

Increased expression of tryptophan and tyrosine tRNAs elevates stop codon readthrough of reporter systems in human cell lines.

Petra Beznosková^{1*}, Laure Bidou^{2,3}, Olivier Namy³ and Leoš Shivaya Valášek^{1*}

¹Laboratory of Regulation of Gene Expression, Institute of Microbiology ASCR, Videnska 1083, 142 20, Prague, the Czech Republic

² Sorbonne Universités, Paris, France.

³ Université Paris-Saclay, CEA, CNRS, Institute for Integrative Biology of the Cell (I2BC), 91198, Gif-sur-Yvette, France.

* To whom correspondence should be addressed. valasekl@biomed.cas.cz
p.beznoskova@seznam.cz
Tel: +420 241 062 288
Fax: +420 241 062 665

Running title: human tRNAs regulate readthrough

SUPPLEMENTAL INFORMATION

EXPERIMENTAL PROCEDURES

Construction of plasmids

List of all plasmids, PCR primers and Genestrings used throughout this study can be found in Tables S1, S2 and S3, respectively.

PBB166 (=SW148 lacking *KpnI* site) was created by altering SW148 plasmid by digestion with *KpnI*, trimming by T4 DNA polymerase and subsequent self-ligation.

PBB168 was created by inserting the undigested PCR product obtained with primers PB159 and PB160 using vector Addgene53188 as template into *SspI* digested PBB166.

PBB170 was created by inserting the *SalI-NotI* digested fragment from SW174 into *SalI-NotI* digested PBB168.

PBB171 was created by inserting the *SalI-NotI* digested fragment from SW149 into *SalI-NotI* digested PBB168.

PBB171 was created by inserting the *SalI-NotI* digested fragment from SW150 into *SalI-NotI* digested PBB168.

PBB180 was created by inserting the *Apal/KpnI* fragment from annealed primers PB162 and PB167 into *Apal/KpnI* digested PBB168.

PBB185 was created by inserting the *SalI-NotI* digested fragment from PBB170 into *SalI-NotI* digested PBB180.

PBB220 was created by inserting the *SalI-NotI* digested fragment from PBB171 into *SalI-NotI* digested PBB180.

PBB184 was created by inserting the *Apal/KpnI* fragment from annealed primers PB162 and PB167 into *Apal/KpnI* digested PBB172.

PBB181 was created by inserting the *SalI-NotI* digested fragment from PBB168 into *SalI-NotI* digested PBB187.

PBB187 was created by inserting the *Apal/KpnI* fragment from annealed primers PB164 and PB169 into *Apal/KpnI* digested PBB172.

PBB182 was created by inserting the *Apal/KpnI* fragment from annealed primers PB166 and PB171 into *Apal/KpnI* digested PBB168.

PBB189 was created by inserting the *SalI-NotI* digested fragment from PBB172 into *SalI-NotI* digested PBB182.

PBB204 was created by inserting the *SalI-NotI* digested fragment from PBB168 into *SalI-NotI* digested PBB198.

PBB206 was created by inserting the *SalI-NotI* digested fragment from PBB170 into *SalI-NotI* digested PBB198.

PBB207 was created by inserting the *SalI-NotI* digested fragment from PBB171 into *SalI-NotI* digested PBB198.

PBB198 was created by inserting the *Apal/KpnI* fragment from annealed primers PB177 and PB178 into *Apal/KpnI* digested PBB172.

PBB288 was created by inserting the *PmeI* digested fragment from Genestring SG7 into *PmeI* digested PBB207.

PBB292 was created by inserting the *PmeI* digested fragment from Genestring SG8 into *PmeI* digested PBB180.

PBB214 was created by inserting the *SalI-NotI* digested fragment from PBB168 into *SalI-NotI* digested PBB196.

PBB196 was created by inserting the *Apal/KpnI* fragment from annealed primers PB175 and PB176 into *Apal/KpnI* digested PBB172.

PBB313 was created by inserting undigested fragment from Genestring SG16 into *SspI* digested PBB166.

PBB314 was created by inserting undigested fragment from Genestring SG17 into *SspI* digested PBB166.

PBB315 was created by inserting undigested fragment from Genestring SG18 into *SspI* digested PBB166.

PBB316 was created by inserting undigested fragment from Genestring SG19 into *SspI* digested PBB166.

PBB317 was created by inserting undigested fragment from Genestring SG20 into *SspI* digested PBB166.

PBB318 was created by inserting undigested fragment from Genestring SG21 into *SspI* digested PBB166.

PBB319 was created by inserting undigested fragment from Genestring SG22 into *SspI* digested PBB166.

PBB320 was created by inserting undigested fragment from Genestring SG23 into *SspI* digested PBB166.

PBB336 was created by inserting *PmeI* digested fragment from Genestring SG24 into *SspI* digested PBB166.

PBB337 was created by inserting *PmeI* digested fragment from Genestring SG25 into *SspI* digested PBB166.

PBB338 was created by inserting *PmeI* digested fragment from Genestring SG26 into *SspI* digested PBB166.

PBB293 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3051.

PBB293 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3051.

PBB306 was created by inserting *SalI/BamHI* digested fragment from Genestring SG9 into *SalI/BamHI* digested GL3051.

PBB307 was created by inserting *SalI/BamHI* digested fragment from Genestring SG10 into *SalI/BamHI* digested GL3051.

PBB308 was created by inserting *SalI/BamHI* digested fragment from Genestring SG11 into *SalI/BamHI* digested GL3051.

PBB309 was created by inserting *SalI/BamHI* digested fragment from Genestring SG12 into *SalI/BamHI* digested GL3051.

PBB310 was created by inserting *SalI/BamHI* digested fragment from Genestring SG13 into *SalI/BamHI* digested GL3051.

PBB311 was created by inserting *SalI/BamHI* digested fragment from Genestring SG14 into *SalI/BamHI* digested GL3051.

PBB295 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3052.

PBB294 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3051.

PBB296 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3052.

PBB371 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3101.

PBB373 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3102.

PBB372 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3101.

PBB374 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3102.

PBB297 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3095.

PBB299 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3096.

PBB298 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3095.

PBB300 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3096.

HR4 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3107.

HR12 was created by inserting *SalI/BamHI* digested fragment from Genestring SG7 into *SalI/BamHI* digested GL3108.

HR5 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3107.

HR13 was created by inserting *SalI/BamHI* digested fragment from Genestring SG8 into *SalI/BamHI* digested GL3108.

PBB271 was created by inserting *SalI/SacI* digested fragment from Genestring SG1 into *SalI/SacI* digested PBB168.

PBB272 was created by inserting *SalI/SacI* digested fragment from Genestring SG2 into *SalI/SacI* digested PBB168.

PBB273 was created by inserting *SalI/SacI* digested fragment from Genestring SG3 into *SalI/SacI* digested PBB168.

PBB274 was created by inserting *SalI/SacI* digested fragment from Genestring SG4 into *SalI/SacI* digested PBB168.

PBB275 was created by inserting *SalI/SacI* digested fragment from Genestring SG1 into *SalI/SacI* digested PBB207.

PBB276 was created by inserting *SalI/SacI* digested fragment from Genestring SG2 into *SalI/SacI* digested PBB207.

PBB277 was created by inserting *SalI/SacI* digested fragment from Genestring SG3 into *SalI/SacI* digested PBB207.

PBB278 was created by inserting *SalI/SacI* digested fragment from Genestring SG4 into *SalI/SacI* digested PBB207.

PBB219 was created by inserting the *SalI-NotI* digested fragment from PBB177 into *SalI-NotI* digested PBB172.

PBB234 was created by inserting the *SalI-NotI* digested PCR product obtained with primers PB43 and PB187 using PBB219 as template into *SalI-NotI* digested PBB219.

PBB223 was created by inserting the *SalI-NotI* digested fragment from PBB177 into *SalI-NotI* digested PBB207.

PBB235 was created by inserting the *SalI-NotI* digested fragment from PBB234 into *SalI-NotI* digested PBB207.

PBB301 was created by inserting *PmeI* digested fragment from Genestring SG7 into *SmaI* digested PBB321 (p53BS-luci).

PBB407 was created by Site-directed mutagenesis of pCMV-p53-WT (PBB395) using primers PB212 and PB213.

PBB400 was created by Site-directed mutagenesis of pCMV-p53-WT (PBB395) using primers PB216 and PB217.

PBB301 was created by inserting two *PmeI* digested fragments from Genestring SG7 into *SmaI* digested PBB321 (p53BS-luci).

PBB332 was created by inserting *PmeI* digested fragment from Genestring SG8 into *SmaI* digested PBB321 (p53BS-luci).

PBB380 was created by inserting *SalI/BamHI* digested fragment from PBB375 into *SalI/BamHI* digested GL3101.

PBB382 was created by inserting *SalI/BamHI* digested fragment from PBB375 into *SalI/BamHI* digested GL3102.

REFERENCES

1. Grentzmann, G., Ingram, J.A., Kelly, P.J., Gesteland, R.F. and Atkins, J.F. (1998) A dual-luciferase reporter system for studying recoding signals. *RNA*, **4**, 479-486.
2. Loughran, G., Howard, M.T., Firth, A.E. and Atkins, J.F. (2017) Avoidance of reporter assay distortions from fused dual reporters. *RNA*, **23**, 1285-1289.
3. Floquet, C., Deforges, J., Rousset, J.P. and Bidou, L. (2011) Rescue of non-sense mutated p53 tumor suppressor gene by aminoglycosides. *Nucleic Acids Res*, **39**, 3350-3362.
4. Chan, P.P. and Lowe, T.M. (2016) GtRNADB 2.0: an expanded database of transfer RNA genes identified in complete and draft genomes. *Nucleic Acids Res*, **44**, D184-189.
5. Chan, P.P. and Lowe, T.M. (2009) GtRNADB: a database of transfer RNA genes detected in genomic sequence. *Nucleic Acids Res*, **37**, D93-97.
6. Firth, A.E., Wills, N.M., Gesteland, R.F. and Atkins, J.F. (2011) Stimulation of stop codon readthrough: frequent presence of an extended 3' RNA structural element. *Nucleic Acids Res*, **39**, 6679-6691.
7. Napthine, S., Yek, C., Powell, M.L., Brown, T.D. and Brierley, I. (2012) Characterization of the stop codon readthrough signal of Colorado tick fever virus segment 9 RNA. *RNA*, **18**, 241-252.

SUPPLEMENTAL TABLES

Table S1. Plasmids used in this study.

Plasmid	Description	Source of reference
SW148	p2luci_UAG-TMV	(1)
SW147	p2luci_UAA-TMV	(1)
SW149	p2luci_UGA-TMV	(1)
SW150	p2luci_CAG-TMV	(1)
PBB166	SW148noKpnI	This study
PBB168	p2luci-UAG-TMV-U6	This study
PBB170	p2luci-UAA-TMV-U6	This study
PBB171	p2luci-UGA-TMV-U6	This study
PBB172	p2luci-CAG-TMV-U6	This study
PBB180	p2luci-UAG-TMV-U6-tY01	This study
PBB185	p2luci-UAA-TMV-U6-tY01	This study
PBB220	p2luci-UGA-TMV-U6-tY01	This study
PBB184	p2luci-CAG-TMV-U6-tY01	This study
PBB181	p2luci-UAG-TMV-U6-tQ01	This study
PBB187	p2luci-CAG-TMV-U6-tQ01	This study
PBB182	p2luci-UAG-TMV-U6-tQ01s	This study
PBB189	p2luci-CAG-TMV-U6-tQ01s	This study
PBB204	p2luci-UAG-TMV-U6-tW01	This study
PBB206	p2luci-UAA-TMV-U6-tW01	This study
PBB207	p2luci-UGA-TMV-U6-tW01	This study
PBB198	p2luci-CAG-TMV-U6-tW01	This study
PBB288	p2luci-UGA-TMV-U6-tW01-U6-tW01	This study
PBB292	p2luci-UAG-TMV-U6-tY01-U6-tY01	This study
PBB214	p2luci-UAG-TMV-U6-tY02	This study
PBB196	p2luci-CAG-TMV-U6-tY02	This study
PBB313	p2luci_UAG-TMV_tY03	This study
PBB314	p2luci_UAG-TMV_tY04	This study
PBB315	p2luci_UAG-TMV_tY05	This study
PBB336	p2luci_UAG-TMV_tY05b	This study
PBB337	p2luci_UAG-TMV_tY05c	This study
PBB338	p2luci_UAG-TMV_tY05d	This study
PBB316	p2luci_UAG-TMV_tY06	This study
PBB317	p2luci_UAG-TMV_tY07	This study
PBB318	p2luci_UAG-TMV_tY08	This study
PBB319	p2luci_UAG-TMV_tY09	This study
PBB320	p2luci_UAG-TMV_tY10	This study
GL3051	pSGDluc_UGA-AQP4	(2)
GL3052	pSGDluc_UGG-AQP4	(2)

PBB293	pSGDluc_UGA-AQP4-U6-tW01	This study
PBB306	pSGDluc_UGA-AQP4-U6-tW02	This study
PBB307	pSGDluc_UGA-AQP4-U6-tW03	This study
PBB308	pSGDluc_UGA-AQP4-U6-tW04	This study
PBB309	pSGDluc_UGA-AQP4-U6-tW05	This study
PBB310	pSGDluc_UGA-AQP4-U6-tW06	This study
PBB311	pSGDluc_UGA-AQP4-U6-tW07	This study
PBB295	pSGDluc_UGG-AQP4-U6-tW01	This study
PBB294	pSGDluc_UGA-AQP4-U6-tY01	This study
PBB296	pSGDluc_UGG-AQP4-U6-tY01	This study
GL3101	pSGDluc_UGA-MDH1	(2)
GL3102	pSGDluc_UGG-MDH1	(2)
PBB371	pSGDluc_UGA-MDH1-U6-tW01	This study
PBB373	pSGDluc_UGG-MDH1-U6-tW01	This study
PBB372	pSGDluc_UGA-MDH1-U6-tY01	This study
PBB374	pSGDluc_UGG-MDH1-U6-tY01	This study
GL3095	pSGDluc_UGA-ACP2-2	(2)
GL3096	pSGDluc_UGG-ACP2-2	(2)
PBB297	pSGDluc_UGA-ACP2-2-U6-tW01	This study
PBB299	pSGDluc_UGG-ACP2-2-U6-tW01	This study
PBB298	pSGDluc_UGA-ACP2-2-U6-tY01	This study
PBB300	pSGDluc_UGG-ACP2-2-U6-tY01	This study
GL3107	pSGDluc_UGA-C	(2)
GL3108	pSGDluc_UGG-C	(2)
HR4	pSGDluc_UGA-C-U6-tW01	This study
HR12	pSGDluc_UGG-C-U6-tW01	This study
HR5	pSGDluc_UGA-C-U6-tY01	This study
HR13	pSGDluc_UGG-C-U6-tY01	This study
PBB271	p2luci-Sindbis_UGG-U6	This study
PBB272	p2luci-Sindbis_UGA-U6	This study
PBB273	p2luci-CTFV_UGG-U6	This study
PBB274	p2luci-CTFV_UGA-U6	This study
PBB275	p2luci-Sindbis_UGG-U6-tW	This study
PBB276	p2luci-Sindbis_UGA-U6-tW	This study
PBB277	p2luci-CTFV_UGG-U6-tW	This study
PBB278	p2luci-CTFV_UGA-U6-tW	This study
PBB219	p2luci-UGA-CTAG-U6	This study
PBB234	p2luci-CAG-CTAG-U6	This study
PBB223	p2luci-UGA-CTAG-U6-tW01	This study
PBB235	p2luci-CAG-CTAG-U6-tW01	This study
PBB301	p53BS-luci_U6-tW01-Smal-a	This study
PBB321	p53BS-luci	(3)

PBB394	pCMV-LacZ	(3)
PBB395	pCMV-p53-WT	(3)
PBB406	pCMV-p53-W53UAG	(3)
PBB407	pCMV-p53-W53UGA	This study
PBB400	pCMV-p53-W146UGA	This study
PBB330	p53BS-luci_2x_U6-tW01-SmaI-a	This study
PBB332	p53BS-luci-U6-tY01	This study
PBB375	tet-U6-tW01	Invitrogen
PBB380	pSGDluc_UGA-MDH1-tetU6-tW01	This study
PBB382	pSGDluc_UGG-MDH1-tetU6-tW01	This study

Table S2. Primers used in this study.

Primer name	Primer sequence (5' to 3')
PB159	ATGGCCAAAGGTCGGGCAGGAAGAG
PB160	AGGTACCACGCGTGGGCCCGGTGTTTCGTCCTTTCCA C
PB162	AATAAGGGCCCCCTTCGATAGCTCAGCTGGTAGAGCGG AGGACTGTAGATCCTTAGGtCGCTGGTTCGATTCCGGCT CGAAGGAGGTACCAATAA
PB167	TTATTGGTACCTCCTTCGAGCCGGAATCGAACCAGCGA CCTAAGGATCTACAGTCCTCCGCTCTACCAGCTGAGCT ATCGAAGGGGGCCCTTATT
PB164	AATAAGGGCCCCGGTTCATGGTGTAATGGTTAGCACTC TGGACTCTGAATCCAGCGATCCGAGTTCAAATCTCGGT GGAACCTGGTACCAATAA
PB169	TTATTGGTACCAGGTTCCACCGAGATTTGAACTCGGATC GCTGGATTCAGAGTCCAGAGTGCTAACCATTACACCAT GGAACCGGGCCCTTATT
PB166	AATAAGGGCCCCGGTTCATGGTGTAATGGTTAGCACTC TGGACTCTAAATCCAGCGATCCGAGTTCAAATCTCGGT GGAACCTGGTACCAATAA
PB171	TTATTGGTACCAGGTTCCACCGAGATTTGAACTCGGATC GCTGGATTTAGAGTCCAGAGTGCTAACCATTACACCAT GGAACCGGGCCCTTATT
PB177	AATAAGGGCCCCGACCTCGTGGCGCAACGGTAGCGCGT CTGACTCCAGATCAGAAGGCTGCGTGTTTCGAATCACGT CGGGGTCAGGTACCAATAA
PB178	TTATTGGTACCTGACCCCGACGTGATTCGAACACGCAG CCTTCTGATCTGGAGTCAGACGCGCTACCGTTGCGCCA CGAGGTTCGGGCCCTTATT
PB175	AATAAGGGCCCCCTTCAATAGTTCAGCTGGTAGAGCAG AGGACTATAGGTCCTTAGGTTGCTGGTTCGATTCCAGC TTGAAGGAGGTACCAATAA
PB176	TTATTGGTACCTCCTTCAAGCTGGAATCGAACCAGCAAC CTAAGGACCTATAGTCCTCTGCTCTACCAGCTGAACTAT TGAAGGGGGCCCTTATT
PB43	CTCGAAGCGGCCGCTCTAGAATTACAC
PB187	CAAATGTCGACGTGCGATCAGCTAGTCGGATCCTTCAA CTTCCCTGAGCTCG
PB212	CGATATTGAACAATGATTCAGTGAAGACCCA
PB213	TGGGTCTTCAGTGAATCATTGTTCAATATCG
PB216	CCCTGTGCAGCTGTGAGTTGATTCCACACCC
PB217	GGGTGTGGAATCAACTCACAGCTGCACAGGG
WARS_qPCR-f	GCTCATTGTTTCGGTTTGGGA

WARS_qPCR-r	GAAGTGGTGTGGTCTTTGG
YARS_qPCR-f	CTGAGGTGGTAAAGCAGGT
YARS_qPCR-r	AATACTCTTCATCCAAAGCCTG
eRF1_qPCR-f	GGTCTGGTTGTATACTGTGG
eRF1_qPCR-r	TGTAAGAGCCTCTGTATGGA
18S_qPCR-f	CCGCAGCTAGGAATAATG
18S_qPCR-r	CCGGTCCAAGAATTTAC
ALAS_qPCR-f	CCACTGGAAGAGCTGTGTGATGTG
ALAS_qPCR-r	GCGATGTACCCTCCAACACAACC
SPIKE_qPCR-f	CGAAATGAGAGCCAAGTGG
SPIKE_qPCR-r	ATGCAATTTAGATCCATTATGAGG

Table S3. Genestrings (GeneArt™ Strings™ DNA Fragments by Invitrogen) used in this study.

Name	DNA sequence (5' to 3')
SG1	GTTCTCAAAAATGAACAAATGTCGACGTA CTGGGTAAGGTGGGTACATATTTTCGACGGACACAGGCC CTGGGCACTTGCAAAGAAGTCCGTTCTGCAGAACCAG CTTACAGAACCGACCTTGGAGCGCAATGTCCTGGAAAG AATTCATGCCCGGTGCTCGACACGTGAAAGAGGAAC AACTCAAACCTCAGGTACCAGATGATGCCACCGAAGCC AACAAAAGTAGGTACGGATCCTTCAACTTCCCTGAGCT CGAAGACGCCAAAAACATAAAG
SG2	GTTCTCAAAAATGAACAAATGTCGACGTA CTGGGTAAGGTGGGTACATATTTTCGACGGACACAGGCC CTGGGCACTTGCAAAGAAGTCCGTTCTGCAGAACCAG CTTACAGAACCGACCTTGGAGCGCAATGTCCTGGAAAG AATTCATGCCCGGTGCTCGACACGTGAAAGAGGAAC AACTCAAACCTCAGGTACCAGATGATGCCACCGAAGCC AACAAAAGTAGGTACGGATCCTTCAACTTCCCTGAGCT CGAAGACGCCAAAAACATAAAG
SG3	GTTCTCAAAAATGAACAAATGTCGACGTGTTGGCGGTG TTGGCCTGGTGAGCTGTACCTCATTAACATCGAGAGTG CAGGCTTTCGTGAAGGAGCTTCAGCGGCTGCGGCTTAC CAGGCGGGCTTTTTGCGTGGATCCTTCAACTTCCCTGA GCTCGAAGACGCCAAAAACATAAAG
SG4	GTTCTCAAAAATGAACAAATGTCGACGTGTTGACGGTGT TGGCCTGGTGAGCTGTACCTCATTAACATCGAGAGTGC AGGCTTTCGTGAAGGAGCTTCAGCGGCTGCGGCTTACC AGGCGGGCTTTTTGCGTGGATCCTTCAACTTCCCTGAG CTCGAAGACGCCAAAAACATAAAG
SG7	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAATTAATGTTTTAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GACCTCGTGGCGAACGGTAGCGCGTCTGACTCCAGA TCAGAAGGCTGCGTGTTTGAATCACGTGCGGGTTCAGGT ACCTATTGAAAAATTTTTGGAACCGGGGTGACGTTT AACTATTGAAAAAGGAAGAGTATG
SG8	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT

	<p>CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGTTTCGATTCCGGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG</p>
SG9	<p>CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GACCTCGTGGCGCAACGGTAGCGCGTCTGACTCCAGA TCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTTCAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG</p>
SG10	<p>CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GACCTCGTGGCGCAATGGTAGCGCGTCTGACTCCAGAT CAGAAGGTTGCGTGTTCAAGTCACGTCGGGGTTCAGGTA CCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTTA AACTATTGAAAAAGGAAGAGTATG</p>
SG11	<p>CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GGCCTCGTGGCGCAACGGTAGCGCGTCTGACTCCAGA</p>

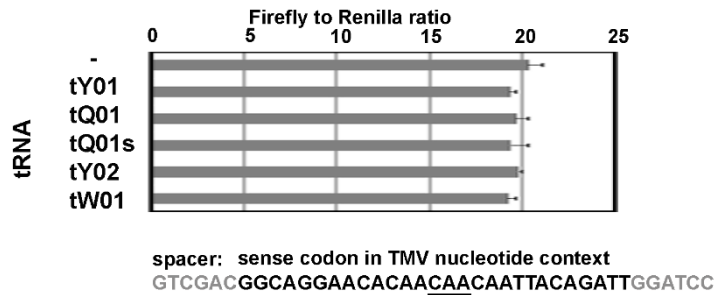
	TCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTCGACGTTT AAACTATTGAAAAAGGAAGAGTATG
SG12	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GACCTCGTGGCGCAACGGCAGCGCTCTGACTCCAGA TCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTCGACGTTT AAACTATTGAAAAAGGAAGAGTATG
SG13	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GGCAGAGGAGGGTGCAGTTGGCAGCCTGTCCAAGTCC AGCACGGTTGGAGCACAGGATTTAGAATGGGATGGTCC TGGGTTCAAACCCAGCTGCGCCCGGTACCTATTGAAA AATTTTTTGAACCCGGGGTCGACGTTTAACTATTGAA AAAGGAAGAGTATG
SG14	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GGAAGGATGGGGCCAAGCTGGAAAGCCTGTGGGCTCC ACAGTCATGTGCCTGGGTTCAATTTCCAGTTCTGCATG GTACCTATTGAAAAATTTTTTGAACCCGGGGTCGACGT TTAACTATTGAAAAAGGAAGAGTATG
SG16	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG

	ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGATTCCAGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AAACTATTGAAAAAGGAAGAGTATG
SG17	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGTTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGATTCCGGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AAACTATTGAAAAAGGAAGAGTATG
SG18	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGTTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGAATCCGGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AAACTATTGAAAAAGGAAGAGTATG
SG19	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC TCTTCAATAGCTCAGCTGGTAGAGCGGAGGACTGTAGA TTCTTAGGTGCTGGTTTGATTCCGACTTGGAGAGGGTA

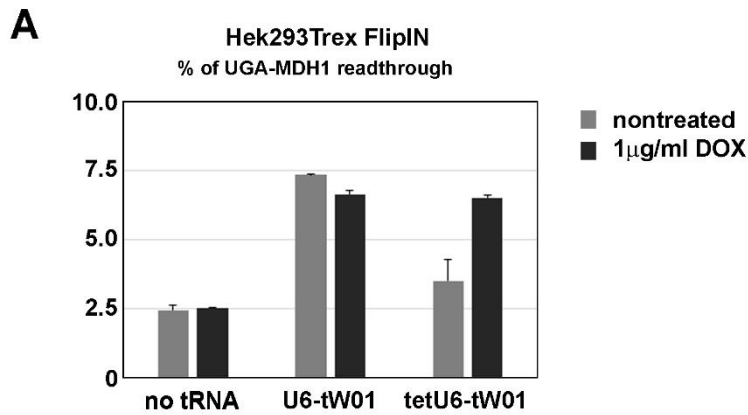
	CCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTTAACTATTGAAAAAGGAAGAGTATG
SG20	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAATTATGTTTTAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCAATTCCGGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTTAACTATTGAAAAAGGAAGAGTATG
SG21	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAATTATGTTTTAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GGGGGTATAGCTCAGGGCTAGAGCTTTTTGACTGTAGA GCAAGAGGTCCCTGGTTCAATCCAGGTTCTCCCTGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTTAACTATTGAAAAAGGAAGAGTATG
SG22	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAATTATGTTTTAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC TCAATTATAGCTCAGTGGTAGAGCATTAACTGTAGATC AAGAGGTCCCTGGATCAACTCTGGGTGCCCCCTTTAAG GTACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTTAACTATTGAAAAAGGAAGAGTATG
SG23	CTGATAAATGCTTCAATAATAGTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT

	AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC GTCAGTGTTCACAACGGTTAAGTGAAGAGGCTGTAAA CCCAGACTGGATGGGTTCATTCCCATCTCTGCCGGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG
SG24	CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CTTTCGATAGCTCAGTTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGAATCCGGCTCGAAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG
SG25	CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGTTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGAATCCGGCTCGGAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG
SG26	CTGATAAATGCTTCAATAATAGTTTTAAACGGATCCTGGC CAAAGGTCGGGCAGGAAGAGGGCCTATTTCCCATGATT CCTTCATATTTGCATATACGATACAAGGCTGTTAGAGAG ATAATTGGAATTAATTTGACTGTAAACACAAAGATATTAG TACAAAATACGTGACGTAGAAAGTAATAATTTCTTGGGT AGTTTGCAGTTTTAAAATTATGTTTTAAAATGGACTATCA TATGCTTACCGTAACTTGAAAGTATTTTCGATTTCTTGGC TTTATATATCTTGTGGAAAGGACGAAACACCGGGGCC CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGA TCCTTAGGTCGCTGGTTCGAATCCGGCTCGGAGGAGGT ACCTATTGAAAAATTTTTTGAACCCGGGGTTCGACGTTT AACTATTGAAAAAGGAAGAGTATG

SUPPLEMENTARY FIGURES AND FIGURE LEGENDS



Supplementary Figure 1. Overexpression of human tRNAs do not affect Firefly to Renilla ratio for a reporter bearing the sense codon (CAA). The HEK293T cells were transfected with sense codon control reporters containing the U6-tRNAbox with or without indicated tRNAs and processed for luciferase activity measurements as described in Materials and Methods.



B

```

1  ctgataaatg cttcaataat agtttaaacg gatcctggcc aaaggtcggg caggaagagg gctatattcc catgatttcc tcataatttcg atatacgata
   gactatttac gaagttatta tcaaatttgc ctaggaccgg ttocagcccc gtccttctcc cggataaagg gtaactaagg agtataaacg tatatgcat

101 caaggctggt agagagataa ttggaattaa ttgactgta aacacaaaga tattagtaca aaatacgtga cgtagaaagt aataattttc tgggtagttt
   gttccgacaa tctctctatt aaccttaatt aaactgacat ttgtgtttct ataatoatgt tttatgcaat gcacttttca ttattaaaga accoatcaaa

201 gcagttttaa aattatggtt taaatggac tatcatatgc ttaccgtaac tgaaagtcc ctatcagtga tagagattat atatctccct atcagtgata
   cgtcaaaatt ttaatacaaa attttacctg atagtatacg aatggcattg aacttccagg gatagtcact atctctaata tatagagga tagtcaactat
                                     PSE O2 TATA O2

301 gagaccggg cccgaacctg tgggccaacg gtagcgctc tgactcaga tcagaaggct gcgtgttga atcacgtcg ggtoaggtac ctattgaaaa
   ctctgggccc gggctggagc accgcttg catcggcag actgaggtct agtcttccga cgcacaagct tagtgcagcc ccagtccatg gataactttt
   +1 tW01

401 attttttgga acccggggtc gacgtttaa ctattgaaaa aggaagagta tg
   taaaaaacct tgggcccag ctgcaaattt gataactttt tcttctcat ac

```

Supplementary Figure 2. Doxycycline inducible TetU6-tW01 expression in HEK293Trex cells increases SC-RT of the UGA-MDH1 reporter. (A) Readthrough measurements using UGA-MDH1 reporter with no tRNA control (no tRNA) and tryptophan tRNA under either the U6 promoter (U6-tW01) or the doxycycline inducible U6 promoter (tetU6-tW01) in the Hek293Trex FlipIN cells. The cells were transfected with stop codon constructs (GL3101, PBB371, PBB380) or their respective sense codon controls (GL3102, PBB373, PBB382) and treated with or without doxycycline 16h prior the readthrough measurement. **(B)** The sequence of tetU6-tW01 cassette including the U6 promoter features as in Figure 1 with insertions of two O2 repressor binding sequences and tW01 tryptophan tRNA. The PSE, TATAbox, +1 position, tRNA and O2 sequences are indicated in red.

polymorphism. (C) The UAG–TMV context readthrough measurements with overexpression of the indicated Tyr tRNA iso-decoders under control of U6 in PBB168 (for UAG) or its sense control PBB172.

A

```

> tY01 Homo_sapiens_chr14.trna16-TyrGTA _ Tyr (GTA) 94 bp Sc: 74.71
CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGATCCTTAGGTCGCTGGTTCGATTCCGGCTCGAAGGA

> tY06 Homo_sapiens_chr8.trna12-TyrGTA _ Tyr (GTA) 88 bp Sc: 46.11
TCTTCAATAGCTCAGCTGGTAGAGCGGAGGACTGTAGATTCTTAGGT_GCTGGTTTGATTCCGACTTGGAGAG

> tY08 Homo_sapiens_chr7.trna9-TyrGTA _ Tyr (GTA) 73 bp Sc: 49.38
GGGGGTATAGCTCAGGGCTAGAGCTTTTGTACTGTAGAGCAAGAGGTCCTGGTTCAAATCCAGGTTCTCCCT

> tY09 Homo_sapiens_chr7.trna24-TyrGTA _ Tyr (GTA) 76 bp Sc: 33.50
TCAATTATAGCTCAGTGGTAGAGCATTAACTGTAGATCAAGAGGTCCTGGATCAACTCTGGGTGCCCCCTTT
AA

> tY10 Homo_sapiens_chr15.trna5-TyrGTA _ Tyr (GTA) 73 bp Sc: 24.21
GTCAGTGTGCACAACGGTTAAGTGAAGAGGCTGTAAACCCAGACTGGATGGGTTCAAATCCCATCTCTGCCG

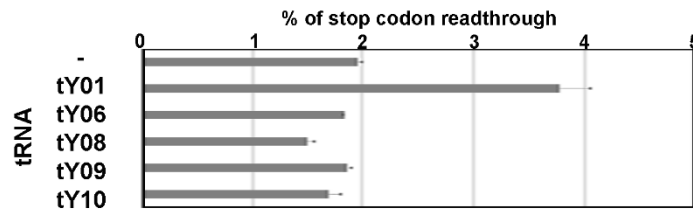
```

B

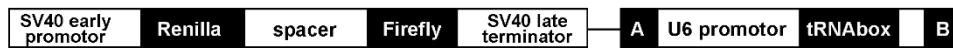
```



> tY01 CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTAGATCCTTAGGTCGCTGGTTCGATTCCGGCTCGAAGGA
> tY06 TCTTCAATAGCTCAGCTGGTAGAGCGGAGGACTGTAGATTCTTAGGTXGCTGGTTTGATTCCGACTTGGAGAG
> tY08 GGGGGTATAGCTCAGGGCTAGAGCTTTTGTACTGTAGAGCAAGAGGTCCTGGTTCAAATCCAGGTTCTCCCT
> tY09 TCAATTATAGCTCAGTGGTAGAGCATTAACTGTAGATCAAGAGGTCCTGGATCAACTCTGGGTGCCCCCTTTAA
> tY10 GTCAGTGTGCACAACGGTTAAGTGAAGAGGCTGTAAACCCAGACTGGATGGGTTCAAATCCCATCTCTGCCG

```

C

Supplementary Figure 4. Bioinformatically predicted “low tRNA score” tyrosine tRNA pseudogenes do not increase UAG readthrough. (A) tY01 and four other tyrosine tRNA sequences retrieved from the GtRNAdb (4,5) (deprived of their computationally predicted introns) that were experimentally shown not to increase the UAG readthrough. (B) The ClustalX2 alignment of the tRNA sequences from panel A and a high tRNA score tY01. (C) The UAG–TMV context readthrough measurements with overexpression of tY01 and the indicated Tyr tRNA pseudogenes under control of U6 in PBB168 (for UAG) or its sense control PBB172.

A**B**

tRNA	>gene name sequence	% of UAG readthrough
-	none	1.96 ± 0.03
 tY01	>Homo_sapiens_chr14.trna16-TyrGTA CCTTCGATAGCTCAGCTGGTAGAGCGGAGGACTGTA GATCCTTAGGTCGCTGGTTCGATTCCGGCTCGAAGGA	3.61 ± 0.11
 tY02	>Homo_sapiens_chr2.trna14-TyrATA CCTTCAATAGTTCAGCTGGTAGAGCAGAGGACTATA GGTCCTTAGGTTGCTGGTTCGATTCCAGCTTGAAGGA	1.91 ± 0.05

Supplementary Figure 5. The singleton human tyrosine tRNA with the ATA anticodon does not increase UAG readthrough (A) Schematic of the readthrough reporter containing the U6-tRNAbox cassette. For details please refer to the main text. **(B)** Readthrough measurements using the UAG reporter with or without the indicated human tRNAs in the HEK293T cells. The schematics of base-pairing between individual tRNAs and the UAG stop codon is shown to the right of the name of each construct, followed by the gene identity with the tRNA sequence and the relative percentage of the UAG readthrough. The tRNA sequences were individually placed under the U6 promoter of the PBB168 reporter or its sense control PBB172 reporter (see Figure 1).

A

```

> tW01 Homo_sapiens_chr12.trna6-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGCTGCGTGTTCCGAATCACGTCGGGGTCA

> tW02 Homo_sapiens_chr17.trna6-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

> tW02 Homo_sapiens_chr6.trna168-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

> tW02 Homo_sapiens_chr6.trna170-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

> tW03 Homo_sapiens_chr17.trna12-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAATGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAAGTCACGTCGGGGTCA

> tW04 Homo_sapiens_chr17.trna39-TrpCCA_ Trp (CCA) 72 bp
GECCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

> tW05 Homo_sapiens_chr7.trna1-TrpCCA_ Trp (CCA) 72 bp
GACCTCGTGGCGCAACGGCAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

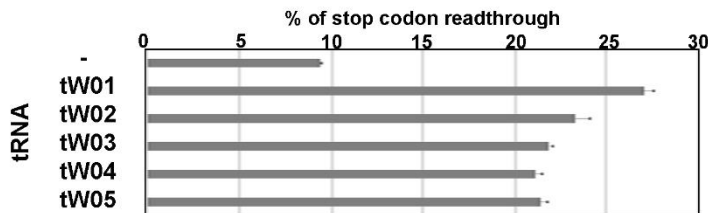
```

B

```

* * * * *
> tW01 GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGCTGCGTGTTCCGAATCACGTCGGGGTCA
> tW02 GACCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA
> tW03 GACCTCGTGGCGCAATGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAAGTCACGTCGGGGTCA
> tW04 GGCCTCGTGGCGCAACGGTAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA
> tW05 GACCTCGTGGCGCAACGGCAGCGCTCTGACTCCAGATCAGAAGGTTGCGTGTTCAAATCACGTCGGGGTCA

```

C

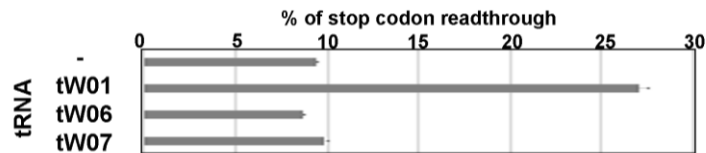
Supplementary Figure 6. The single nucleotide polymorphism in human “high tRNA score” tryptophan iso-decoders does not impact the readthrough efficiency. **(A)** Sequences of all available tryptophan tRNAs retrieved from the GtRNAdb (4,5). **(B)** The ClustalX2 alignment of the tRNA sequences from panel A. Single nucleotide polymorphisms (all are unique) are marked by a red asterisk on the top of sequences. **(C)** The UGA–AQP4 context readthrough measurements with overexpression of the indicated Trp tRNA iso-decoders under control of U6 in GL3051 (for UGA) or its sense control GL3052..

A

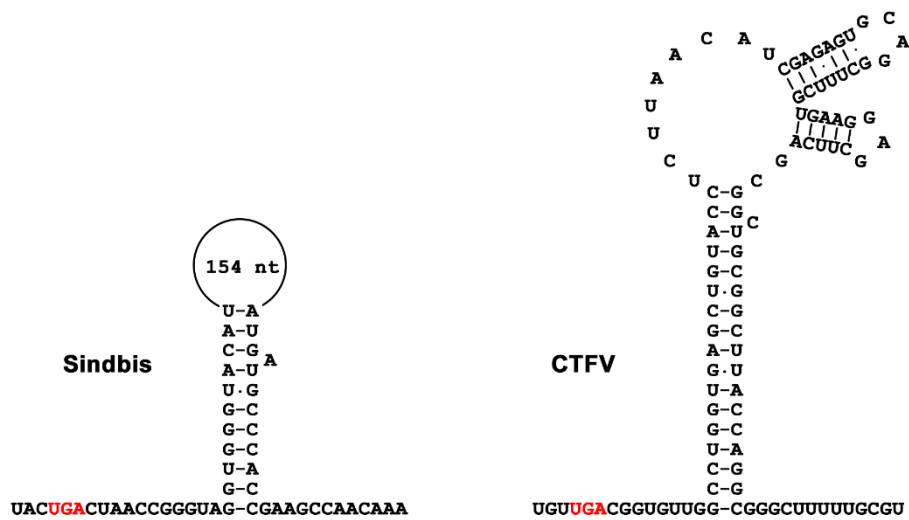
> tW01 Homo_sapiens_chr12.trna6-TrpCCA _ Trp (CCA) 72 bp Sc: 71.65
GACCTCGTGGCGCAACGGTAGCGCGTCTGACTCCAGATCAGAAGGcTGC GTTTCGAATCACGTCGGGGTCA

> tW06 Homo_sapiens_chr9.trna3-TrpCCA _ Trp (CCA) 99 bp Sc: 23.38
GGCAGAGgagGGTGCAGTTGGCAGCCTGTCCAAGTCCAGcacggttgagcacaggattagaATGGGATGGtCCTGG
GTCAAACCCAGCTGCGCCC

> tW07 Homo_sapiens_chr11.trna19-TrpCCA _ Trp (CCA) 74 bp Sc: 21.17
GGAAGGATggGGCCAAGCTGGAAAGCCTGTGGGCTCCACAGTCATGTGCCTGGGTTCAAT
TCCAGTTCTGCAT

B

Supplementary Figure 7. Bioinformatically predicted “low tRNA score” tryptophan tRNA pseudogenes do not increase UAG readthrough. (A) tW01 and two other tryptophan tRNA sequences retrieved from the GtRNadb (4,5) that were experimentally shown not to increase the UGA readthrough. (B) The UGA-AQP4 context readthrough measurements with overexpression of tW01 and the indicated Trp tRNA pseudogenes under control of U6 in GL3051 (for UGA) or its sense control GL3052.



Supplementary Figure 8. Schematics of the structural features of the Sindbis virus (SINV) (6) and Colorado thick fever virus (CTFV) (7) increasing the UGA readthrough. For details see the main text and Figure 5A.