# THE LANCET Infectious Diseases

## Supplementary appendix

This appendix formed part of the original submission and has been peer reviewed. We post it as supplied by the authors.

Supplement to: Ramanathan M, Ferguson ID, Miao W, Khavari PA. SARS-CoV-2 B.1.1.7 and B.1.351 spike variants bind human ACE2 with increased affinity. *Lancet Infect Dis* 2021; published online May 19. https://doi.org/10.1016/S1473-3099(21)00262-0.

#### **Materials and Methods**

#### **Recombinant Proteins**

Human ACE2 protein was purchased from Sino Biological (SinoBiological, catalog#10108-H05H) and re-suspended in PBS-T to obtain 4.54uM stock concentration. Recombinant Spike Receptor Binding Domains were ordered from BEI resources. Protein quality was determined using a Tycho NT.6 (**Figures S4 & S5**) (NanoTemper Technologies). Human hnRNPC was purchased from Abnova (catalog# H00003183P01S.)

#### Microscale Thermophoresis

Spike RBD proteins were labeled using Monolith His-Tag Labeling Kit RED-tris-NTA (NanoTemper Technologies) following manufacturer protocol at 2:1 protein to dye ratio. Briefly, 100 uL of 200nM Spike RBD (Hu-1 or B.1.1.7 or B.1.351 RBD) was mixed with 100 uL of 100nM dye for 30 minutes at room temperature, followed by centrifugation for 10 minutes at 15000g at 4C. A series of sixteen 1:1 dilutions of hACE2 were prepared, with 2.27uM the highest concentration. Each dilution of hACE2 was mixed with one volume of Spike labeling reaction mix prior to incubation for 5 minutes at room temperature. Mixed samples were loaded into Monolith NT.115 Capillaries (NanoTemper Technologies). As a specificity control, hnRNPC was utilized instead of hACE2; MST showed no binding (**Figure S6**). MST was performed using a Monolith NT.115 instrument (NanoTemper Technologies) at room temperature, with 60-80% excitation power and Medium MST power.

#### **Statistical Analyses**

All statistical analyses were performed with GraphPad Prism 8.4 (Graphpad Software Inc.). Student's t-test was performed to compare the binding affinity of SCoV2 and variants with human ACE2 protein.

### Acknowledgements

The following reagent was produced under HHSN272201400008C and obtained through BEI Resources, NIAID, NIH: Spike Glycoprotein Receptor Binding Domain (RBD) from SARS-Related Coronavirus 2, Hu-1 with C-Terminal Histidine Tag, Recombinant from HEK293F Cells, NR-52366. The following reagents were obtained through BEI Resources, NIAID, NIH: Spike Glycoprotein Receptor Binding Domain (RBD) from SARS-Related Coronavirus 2, South Africa Variant (NR-54005) and United Kingdom Variant (NR-54004) with C-Terminal Histidine Tag, Recombinant from HEK293 Cells. This work was supported by the USVA Office of Research and Development I01BX00140908 (P.A.K.), NIH CA142635, AR45192, AR076965 and HG007919 (P.A.K).

Wuhan-Hu-1	1 R V Q P T E S I V R F P N I T N L C P F G E V F N A T R F A S V Y AWNR K R I S N C V A D Y S V L Y N S A S F S T F K C Y G V S P T K	K L N D L C F	74
B.1.1.7	1 RVQPTESIVRFPNITNLCPFGEVFNATRFASVYAWNRKRISNCVADYSVLYNSASFSTFKCYGVSPTK	KLNDLCF	74
B.1.135	1 R V Q P T E S I V R F P N I T N L C P F G E V F NAT R F A S V Y AWNR K R I S N C V A D Y S V L Y N S A S F S T F K C Y G V S P T K	K L N D L C F	74
Wuhan-Hu-1	75 TNVYADS FVIRGDEVRQIAP GQTGKIADYNYKLPDDFTGCVIAWN SNNLDSKVGGNYNYLYRL FRKSM	NLKPFER 1	48
B.1.1.7	75 TNVYADS FVIRGDEVRQIAP GQTGKIADYNYK LPDDFTGCVIAWN SNNLDSKVGGNYNYLYRL FRK SN	NLKPFER 1	48
B.1.135	75 TNVYADS FVIRGDEVRQIAP GQTGNIADYNYKLPDDFTGCVIAWNSNNLDSKVGGNYNYLYRLFRKSM	NLKPFER 1	48
Wuhan-Hu-1	149 DI ST E I YQAG ST P C NG V E G F N C Y F P L Q S Y G F Q P T N G V G Y Q P Y R V V V L S F E L L H A P A T V C G P K K S T N L V	KNKCVN 2	222
B.1.1.7	149 D I ST E I YQAG ST P C NG V G F NC Y F P LQ S Y G F Q P T G V G Y Q P Y R V V V L S F E L L HA P A T V C G P K K S T N L V	VKNKCVN 2	22
B.1.135	149 DI STEIYQAGSTPCNGVKGFNCYFPLQSYGFQPTWGVGYQPYRVVVLSFELLHAPATVCGPKKSTNLV	KNKCVN 2	22
Wuhan-Hu-1	223 F	2	223
B.1.1.7	223 F	2	223
B.1.135	223 F	2	23

**Figure S1**: Sequence and alignment of RBD proteins assayed. The sequence of the parental Hu-1 strain is used as SCoV2 RBD sequence. Mutations seen in B.1.1.7 and B.1.351 are highlighted.



**Figure S2**: Structure of the SARS-CoV-2 RBD (PDB: 6M0J) with B.1.1.7 mutation (N501Y) on RBD colored red. Coloring schema: ACE2 (grey), receptor-binding domain (green)



**Figure S3**: Structure of the SARS-CoV-2 RBD (PDB: 6M0J) with B.1.351 mutation (K417N, E484K, N501Y) on RBD colored red. Coloring schema: ACE2 (grey), receptor-binding domain (green)





**Fig. 1.** Microscale thermophoresis (MST) with 100nM labeled Spike RBD (**A**) Binding curve of SCoV2 and B.1.1.7 RBD with a titration series of human ACE2 (n=3 series of independent replicates)

(**B**) Binding curve of SCoV2 and B.1.351 RBD with a titration series of human ACE2 (n=3 series of independent replicates).

MST data are mean ± SD. Error bars=SD



**Figure S4**: Protein quality of RBD proteins assayed. Tycho NT. 6 (Nanotemper) was used to measure changes in intrinsic fluorescence at 330 nm and 350 nm with increasing temperature. The fluorescence signals were plotted as a first derivative of the 350 nm/330 nm ratio.



**Figure S5**: Protein quality of ACE2. Tycho NT. 6 (Nanotemper) was used to measure changes in intrinsic fluorescence at 330 nm and 350 nm with increasing temperature. The fluorescence signals were plotted as a first derivative of the 350 nm/330 nm ratio.



**Figure S6**: Microscale thermophoresis with 100nM labeled Spike RBD. SCoV2 and hnRNPC with a titration series of human hnRNPC protein shows no binding, confirming SCoV2 does not interact non-specifically (n=2 series of independent replicates).