1 $\mu\text{g}/\text{mL}$ for the last 2 d) to induces ER stress. At 3 d, cell death was quantified in all samples; cell death was limited by CIDE-A-KD; the data represent the mean (from three independent experiment in which each experimental sample had three biological replicates) \pm s.d.; * $p < 0.05$ versus samples without CIDE-A-KD.



Morishita et al., SI Fig. S6

Figure S6. ER stress effect on ER calcium level in PCCL3 thryocytes. One day after plating, PCCL3 cells were transfected with Lipofectamine bearing either 10 nM scrambled oligo (*left-two set of samples*) or siRNA for knockdown of CIDE-A (*right-two set of samples*). Thereafter, all sets of cells were plasmid-transfected (Viafect transfection reagent) to express D4ER in pCDNA3.1. One half of each set of cells were challenged with TUN (0.1 $\mu\text{g}/\text{mL}$), or vehicle alone. At 48 h after TUN addition, transfected cells were washed in PBS plus 2 mM MgCl_2 and 5.5 mM glucose, and underwent fluorescence imaging and FRET analysis using 430 nm excitation and 470/535 nm ratiometric emission. The FRET ratio shown for each cell (acquired using Metafluor) reflects the relative Ca^{++} level in the ER; a lower ratio signifies lower ER calcium. Each point reflects a single cell from a representative experiment (of four independent FRET experiments); mean values \pm s.d. are shown.

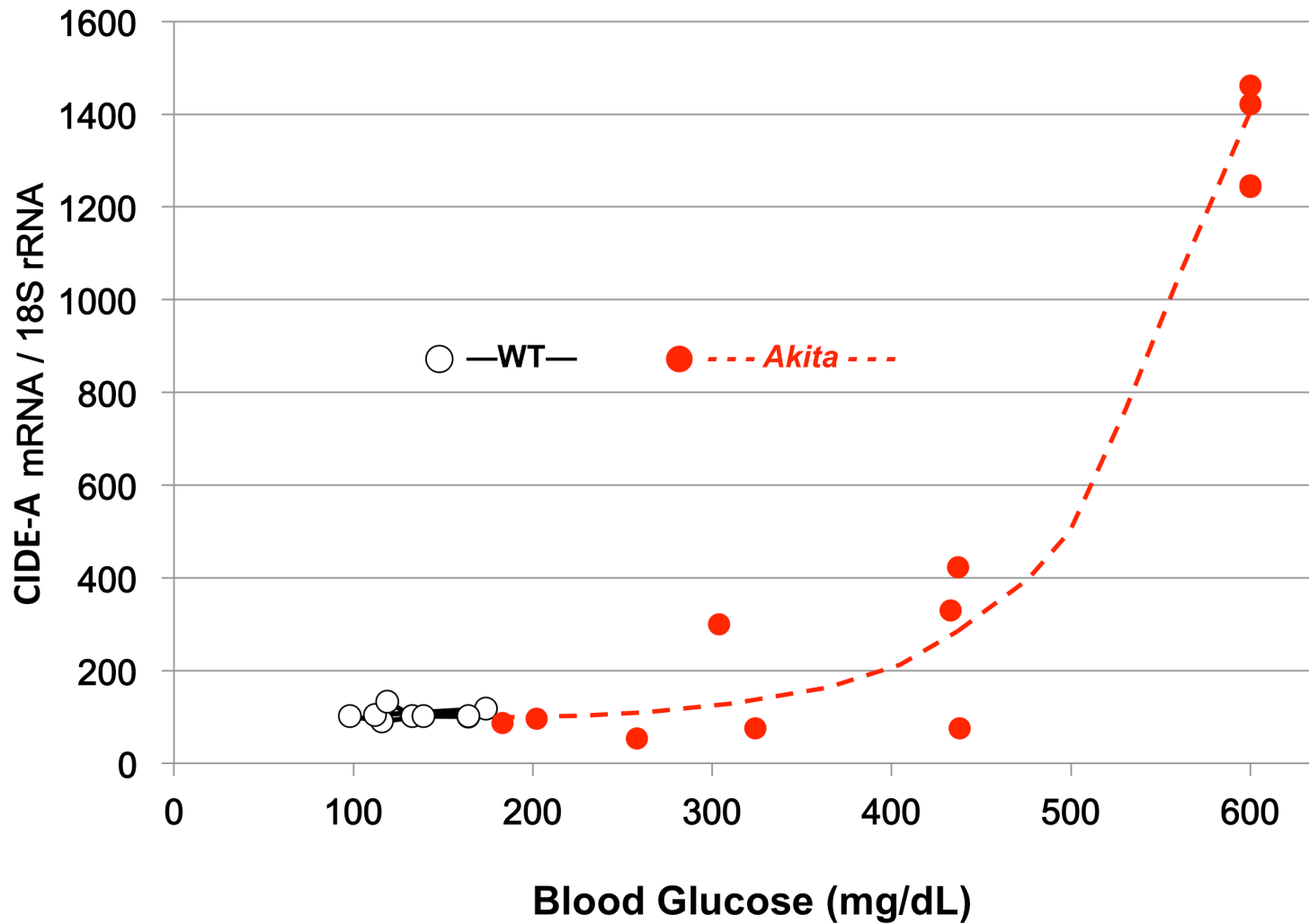


Figure S7. CIDE-A mRNA levels in *Akita* mouse pancreatic islets as a function of progressive dysglycemia. Islets were isolated from male wild-type and *Akita* mice that reflect progression based on random blood glucose. Each point represents a different animal, with CIDE-A mRNA levels normalized to ribosomal 18S RNA.