Figure S1

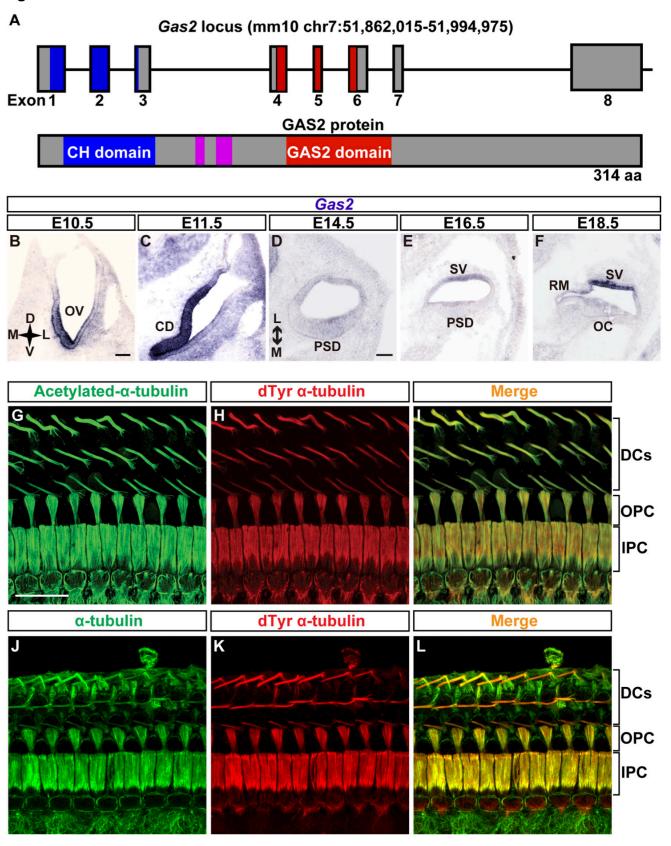


Figure S1. Gas2 is expressed in the developing cochlea (related to Fig.1).

(A) Schematic of the mouse Gas2 gene and encoded protein highlighting the positions of the calponin homology (CH) and GAS2 domains. (B-F) Transverse sections through the inner ear showing the spatial and temporal patterns of Gas2 expression during mouse embryonic development. Scale bar = $100\mu m$ (B, C); $50\mu m$ (D-F). (G-L) Whole mount immunostaining of cochlear preparations at P14 with antibodies against acetylated α -tubulin and dTyr α -tubulin (G-I), and pan α -tubulin and dTyr α -tubulin (J-L). The majority of microtubules in support cells are both acetylated and detyrosinated. Scale bar = $20\mu m$. Abbreviations: cochlear duct (CD), dorsal (D), lateral (L), medial (M), prosensory domain (PSD), organ of Corti (OC), otic vesicle (OV), Reissner's membrane (RM), stria vascularis (SV), ventral (V).

Figure S2

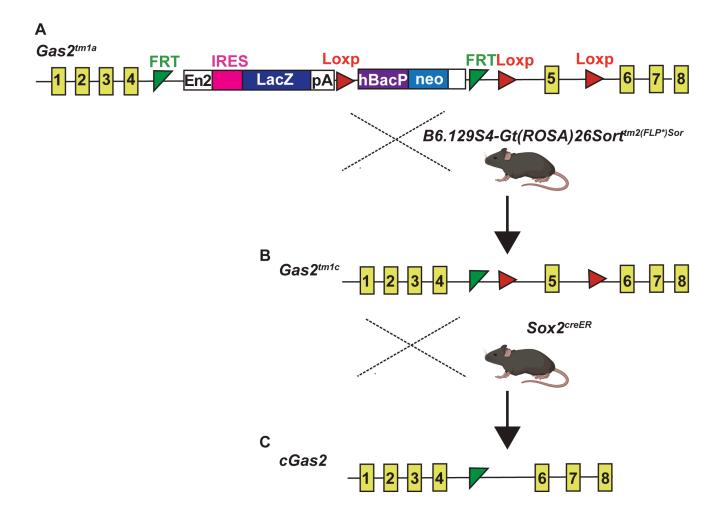


Figure S2. Breeding scheme to generate Gas2 mutant alleles (related to Fig. 2).

(A) Schematic of the *Gas2* knockout first allele (*Gas2*^{tm1a}) generated by the European Conditional Mouse Mutagenesis Program. (B) The *Gas2* floxed allele (*Gas2*^{tm1c}) was generated by crossing *Gas2*^{tm1a/+} mice with a FLPo deletor strain and selecting progeny with loxp sites surrounding exon 5. (C) Conditional *Gas2* knockout mice (*cGas2*) were generated by crossing *Gas2*^{tm1c/tm1c} mice with an inducible *Sox2*^{CreER} line, which selectively deletes *Gas2* in supporting cells upon tamoxifen administration.

Figure S3

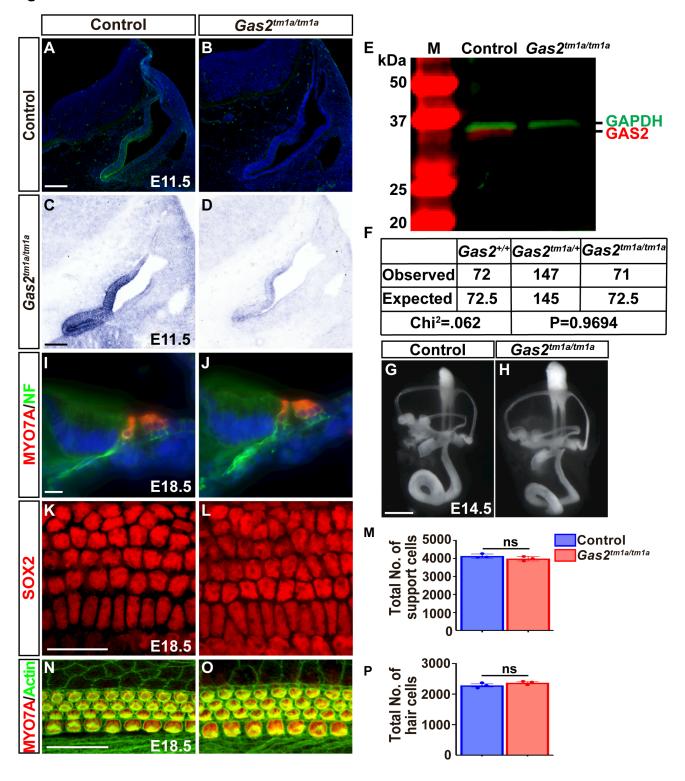


Figure S3. Inner ear development is unaffected by loss of GAS2 (related to Fig. 2). (A-D) Transverse sections through the otic vesicle showing the loss of GAS2 protein (A,B) and Gas2 mRNA (C,D) in Gas2^{tm1a/tm1a} embryos compared to controls (n=3). (E) Western blot showing loss of GAS2 expression (red band) in relation to GAPDH (green band) in cochleae from Gas2^{tm1a/tm1a} embryos compared to controls (P0) (n=3). **(F)** Table displaying the observed genotypes of progeny from *Gas2*^{tm1a/+} intercrosses at the time of weaning, which fit the expected Mendelian ratios. (G-H) Paint fills of control and Gas2^{tm1a/tm1a} inner ears at E14.5 revealed no difference in morphology (n=3). (I-J) Transverse sections through the cochlea stained with markers of hair cells (MYO7A) and spiral ganglia (NF, neurofilament) show normal patterns of innervation between control and Gas2^{tm1a/tm1a} embryos at E18.5. (K-P) Whole mount immunostaining of cochlear preparations show no differences in the number of SOX2 positive supporting cells (K-M), or MYO7A, Phalloidin/F-actin positive hair cells (N-P) between control and Gas2^{tm1a/tm1a} embryos at E18.5. (M,P) Quantification of the total number of hair and supporting cells is presented as mean ±SD (ns = not significant, p=0.2373 (M), p=0.2156 (P), two-sided t-test, n=3). Scale bar = 100µm (A-H), 10µm (I-J), 20µm (K-O).

Figure S4

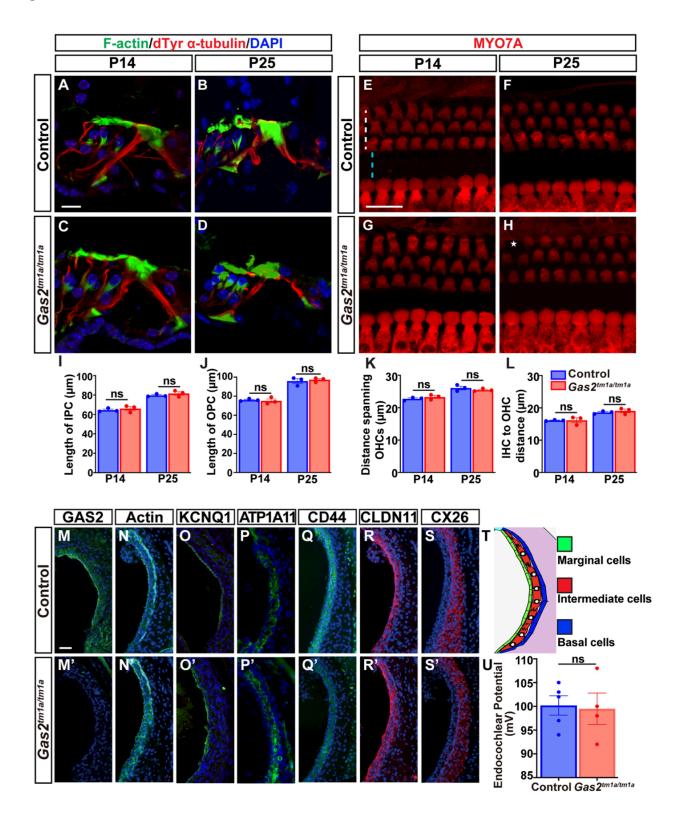


Figure S4. Cellular organization in the organ of Corti and stria vascularis is maintained in postnatal Gas2^{tm1a/tm1a} mice (related to Fig. 2). (A-D) Transverse sections through the cochlea of control and Gas2^{tm1a/tm1a} mice at P14 (A, E) and P25 (B, F) stained for F-actin (green), dTyr α-tubulin (red) and DAPI (blue) to reveal inner and outer pillar cell morphology. (E-H) Whole mount cochlear preparations from control and Gas2^{tm1a/tm1a} mice at P14 (E, G) and P25 (F, H) immunostained for MYO7A (red) to reveal inner and outer hair cell rows. A missing hair cell is indicated (asterisk). (I-J) The lengths of IPCs and OPCs were not significantly different between control and Gas2^{tm1a/tm1a} mice (presented as mean ±SEM, ns=not significant, Student's t-test, n=3). Scale bar = 20µm. (K-L) The distance between OHC rows (dashed white line in E) and the distance between inner and outer hair cell rows (dashed blue line in E) was not significantly different between control and Gas2^{tm1a/tm1a} mice (presented as mean ±SEM, ns=not significant, Student's t-test, n=3). (M-S') Transverse sections through the stria vascularis of control (M-S) and Gas2^{tm1a/tm1a} (M'-S') mice at P25 stained with layer specific markers as schematized in (T). Despite the loss of GAS2 in the marginal cell layer, no other markers were differentially expressed between control and Gas2^{tm1a/tm1a} mice. Scale bar = 25µm. (U) Endocochlear potential measurements between control (n=5) and Gas2^{tm1a/tm1a} (n=4) mice at P56 were not significantly different (presented as mean ±SEM, p=0.8556, two-tailed t-test).

Figure S5

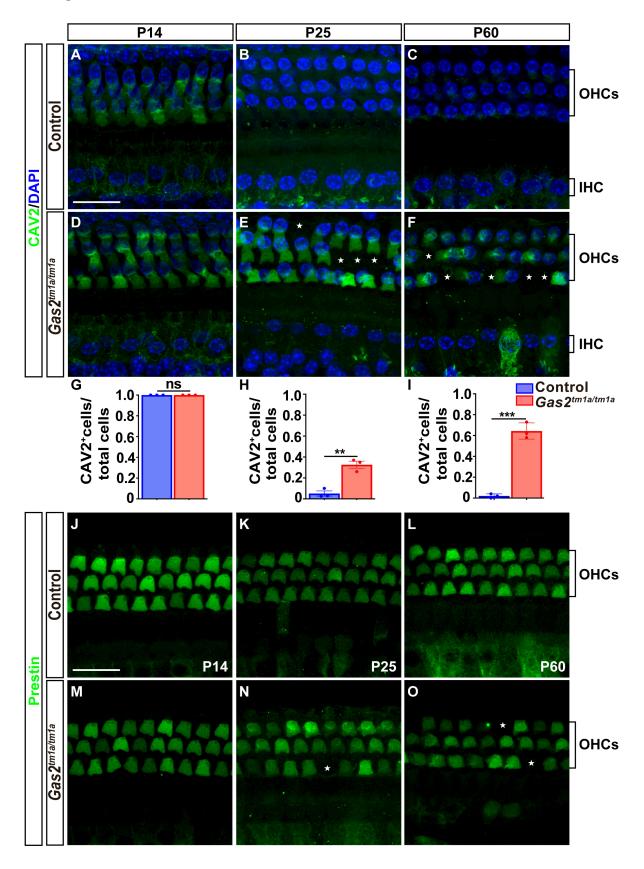


Figure S5. OHCs display increased mechanical stress and maintain Prestin expression in *Gas2^{tm1a/tm1a}* mice (related to Fig. 5). (A-F) Whole mount cochlear preparations from control and *Gas2^{tm1a/tm1a}* mice at P14 (A, D), P25 (B, E) and P60 (C, F) immunostained for Caveolin-2 (CAV2). Missing hair cells are indicated (asterisk). (G-I) Quantification of the ratio of CAV2⁺ cells to the total number of hair cells at P14, P25, P60 presented as mean ±SEM (ns=not significant, **p<0.01, ***p<0.001, two sided t-test, n=3). Whole mount cochlear preparations from control and *Gas2^{tm1a/tm1a}* mice at P14 (J, M), P25 (K, N) and P60 (L, O) showing no difference in the expression of Prestin. Scale bar = 20μm.

Figure S6

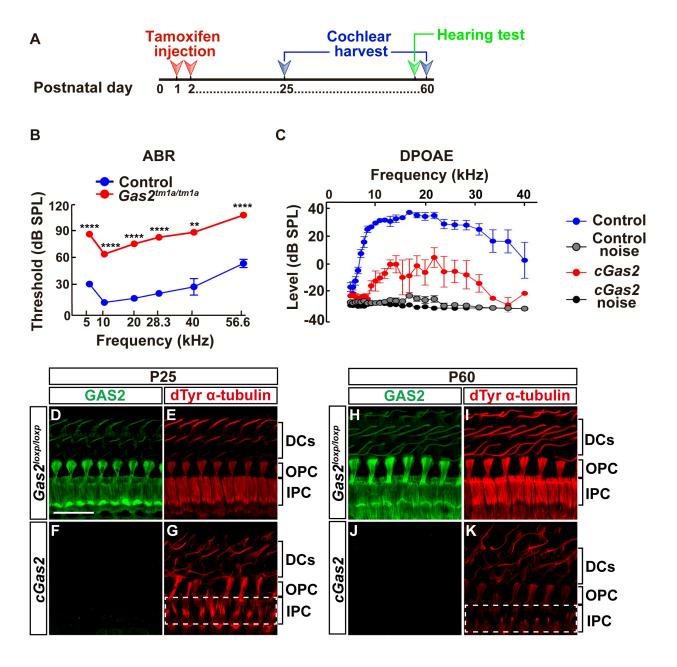


Figure S6. GAS2 is required postnatally in supporting cells for microtubule stability and hearing (related to Fig. 5). (A) Timeline for the generation and analysis of *cGas2* knockout mice. (B) ABR thresholds from control (n=3) and c*Gas2* (n=3) mice at P60 (**p<0.01, ****p<0.0001, multiple T-test with Holm-Sidak method). (C) Reduced DPOAE response in *cGas2* mice at P60 (p<0.05, multiple T-test with Holm-Sidak method, n=3). (D-K) Whole mount cochlear preparations from control and c*Gas2* mice at P25 (D-G) and P60 (H-K) immunostained for GAS2 and detyrosinated α-tubulin. A progressive loss of pillar cell microtubules is observed with age, comparable to that described in *Gas2*^{tm1a/tm1a} mice (Fig. 2G,H,L,M).

Figure S7

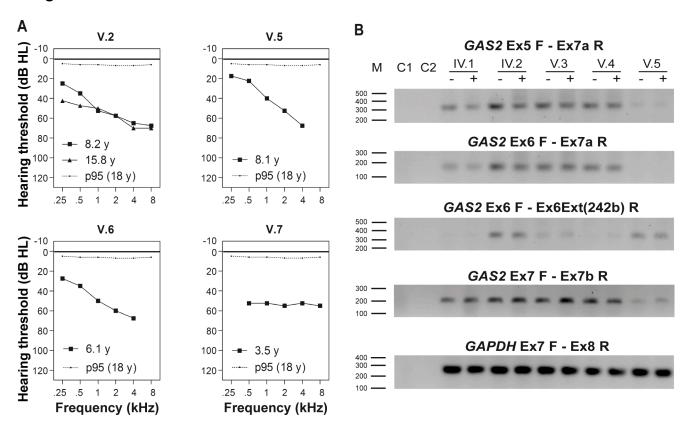


Figure S7. Hearing loss and splicing alterations in subjects with the c.723+1G>A GAS2 variant (related to Fig. 7). (A) Average air conduction thresholds of subjects V2 and V.5-7 with the c.723+1G>A GAS2 variant in the homozygous state. The dotted line represents the 95th percentile of age- and sex-specific hearing level at the age of 18 years. This is the lowest age for which the ISO 7029:2017 standard can be applied. Thresholds of subjects V5 and V6 are based on pure tone audiometry; thresholds of subject V7 are based on conditioned play audiometry. Age in years (y), decibel hearing level (dB HL), kilohertz (kHz). (B) RT-PCR using primer pairs in exon 6 and flanking exons and intronic sequences (referring to transcript NM_005256.3) was performed to detect aberrant splicing associated with the GAS2 c.723+1G>A variant. 100bp marker

(M), no reverse transcriptase in cDNA synthesis step (C1), PCR without cDNA input (C2), cells treated with cycloheximide (+), no cycloheximide treatment (-).

Figure S8.

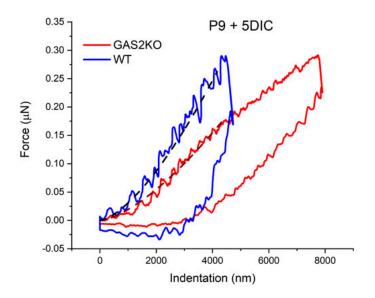


Fig. S8. Representative indentation curves from an atomic force microscopy experiment (related to STAR Methods: Atomic Force Microscopy and Fig. 4).

Representative indentation curves from inner pillar cells taken from the Gas2 knockout (KO) and WT cochlea at P9 + 5DIC. The slope of the indentation (top portion of the curve) is fitted to the Hertz equation, dashed lines, for a spherical indenter to determine the stiffness of the cell.

Table S1. Primer sequences for analysis of human *GAS2* (related to Fig. 7 and Fig. S7)

Fragment	Primer name in Fig. 7 and Fig. S7		Oligonucleotides	RT-PCR size wildtype (bp)	RT-PCR size mutant (bp)
	Primers for amplification	and sequence an	alysis of genomic DNA		
GAS2 exon 6	NA	Forward:	GATGTCTTTGGGGCCTGAAC		
GA32 exon 6	NA	Reverse:	GATCTCCATGGTGTCTTTGGTG	-	-
	Primers for RT-PCR and	d sequence analys	is of <i>GAS2</i> transcript		
GAS2 exon 5 - exon 7	Ex5 F	Forward:	TTCTGCCCCTTCTCCTTCAC	294	
GA32 EXON 3 - EXON 7	Ex7a R	Reverse:	ACACGGGAGATCTGCAGC	254	-
GAS2 exon 6 - exon 7	Ex6 F	Forward:	GTTCTGTGTGGAGCGGCTC	171	
GA32 exoll 6 - exoll 7	Ex7a R	Reverse:	ACACGGGAGATCTGCAGC	1/1	-
GAS2 exon 5 - exon 6 extension	Ex5 F	Forward:	TTCTGCCCCTTCTCCTTCAC		450
c.723+242	Ex6Ext(242b) R	Reverse:	TCACAAGTGGAAAGCCTTCTTC	-	430
GAS2 exon 6 - exon 6 extension	Ex 6 F	Forward:	GTTCTGTGTGGAGCGGCTC		327
c.723+242	Ex6Ext(242b) R	Reverse:	TCACAAGTGGAAAGCCTTCTTC	-	327
GAS2 exon 6 extension c.723+241 -	Ex6Ext(241b) F	Forward:	TGAAGAAGGCTTTCCACTTGTG		
exon 7	Ex7a R	Reverse:	ACACGGGAGATCTGCAGC	-	-
GAS2 exon 6 extension c.724-224 -	Exon6Ext(-224b) F	Forward:	ACAGCAAAGCTGGGTTCTGG		331
exon 7	Ex7a R	Reverse:	ACACGGGAGATCTGCAGC	-	331
CAS2 aver 7 aver 7	Ex7 F	Forward:	TGCTGCACAACAACATGTC	216	
GAS2 exon 7 – exon 7	Ex7b R	Reverse:	GGCAGAGACCACCAAGTAG	216	-
CARRU 7 0 (NNA 002015 T)	NA	Forward:	CTGCACCACCAACTGCTTAG	240	
GAPDH exon 7 - 8 (NM_002046.5)	NA	Reverse:	AGCTCAGGGATGACCTTGC	219	-

	Pr	imers for qPCR of GAS2	cDNA		
C463 2 4	NA	Forward:	ATCCTGGTGCCGAGATTTAGG		
<i>GAS2</i> exon 3 - 4	NA	Reverse:	CACACTTCTCTGGGTTGCTTG	-	-
GAS2 exon 6 - exon 6Ext	NA	Forward:	GTGGGAGAAAAGATCCTCTTC		
GASZ EXON 6 - EXON 6EXT	NA	Reverse:	GAAGTTCCTGCTCTTATCTGTC	-	-
GUSB exon 2 - 3 (NM_000181.4)	NA	Forward:	AGAGTGGTGCTGAGGATTGG		
	NA	Reverse:	CCCTCATGCTCTAGCGTGTC	-	-

GAS2 reference sequence: NM_005256.3. (RT-)PCR was performed with annealing temperatures decreasing from 62.5 to 57.5 °C for 30 cycles. qPCR was performed with an anealing temperature of 60.0 °C.

Table S2. Individual results of newborn hearing screening, otoscopic examination, otoacoustic emissions and audiometry (related to Fig. 7).

Family	Subject	Newborn screening	Age of HL first detected (y)	Otoscopic examination	Otoacousic emissions		P	ГΑ	SF	RT	-	imum 5 (%)		
					TEOA	DPOAE	I/O	Age at hearing test (y)	R	L	R	L	R	L
W19-1278	IV.2	NA	NA	NT	present	present	present	38	8	5	NT	NT	NT	NT
	V.2	NT	1	N	absent	absent	absent	15	58	60	50	42	94	97
	V.3	NA	NA	NT	present	present	present	14	5	5	NT	NT	NT	NT
	V.4	NA	NA	NT	present	present	present	12	2	0	NT	NT	NT	NT
	V.5	Р	1	N	absent	absent	absent	7	58	58	47	NT	93	98
	V.6	F	0	N	absent	absent	absent	5	60	62	45	50	95	95
	V.7	F	0	N	present	absent	absent	3	52	55	NT	NT	NT	NT

Age of onset (AoO), age of onset in years as reported by the subjects. Age at hearing test, the age at which the audiometric data of column 9 to 14 were obtained, in general the last audiogram. If no speech audiometry was performed during the latest pure tone audiometry, the latest audiogram in which both were measured, was selected. TEOAE, transient evoked otoacousic emissions; DPOAE, distortion product otoacoustic emissions; I/O, DPOAE input/output functions; y, years; PTA, pure tone average, mean of 0.5, 1 and 2 kHz air conduction thresholds; SRT, speech reception threshold; SRS, speech recognition score in %; R, right; L, left; NA, not applicable; NT, not tested; N, normal; P, pass; F, fail.

Table S3. Primer sets for genotyping mouse *Gas2* alleles (relates to Fig. S2)

Fragment	Primer name in Supplementary Figure 2	Oligonucleotides		PCR size wildtype (bp)	PCR size mutant (bp)			
	Primers for amplification and sequence analysis of genomic DNA							
	NA	Forward:	GAAGTTCCTATTCCGAAGTTCCT	7076				
5 arm	NA	Reverse:	CTGCCTGTGTTGAATGTTCTCAT	7076	-			
2/ 2/22	NA	Forward:	CCTGCTTGCCGAATATCATGG	6402				
3' arm	NA	Reverse:	ACTTCTGTGGAGTCTCAGCTC	6493	-			
Tm1a/Loxp	NA	Forward:	TTGGATCATATGGAGAGAGCCAT	223	257			
ттальохр	NA	Reverse:	GGGCATATCACAGGCCCATA	223	257			
Tm1c	NA	Forward:	GGCGCATAACGATACCACGA		208			
IIIIC	NA	Reverse:	TATAAGCCGCCTACTGCGAC	-	208			
Gas2-LacZ	NA	Forward:	AGAACAAGCAACATGCGCTC		611			
Gusz-Lacz	NA	Reverse:	GACCTTGGGACCACCTCATC	-	011			
Cara Non	NA	Forward:	CTCCCCTGAACCTGAAACATA		200			
Gas2-Neo	NA	Reverse:	TTATATAAGCCGCCTACTGCGA	-	360			
oCas2 Cro	NA	Forward:	GCGGCATGGTGCAAGTTGAAT		222			
<i>cGas2-</i> Cre	NA	Reverse:	CGTTCACCGGCATCAACGTTT	-	232			